

A French National Survey on Clotting Disorders in Mastocytosis

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Abstract: Mastocytosis is characterized by a clonal mast cell proliferation with organ infiltration and uncontrolled degranulation. Although not characteristic and poorly explained, some patients develop clotting abnormalities.

We retrospectively identified patients with established diagnosis of mastocytosis and related clotting abnormalities (clinical and/or biological) using the national French Reference Centre for Mastocytosis database.

From our cohort of 14 adult patients with clotting abnormalities (median age 46 years [range 26–75]), 4 had a presentation suggestive of a primary hemostasis disorder alone (by their symptoms and/or abnormal clotting tests [PFA, von Willebrand's disease [vWD] screening]) and 10 had a laboratory impairment of secondary hemostasis. Among these, 7 had bleeds characteristic of a coagulation cascade disorder (severe/life-threatening in 5 and mild in 2 patients). Clotting abnormalities were of variable severity, typically related to intense crisis of degranulation, such as anaphylactic reactions, and/or to severe organ infiltration by mast cells. Importantly, classical hemostatic management with platelet transfusion, fresh frozen plasma, or vitamin K infusions

was unsuccessful, as opposed to the use of agents inhibiting mast cell activity, particularly steroids. This illustrates the crucial role of mast cell mediators such as tryptase and heparin, which interfere both with primary (mainly via inhibition of von Willebrand factor) and secondary hemostasis. There was interestingly an unusually high number of aggressive mastocytosis (particularly mast cell leukemia) and increased mortality in the group with secondary hemostasis disorders ($n = 5$, 36% of the whole cohort).

Mast cell degranulation and/or high tumoral burden induce both specific biologic antiaggregant and anticoagulant states with a wide clinical spectrum ranging from asymptomatic to life-threatening bleeds. Hemostatic control is achieved by mast cell inhibitors such as steroids.

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Abbreviations: ASM = aggressive systemic mastocytosis, CEREMAST = centre de référence des mastocytoses (French reference centre for mastocytosis), CM = cutaneous mastocytosis, ISM = indolent systemic mastocytosis, MCAS = mast cell activation symptoms/syndrome, MCL = mast cell leukemia, PFA = platelet function assay, SM = systemic mastocytosis, SM-AHNMD = systemic mastocytosis associated with a clonal hematological non mast cell lineage disease, SSM = smouldering systemic mastocytosis, vWF = von Willebrand factor, vWF: RCo activity = von Willebrand ristocetin cofactor activity.

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INTRODUCTION

Mastocytosis is a rare clonal hematological disorder characterized by (1) the proliferation and accumulation of neoplastic mast cells in ≥ 1 organs (the skin and the bone marrow being predominantly involved) and (2) their unregulated activation/degranulation (ie mast cell mediators release), resulting in mast cell activation symptoms/syndrome (MCAS).¹ The classification of mastocytosis is based on signs of organ infiltration,² with or without organ enlargement (B-findings) and dysfunction (C-findings). It distinguishes localized mastocytosis (cutaneous mastocytosis [CM]) from systemic mastocytosis (SM). Systemic mastocytosis can be indolent (ISM; no B- nor C-findings), smoldering (SSM; ≥ 2 B-findings and no C-findings), aggressive (ASM; ≥ 1 C-finding), and associated with a clonal hematological nonmast cell lineage disease (SM-AHNMD). With regard to prevalence, 880 patients are regularly followed-up at the national French Reference Centre (CEREMAST) located at Necker Children's Hospital. We estimate a total of ~2000 cases of mastocytosis in France with 80% CM or ISM, 15–20% ASM or SM-AHNMD, <1% MCL, and an unknown prevalence of SSM (CEREMAST, unpublished data).

In terms of prognosis, in indolent forms life expectancy is similar to that of the general population but MCAS remains a

cause of an all too often underestimated burden and disability for patients.³ In ASM and SM-AHNMD, life expectancy is significantly reduced with a respective median survival of 6.7 years and 4.4 years, depending on organ dysfunction as well as AHNMD type and progression.^{2,4} Finally, mast-cell leukemia (MCL) usually presents with rapid organ failure as well as florid MCAS, including anaphylactic shock and is still associated with a catastrophic median survival of around 6 months.^{2,5}

In addition to classical symptoms of mast cell degranulation and organ infiltration, clotting disorders have been reported in 13 patients to date, ranging from simple ecchymosis tendency to severe/life-threatening bleeds (epidural hematoma, intracerebral hemorrhage, severe gastrointestinal bleeding, massive postsurgical blood loss, hemorrhagic pleural effusion requiring ventilation).^{6–18} No cases of thrombosis have been reported.

Physiological hemostasis is a dynamic process, traditionally described in 3 steps. Primary hemostasis leads to platelet aggregation and involves the blood vessel, platelets, and von Willebrand factor (vWF). Secondary hemostasis, or coagulation, starts with the interaction between tissue factor, exposed to the blood stream after endothelial disruption, and FVII, leading to the formation of thrombin (FII), which will activate both branches of the coagulation cascade (extrinsic [triggered by the activation of FVII] and intrinsic [triggered by the activation of FXII]), thus increasing its own production (thrombin burst). Thrombin will eventually be transformed into fibrin, which stabilizes the clot. Tertiary hemostasis, or fibrinolysis, prevents excessive clotting formation through fibrin degradation by plasmin.

Based on the detailed description of a series of 14 French patients, we aimed to (1) characterize clotting abnormalities in mastocytosis, (2) suggest underlying mechanisms for these, and (3) appreciate their relevance in clinical practice.

METHODS

We used the CEREMAST (*Centre des référence des Mastocytoses*) national database to retrospectively identify patients with mastocytosis and clotting abnormalities. The use of this computer database to store personal information was authorized by the French National Data Protection Commission; CEREMAST authorization: CNIL: No1445939 (October 29, 2010). Overall, 23 regional centers affiliated to the CEREMAST and homogeneously distributed in France were then contacted to obtain more information on potential cases. Patients aged ≥ 15 years old, with a confirmed diagnosis of mastocytosis (including genetics to determine *CKIT* mutations) according to the World Health Organization (WHO) criteria,¹⁹ with evidence of a current or past medical history of clinical and/or biological clotting abnormalities were included and analyzed. Patients with the following characteristics were excluded from this study: inherited clotting abnormality, suspicion or confirmation of antibodies interfering with coagulation, vitamin K deficiency, treatments directly interfering with clotting, disease-related or other causes of hepatocellular insufficiency at the time of the study, severe renal impairment (with urea >10 mmol/L). Patients for whom data was deemed insufficient were also excluded. Approval was obtained from the Necker Children's University Hospital (Paris, France) ethical committee. Written informed consent was also obtained from all patients whose data are discussed in this paper in accordance with the declaration of Helsinki.

The clinician from CEREMAST verified, for each patient, the type and classification of mastocytosis according to the

WHO criteria¹⁹ and ensured accuracy of the clinical and biological history. To avoid including patients with pre-existing clotting disorders, a bleeding history and, when available, the results of biological tests performed before mastocytosis onset were collected.

In order to provide a well-recognized and uniform tool, we used the Common Terminology Criteria for Adverse Events (CTCAE), version 4.0, to grade bleeding symptoms. Primary hemostasis was assessed by platelet function assay (PFA) for epinephrine, adenosine diphosphate (ADP), and von Willebrand factor (vWF) antigen (Ag) and activity (Rco). We have excluded classical causes of acquired vWF disease, including hypothyroidism, autoimmune disease, monoclonal gammopathy, aortic stenosis, and solid malignant tumors. Secondary hemostasis was assessed by prothrombin time ratio (PT_r), an equivalent of prothrombin time (PT) used in France, expressed in percentage of a reference value (low PT_r corresponds to a high PT, normal $>70\%$) as well as by activated partial thromboplastin time (aPTT) ratio (patient's aPTT/reference aPTT in seconds = aPTT/aPTT ratio or aPTT_r, normal ≤ 1.20), clotting factors levels (F followed by the corresponding roman numeral), and anti-Xa levels. When multiple clotting samples were available, we selected the most abnormal one.

Importantly, note that in view of the retrospective nature of this study, the data available are extremely variable from patient to patient, depending on the degree of detail of clinical records and investigations performed by the clinician who assessed the patient at that time.

For the purpose of clarity, we divided patients into 2 groups depending on whether they predominantly presented with abnormalities of primary hemostasis (group A, #1–4), or secondary hemostasis (group B, #5–14). As discussed later, some patients from group B also had concomitant primary hemostasis abnormalities, but these were not predominant. Tertiary hemostasis was not analyzed due to lack of data.

Literature research was performed using *PubMed* database. Key words included “mast cell”, “mast cell disease,” “mastocytosis,” “clotting,” and “coagulation.” Terms related to specific mast cell mediators and actors of coagulation were also used. Papers were selected from the main results list and from the “related citations” list suggested by PubMed. In addition, we explored the bibliographies of relevant papers.

RESULTS

From the 21 patients with mastocytosis and clotting abnormalities initially identified via the CEREMAST database, $n = 14$ met the study's inclusion criteria. Population characteristics are summarized in Tables 1 and 2. Table 1 particularly summarizes each patient's clinical and biological data with regard to their mastocytosis. Clotting findings including clinical presentation, investigations, management, and response to the latter are presented in Table 2. The symptomatic (mast cell-stabilizing or mediator-targeting drugs) and cytoreductive treatment history, notably the use of steroids, has been clearly mentioned for each patient in Tables 1 and 2.

Group A ($n = 4$) is characterized by an impairment of primary hemostasis, a median age of 35.5 (26–62) years, absent (#1) or mild (#2–#4) bleeding symptoms (maximum CTCAE bleeding grade of 2), a majority of indolent mastocytosis (3/4), no C-findings, no AHNMD and no deaths to date. After adjusting reference ranges to the blood group, the diagnosis of vWF disease was confirmed in #1 (type 1 vWF disease: prolonged PFA, proportionate decrease of vWF Ag and FVIII

TABLE 1. Population Characteristics: Patients Clinical and Laboratory Data With Regard to Their Mastocytosis Type

Patients	Group A—Primary Hemostasis Abnormalities (n = 4, #1–#4)				Group B – Secondary Hemostasis abnormalities (n = 10, #5–#14)									
	Pt 1	Pt 2	Pt 3	Pt 4	Pt 5	Pt 6	Pt 7	Pt 8	Pt 9	Pt 10	Pt 11	Pt 12	Pt 13	Pt 14
Main features	MC –	ISM –	cMCL –	CM –	ISM CLL	ISM –	SSM –	SSM CMML	cMCL –	aMCL –	aMCL –	ASM MDS	ASM MDS/MPs	ASM AML
Sex (mf)	m	M	m	f	m	m	f	m	m	f	m	m	m	m
Age (yrs)	35	26	62	36	47	46	46	69	29	29	36	75	74	73
Age of diagnosis (yrs)	> 5	4	12	3	10	5	26	1.5	> 20	< 1	< 1	< 1	< 1	1
Skin involvement	TMEP	UP	UP	UP	UP	no	no	UP	UP	no	no	TMEP	no	UP
MCAS	–	–	–	–	–	–	–	–	–	–	–	–	–	–
	Anaphylactic shock	–	–	–	–	–	–	–	–	–	–	–	–	–
	Malaise	–	–	–	–	–	–	–	–	–	–	–	–	–
	Angioedema	–	–	–	–	–	–	–	–	–	–	–	–	–
	Flushing/Pruritus	+/+	+/+	+/+	+/+	–	+/–	–	+/–	+/+	+/–	–	–	–
	Diarrhea	+	+	+	–	–	–	+	–	+	–	–	–	–
	Abdominal pain/nausea	+/–	+/–	+/–	+/–	–	+/+	+/+	–	+/–	+/–	–	–	–
	Pollakiuria	–	–	–	–	–	–	–	–	–	–	–	–	–
	Acute urticaria	+	+	–	–	–	–	–	–	–	–	–	–	–
B-findings (n)	0	0	2	0	0	0	2	2	1	2	1	1	1	1
C-findings (n)	0	0	0	0	0	0	0	0	1	4	2	4	3	3
Bone demineralization	+	–	–	–	+	+	–	+	–	–	–	–	–	–
Tryptase (ng/mL) [†]	6	14.9	>200	19.6	75	140	696	248	369	840	74	458	>200	244
Ckit mutation	WT	D816V	D816V	D816V	D816V	D816V	D816V	D816V	D816V	Dup512	D816V	D816V	D816V	D816V
Therapy	HI	HI/H2/AL	HI/H2/ AAC	HI/H2/ AAC	0	HI	HI	HI/H2	HI/H2	HI/H2/AL	HI	HI	HI/AL	HI/H2/ AAC
High dose steroids [‡]	No	No	No	No	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	Yes	na
Cytoreductive therapy	–	–	PKC412 TUA	TKI trial	–	Imatinib	–	PKC412 TUA	PKC412 TUA	–TKI trial	–PKC412 TUA	–TKI trial	Rapid death before therapy	Pt refusal
	–	–	–	–	–	–	–	–	–	–AraC	–5-Aza	–2CDA	–	–
	–	–	–	–	–	–	–	–	–	–5-Aza	–Dasatimib	–Thal	–	–
	–	–	–	–	–	–	–	–	–	–mTOR inh	–AraC	–	–	–
	–	–	–	–	–	–	–	–	–	–mTOR inh	–Allograft	–	–	–
Outcome	Alive	Alive	Alive	Alive	Alive	Alive	Alive	Dead	Alive	Dead	Alive	Dead	Dead	Dead

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TABLE 2. Patients Clinical and Laboratory Data With Regard to Their Bleeding History and Clotting Investigations

Patient/gender/age	Group A—Primary Hemostasis Abnormalities (n = 4, patients 1–4)			
	Pt 1/M/37	Pt 2/M/26	Pt 3/M/62	Pt 4/F/38
Mastocytosis type	CM	ISM	cMCL	CM
AHNMD	—	—	—	—
Tryptase level	6	14.9	>200	19.6
Clinically detectable acute MCAS*	No	No	No	No
B-findings	0	0	2	0
C-findings	0	0	0	0
Bleeding symptoms	Prolonged bleedings after phlebotomy	Recurrent epistaxis	Recurrent severe epistaxis (cauterization)	- Postsurgical ophthalmological bleeding. - Metrorrhagia. - Spontaneous large ecchymosis and subcutaneous hematomas of the thighs. - Major and persistent ecchymosis/hematomas after breast implants insertion
Bleeding grade [†]	1	1	2	2
PLT	209	230	244	180
PTT	97%	86%	93%	89%
APTT	0.84	↑ 1.21	↑ 1.24	↑ 1.23
Other biological results	- PFA ADP ↑ 190** - PFA adrenaline ↑ 256** - vWF Ag ↓ 32% - FVIII ↓ 51% - DDAVP+ with vWF Ag 123% and FVIII 172%	- FIX 91% - FXI 80% - FXII 82% - vWF:RCO activity ↓ 53% - vWF Ag 72%	- No circulating anticoagulant - FXI ↓ 59% - PFA adrenaline ↑ 178** - PFA ADP ↑ 138** - vWF Ag 80% - vWF:RCO activity ↓ 58%	- FVIII 94% - FIX 133% - FXI 134% - FXII 102% - Normal bleeding time - vWF Ag 51% - vWF:RCO activity 51%
Blood group	A	O	A	O
Long-term treatment	HI	HI/H2/AL	HI/H2/AAC/PKC412 (TUA)	HI/H2/AAC/TKI trial
Treatment in the acute phase	No	No	No	No
Evolution of clotting disorder	- Stable - vWF-acquired type disease	- Fluctuating aPTT; usually N - vWF-acquired type disease	- Fluctuating aPTT; sometimes N - vWF-acquired type disease	Spontaneously resolved
Status	Alive	Alive	Alive	Alive

Patient/gender/age	Group B—Secondary Hemostasis Abnormalities (n = 10, patients 5–14)			
	Pt 5/M/47	Pt 6/M/46	Pt 7/F/46	Pt 8*/M/69
Mastocytosis type	ISM	ISM	SSM	SSM
AHNMD	B-CLL	—	—	CMML
Clinically detectable acute MCAS*	75	140	696	248
B-findings	0	0	2	2
C-findings	0	0	0	0
Bleeding symptoms	- Anaphylactic reactions with dyspnoea, flush - No	- Anaphylactic shock - Diffuse cutaneous and at puncture sites - UGI (duodenal)	- Anaphylactic reaction - UGI (gastric ulcer)	- Acute MCAS with intense digestive symptoms and malaise - UGI (rectorrhagia) - UGI (melena, gastric ulcer, OV)
Bleeding grade [†]	na	2	3	3
PLT	N	↓ 22	↓ 137	N
PTT	↓ 40%	↓ 53%	↓ 46%	↓ 37%
		Unreliable		↓ 53%
				na
				↓ 80
				↓ 42%

Group B – Secondary Hemostasis Abnormalities (n = 10, patients 5–14)

Patient/gender/age	Pt5/M/47	Pt6/M/46	Pt7/F/46	Pt8 ^f /M/69	Pt 9 ^{**} /M/29
aPTT _r	>3.53	Unrecordable	0.97	↑ 1.75	↑ 1.96
Other biological results	- FV 107% - ↑ anti-Xa 0.56 U/ml	- Fg unrecordable - ↑ anti-Xa2.5 U/ml - ATIII ↓ 53%	- Fg 2.4 - FII ↓ 63% - FV ↓ 24% - FVII+X ↓ 43%	- FV ↓ 37% - FII ↓ 64% - FV ↓ 59% - FVII ↓ 44% - FX ↓ 57%	- FII ↓ 25% - FV ↓ 51% - FVII+FX ↓ 36% - Fg 3.7 g/L
Blood group	A	B	O	O	O
Long-term treatment	No (patient refusal)	HI/Imatinib	HI	HI/H2/PKC412 TUA	HI/H2/PKC412 TUA
Treatment in the acute phase	- ICU admission - HI/H2 - Steroids bolus ^s (1 st episode)	- ICU admission - FFP: ineffective - Fg: ineffective - Protamine: low effect - RBC transfusion - Steroids bolus ^s	- HI/H2 - Steroids bolus ^s	- ICU admission - Vit K: ineffective - RBC transfusion - PLT transfusion	- HI: ineffective - Steroids bolus ^s
Evolution of clotting disorder	Completely normalized alongside MCAS resolution	Normalized alongside MCAS resolution except persisting prolonged aPTT _r 1.40	Normalized alongside MCAS resolution	Completely normalized alongside MCAS resolution	Improved with steroids then normalized with PKC412 concomitantly with IMR on MCL
Status	Alive	Alive	Alive	Dead (septic shock in the context of CMML acutisation with blastosis)	Alive

Group B – Secondary Hemostasis Abnormalities (n = 10, patients 5–14)

Patient/gender/age	Pt 11 ^f /M/36	Pt 12 ^f /M/75	Pt 13 ^{ss} /M/74	Pt 14 ^{ll} /M/73
Mastocytosis type	aMCL	ASM	ASM	ASM
AHNMD	-	MDS	MDS/MPS	AML (2011)
Trypsin level	840	458	>200	244
Clinically detectable acute MCAS	Intense crisis of mast cell degranulation	No	No	No
B-finders	2	1	1	1
C-finders	4	4	3	3
Bleeding symptoms	Anaphylactic reaction with flush	UGI (massive melena)	na	Data missing
Bleeding grade ^f	Menometrorrhagia	3	na	na
PLT	↓ 66 [chronic]	↓ 41	↓ 120	↓ 138
PTr	↓ 60%	↓ 61%	Slight ↓ 68%	↓ 62%
aPTT _r	↑ 2.82	↑ 1.46	↑ 1.60	↑ 1.39
Other biological results	- TTr > 3 - Fg 1.4 g/L - FII 87% - FV ↓ 27% - FVII+X ↓ 58% - Anti-Xa ↑ 0.15 U/ml	- FV 70% - FVII 90% - FIX and FXIII N - vWF activity 86% - vWF Ag 111% - PFA ADP ↑ 139** - Other aggregation tests N	- FII ↓ 68% - FV 70% - FVII+X ↓ 42% - ELT N	- FII ↓ 52%* - FV ↓ 63% - FVII ↓ 62% - FX ↓ 39% - FVIII 131% - FIX 74% - FXI ↓ 49% - FXII 64%
Blood group	AB	A	O	O
Long term treatment	- HI/H2 - Several lines of cytoreductive therapy (Table 1)	- HI/H2 - Several lines of cytoreductive therapy (Table 1)	- HI - Steroids in view to treat ASM	- HI - No cytoreductive therapy due to patient refusal - Supportive care

Group B—Secondary Hemostasis Abnormalities (n = 10, patients 5–14)

Patient/gender/age	Pt 10/F/29	Pt 11 [†] /M/36	Pt 12 ^{††} /M/75	Pt 13 ^{§§} /M/74	Pt 14 /M/73
Treatment in the acute phase	- HI - Steroids bolus [§]	- Decompressive craniectomy - Steroids bolus [§]	- RBC transfusion - PLT transfusion	No	- No specific treatment due to patient refusal - Supportive care including RBC & PLT transfusions
Evolution of clotting disorder	Normalized alongside MCAS resolution (despite MCL progression) except for persisting FVII+X 50% Dead from aMCL	Completely normalized alongside MCAS resolution within 2 days of steroids Alive	Biological (clotting and PLT) and clinical (bleeding) remission under steroids Dead from ASM progression	Normalized with concomitant resolution of C-findings under steroids Dead from ARDS	Progressive worsening until death Dead from ASM and AML progression
Status					

AAC = antiallergic cromones (disodium cromoglycate), AHNMD = associated clonal hematological nonmast cell lineage disease, AL = antileukotrienes (montelukast), aMCL = acute mast-cell leukemia, AML = acute myeloblastic leukaemia, Anti-Xa = anti-Xa activity, ARDS = acute respiratory distress syndrome, ASM = aggressive systemic mastocytosis, ATIII = antithrombin III, CLL = chronic lymphocytic leukemia, CM = cutaneous mastocytosis, cMCL = chronic mast-cell leukemia, CMML = chronic myelomonocytic leukemia, DDAVP+ = positive response to the desmopressin test, ELT = euglobulinlisis time, F = female, FFP = fresh frozen plasma, Fg = fibrinogen, H/SMG = hepato-splenomegaly, H1/H2 = histamine blockers of type 1 or 2, ICU = intensive care unit, IMR = incomplete major response, ISM = indolent systemic mastocytosis, LGI = lower gastrointestinal, M = male, MCAS = mast cell activation symptoms/syndrome, N = normal, TKI = tyrosine kinase inhibitor, TTr = thrombin time ratio, TUA = transitory use authorization (PKC412 therapy under cover of a TUA was approved in France in Summer 2012), UGI = upper gastrointestinal, Vit K = Vitamin K, vWF antigen = vWF Ag, WT = *c-Kir* wild type. Arrows indicate an abnormal decrease (↓) or increase (↑) of a specific value.
Normal values: aPTTr <1.20; FII >70%; FV >60%; FVIII >60%; FVIII:C activity >50%; FX >65%; FXI >65%; FXII >50%; PFA ADP <120; PFA adrenaline <170; PLT: 150–300 × 10³ /μL; PT >70%; Trypsin < 13.5 ng/mL; vWF Ag: 50–160%; vWF: RCo activity 60–160; (vWF: RCo activity/vWF Ag) ratio >0.70.
 *Clinically detectable acute MCAS that means sudden crisis of degranulation with more intense MCAS than usual occurring at the time of bleeding and/or clotting tests abnormalities.
[†]The grading of bleedings was inspired of the Common Terminology Criteria for Adverse Events (CTCAE), version 4.0.
[‡]In patients 2 and 3, clotting tests were not assessed during the bleeding episodes.
[§]Steroids bolus corresponds to high-dose dexamethasone (40 mg/day for 4 days) or equivalent.
^{||}Patient 6 was previously reported by French colleagues [16]. Note that imatinib was later stopped due to D816V mutation and that aPTTr remained chronically prolonged.
[¶]Patient 7 was diagnosed for years with SSM and presented fluctuating H/SMG without PHT and fluctuating bicytopenia but to date, she never acquired C-findings.
^{**}Patient 8, initially diagnosed with an SSM, rapidly worsened with acquisition of 2 C-findings including symptomatic PHT (ascites, esophageal, and perigastric varices).
^{***}Patient 9 presented for years very frequent and intense degranulation crisis associated with fluctuating hepato-splenomegaly, central anemia, and clotting tests deterioration. Clotting would be the most disturbed during anaphylactic reactions. Only high-dose steroids initially allowed control of MCAS and improvement of anemia and clotting tests (failure of cladribine and alpha interferon). However, the sensitivity of steroids started decreasing leading to PKC412 therapy under cover of a TUA.
^{††}Patient 11: note that clotting tests reported in the table were from day 3 after the diagnosis of SDH. Platelet aggregation studies were also performed later on, while the patient was asymptomatic. They showed a normal aggregation with ADP 2.5, 5.0, and 10.0 memo/L, with collagen, arachadonic acid, and with ristocetin 0.5 and 1.2 mg/mL. Aggregation was delayed with epinephrine 5 memo/L.
^{‡‡}Patient 12: despite profuse intestinal hemorrhage, the gastroscopy, colonoscopy, and videocapsule only revealed a single rectal ulcer. Systematic colic biopsies revealed mast cell infiltration. Steroids led to rapid control of the hemorrhage, clotting, and PLT levels.
^{§§}Patient 13 died from SDR A in context of terminal phase of chronic obstructive pulmonary disease.
^{|||}Patient 14, initially diagnosed with an unclassified SM, progressively acquired 3 C-findings (ASM) and developed an AML. Clotting tests were initially only mildly altered, but deteriorated alongside disease progression until death (the patient refused all cytoreductive treatment). *Data used to calculate medians of PTR and aPTTr for this patient.

levels), and strongly suspected in #3 (type 2 A or B vWF disease: prolonged PFA, borderline vWF:RCo activity/vWF antigen, normal vWF antigen). Interestingly, #2 and #4 were blood group O, which is typically associated with lower levels of vWF antigen. The clinical presentation of these 2 patients was very suggestive of impairment of primary hemostasis.

In contrast, group B (n = 10) is characterized by an impairment of secondary hemostasis, a median age of 46.5 (29–75) years, and more frequent, more severe, sometimes life-threatening bleeds. Seven out of 9 patients had bleeding symptoms (data missing in n = 1), which were particularly severe (CTCAE grade ≥3) in 5 patients. These included 4 gastrointestinal (#6–#8, #12) and 1 intracranial (#11) bleeds. A majority of patients from group B had an aggressive type of mastocytosis (associated with AHNMD in 5/10) and a significantly reduced survival with 5 deaths among 10 patients and a median survival time of 2.5 years (Fig. 1).

Most of the patients from group B had severe episodes of mast cell degranulation (7/10, #5–#11), and/or high mast cell organ infiltration (8/10, #7–#14) illustrated by B and C-findings with a median of 1.5 C-findings (0–4). Importantly, a median PTR of 53% (out of range [low]–64) and a median aPTR of 1.75 (1.03–out of range [high]) were recorded in patients with MCAS (n = 7, #5–#11), which suggests a more severe hemostasis impairment profile than that of patients without MCAS (n = 3, #12–#14) who exhibited a median PTR of 64% (43–68) and a median aPTR of 1.39 (1.00–1.97).

Tryptase levels were significantly higher in group B compared to in group A (P = 0.0235, nonparametric Wilcoxon test, Figure 2). Moreover, the prevalence of CM/ISM, SSM, ASM/SM-AHNMD, and MCL was, respectively, 75% (3/4), 0%, 0%, 25% (1/4) in group A versus 10% (1/10), 10% (1/10), 50% (5/10), 30% (3/10) in group B where an increased mortality rate was recorded (n = 5, 36% of the overall population, Figure 1).

Classical management of bleeding including vitamin K infusion, platelet transfusions, and fresh frozen plasma appeared ineffective to achieve hemostasis (#6, #8, #12). By contrast, clotting tests and bleeding symptoms dramatically improved and/or normalized alongside subsidence of the degranulation crisis. The latter either occurred spontaneously (#4) or by targeting blockade of mast cell activation with agents such as antihistamines and, more effectively, high dose of steroids (#5–#13). Failure to control mast cell hyperactivity and aggressive

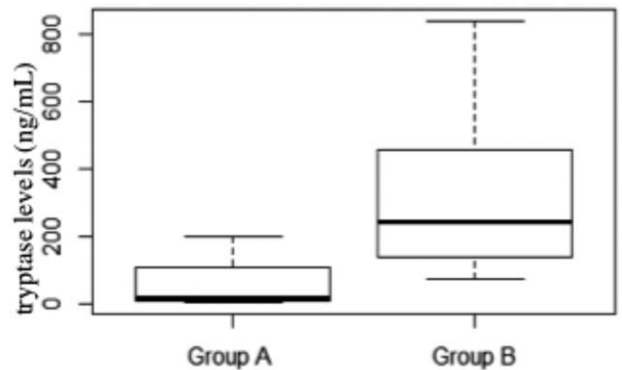


FIGURE 2. Comparison of tryptase levels (ng/mL) in groups A and B (nonparametric Wilcoxon test).

proliferation (C-findings) led to exponential deterioration of coagulation tests and death from the disease (#14).

DISCUSSION

This study is the first detailed description of several types of clotting abnormalities associated with mastocytosis. Although rare, they can be severe and must be promptly recognized to allow appropriate management. The spectrum of mastocytosis-related clotting disorders includes acquired vWF disease, acquired disorders of the coagulation cascade, and probably of fibrinolysis.

Only few previous case reports had already pointed out clotting impairment in the context of mastocytosis.^{6–18} However, the disparity in the population characteristics, the lack of systematic clinical/laboratory data, and of accurate mastocytosis classification mean that a clear hemostatic analysis from those reports remains poor. From our series, the description of our cases and the therapeutic implications of the background disease on the acquired hemostasis disorders shed new light on the significance and mechanisms involved in clotting disorders in the context of mastocytosis.

The mucocutaneous bleeds observed in group A are characteristic of abnormalities in primary hemostasis and reflect the role of von Willebrand factor in platelet adhesion and aggregation. This can be impaired by heparin, a substance present in mast cells, and released during degranulation. Indeed, heparin can bind to vWF, therefore preventing platelet adhesion via GPIb, and platelet aggregation via GPIb and GPIIb/IIIa.²⁰ This may explain why patients with low baseline vWF levels (either secondary to vWF disease or to blood group O) are more prone to mucocutaneous bleeds during mast cell activation syndromes. Therefore, by checking the blood group and testing for vWF disease in patients with mastocytosis, clinicians could potentially identify those at higher risk of bleeding and provide education on prophylactic measures, as well as the plan for possible complications following invasive procedures (for example, #4 developed extensive mammary hematomas after breast implant).

Impairment of primary hemostasis may coexist and be concealed by abnormalities in the coagulation cascade, particularly in subjects who are blood group O (60% of patients in group B, versus a prevalence of 43% in the normal French population according to the *Institut National de la Transfusion Sanguine* [INTS]). Patients #5 and #6 illustrate this particularly

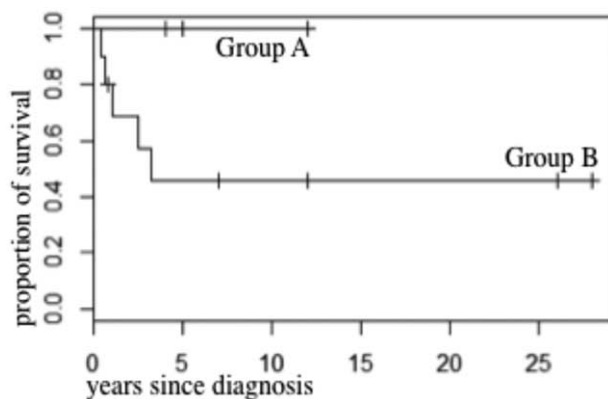


FIGURE 1. Survival curves in groups A and B (Kaplan–Meier analysis).

well as they developed hematomas/conjunctival bleeds, which are characteristic of primary hemostasis abnormalities.

The abnormalities of secondary hemostasis observed in group B were more pronounced in acute episodes of mast cell degranulation: patients with MCAS with or without organ infiltration (ie B and/or C-findings) had more bleeding symptoms than those where no clinically detectable MCAS was reported (Table 2). Bleeds in group B tended to be internal and therefore characteristic from abnormalities of the coagulation cascade (as opposed to mucocutaneous bleeds, which are often related to abnormalities in primary hemostasis). They were typically severe and life-threatening (4 gastrointestinal [#6–#8, #12] and 1 intracranial [#11]). The major role of MCAS exacerbations is further demonstrated by the rapid resolution of bleeding symptoms and/or clotting tests abnormalities as the MCAS subsides, even in cases where they were initially very severe.

This has major therapeutic implications in clinical practice and explains why standard hemostatic management with vitamin K, fresh frozen plasma, or platelet transfusion is insufficient. Indeed, to overcome mast cell hyperactivity and subsequent uncontrolled degranulation leading to an anticoagulant state, it is crucial that mast cell mediator blockers and mast cell stabilizers are used. We suggest the use of steroids in the acute setting and of maintenance treatment in the chronic setting for patients who have persistent symptoms of degranulation.

Regarding the role of mast cell mediators released during degranulation, tryptase, heparin, and antithrombin III are the 3 main molecules interfering with standard coagulation tests. The prolonged aPTT observed in our patients could result from several intricate mechanisms: the activating effect of tryptase on fibrinogenolysis^{21–25} and on kininogenolysis,^{26,27} the overall anticoagulant role of heparin through upregulation of thrombomodulin, which activates protein C and S pathways,^{28,29} and the release of antithrombin III.^{30,31} Interestingly, all these mechanisms can only account for prolonged PT secondary to an anticoagulant activity of the common pathway, as there is no known anticoagulant interaction between mast cell mediators and the extrinsic pathway. Therefore, we should expect PT to be prolonged only in the presence of prolonged aPTT, which is not the case (#7, #12), suggesting that other unknown mechanisms are involved.

Finally, the overrepresentation of aggressive mastocytosis types, increased mortality rate and higher tryptase levels among patients with abnormalities of secondary hemostasis raises a question about the role of standard clotting tests in mastocytosis. Although the disease by itself, and particularly AHNMD, can impair coagulation, this seems unlikely here given the close relationship between clotting, episodes of degranulation, and response to steroids. As a consequence, and pending further research, targeted coagulation history and standard blood tests (PT, aPTT) might become relevant markers of poor prognosis in this population, as they would reflect the increasingly uncontrolled clonal activation and proliferation of mast cells.

The main limitations of this study reside in its retrospective analysis, the small number of cases, the delays between the onset of symptoms and blood testing, the lack of systematic platelet aggregation tests, and the intrinsic limitations of coagulation tests (a constantly evolving process). The overall clinical picture and transient character suggest nevertheless that these abnormalities are related to mast cell activity rather than to a pre-existing factor deficiency. We are unable, at this stage, to fully understand the disparity between clinical and laboratory

pictures (which could be partly related to under-diagnosis and to the above limitations).

CONCLUSION

This study suggests that mastocytosis can be a cause of acquired clotting abnormalities, particularly during mast cell activation and degranulation. We suggest that these patients are tested for blood group O and vWF disease as these increase the risk of mucocutaneous bleed during mast cell degranulation. Severe, potentially life-threatening bleeds were described in half of the patients with impairment of the coagulation cascade. Mast cell disease-controlling drugs, particularly steroids, were the only effective way to achieve hemostasis. A larger and prospective study systematically assessing clotting in patients with mastocytosis would be required to confirm whether in the future, standard or specific clotting tests could be used as a marker of mast cell activity and potentially of poor disease prognosis.

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REFERENCES

1. Valent P. Mast cell activation syndromes: definition and classification. *Allergy*. 2013;68:417–424.
2. Valent P, Akin C, Arock M, et al. Definitions, criteria and global classification of mast cell disorders with special reference to mast cell activation syndromes: a consensus proposal. *Int Arch Allergy Immunol*. 2012;157:215–225.
3. Hermine O, Lortholary O, Leventhal PS, et al. Case-control cohort study of patients' perceptions of disability in mastocytosis. *PLoS One*. 2008;3:e2266.
4. Cohen S, Skovbo S, Vestergaard H, et al. Epidemiology of systemic mastocytosis in Denmark. *Br J Haematol*. 2014;166:521–528.
5. Georjin-Lavialle S, Lhermitte L, Dubreuil P, et al. Mast cell leukemia. *Blood*. 2013;121:1285–1295.
6. Adler SN, Klein RALD. Bleeding after liver biopsy in a patient with systemic mastocytosis and malabsorption. *J Clin Gastroenterol*. 1985;7:350–353.
7. Arias M, Villalba C, Requena I, et al. Acute spinal epidural hematoma and systemic mastocytosis. *Spine*. 2004;29:E161–163.
8. Black JS, Feldmann MLS. Systemic mastocytosis as a rare cause of diffuse gastrointestinal hemorrhage. *Am Surg*. 2009;75:429–430.
9. Arrighi F. Diffuse cutaneous mastocytosis of bullous and hemorrhagic form in a young child. *Bull Soc Fr Dermatol Syphiligr*. 1968;75:603–604.
10. Hansen U, Wiese RKJ. Shock and coagulation disorders in systemic mastocytosis. *Dtsch Med Wochenschr*. 1994;119:1231–1234.
11. Nishio M, Kuroda S, Takeda A, et al. A case of mastocytoma associated with disseminated intravascular coagulation (DIC). *Rinsho Ketsueki*. 1982;23:762–767.
12. Poterack CD, Sheth KJ, Henry DPEC. Shock in an infant with bullous mastocytosis. *Pediatr Dermatol*. 1989;6:122–125.
13. Qizilbash ALIH, Frpc C. Gastrointestinal hemorrhage in mastocytosis. *Can Med Assoc J*. 1980;122:311–314.
14. Simone JVHW. Bullous urticaria pigmentosa with bleeding. *J Pediatr*. 1971;78:160–166.

15. Smith TF, Welch TR, Allen JBSJ. Cutaneous mastocytosis with bleeding: probable heparin effect. *Cutis*. 1987;39:241–344.
16. Sucker C, Mansmann G, Steiner S, et al. Fatal bleeding due to a heparin-like anticoagulant in a 37-year-old woman suffering from systemic mastocytosis. *Clin Appl Thromb Hemost*. 2008;14:360–364.
17. Buchholz WA. A case contribution to hemorrhagic mastocytosis. *Z Haut Geschlechtskr*. 1969;44:113–120.
18. Koenig M, Morel J, Reynaud J, et al. An unusual cause of spontaneous bleeding in the intensive care unit—mastocytosis: a case report. *Cases J*. 2008;1:100.
19. Valent P, Horny HP, Escribano L, et al. Diagnostic criteria and classification of mastocytosis: a consensus proposal. *Leuk Res*. 2001;25:603–625.
20. Lassila R, Lindstedt K, Kovanen PT. Native macromolecular heparin proteoglycans exocytosed from stimulated rat serosal mast cells strongly inhibit platelet–collagen interactions. *Arterioscler Thromb Vasc Biol*. 1997;17:3578–3587.
21. Ren S, Lawson AE, Carr M, et al. Human tryptase fibrinogenolysis is optimal at acidic pH and generates anticoagulant fragments in the presence of the anti-tryptase monoclonal antibody B12. *J Immunol*. 1997;159:3540–3548.
22. Thomas VA, Wheelless CJ, Stack MS, et al. Human mast cell tryptase fibrinogenolysis: kinetics, anticoagulation mechanism, and cell adhesion disruption. *Biochemistry*. 1998;37:2291–2298.
23. Huang C, Wong GW, Ghildyal N, et al. The tryptase, mouse mast cell protease 7, exhibits anticoagulant activity in vivo and in vitro due to its ability to degrade fibrinogen in the presence of the diverse array of protease inhibitors in plasma. *J Biol Chem*. 1997;272:31885–31893.
24. Ghildyal BN, Friend DS, Stevens RL, et al. Fate of two mast cell tryptases in V3 mastocytosis and normal BALB/c mice undergoing passive systemic anaphylaxis: prolonged retention of exocytosed mMCP-6 in connective tissues, and rapid accumulation of enzymatically active mMCP-7 in the blood. *J Exp Med*. 1996;184:1061–1073.
25. Prieto-García A, Zheng D, Adachi R, et al. Mast cell restricted mouse and human tryptase-heparin complexes hinder thrombin-induced coagulation of plasma and the generation of fibrin by proteolytically destroying fibrinogen. *J Biol Chem*. 2012;287:7834–7844.
26. Maier M, Spragg JSL. Inactivation of human high molecular weight kininogen by human mast cell tryptase. *J Immunol*. 1983;130:2352–2356.
27. Proud D, Siekierski ESBG. Identification of human lung mast cell kininogenase as tryptase and relevance of tryptase kininogenase activity. *Biochem Pharmacol*. 1988;37:1473–1480.
28. Hirokawa K, Aoki N. Up-regulation of thrombomodulin by activation of histamine H1-receptors in human umbilical-vein endothelial cells in vitro. *Biochem J*. 1991;276 (Pt 3):739–743.
29. Bombeli T, Mueller MHA. Anticoagulant properties of the vascular endothelium. *Thromb Haemost*. 1997;77:408–423.
30. Alter SC, Kramps JA, Janoff A, et al. Interactions of human mast cell tryptase with biological protease inhibitors. *Arch Biochem Biophys*. 1990;276:26–31.
31. Oscarsson LG, Pejler GLU. Location of the antithrombin-binding sequence in the heparin chain. *J Biol Chem*. 1989;264:296–304.