

Cancer cells mimic *in vivo* spatial-temporal cell-cycle phase distribution and chemosensitivity in 3-dimensional Gelfoam[®] histoculture but not 2-dimensional culture as visualized with real-time FUCCI imaging

Shuya Yano^{1,2,3}, Shinji Miwa^{1,2}, Sumiyuki Mii^{1,2}, Yukihiko Hiroshima^{1,2}, Fuminaru Uehara^{1,2}, Hiroyuki Kishimoto³, Hiroshi Tazawa³, Ming Zhao¹, Michael Bouvet², Toshiyoshi Fujiwara³, and Robert M Hoffman^{1,2,*}

¹AntiCancer, Inc.; San Diego, CA USA; ²Department of Surgery; University of California San Diego; San Diego, CA USA; ³Department of Gastroenterological Surgery; Okayama University; Graduate School of Medicine; Dentistry and Pharmaceutical Sciences; Okayama, Japan

Keywords: cell cycle, chemotherapy, FUCCI, Gelfoam[®], GFP, imaging, RFP, stomach cancer, Three dimensional-histoculture culture

The phase of the cell cycle can determine whether a cancer cell can respond to a given drug. We previously reported monitoring of real-time cell cycle dynamics of cancer cells throughout a live tumor, intravitaly in live mice, using a fluorescence ubiquitination-based cell-cycle indicator (FUCCI). Approximately 90% of cancer cells in the center and 80% of total cells of an established tumor are in G₀/G₁ phase. Longitudinal real-time imaging demonstrated that cytotoxic agents killed only proliferating cancer cells at the surface and, in contrast, had little effect on quiescent cancer cells, which are the vast majority of an established tumor. Moreover, resistant quiescent cancer cells restarted cycling after cessation of chemotherapy. These results suggested why most drugs currently in clinical use, which target cancer cells in S/G₂/M, are mostly ineffective on solid tumors. In the present report, we used FUCCI imaging and Gelfoam[®] collagen-sponge-gel histoculture, to demonstrate in real time, that the cell-cycle phase distribution of cancer cells in Gelfoam[®] and *in vivo* tumors is highly similar, whereby only the surface cells proliferate and interior cells are quiescent in G₀/G₁. This is in contrast to 2D culture where most cancer cells cycle. Similarly, the cancer cells responded similarly to toxic chemotherapy in Gelfoam[®] culture as *in vivo*, and very differently than cancer cells in 2D culture which were much more chemosensitive. Gelfoam[®] culture of FUCCI-expressing cancer cells offers the opportunity to image the cell cycle of cancer cells continuously and to screen for novel effective therapies to target quiescent cells, which are the majority in a tumor and which would have a strong probability to be effective *in vivo*.

Introduction

The phase of the cell cycle can determine whether a cancer cell can respond to a given drug. Sakaue-Sawano et al. have demonstrated that the cell cycle phase in viable cells can be visualized using a fluorescent ubiquitination-based cell-cycle indicator (FUCCI) system.¹ We previously imaged real-time cell cycle dynamics of cancer cells throughout a live tumor intravitaly using FUCCI. Approximately 90% of cancer cells in the center and 80% of total cells of an established tumor are in G₀/G₁ phase. FUCCI imaging demonstrated that cytotoxic agents killed only proliferating cancer cells at the surface and, in contrast, had little effect on quiescent cancer cells, which are the vast majority of an established tumor. Moreover, resistant quiescent cancer cells

restarted cycling after the cessation of chemotherapy. However, *in vivo* systems are not amenable to continuous, long-term imaging, which can be critical for studying the cell cycle and its relationship to tumor behavior. *In vivo*-like culture systems can be of important use for long-term imaging of cancer-cell behavior.²

Collagen-sponge-gel histoculture was developed by Leighton in the 1950s.³ Sponge-gel histoculture enables cancer cells to form 3-dimensional structures. For example, Leighton observed that when C3HBA mouse mammary adenocarcinoma cells were grown on sponge-gel histoculture, the cells aggregated similar to the original *in vivo* tumor. Distinct structures were formed within the tumors such as lumina and stromal elements, with the glandular structures similar to the original tumor.⁴

*Correspondence to: Robert M Hoffman; Email: all@anticancer.com

Submitted: 10/23/2014; Accepted: 12/15/2014

<http://dx.doi.org/10.1080/15384101.2014.1000685>

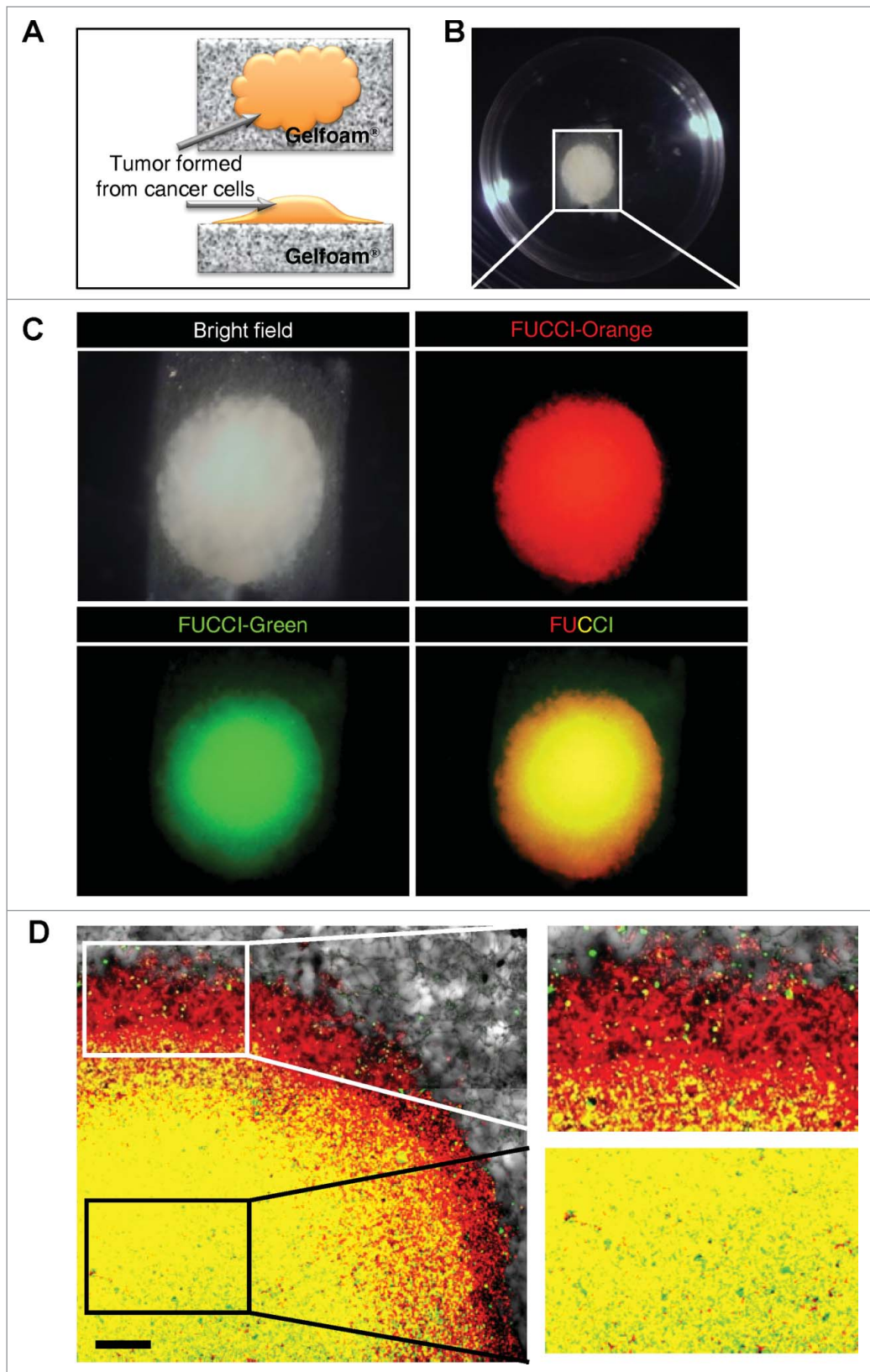


Figure 1. Gelfoam® histoculture of FUCCI-expressing cancer cells. **(A)** Schema of FUCCI-expressing MKN45 stomach cancer cells forming a tumor on Gelfoam®. **(B)** Macroscopic appearance of the tumor formed on Gelfoam® histoculture. **(C)** Macro images of a tumor formed on Gelfoam® demonstrating FUCCI fluorescence. **(D)** FUCCI-expressing cancer cells in the tumor formed on Gelfoam®. Images at the single-cell level were acquired by confocal laser-scanning microscopy. High magnification images ($\times 10$) of an invading area of the tumor (upper right) and a non-invading area (lower right) of the tumor on Gelfoam®.

We have shown that in contrast to Gelfoam® histoculture, in Matrigel culture, cancer cells formed colonies but no other structures. The behavior of human 143B osteosarcoma cells on Gelfoam® in culture was remarkably different from those of these cells in monolayer culture or in Matrigel. Tissue-like structures were observed only in Gelfoam® culture. A flexible structural substrate such as Gelfoam® provides a more in vivo-like culture condition than monolayer culture or Matrigel.⁵

We previously demonstrated, using FUCCI imaging, real-time visualization of the cell cycle kinetics of invading cancer cells in Gelfoam® histoculture. Cancer cells in G_0/G_1 phase in Gelfoam® histoculture migrated more rapidly and further than the cancer cells in $S/G_2/M$ phase. After

entry into $S/G_2/M$ phases, cancer cells ceased migrating and restarted migrating after division when the cells re-entered G_0/G_1 . Migrating cancer cells were resistant to cytotoxic chemotherapy, since they were mostly in G_0/G_1 , where cytotoxic chemotherapy is not effective.

In the present report, we compared spatial-temporal cell-cycle dynamics and chemosensitivity of cancer cells forming tumors on Gelfoam[®] with cancer cells growing in tumor spheres and on monolayers on plastic, as well as *in vivo*.

Results and Discussion

Gelfoam[®] histoculture of cancer cells

FUCCI-expressing MKN45 cells formed tumors after seeding in Gelfoam[®] histoculture. The cancer cells forming tumors on Gelfoam[®] brightly expressed either mK02-hCdt1 (green fluorescence) or mAG-hGem (orange-red fluorescence), which report the phases of the cell cycle, $S/G_2/M$ and G_0/G_1 , respectively (Fig. 1).

Comparison of cell-cycle-phase distribution of FUCCI-expressing MKN45 cells cultured in monolayer, sphere, Gelfoam[®], and *in vivo*

In monolayer culture, in both the central and edge areas, approximately 50% of the cells were in $S/G_2/M$. In tumor spheres, most of the cells were in G_0/G_1 at both the surface and center. In both *in vivo* and in Gelfoam[®] histoculture, the majority of the surface cells of the tumor were in $S/G_2/M$. In contrast, in the central area of the tumor, only

approximately 10% of the cells were in $S/G_2/M$ (Fig. 2). A comparison was made of the cell-cycle phase distribution in a subcutaneous tumor, liver tumor and Gelfoam[®], all formed from FUCCI-expressing MKN45 stomach cancer cells. At the early

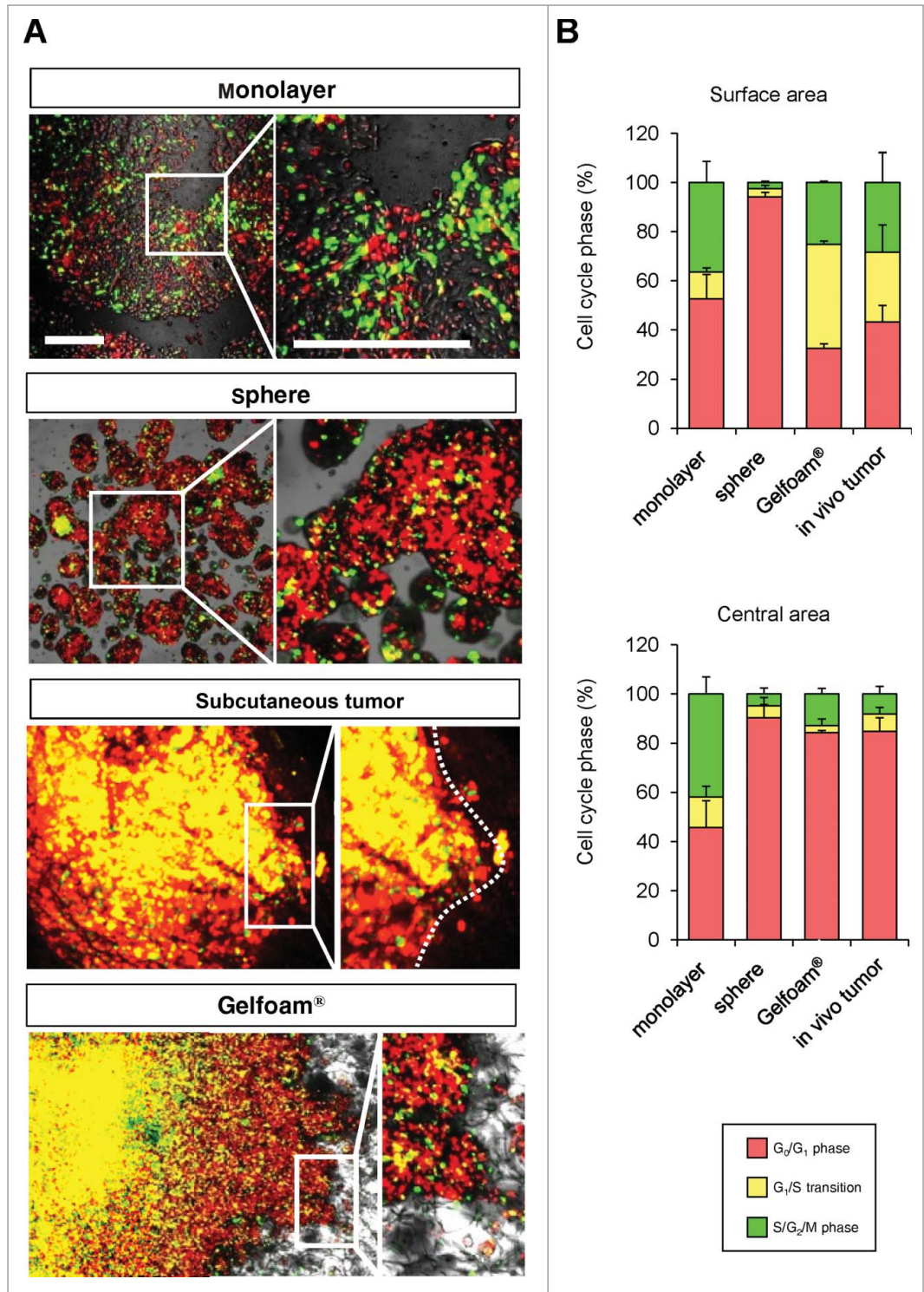


Figure 2. For figure legend, see page 811.

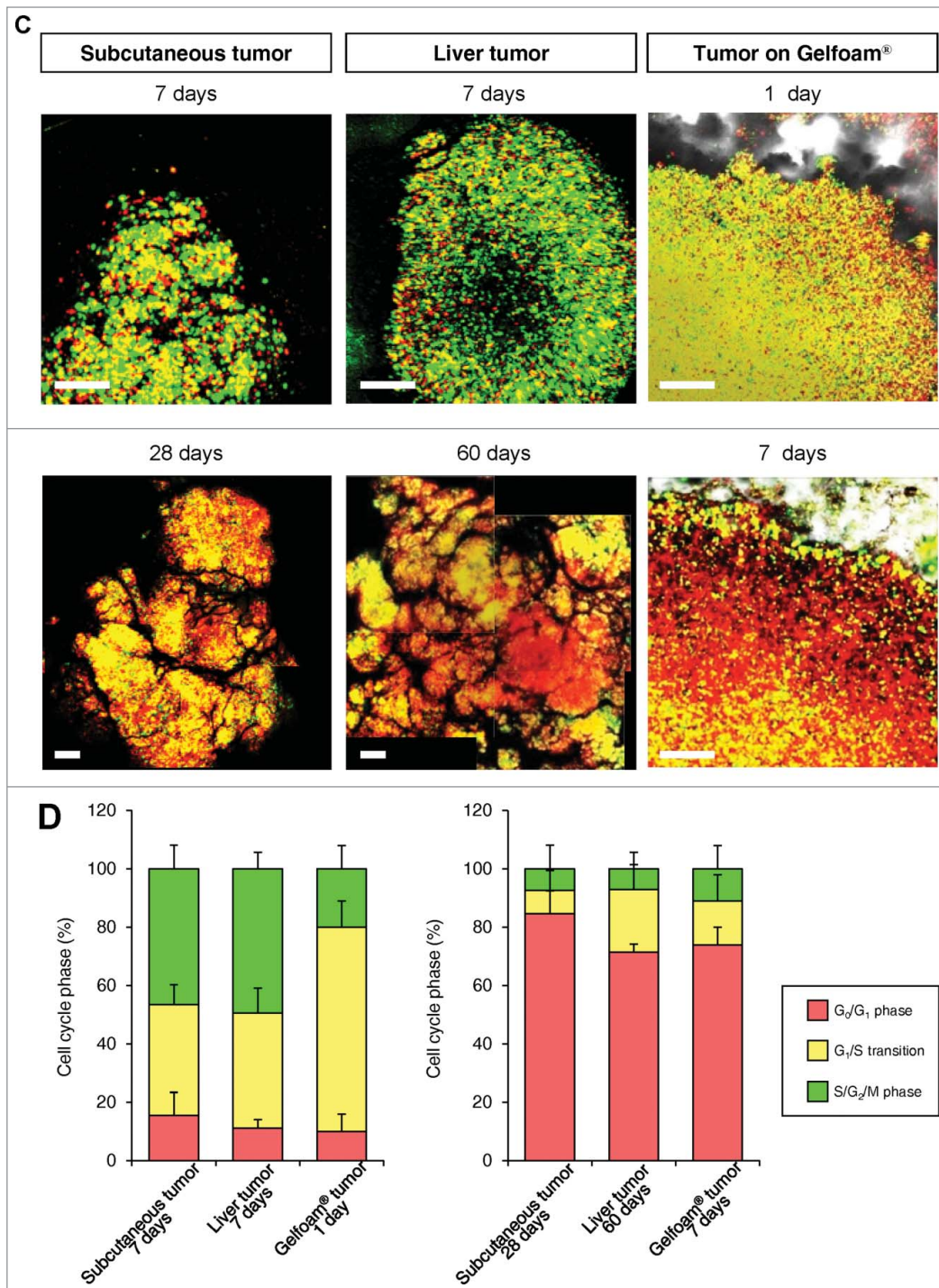


Figure 2. Comparison of cell cycle phase distribution of FUCCI-expressing MKN45 cells cultured as monolayers, and on Gelfoam®, and *in vivo*. (A) Representative images of FUCCI-expressing MKN45 cells cultured as monolayers, as spheres on agar and Gelfoam®, and *in vivo*. (B) Histograms show the cell-cycle phase distribution in the central area and invading area of the cultures and *in vivo*. Cancer cells in monolayer culture divide continuously. Cancer cells on agar aggregate and mostly remain in G₀/G₁ phase. Cancer cells on and in Gelfoam® have an *in vivo*-like cell cycle distribution. (C) Representative time-course images of FUCCI-expressing subcutaneous tumor (left), FUCCI-expressing liver tumor (middle), and tumor-formed from FUCCI-expressing cancer cells on Gelfoam® (right) at early and later stages. (D) Histograms show the cell-cycle phase distribution in early-stage and late-stage tumors growing subcutaneously, or in the liver or on Gelfoam®. Scale bars; 500µm.

stages of each tumor, whether subcutaneous or in the liver, or on Gelfoam®, approximately 90% of the cells were in S/G₂/M. In contrast as each tumor matured, approximately 80% of the cells were in G₀/G₁. The early-stages and mature-stage cell-cycle-phase distribution was very similar for each tumor, subcutaneous, liver and on Gelfoam® (Fig. 2).

Cancer cells in Gelfoam® histoculture and *in vivo* tumors have similar 3-dimensional-spatial cell-cycle phase distribution

In both tumors *in vivo* and in Gelfoam® culture, cancer cells were proliferating only near the surface of the tumor. The majority of cancer cells were in S/G₂/M both subcutaneous tumors and in Gelfoam®, as deep as 500–600 µm from the surface. At deeper levels, the vast majority of the cells were in G₀/G₁ in both tumors and on Gelfoam®. At greater depths, approximately 20% of the cells in the *in vivo* liver tumor were in S/G₂/M and in Gelfoam® histoculture, approximately 10% of the cells were in S/G₂/M, with the other cells in G₀/G₁ in both the subcutaneous tumor and on Gelfoam® (Fig. 3).

Cancer cells on Gelfoam®, but not 2D culture, have the same cell cycle response to cytotoxic agents as *in vivo* tumors

Mostly chemotherapy agents targets only proliferating cancer cells and have

little effect on quiescent cancer cells. In monolayer culture, chemotherapy blocked cancer cells in G₂/M phase. In sphere culture, chemotherapy had little effect since most cancer cells were in G₀/G₁, where they remained after chemotherapy. In Gelfoam[®] histoculture and the subcutaneous tumor, chemotherapy targeted only proliferating cancer cells and had little effect on quiescent cancer cells, which were the majority of the cells. In both the subcutaneous tumors and the tumors in Gelfoam[®] histoculture, chemotherapy killed the surface proliferating cells, but the remaining cells were blocked in G₀/G₁ and resistant to chemotherapy (Fig. 4 and Video S1).

Cancer cells on Gelfoam[®] have similar spatial-temporal recovery from cisplatin and paclitaxel treatment as *in vivo* tumors, in contrast to 2D monolayer and sphere culture

In 2D monolayer culture, approximately 50% of the cells were in S/G₂/M before chemotherapy and after chemotherapy, approximately 90% of the cells were in S/G₂/M. In sphere culture, approximately 10% of the cells were in S/G₁/M before chemotherapy, and after chemotherapy, almost 100% of the cells were in G₀/G₁. In Gelfoam[®] histoculture, approximately 40% of the cells were in S/G₀/M before chemotherapy and during chemotherapy, almost 100% of the cancer cells were in G₀/G₁ and after termination of chemotherapy approximately 20% of the cancer cells re-entered S/G₂/M, mostly at the surface, for both cisplatin and paclitaxel. In the subcutaneous tumor, before chemotherapy approximately 30% of the cells were in S/G₂/M. During chemotherapy, almost 100% of the cancer cells were in G₀/G₁. After termination of chemotherapy, approximately 20% of the cancer cells re-entered S/G₂/M for both cisplatin and paclitaxel, mostly at the surface and very similar to tumors on Gelfoam[®] (Fig. 5). After the cessation of chemotherapy, the cancer cells at the surface of the tumor resumed cycling in both the liver and on Gelfoam[®].

FUCCI imaging demonstrated that cancer cells in monolayer culture (2D culture) and spheres on agar behave very differently

than cancer cells *in vivo* or on Gelfoam[®]. Gelfoam[®] tumor histoculture provides an *in vivo*-like microenvironment for cancer

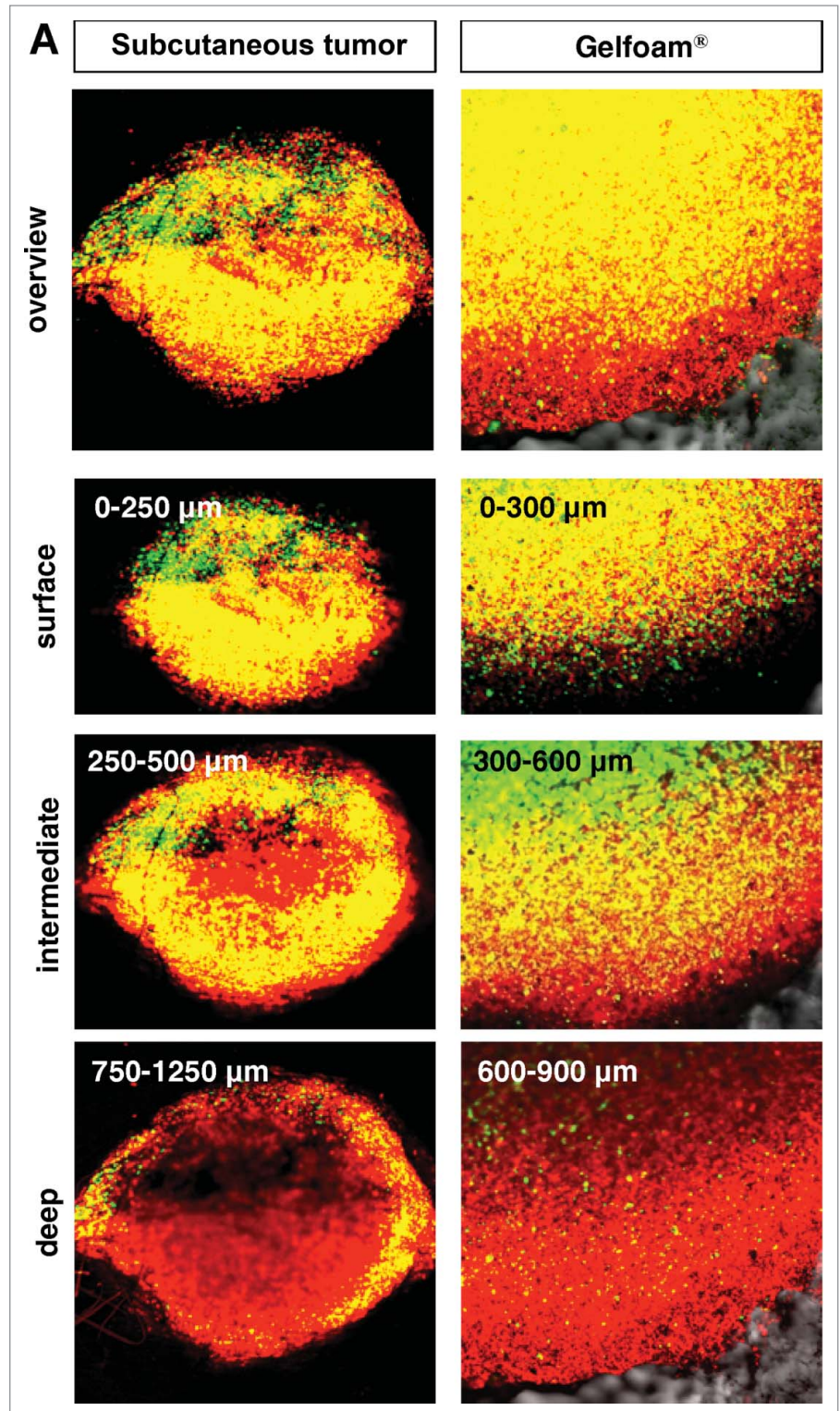


Figure 3. For figure legend, see page 813.

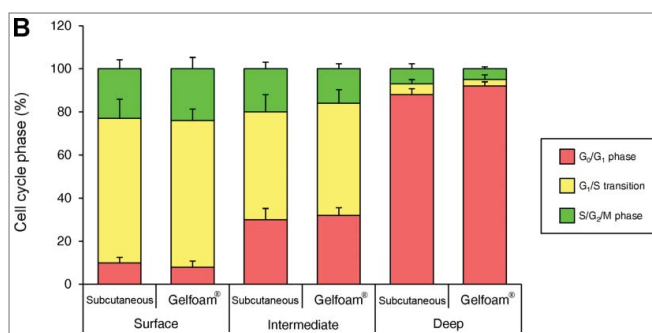


Figure 3. Gelfoam[®] histocultured tumor and subcutaneous FUCCI-expressing tumors have similar 3-dimensional spatial-temporal cell-cycle phase distribution. (A) Representative images of FUCCI-expressing MKN45 cells in a tumor in the liver and tumor-formed on Gelfoam[®] at the indicated depths. (B) Histograms show the cell-cycle distribution at the surface, intermediate area, and deep area of tumors in the liver and on Gelfoam[®]. Scale bars; 500 μ m.

cells enabling the cancer cells to mimic the spatial-temporal cell-cycle phase distribution of tumors *in vivo*.⁵

FUCCI imaging demonstrated that cancer cells on monolayer culture and spheres on agar have a different response to chemotherapy from cancer cells in a tumor *in vivo*. Gelfoam[®]-histocultured tumors, however, had a similar response to chemotherapy as *in vivo* tumors, suggesting that the combination of FUCCI imaging and Gelfoam[®] provides a new platform for developing and evaluating anticancer agents, as well as studying basic cancer biology, such as the cell cycle.⁸

An important feature of Gelfoam[®] histoculture is that it is a very convenient system for imaging long-term experiments. The possibility of long-term imaging, provided by Gelfoam[®] histoculture of FUCCI-expressing cancer cells, described in the present report, is especially important when studying the relationship of cell cycle phase to cancer-cell migration and invasion.^{6,7}

Previously-developed concepts and strategies of highly-selective tumor-targeting⁹⁻¹⁶ can take advantage of the long-term Gelfoam[®] histoculture of FUCCI-expressing cancer cells described in the present report.

Materials and Methods

Cells

MKN45 is a poorly-differentiated stomach adenocarcinoma-derived from a liver metastasis of a patient. The cells were grown in RPMI 1640 medium with 10% fetal bovine serum and penicillin/streptomycin.^{2,17}

Establishment of MKN45 cells stably transfected with FUCCI-vector plasmids

For cell cycle-phase visualization, the FUCCI (fluorescent ubiquitination-based cell cycle indicator) expression system

was used.¹ Plasmids expressing mKO2-hCdt1 (green fluorescent protein) or mAG-hGem (orange-red fluorescent protein) were obtained from the Medical & Biological Laboratory (Nagoya, Japan). Plasmids expressing mKO2-hCdt1 were transfected into MKN45 cells using Lipofectamine[™] LTX (Invitrogen, Carlsbad, CA). The cells were incubated for 48 h after transfection and were then trypsinized and seeded in 96-well plates at a density of 10 cells/well. In the first step, cells were sorted into green (S, G₂, and M phase) cells using a FACSaria cell sorter (Becton Dickinson, Franklin Lakes, NJ). The first-step-sorted green-fluorescent cells were then re-transfected with mAG-hGem and then sorted by orange fluorescence.^{1,2,6}

Gelfoam[®] histoculture

Sterile Gelfoam[®] sponges (Pharmacia & Upjohn, Kalamazoo, MI), prepared from porcine skin, were cut into 1 cm cubes. The Gelfoam[®] cubes were placed in 6-well tissue-culture plates. RPMI 1640 medium was added and Gelfoam[®] was incubated at 37°C in order that the Gelfoam[®] absorbed the medium. Cancer cells (1×10^6) expressing FUCCI were then seeded on top of the hydrated Gelfoam[®] and incubated for 1 h. Medium was carefully added up to the top of the Gelfoam[®]. Cells were incubated at 3°C in a humidified incubator with 5% CO₂.^{5,6,18-21}

Animal experiments

Athymic *nu/nu* nude mice (AntiCancer, Inc., San Diego, CA) were maintained in a barrier facility under HEPA filtration and fed with autoclaved laboratory rodent diet (Teklad LM-485; Harlan Labs, Hayward, CA). All animal studies were conducted in accordance with the principles and procedures outlined in the National Institute of Health Guide for the Care and Use of Animals under Assurance Number A3873-1.

Tumor models

All animal procedures were performed under anesthesia using s.c. administration of a ketamine mixture (10 μ l ketamine HCl, 7.6 μ l xylazine, 2.4 μ l acepromazine maleate, and 10 μ l PBS) (Henry-Schein, Melville, NY). FUCCI-expressing MKN45 cells were harvested by brief trypsinization. Single-cell suspensions were prepared at a final concentration of 2×10^5 cells/5 μ l Matrigel (Becton Dickinson). After laparotomy of 5-week-old female nude mice, the mouse liver was exteriorized and the cancer cells subserosally injected directly into the left lobe of the liver using a 31-gauge needle. After cancer cell implantation, the abdominal wall of mice was closed with 6-0 sutures.² In order to obtain subcutaneous tumors, FUCCI-expressing MKN45 cells (5×10^6 /mouse) were injected in the flank of 5-week-old female nude mice (under the ketamine-mixture anesthesia previously described).

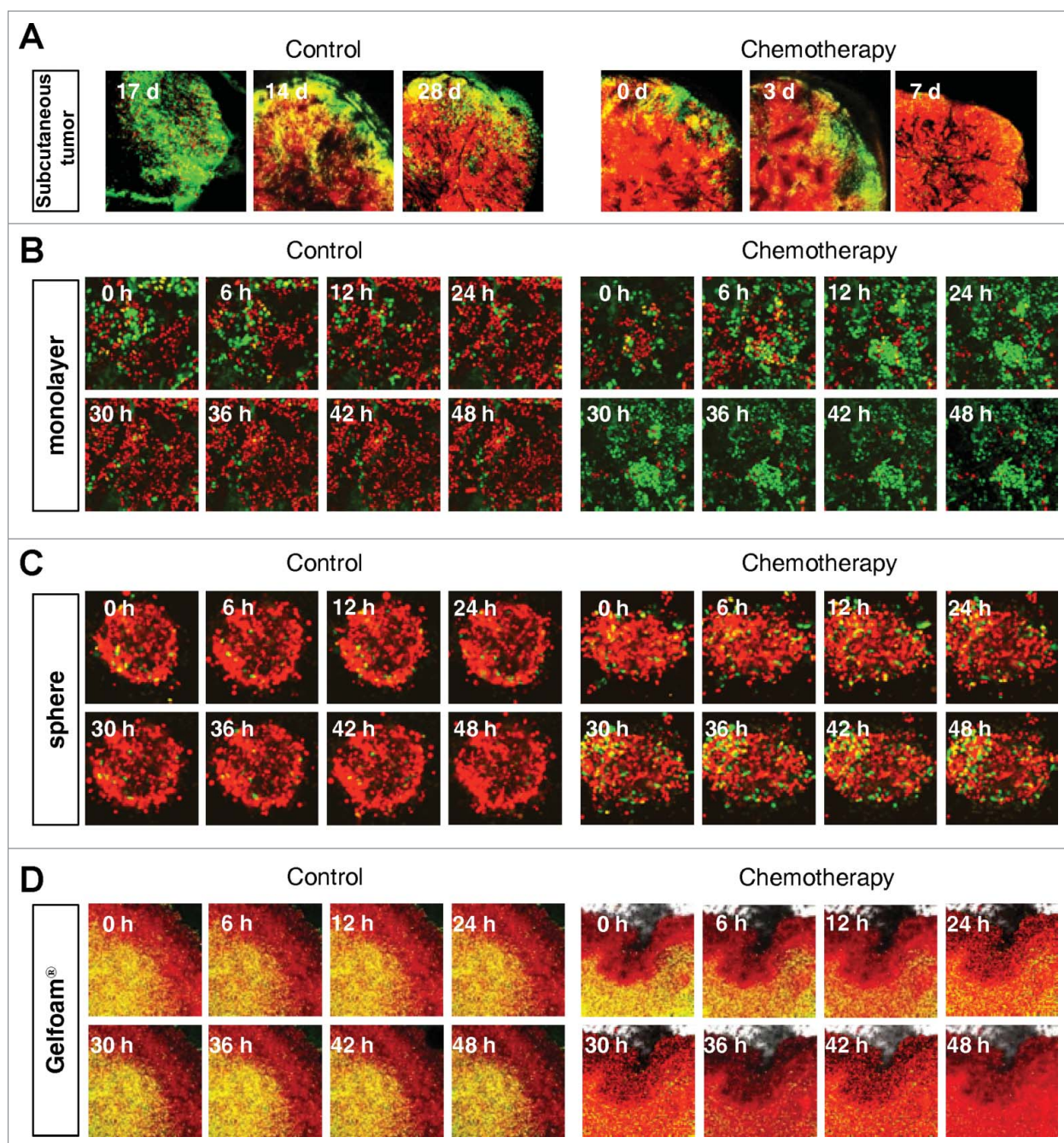


Figure 4. For figure legend, see page 816.

Confocal laser microscopy

The liver, with a Fucci-expressing tumor, was exteriorized and a cover glass was gently put on the liver, which inhibited vibration caused by heartbeat and respiratory movement. Subcutaneous tumors, and spheres, 2D monolayer and Gelfoam® cultures with Fucci-expressing cancer cells were directly imaged by confocal microscopy. Confocal laser scanning microscopy (CLSM) was performed using the

FV-1000 (Olympus Corp., Tokyo, Japan) with 2-laser diodes (473 nm and 559 nm). A $4 \times$ (0.20 numerical aperture immersion) objective lens and $20 \times$ (0.95 numerical aperture immersion) objective lens (Olympus) were used. 800×800 pixels and $1.0\text{-}\mu\text{m}$ z steps were scanned, which took 1–2 s per section, with 6–8 min per full 3D scan. Scanning and image acquisition were controlled by Fluoview software (Olympus).²

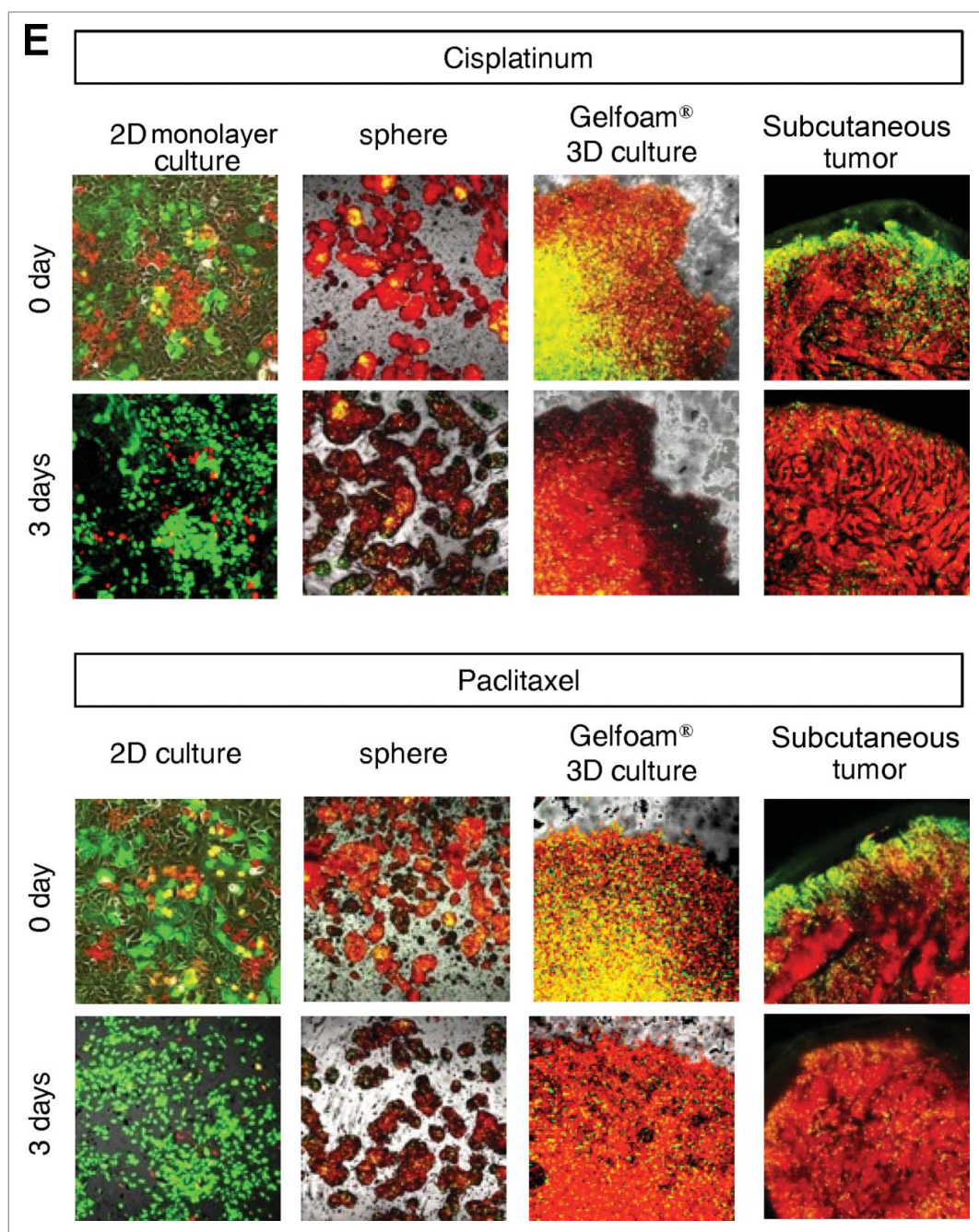


Figure 4. For figure legend, see page 816.

3D image analysis

The tracing data were imported to Volocity 6.0 version (Perkin Elmer, Waltham, MA), where all further analysis was performed.²

Statistical analysis

Data are shown as means \pm SD.

Author Contribution

SY and RMH conceived the idea for this project. SY and RMH designed all experiments and wrote the manuscript. SY, SM, SM, YH and FU performed all experiments. HK, HT, MZ, MB, and TF provided crucial ideas and helped with data interpretation. HT provided special technical assistance.

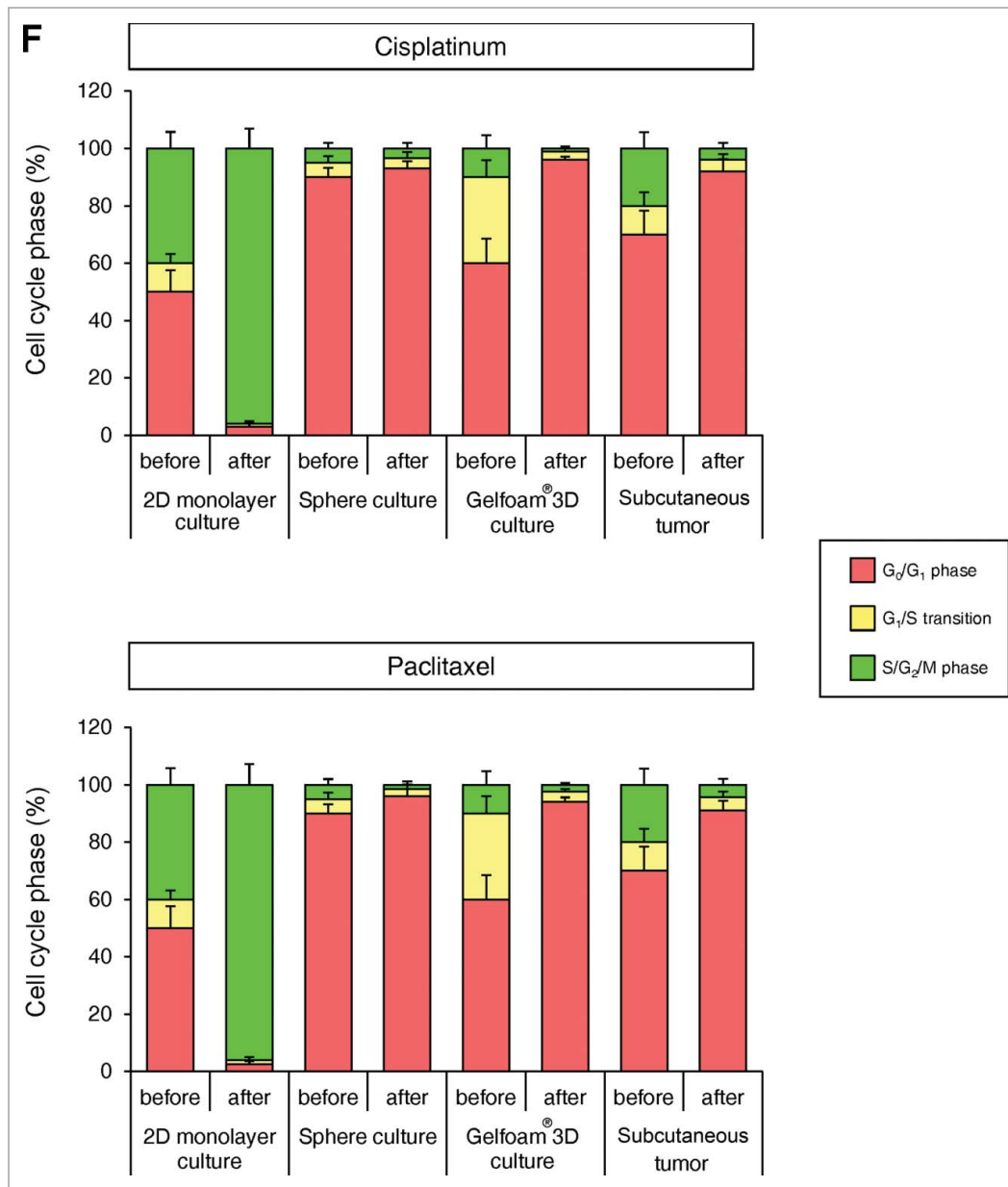


Figure 4. FUCCI-expressing cancer cells on Gelfoam[®] have the same cell cycle response to cytotoxic agents as subcutaneous tumors. Time-course imaging of FUCCI-expressing cancer cells (A) subcutaneous, (B) in monolayer culture, (C) in tumor spheres on agar and (D) on Gelfoam[®], before and after chemotherapy. (E) Representative images of FUCCI-expressing cancer cells in monolayer culture, spheres on agar, on Gelfoam[®], and subcutaneous tumors, before and after chemotherapy with cisplatin or paclitaxel. In monolayer culture, chemotherapy blocked cancer cells in S/G₂/M phase. Chemotherapy had little effect on quiescent tumor spheres. In contrast, tumors on Gelfoam[®] histoculture and subcutaneous tumors had a similar initial response to chemotherapy with cells becoming blocked in G₀/G₁. (F) Histograms of cell-cycle phase distribution before and after chemotherapy of 2D monolayer, sphere and Gelfoam[®] cultures and subcutaneous tumors. (Also see **Video S1**).

Disclosure of Potential Conflicts of Interest

MZ is an employee of AntiCancer Inc. SY, SM, SM, YH, FU, HK and RMH are or were unsalaried associates of Anti Cancer Inc. There are no other competing financial interests.

Funding

This study was supported in part by National Cancer Institute grant CA132971.

Dedication

This paper is dedicated to the memory of A.R. Moossa, MD.

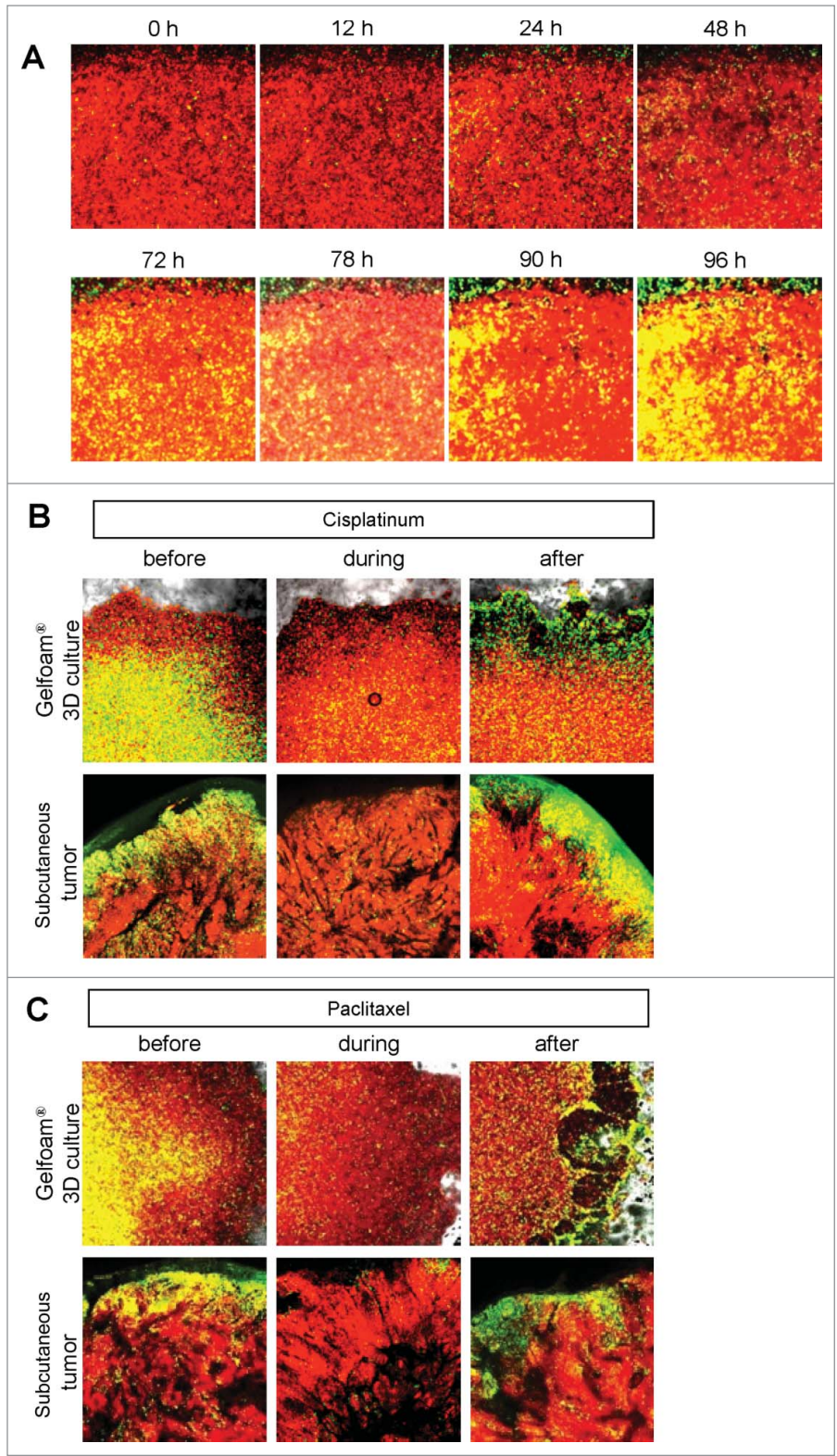


Figure 5. For figure legend, see page 818.

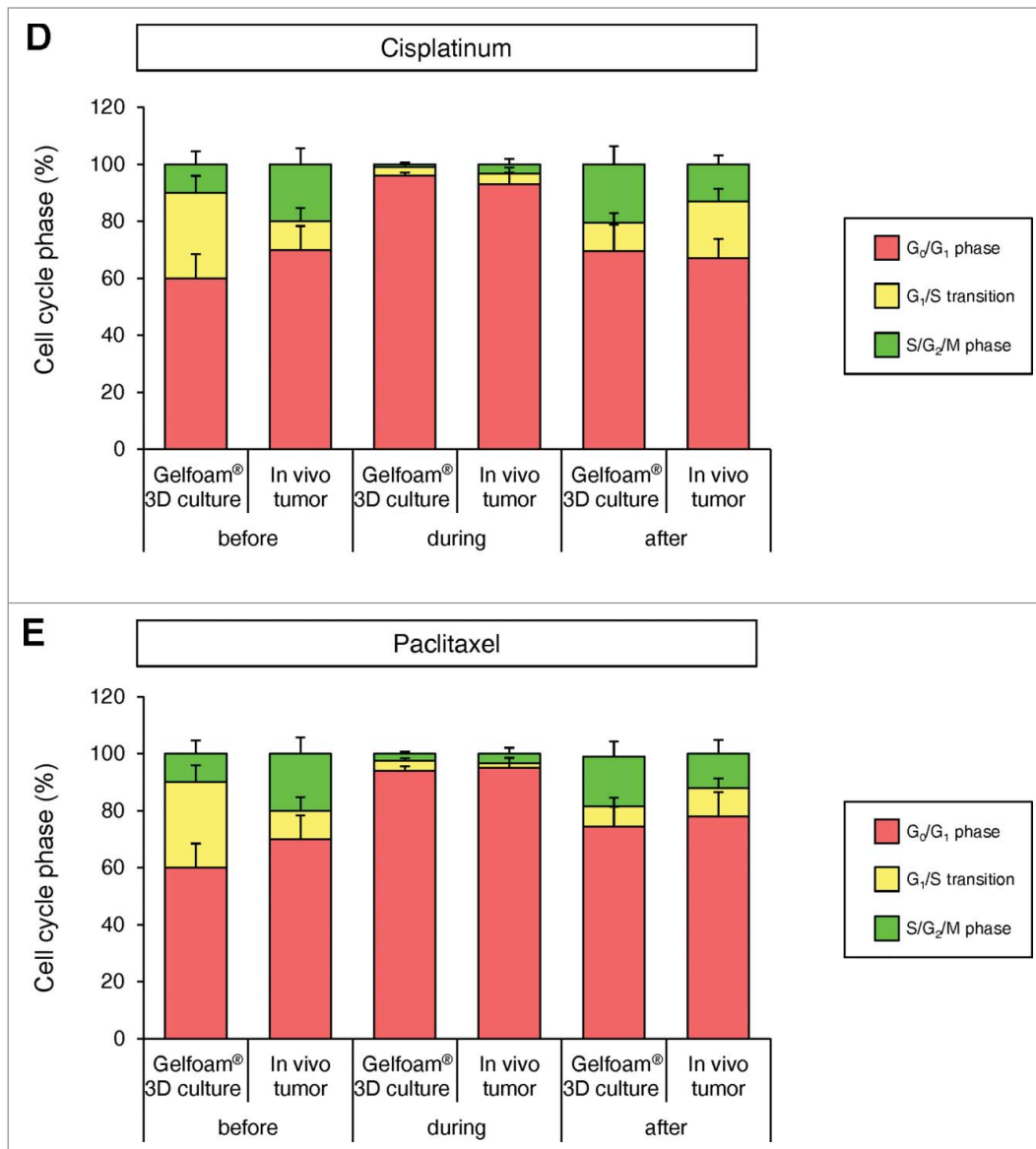


Figure 5. FUCCI-expressing cancer cells on Gelfoam® have a similar recovery pattern from cisplatin and paclitaxel as subcutaneous tumors. (A) Time-course imaging (96 hours) of FUCCI-expressing MKN45 cells histocultured on Gelfoam® after cisplatin chemotherapy. (B) Representative images of FUCCI-expressing cancer cells on Gelfoam® and subcutaneous tumors before and after recovery from chemotherapy with (B, D) cisplatin or (C, E) paclitaxel. G₀/G₁ quiescent cancer cells on Gelfoam® and in subcutaneous tumors are resistant to chemotherapy and can restart proliferation after treatment is terminated. Scale bars; 500 μ m.

References

- Sakaue-Sawano A, Kurokawa H, Morimura T, Hanyu A, Hama H, Osawa H, Kashiwagi S, Fukami K, Miyata T, Miyoshi H, et al. Visualizing spatiotemporal dynamics of multicellular cell cycle progression. *Cell* 2008; 132:487-98; PMID:18267078; <http://dx.doi.org/10.1016/j.cell.2007.12.033>
- Yano S, Zhang Y, Miwa S, Tome Y, Hiroshima Y, Uehara F, Yamamoto M, Suetsugu A, Kishimoto H, Tazawa H, et al. Spatial-temporal FUCCI imaging of each cell in a tumor demonstrates locational dependence of cell cycle dynamics and chemoresponsiveness. *Cell Cycle* 2014; 13:2110-9; PMID:24811200; <http://dx.doi.org/10.4161/cc.29156>
- Leighton J. A sponge matrix method for tissue culture; formation of organized aggregates of cells in vitro. *J Natl Cancer Inst* 1951; 12:545-61; PMID:14889259
- Leighton J, Justh G, Esper M, Kronenthal RL. Collagen coated cellulose sponge: Three-dimensional matrix for tissue culture of Walker tumor 256. *Science* 1967; 155:1259-61; PMID:6018647; <http://dx.doi.org/10.1126/science.155.3767.1259-a>
- Tome Y, Uehara F, Mii S, Yano S, Zhang L, Sugimoto N, Maehara H, Bouvet M, Tsuchiya H, Kanaya F, et al. 3-dimensional tissue is formed from cancer cells *in vitro* on Gelfoam®, but not on Matrigel™. *J Cell Biochem* 2014; 115:1362-7; PMID:24497277; <http://dx.doi.org/10.1002/jcb.24780>
- Yano S, Miwa S, Mii S, Hiroshima Y, Uehara F, Yamamoto M, Kishimoto H, Tazawa H, Bouvet M, Fujiwara T, et al. Invading cancer cells are predominantly in G₀/G₁ resulting in chemoresistance demonstrated by real-time FUCCI imaging. *Cell Cycle* 2014; 13:953-60; PMID:24552821; <http://dx.doi.org/10.4161/cc.27818>
- Kagawa Y, Matsumoto S, Kamioka Y, Mimori K, Naito Y, Ishii T, Okuzaki D, Nishida N, Maeda S, Naito A, et al. Cell cycle-dependent Rho GTPase activity dynamically regulates cancer cell motility and invasion in vivo. *PLoS One* 2013; 8:e83629; PMID:24386239; <http://dx.doi.org/10.1371/journal.pone.0083629>
- Yano S, Li S, Han Q, Tan Y, Bouvet M, Fujiwara T, Hoffman RM. Selective methionine-induced trap of cancer cells in S/G₂ phase visualized by FUCCI imaging confers chemosensitivity. *Oncotarget* 2014; 5:8729-36.
- Blagosklonny MV. How cancer could be cured by 2015. *Cell Cycle* 2005; 4:269-78; PMID:15655345

10. Blagosklonny MV. Tissue-selective therapy of cancer. *Br J Cancer* 2003; 89:1147-51; PMID:14520435; <http://dx.doi.org/10.1038/sj.bjc.6601256>
11. Blagosklonny MV. Matching targets for selective cancer therapy. *Drug Discov Today* 2003; 8:1104-7; PMID:14678733; [http://dx.doi.org/10.1016/S1359-6446\(03\)02806-X](http://dx.doi.org/10.1016/S1359-6446(03)02806-X)
12. Blagosklonny MV. "Targeting the absence" and therapeutic engineering for cancer therapy. *Cell Cycle* 2008; 7:1307-12; PMID:18487952; <http://dx.doi.org/10.4161/cc.7.10.6250>
13. Blagosklonny MV. Teratogens as anti-cancer drugs. *Cell Cycle* 2005; 4:1518-21; PMID:16258270; <http://dx.doi.org/10.4161/cc.4.11.2208>
14. Blagosklonny MV. Treatment with inhibitors of caspases, that are substrates of drug transporters, selectively permits chemotherapy-induced apoptosis in multidrug-resistant cells but protects normal cells. *Leukemia* 2001; 15:936-41; PMID:11417480; <http://dx.doi.org/10.1038/sj.leu.2402127>
15. Blagosklonny MV. Target for cancer therapy: proliferating cells or stem cells. *Leukemia* 2006; 20:385-91; PMID:16357832; <http://dx.doi.org/10.1038/sj.leu.2404075>
16. Blagosklonny MV. Cancer stem cell and cancer stem-loids: from biology to therapy. *Cancer Biol Ther* 2007; 6:1684-90; PMID:18344680; <http://dx.doi.org/10.4161/cbt.6.11.5167>
17. Yano S, Tazawa H, Hashimoto Y, Shirakawa Y, Kuroda S, Nishizaki M, Kishimoto H, Uno F, Nagasaka T, Urata Y, et al. A genetically engineered oncolytic adenovirus decoys and lethally traps quiescent cancer stem-like cells into S/G2/M phases. *Clin Cancer Res* 2013; 19:6495-505; PMID:24081978; <http://dx.doi.org/10.1158/1078-0432.CCR-13-0742>
18. Freeman A, Hoffman RM. 1986. In vivo-like growth of human tumors in vitro. *Proc Natl Acad Sci USA* 1986; 83:2694-8; PMID:3458228; <http://dx.doi.org/10.1073/pnas.83.8.2694>
19. Hoffman RM. Histocultures and their use. In: *Encyclopedia of life sciences*. Chichester: John Wiley and Sons Ltd. Published Online. 2010; DOI: 10.1002/9780470015902.a0002573.pub2
20. Hoffman RM. Tissue culture. In: *Brenner's Encyclopedia of Genetics*. 2nd edition. Elsevier, 2013; 73-76.
21. Mii S, Duong J, Tome Y, Uchugonova A, Liu F, Amoh Y, Saito N, Katsuoka K, Hoffman RM. The role of hair follicle nestin-expressing stem cells during whisker sensory-nerve growth in long-term 3-D culture. *J Cell Biochem* 2013; 114:1674-84; PMID:23444061; <http://dx.doi.org/10.1002/jcb.24509>