

Research article

Effects of prandial challenge on triglyceridemia, glycemia, and pro-inflammatory activity in persons with chronic paraplegia

Dennis Ellenbroek^{1*}, Jochen Kressler^{2*}, Rachel E. Cowan^{2,3}, Patricia A. Burns², Armando J. Mendez^{4,5}, Mark S. Nash^{2,3,6}

¹Department of Physiology, Radboud University Nijmegen Medical Centre, Nijmegen, The Netherlands, ²The Miami Project to Cure Paralysis, University of Miami, Miami, FL, USA, ³Department of Neurological Surgery, University of Miami, Miami, FL, USA, ⁴Department of Medicine, University of Miami, Miami, FL, USA, ⁵Diabetes Research Institute, University of Miami, Miami, FL, USA, ⁶Department of Rehabilitation Medicine, Miller School of Medicine, University of Miami, Miami, FL, USA

Context/Objective: Exaggerated postprandial lipemia has been reported after spinal cord injury (SCI). We examined metabolite and accompanying pro-inflammatory biomarker responses to repeat feeding of typical high-fat meals in individuals with chronic paraplegia.

Design: Descriptive trial.

Methods: Metabolites (triglycerides, glucose, and insulin) and inflammatory biomarkers (interleukin-6 and high-sensitivity C-reactive protein (hsCRP)) were measured under fasting conditions in 11 recreationally active individuals with chronic (>1 year) paraplegia. Subjects received high-fat meals at time point 0 and again at minute 240. Antecubital venous blood was obtained at time points -30 (fasting), 0 (first meal), 30, 60, 90, 120, 240 (second meal), 360, and 480 minutes. Correlations were examined among the study variables. Exploratory subgroup analysis was performed for subjects with levels of postprandial glucose greater than >200 mg/dl.

Results: Triglycerides showed a significant rise 4 hours after eating. Basal inflammatory markers were elevated, and did not undergo additional change during the testing. Additionally, subjects with excessive postprandial glucose responses showed higher hsCRP levels than those having typical glucose responses both for fasting (11.8 ± 6.5 vs. 2.9 ± 2.7 mg/l, $P = 0.064$) and postprandial (11.1 ± 4.9 vs. 3.7 ± 3.8 mg/l, $P = 0.018$) values.

Conclusions: Despite elevations in metabolic response markers, inflammatory markers did not change significantly after consumption of population-representative (i.e. hypercaloric) mixed-nutrient meals. Levels of fasting CRP in the high-risk range are consistent with other reports in persons with SCI and continue to pose concern for their cardiovascular disease risk. The possible association between postprandial metabolic responses and inflammatory states warrants further investigation to identify individual component risks for this secondary health hazard.

Keywords: Feeding, Meal challenge, Glucose, Lipids

Introduction

Persons living with a spinal cord injury (SCI) are reported to undergo accelerated rates of organ system

aging,^{1,2} and in the process sustain an increased risk for early all-cause cardiovascular disease (CVD).³ CVDs are now the primary cause of morbidity in those older than age 60, or living more than 30 years with SCI.⁴ Causes for this increased risk have yet to be established, although well-established CVD risks of physical inactivity,⁵ overweight,⁶ impaired fasting

*These authors have contributed equally to this work.
Correspondence to: Jochen Kressler, The Miami Project to Cure Paralysis, University of Miami, 1095 NW 14th Ter, Miami, FL 33136, USA.
Email: jkressler@med.miami.edu

glucose or diabetes,^{6,7} elevated pro-atherogenic inflammatory cytokines,^{8,9} and imprudent diet^{6,10} have all been suggested as component hazards. Evidence confirms the tendency for these individual component risks to cluster after SCI,¹¹ making accelerated CVD a credible health hazard.

Among the most common of CVD risks reported after an SCI is a fasting dyslipidemia, which is typically associated with low blood levels of the cardioprotective high-density lipoprotein cholesterol (HDL-C).^{3,12,13} While longstanding fasting lipid disorders are often considered a prerequisite for atherogenesis, and the CVD hazard imposed by low HDL-C is forcefully expressed in authoritative risk guidelines,¹⁴ almost half of individuals sustaining 'hard' CVD events (i.e. myocardial infarction, sudden death, or stroke) have fasting blood lipid profiles that would not predict these episodes.¹⁵ This observation has caused investigators to search for more hidden risks as potential instigators for CVD.

While assessment of lipids in the fasting state is a recommended approach for clinical CVD risk assessment,¹⁴ the fact remains that humans primarily live in the *fed*, not the fasted condition. For this reason, considerable interest has been generated in the postprandial period when levels of glucose and triglycerides (TGs) become predictably elevated.¹⁶ Postprandial TG elevation in particular poses risks for oxidation to TG-rich lipoproteins, which then through remnant lipoproteins hasten atheroma formation.¹⁷ Recent evidence also suggests that the postprandial period is associated with increased cytokine activity, which may be associated with TG appearance or delayed removal, as well as persistently elevated glycemia.¹⁸

Nash *et al.*¹⁹ have previously reported an exaggerated postprandial hypertriglyceridemia in persons with SCI. Others reported similar postprandial responses between person with SCI and AB controls using the same prandial challenge of premium ice cream and heavy whipping cream.²⁰ While this particular nutrient intake is commonly used in clinical studies examining prandial challenge, its composition represents a 'high fat load' to a greater extent than a typical meal consumed by persons in real life. Additionally, both studies examined a 6-hour interval ensuing a single feeding. In this study we adopted a more representative approach using a fast food breakfast, followed four hours later by re-feeding with a fast-food lunch. This method of prandial testing has been reported,²¹ better matches the recently reported macronutrient intake that is typical of persons with SCI,⁶ and elevates postprandial TGs, carbohydrates, and pro-inflammatory cytokines^{18,22} and is more in line with general eating

habits of most people. Thus, the purpose of this study was to examine the degree to which the postprandial period in persons with SCI was associated with prolonged elevation of serum TG and glucose, and whether these responses are associated with elevated levels of pro-atherogenic inflammatory cytokines.

Methods

Subjects

Study participants were nine men and two women aged 20–53.6 years with chronic motor-complete SCI (ASIA Impairment Scale A-B) between the T4 and L1 levels. Participants were healthy, community-dwelling individuals with a SCI for longer than 1 year. Exclusion criteria were participation in an endurance training program or a self-initiated program of exercise conditioning; habitual use of steroidal or non-steroidal anti-inflammatory drugs; use of lipid-lowering agents or anti-hyperglycemics; and use of antioxidant vitamins or nutrient supplements. Data from one participant were excluded because of pronounced postprandial hyperinsulinemia suggesting profound insulin resistance. Informed consent was obtained in accordance with Institutional Review Board guidelines. Descriptive characteristics of the study subjects are shown in Table 1.

Anthropometric measurements

Body mass was determined by weighing participants in their wheelchair on a calibrated electronic scale and then subtracting the wheelchair mass. Height was determined by participant report. Body mass index (BMI) was calculated as the quotient of body mass (kg) and the square of height (meters). Body composition (including total body fat and abdominal fat) was determined by a dual-energy X-ray absorptiometry as previously described in persons with SCI by Spungen *et al.*²³

Phlebotomy and postprandial testing

Study participants refrained from caffeine and alcohol intake for 24 hours before testing, and began the procedure following a 10-hour overnight fast. Preparation for phlebotomy included antiseptic placement in a superficial arm vein of a 21-gauge × 1-inch Teflon

Table 1 Descriptive characteristics of the study subjects

| | Range | Mean ± SD |
|----------------------------|-----------|-------------|
| Age (years) | 20–54 | 39 ± 13 |
| Level of injury | T4–L1 | N/A |
| Duration of injury (years) | 1.1–33.7 | 10.8 ± 10.5 |
| Body mass (kg) | 60–113 | 83 ± 19 |
| Body mass index | 22.0–44.8 | 29.3 ± 6.7 |
| Body fat (%) | 24–59 | 38 ± 9 |

Table 2 Macronutrient composition of the test meals

| | Kilocalories (kcal) | Fat | | Carbohydrate | | Protein (g) |
|-------------|---------------------|-----|----|--------------|------|-------------|
| | | g | % | g | % | |
| First meal | 820 | 31 | 34 | 119 | 58 | 15 |
| Second meal | 1190 | 45 | 34 | 163 | 54.7 | 33 |

catheter (Jelco, Smiths Medical, London, UK), which was capped with a multi-sample port and kept patent with sterile physiological (0.9%) saline. Blood samples were obtained 30 minutes before the test meal was consumed (−30 minutes), immediately following food intake (0 minute) and at the following time points thereafter: 30, 60, 90, 120, 180, 240, 300, 360, 420, and 480 minutes.

The first test meal consisted of a fast-food breakfast containing a sausage-egg-muffin sandwich, hash brown potato, and an oral glucose tolerance test drink containing 75 g of dextrose, which was consumed over a 15-minute period. A similar approach to prandial testing using fast-foods has been reported.²¹ After obtaining the blood sample at the 4-hour time point, participants were fed a second fast-food meal consisting of a quarter pound (pre-cooking weight) hamburger with one slice of American cheese, a medium order of French fried potatoes, and another oral glucose tolerance test drink. Macronutrient composition of the meals is shown in Table 2.

Blood samples were collected in clot lysis activator (serum) and citrated (plasma) Vacutainer tubes. To isolate the serum, the blood samples were centrifuged for 15 minutes at 1500 g. Serum TG and high-sensitivity C – reactive protein (CRP) were measured using the Roche Cobas 6000 Analyzer (Roche Diagnostic Systems, Indianapolis, IN) using manufacturer reagents and procedures. Plasma interleukin (IL)-6 samples were assayed in duplicate using an enzyme-linked immunosorbent assay kit (Quantikine HS6000, R&D System, Minneapolis, MN, USA) following manufacturer's instructions.

Data analysis

Values are presented as mean ± standard deviation. Correlations among blood markers of metabolic and inflammatory responses for area under the curve (AUC) of postprandial time points (i.e. 0–480 minutes), were assessed with the Pearson product–moment correlation coefficient. Changes for each measure across time points were analyzed with univariate analysis of variance with repeated measures, followed by *post hoc* analysis without adjustment. Level of significance for all analysis was set *a priori* at $\alpha = 0.05$. To further explore the influence of prandial glycemia on pro-

inflammatory responses, study participants were stratified into two groups based on postprandial glucose levels with a cutoff threshold at ≥ 200 mg/dl (at any time point). This post-load glycemia level represents an American Diabetes Association standard for provisional diagnosis of diabetes mellitus.²⁴ The grouping variable was added as a between-subjects factor in the repeated measure analysis described above.

Results

Response to feeding

Glucose rose significantly with each feeding and returned to baseline values within 4 hours after the first meal, but not the second. Insulin tracked the changes in postprandial glucose but remained slightly elevated over baseline values (Fig. 1A). TG levels were significantly higher 4 hours after the first feeding, and were further elevated 2 and 4 hours after the second feeding (Fig. 1B).

Changes in inflammatory markers following the feedings were small and did not reach statistical significance for the assessed time period (Fig. 1C).

Correlations

The correlation matrix for all inflammatory biomarkers (IL-6 and CRP) and metabolic response markers (TG, glucose and insulin) is presented in Table 3.

Exploratory analysis

Exploratory analysis revealed two subgroups of participants. One subgroup experienced an exaggerated glucose response (EGR) reaching glucose levels > 200 mg/dl during the testing, while another subgroup had a more typical glucose response (TGR), i.e. glucose levels remained < 200 mg/dl. Significant interactions for these subgroups in glucose and insulin response over time were observed, and simple effects for each subgroup are presented in Fig. 2A. In addition, basal CRP values were significantly higher in the EGR vs. the TGR group (EGR = 11.2 ± 4.08 TGR = 3.61 ± 4.08 P = 0.016, Fig. 2C).

Discussion

The key finding of this study is that postprandial glucose, insulin, and TGs all increased in response

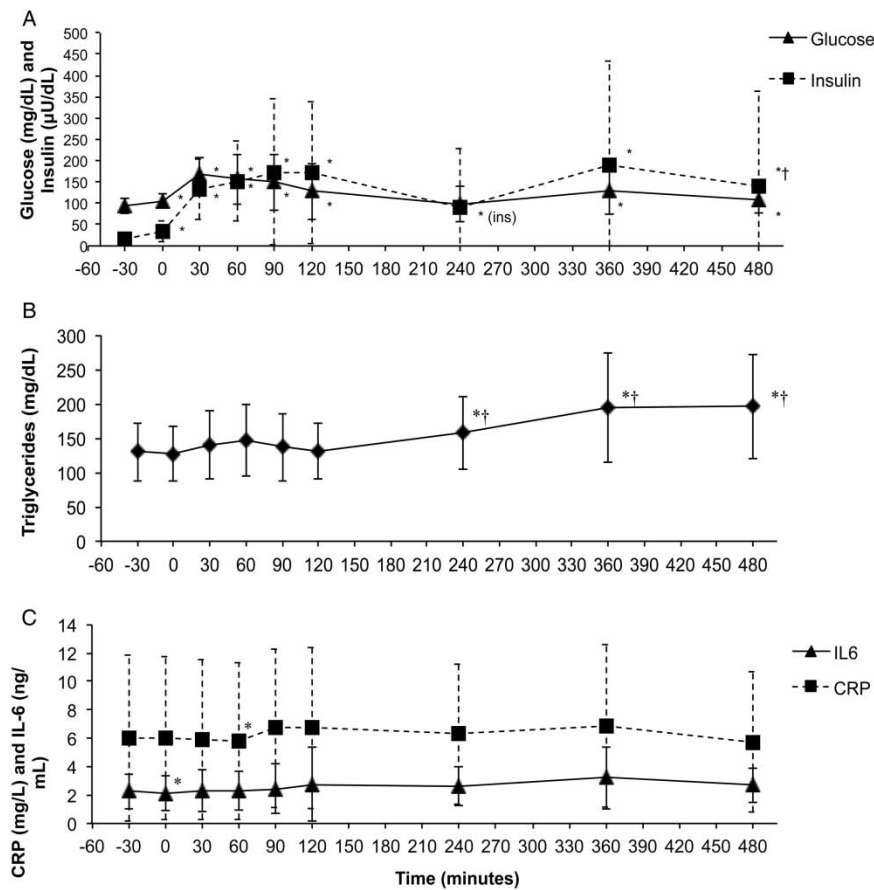


Figure 1 Metabolic and Inflammatory response markers following ingestion of high-fat meals in persons with SCI. Meals were consumed at 0 and 240 minutes. Values are mean ± SD. *Significant difference compared with -30 minutes (P < 0.05). †Significant difference compared with 240 minutes (P < 0.05).

to multiple feedings with population-representative (i.e. hypercaloric) mixed-nutrient meals, while biomarkers of inflammation did not significantly change over time.

All-cause CVD represents an accelerated health risk for all persons with chronic SCI,^{25,26} although the basis for hastening of disease remains obscure. While the component risk of fasting dyslipidemia has been extensively documented in the population,^{3,12,27} more recent interest has focused on the postprandial period,

where excessive prandial challenge is known to stimulate lipid oxidation, remnant lipoprotein evolution, and ultimately atheroma formation.^{17,28,29} Inflammation plays an important role in this process, and recent evidence suggests that the postprandial period is associated with increased cytokine activity,³⁰ which is possibly related to TG appearance or delayed clearance. Exaggerated postprandial TG response to a single dosing of a high-fat liquid meal (92% of calories derived from fat) has been reported in persons with SCI.¹⁹ Others reported TG levels for person with SCI that while on average higher at all measurement time points were not statistically different from AB controls and also showed a similar postprandial response between the two groups.²⁰ As a high-fat liquid nutrient challenge is unrepresentative of a typical diet and feeding regimen, we therefore administered two fast food meals (4 hours apart), which is similar in caloric level and macronutrient composition to intake for this population.⁶ This study was the first to use a meal that represented more typical composition, and an anticipated re-feeding four hours later.

Table 3 Correlation matrix for postprandial AUC values

| | | Glucose | Insulin | CRP | IL-6* |
|---------------|---|---------|---------|-------|---------|
| Triglycerides | r | 0.359 | 0.269 | 0.363 | -0.053 |
| | P | 0.252 | 0.398 | 0.246 | 0.871 |
| Glucose | r | 1 | 0.661† | 0.197 | -0.596† |
| | P | | 0.019 | 0.540 | 0.041 |
| Insulin | r | | 1 | 0.149 | -0.433 |
| | P | | | 0.643 | 0.160 |
| CRP | r | | | 1 | 0.025 |
| | P | | | | 0.940 |

*Statistically significant at P < 0.05; CRP, C-Reactive protein; IL-6, interleukin-6.

†Values were Ln transformed when departing from normality.

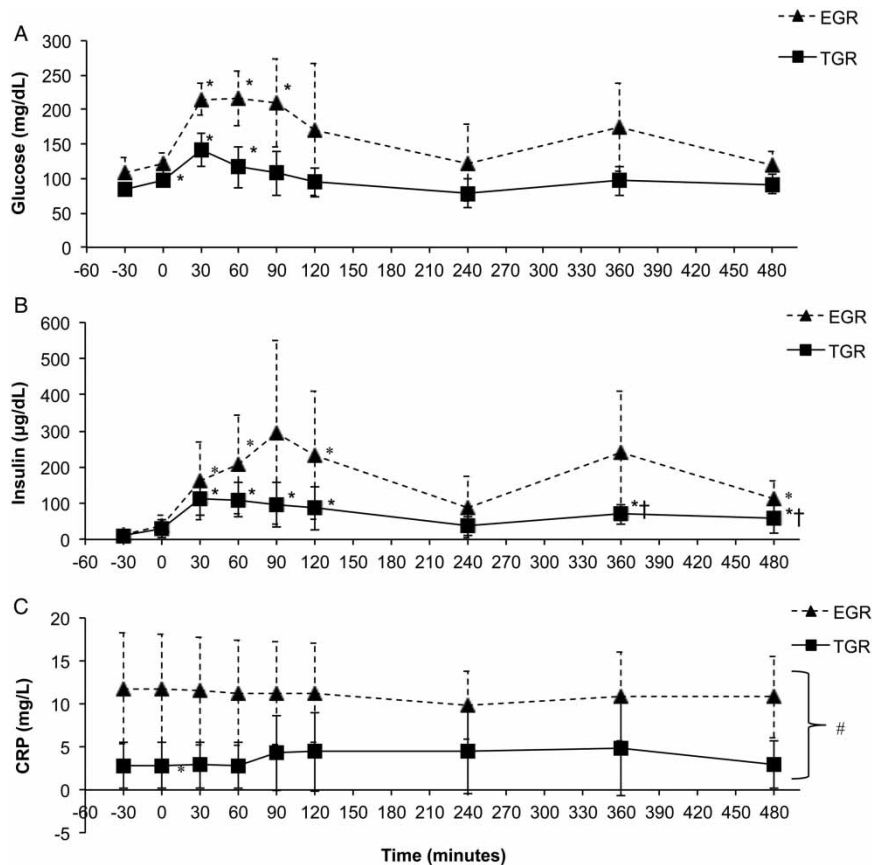


Figure 2 Glucose (A), insulin (B), and CRP (C) responses to ingestion of high-fat meals in persons with SCI. Subjects were stratified by exaggerated glucose response (EGR) and typical glucose response (TGR). Meals were consumed at 0 and 240 minutes. Values are mean \pm SD. *Significant difference compared to -30 minutes ($P < 0.05$). †Significant difference compared to 240 minutes ($P < 0.05$). #Significant main effect for group ($P < 0.05$).

Metabolic response markers examined in this study adhered to expected tendencies with glucose rising post-challenge and returning to baseline values within 4 hours (Fig. 1A). Insulin response generally followed the same trajectories as those observed for glucose although values remained slightly elevated over baseline levels 4 hours after initial feeding (Fig. 1A). TG levels were significantly increased 240 minutes after the first feeding and were further elevated at 360 minutes and at 480 minutes (i.e. 2 and 4 hours after the second feeding, Fig. 1B). The delayed appearance of plasma TG is an anticipated feature of food intake, and is expected to occur more slowly than glucose appearance. The response to the first feeding is consistent with previous reports^{20,31,32} while the response to the second finding presents novel information. Others have observed excessive rises in TG after 120 minutes compared with control in patients with SCI¹⁹ premature CVD³³ or obesity.³⁴ While the change in TG levels did not reach statistical significance until 240 minutes after the first meal, changes observed after 120 minutes did not appear excessive, and after the second feeding

there was no further increase from 360 to 480 minutes. However, due to lack of a control group this is not conclusive evidence for the lack of an excessive postprandial TG. Moreover, the extended TG elevation accompanying a second feeding may delay TG removal necessary to lessen oxidation to TG-rich lipoproteins, whose remnant lipoproteins carry high risk for atherosclerosis the longer they remain in general circulation.³⁵

Postprandial TG responses of persons with SCI have been reported to be related to visceral abdominal fat,²⁰ although this observation was based upon a feeding challenge that was both liquid in composition and abnormally high in saturated fat. Others have reported similar associations for non-disabled individuals.^{30,36,37} To the contrary, no such associations between TG and either BMI or body mass were observed in this study, although the test meal was both lower in total fat than previously studied and solid in composition. We believe this explains the slower rise in serum TG than previously reported in persons with SCI undergoing prandial challenge.^{19,20} While our postprandial TG and glucose responses appeared to be independent of

anthropometrics (although this did not include direct measures of visceral fat), insulin AUC was significantly associated with both BMI and body mass. Similar significant correlations between insulin response and visceral fat or waist-hip ratio have been reported in non-disabled individuals having wide ranges of body composition.^{36,37}

In this study we found no significant IL-6 and CRP responses to prandial challenge. Others³⁸ have also reported no significant changes in IL-6 and CRP after a high fat meal (51 g of fat) but saw increased values for IL-1 β 6–8 hours post-challenge. Still others have identified significant increases in IL-6 using a 50–65 g fat intake challenge in persons with premature CVD, type 2 diabetes compared with healthy controls.^{18,33} Similarly, metabolic disease states such as insulin resistance³⁹ and the metabolic syndrome⁴⁰ are associated with elevated CRP levels in persons with SCI. Despite the non-significant increases in IL-6, IL-6, and glucose postprandial responses were negatively correlated (Table 1). This is contrary to some reports that identify correlations between elevated fasting^{41,42} and post-challenge⁴³ glucose and CRP levels even absent of overt metabolic disease. Insulin, on the other hand, has been reported to have direct or indirect anti-inflammatory properties, most likely via a glucose lowering effect.^{29,44–46} Our data did show a negative (though non-significant) correlation of insulin and IL-6 ($r = -0.433$, $P = 0.160$) but does not support a pro-inflammatory effect of glucose in this sample. Future studies employing more complex analysis (i.e. beyond simple correlations) will have to detangle the interrelations among metabolic and inflammatory response markers in this population.

As a consequence of the large variability, we observed in baseline and post-feeding glucose and insulin values an exploratory analysis was performed that identified two subsets of participants with postprandial glucose levels greater than 200 mg/dl (EGR, $n = 4$) or less (TGR, $n = 7$). The EGR subset was significantly older and had higher BMI compared with TGR (Table 4). Significant interactions for these groups in glucose and insulin response over time were observed and simple effects for each group are presented in (Fig. 2A). No group \times time interaction was observed for either inflammatory biomarker, or for TG. However, CRP (but not IL-6) values were consistently higher for the EGR group in line with the known association between glucose and CRP^{41–43} (but not IL-6). It is noteworthy that both subgroups had average CRP values in the high-risk range CVD according to the American Heart Association⁴⁷ but levels were almost three times greater in the EGR versus TGR demonstrating a

Table 4 Descriptives for postprandial glucose response groups

| | Postprandial glucose response group | | Independent sample <i>t</i> -test <i>P</i> |
|----------------------------|-------------------------------------|------------------------|---|
| | EGR (<i>n</i> = 4) | TGR (<i>n</i> = 7) | |
| Age (years) | 49 \pm 4 | 32 \pm 12 | 0.014* |
| Level of injury | T5–L1 | T4–10 | N/A |
| Duration of injury (years) | 14.4 \pm 15 | 8 \pm 6 | 0.345 |
| Body mass (kg) | 89 \pm 21 | 78 \pm 19 | 0.349 |
| Body mass index | 34 \pm 7 | 26 \pm 5 | 0.037* |
| Body fat (%) | 42 \pm 10 | 33 \pm 7 | 0.082 |
| HOMA-IR | 4.8 \pm 3.4 | 2.4 \pm 2.0 | 0.142 |

Values are mean \pm SD; HOMA-IR, homeostatic model assessment-insulin resistance.

*Statistically significant at $P < 0.05$.

possible compounding effect of SCI and postprandial hyperglycemia. Interestingly, only two of the participants in the EGR and 1 in the TGR exhibited fasting hyperglycemia (i.e. >100 mg/dl) suggesting that postprandial glucose response may be a more sensitive predictor of inflammation than fasting glucose levels in this population. A sustained elevation of CRP concentration as observed in this study has been reported by other investigators examining persons with chronic SCI^{8,9,47,48} and has been reviewed by several authors.^{8,11} This feature of pro-inflammatory activity is a recognized risk factor for, and potential instigator of cardiovascular and pulmonary disease,^{49–51} and is elevated in non-disabled individuals with insulin resistance and MetS, the latter showing correlation between severity of insulin resistance and inflammatory stress biomarkers.^{52–55} As these conditions are seen at elevated prevalence after SCI the associations among cardioendocrine control, inflammation, and long-term health prognosis is worthy of additional study.

Study limitations

Several factors limit this study findings. The dataset was derived from a relatively small test population, did not have a control group and as a first study we did not place limits on body mass. This made isolation of individual component risks challenging, but also made significant findings more compelling. Clinically the participants were free of infection and musculoskeletal injury that might otherwise elevate inflammatory biomarkers, although occult or subclinical conditions cannot be entirely ruled out. The possibility that subclinical bacteria and musculoskeletal injury contribute to CVD risk after SCI – beyond that sustained from vascular depots – has been suggested, and remains a potentially novel non-cardiac/vascular CVD risk in this

population. This study did not individualize caloric intake based upon participant body mass, although without prescriptive diet this would have been impractical in a mixed-nutrient test meal.

Conclusions

Persons with SCI presented with fasting levels of CRP in the high-risk range that changed little with feeding and pose concern for their CVD risk. CRP values were even higher for those who exhibited an exaggerated blood glucose response to feeding. The possible association between postprandial metabolic responses and inflammatory states warrants further investigation to identify individual component risks for this secondary health hazard.

Disclaimer statements

Contributors There are no contributors beyond the authors.

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Conflicts of interest We certify that no party having a direct interest in the results of the research supporting this article has or will confer a benefit on us or on any organization with which we are associated.

Ethics approval Signed informed consent was obtained from all subjects before the start of the study, which was approved by the University of Miami Medical Sciences Subcommittee for the Protection of Human Subjects.

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