• CLINICAL RESEARCH •

Impact of endoscopically minimal involvement on IL-8 mRNA expression in esophageal mucosa of patients with non-erosive reflux disease

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Abstract

AIM: Little has been known about the pathogenesis of nonerosive reflux disease (NERD). Recent studies have implicated interleukin 8 (IL-8) in the development and progression of gastroesophgeal reflux disease (GERD). The purpose of this study was to determine IL-8 RNA expression levels in NERD patients with or without subtle mucosal changes.

METHODS: We studied 26 patients with NERD and 13 asymptomatic controls. Biopsy sample was taken from the esophagus 3 cm above the gastroesophageal junction and snap frozen for measurement of IL-8 mRNA levels by realtime quantitative polymerase chain reaction (PCR). We also examined mRNA expression of IL-8 receptors, CXCR-1 and -2 by reverse transcriptase PCR. The patients were endoscopically classified into grade M (mucosal color changes without visible mucosal break) and N (neither minimal involvement nor mucosal break) of the modified Los Angeles classification.

RESULTS: The relative IL-8 mRNA expression levels were significantly higher in esophageal mucosa of NERD patients than those of the controls. There was a significant difference in IL-8 mRNA levels between grade M and N. The CXCR-1 and -2 mRNAs were constitutively expressed in esophageal mucosa.

CONCLUSION: Our results suggest that high IL-8 levels in esophageal mucosa may be involved in the pathogenesis of NERD through interaction with its receptors. NERD seems to be composed of a heterogeneous population in terms of not only endoscopically minimal involvement but also immune and inflammatory processes.

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INTRODUCTION

Gastroesophageal reflux disease (GERD) is one of the most common chronic disorders in modern humans. In the United States, 44 % of the adult populations reported experiencing heartburn at least once a month, 14 % on a weekly basis, and 7 % daily^[1]. Esophageal erosions are the characteristic lesions of GERD seen on endoscopy^[2]. A small number of GERD patients develop stricture, Barrett's esophagus and adenocarcinoma of the esophagus. In fact, the majority of GERD patients have endoscopically normal-appearing esophageal mucosa; this group is termed non-erosive reflux disease (NERD) or endoscopy-negative reflux disease^[2,3].

The Los Angeles (LA) classification is widely used for endoscopic assessment of GERD^[4]. The slightest degree of esophagitis, i.e., grade A, is defined as one or more mucosal breaks confined to the mucosal folds, each no longer than 5 mm. Accordingly, this classification scheme ignores subtle mucosal damage in the absence of mucosal breaks. In this regard, Hoshihara *et al.*^[5,6] have proposed a modified LA system, in which grade O was subdivided into M and N, based on the concept of mucosal color changes. Thus, NERD patients can be classified into two subgroups (grade M and N) based on minimal esophageal involvement during endoscopy.

Recently, several studies have shown that mucosal immune and inflammatory responses, characterized by specific cytokine and chemokine profiles, may determine the diversity of esophageal phenotypes of GERD^[7-9]. Of note, Fitzgerald *et al.*^[7] reported significantly higher expression levels of interleukin 8 (IL-8) messenger ribonucleic acid (mRNA) in patients with reflux esophagitis (RE), assessed by competitive reverse transcriptase polymerase chain reaction (RT-PCR), compared with subjects with non-inflamed or Barrett's esophagus. Studies from our laboratories have also demonstrated high IL-8 protein levels in esophageal biopsy samples of patients with erosive esophagitis by enzyme linked immunosorbent assay (ELISA)^[8]. Furthermore, we also showed significantly high mucosal IL-8 production, which paralleled the endoscopic severity of RE^[8]. However, little has been known about the role of IL-8 in NERD.

The aim of the present study was to assess esophageal expression levels of IL-8 mRNA in NERD patients by quantitative real-time PCR procedure, with special reference to the difference between grade M and N subgroups of the modified LA scheme.

MATERIALS AND METHODS

Subjects and samples

We studied 26 patients with NERD and endoscopically confirmed normal-appearing esophageal mucosa who visited

the Outpatient Department between August 2002 and July 2003. They included 19 men and 7 women, aged between 28 and 80 years (mean, 62.0 years). The diagnosis of GERD was made with more than 6 points in the questionnaire for the diagnosis of reflux disease (QUEST) described by Carlsson et al^[10]. None of these patients had been treated with nonsteroidal anti-inflammatory drugs, proton pump inhibitors, histamine H₂-receptor antagonists, anti-cholinergic agents or antibiotics within 4 weeks prior to the present study. Furthermore, patients with severe concomitant diseases, prior esophageal or gastric surgery, peptic ulcer diseases and comorbid conditions that might interfere with esophageal or gastric motility including diabetes mellitus, systemic sclerosis and neurological disorders were excluded. As a control group, we recruited 13 asymptomatic subjects with no hiatal hernia or any lesions in the esophagus, stomach and duodenum at endoscopy for a health check-up.

In each case, a biopsy specimen was obtained from the esophageal mucosa, 3 cm above the gastroesophageal junction^[8], snap-frozen in an ethanol-dry ice mixture for quantitative analysis of IL-8 mRNA expression and stored at -80 $^{\circ}$ C until use.

Endoscopic assessment of NERD

NERD was endoscopically classified into grade M and N in accordance with the modified Los Angeles (LA) classification system proposed by Hoshihara *et al*^[5,6]. The criteria were: grade M represents minimal changes (irregular redness or whiteness) without any mucosal breaks and grade N represents esophageal mucosa with neither the minimal changes nor mucosal injury. In addition, we also evaluated the presence of hiatal hernia by endoscopy^[11].

Real-time quantitative PCR

Total RNA from the biopsy samples was extracted using a commercial kit according to the instructions provided by the supplier (Isogen, Nippon Gene Co., Toyama, Japan). One μ g of total RNA was reversely transcribed into complementary DNA (cDNA) in a volume of 25 μ l with MuLV reverse transcriptase and random hexamers (both from PE Applied Biosystems, Warrington, UK).

Real-time PCR measurement of IL-8 cDNA was performed in the ABI PRISM 7700 sequence detector (PE Applied Biosystems) with TaqMan assay. The primers and probe sequences for IL-8 were synthesized (PE Applied Biosystems) as described previously^[12]: IL-8 forward primer, 5' -CTCTTGGCAGCCTTCCTGATT-3', reverse primer, 5' -TATGCACTGACATCTAAGTTCTTTAGCA-3' and probe, 5' -CTTGGCAAAACTGCACCTTCACACAGA-3', labeled with the reporter dye 6-carboxyfluorescein at the 5' end and quencher dye 6-carboxytetramethylrhodamine at the 3' end. PCR was performed in a total volume of 50 µl of each amplification mixture containing 1 µl of each RT product, 25 µl of 2×Universal Master Mix (PE Applied Biosystems), 200 nM IL-8 forward and reverse primers, 100 nM fluorogenic probe. Thermal cycling was initiated with at 50 °C for 2 min, followed by a first denaturation step at 95 °C for 10 min, and followed by 50 cycles of at 95 $^{\circ}$ C for 15 s and at 60 $^{\circ}$ C for 1 min.

The *tubulin alpha 3* gene cDNA (internal control) was quantified in the same machinery using SYBR Green PCR Core reagents kit (PE Applied Biosystems). The primers used were: forward, 5' -AGATCATTGACCTCGTGTTGGA-3' and reverse, 5' -ACCAGTTCCCCCACCAAAG-3', which correspond to nucleotides 437-458 and 537-519, respectively (*TUBA3*, GenBank accession number 17986282). PCR was performed in a total volume of 25 µl of each amplification mixture containing 1 µl of each RT product, 3 µl of 25 mM MgCl₂, 2.5 µl of 10×SYBR Green buffer, 2 µl of dNTP Mix (5 mM adenosine, deoxycytosine and deoxyguanosine triphosphate and 2.5 mM deoxyuridine triphosphate), 0.625 U AmpliTaq Gold polymerase, 0.125 U AmpErase and 100 nM tubulin alpha 3 forward and reverse primers. Thermal cycling was initiated at 50 $^{\circ}$ C for 2 min, followed by a first denaturation step at 95 $^{\circ}$ C for 10 min, and continued with 40 cycles of at 95 $^{\circ}$ C for 15 s and at 59 $^{\circ}$ C for 1 min.

Each assay included a standard curve, a no-template control and cDNA samples in triplicate. The standard curve was generated by serial 5-fold dilutions of pooled cDNA obtained from gastric tissues that were found to contain high levels of mRNAs of both genes. Contents of the tubulin alpha 3 and IL-8 cDNAs were expressed in arbitrary units calculated according to the standard curve. The relative expression level of IL-8 was expressed as the ratio of IL-8/tubulin alpha 3 in arbitrary units^[13].

RT-PCR

Based on the technique described previously^[14] with slight modification, the target sequence of CXCR-1 mRNA was amplified through 35 cycles, each consisting of denaturation at 94 °C for 30 sec, annealing at 53 °C for 30 sec and extension at 72 °C for 30 min, followed by a final extension at 72 °C for 5 min with specific primers (forward, 5' -CAGATCCACAGA-TGTGGGAT-3' and reverse, 5' -TCCAGCCATTCACCTTG-GAG-3') using an RT-PCR kit (Takara Shuzo Co., Otsu, Japan). Similarly, CXCR-2 mRNA expression was detected under the following conditions: amplification through 35 cycles, each consisting of denaturation at 94 °C for 30 sec, annealing at 60 °C for 30 sec and extension at 72 °C for 30 min, followed by a final extension at 72 °C for 5 min with specific primers (forward, 5' -AGCTGCTCTTCTGGAGGTGT-3' and reverse, 5' -TTAGAGAGTAGTGGAAGTGTGC-3')^[14]. A 10µl aliquot of each PCR product was analyzed by electrophoresis on 2 % agarose gel containing ethidium bromide, and the bands were examined under ultraviolet light for the presence of amplified DNA. Glyceraldehyde-3-phosphate dehydrogenase (G3PDH) gene transcript was also amplified as described previously^[15], and used as an internal control of the processed RNA for each preparation.

Detection of Helicobacter pylori infection

H pylori status was assessed by serology (anti-*H pylori* Immunoglobulin G antibody, HEL-p TEST, Amrad Co., Melbourne, Australia), rapid urease test (Helicocheck, Otsuka Pharmaceutical Co., Tokushima, Japan) and histopathology (hematoxylin-eosin and Giemsa staining) using additional biopsy specimens obtained during endoscopy from the antrum within 2 cm of the pyloric ring and the corpus along the greater curvature. Patients were considered positive for *H pylori* infection when at least two of these examinations yielded positive results. On the other hand, patients were defined as *H pylori*-negative if all the test results were negative^[16].

All the samples were obtained with written informed consent of the patients prior to their inclusion in this study, in accordance with the Helsinki Declaration.

Statistical analysis

Statistical analyses were performed using Fisher's exact, χ^2 , Student's *t*, Mann-Whitney U, and Kruskal-Wallis tests, whenever appropriate. A *P* value of less than 0.05 was accepted as statistically significant. Data are expressed as mean \pm standard deviation (SD).

RESULTS

Patient demographics

According to the modified LA system, 14 patients were classified as grade M and 12 as grade N. There were no

significant differences in age, gender, current tobacco use, alcohol intake, body mass index, the presence of hiatus hernia and *H pylori* status among the patients with grade M and N and the controls (Table 1). None had such complications as stricture, bleeding and columnar-lined esophagus. The overall incidence of *H pylori* infection in our series was 51.3 %.

Table 1 Baseline characterisitics of the enrolled subjects

	Control group - n=13	Nonerosive reflux disease group	
		Grade M ^a n=14	Grade N ^a n=12
Mean age, yr, (range)	61.6(39-80)	58.7(28-75)	62.5 (33-75)
Male/female	8/5	11/3	8/4
Smoker	53.8 %(7/13)	28.6 %(4/14)	33.3 %(4/12)
Alcohol drinker	46.2 %(6/13)	57.1 %(8/14)	33.3 %(4/12)
Hiatal hernia	0 %(0/13)	35.7 %(5/14)	50.0 %(6/12)
H pylori infection	53.8 %(7/13)	57.1 %(8/14)	41.7 %(5/12)

According to the modified Los Angeles system.

Relative expression levels of IL-8

We confirmed that both RT-PCR procedures for IL-8 and tubulin alpha 3 yielded 87- and 101-base pair (bp) specific bands, respectively (data not shown). As a whole, NERD patients had significantly higher expression levels of IL-8 than the controls (Figure 1, P<0.05). The expression levels of IL-8 in esophageal mucosa of grade M patients with NERD were significantly higher than those of grade N patients (P<0.05, Figure 2). In addition, the expression levels of IL-8 were higher in grade M than control group (P<0.01, Figure 2), but not significantly different between NERD-grade N and control group.



Figure 1 Relative interleukin 8 (IL-8) mRNA expression levels assessed by real-time quantitative polymerase chain reaction in patients with non-erosive reflux disease (NERD) and asymptomatic controls.



Figure 2 The expression levels of IL-8 assessed by real-time quantitative polymerase chain reaction in patients with non-erosive reflux disease (NERD) of grade M and N and asymptomatic controls.

Expression of CXCR-1 and CXCR-2 genes in esophageal mucosa

We identified the *CXCR-1* and -2 gene-specific products as 257- and 1 154-bp bands, respectively, by RT-PCR (Figure 3). These mRNAs were constitutively expressed in each subject examined, irrespective of the presence of GERD-related symptoms or endoscopic grading. We confirmed that RT-PCR procedures for *G3PDH* housekeeping gene expression yielded 983 bp specific bands (data not shown).



Figure 3 CXCR-1 and -2 mRNA transcripts were detected as 257 and 1 154 base pair (bp) bands with reverse transcriptionpolymerase chain reaction, respectively. Lane N; negative control, lane N'; RT (-) negative control, lane 1; asymptomatic normal control, lane 2; patient with NERD grade M, lane 3; patient with NERD grade N.

DISCUSSION

Several lines of evidence indicated that a prototype of CXC chemokine, IL-8, played a crucial role in the development and progression of erosive esophagitis^[7,8]. In the present study, we focused on the implication of this potent inflammatory mediator in NERD. We found significantly higher expression levels of IL-8 mRNA in esophageal mucosa of patients with NERD than those in asymptomatic controls, suggesting that IL-8 is implicated in the pathogenesis of this incipient form of GERD.

The striking finding of this study was that NERD patients classified as grade M subgroup based on the modified LA system had significantly higher expression levels of IL-8 mRNA compared to those of grade N. IL-8 mediated the recruitment of neutrophils into sites of inflammation^[17]. In addition, this potent chemoattractant acted on neutrophils to respiratory burst and release of a variety of reactive oxygen species (ROS), leading to tissue damage^[18]. In our previous work employing ELISA, we found a significant association between the presence of intraepithelial neutrophils and increased IL-8 levels in esophageal mucosa of patients with RE^[13]. Although we did not perform histopathological evaluation in the current study, elevated mucosal IL-8 expression may be involved even in such subtle mucosal changes as seen in the grade M subcategory, probably through its action on neutrophils, thus triggering chemotaxis and generating harmful ROS. This result also highlighted the possibility that NERD patients could encompass diverse subpopulations in terms of immune and inflammatory reactions. Further studies on the implication of other members of chemokines and proinflammatory cytokines can shed light on our understanding of the mechanisms underlying this poorly studied disorder.

In the present study, we demonstrated constitutive mRNA expression of CXCR-1 and -2 in esophageal biopsy samples by RT-PCR procedure. To date, CXCR-1 and -2 are two distinct receptors for IL-8^[19]. It is suggested that the increased IL-8 may facilitate trafficking of neutrophils into the mucosa

affected by GERD process through the interaction with these receptors. Again, recent data from our laboratories showed a significant correlation between IL-8 protein levels and basal layer hyperplasia as well as papillary elongation in patients with RE^[13]. IL-8 also exerted mitogenic actions directly or by binding to its receptors on epithelial cells^[20,21]. Taken together, it is possible that IL-8, together with other cytokines as well as growth factors^[14,22], could contribute to epithelial cell proliferation even in NERD, and could be eventually linked to carcinogenesis. Again, unlike CXCR-1, CXCR-2 is not specific for IL-8 and can bind to other chemokines such as growth related oncogene α, but it has 2- to 5-fold higher affinity for IL-8 than CXCR-1^[19]. Further studies on the distribution of diverse IL-8 receptors and the receptor-mediated signaling pathway may help to elucidate the pathogenesis of GERD via IL-8 action.

In conclusion, our study demonstrated significantly enhanced expression of IL-8 mRNA level in NERD by realtime PCR technology. The interaction of IL-8 with CXCR-1 and -2 is likely to be involved in the pathogenesis of NERD. We also showed a significant difference in IL-8 mRNA levels between grade M and N subgroups of the modified LA classification, indicating the heterogeneity of NERD patients both immunologically and endoscopically.

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