Hindawi Publishing Corporation Mediators of Inflammation Volume 2015, Article ID 524291, 6 pages http://dx.doi.org/10.1155/2015/524291

Research Article

Urinary Malondialdehyde Is Associated with Visceral Abdominal Obesity in Middle-Aged Men

Sun Min Lee,¹ Young Hye Cho,² Sang Yeoup Lee,² Dong Wook Jeong,² A Ra Cho,² Jeong Suk Jeon,² Eun-Ju Park,² Yun Jin Kim,³ Jeong Gyu Lee,³ Yu Hyeon Yi,³ Young Jin Tak,³ Hye Rim Hwang,³ Seung-Hun Lee,³ and Junehee Han⁴

Correspondence should be addressed to Young Hye Cho; younghye82@naver.com

Received 20 May 2015; Accepted 25 August 2015

Academic Editor: Doan T. M. Ngo

Copyright © 2015 Sun Min Lee et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

The purpose of the present study was to investigate multiple anthropometric parameters used to evaluate obesity, particularly visceral abdominal fat area, and various metabolic parameters including malondialdehyde (MDA) as an oxidative stress marker. We evaluated various measures of obesity, including body mass index (BMI), waist circumference (WC), sagittal abdominal diameter, fat percentages using dual-energy X-ray absorptiometry, visceral fat area (VFA), subcutaneous fat area, multiple biomarkers related to metabolic disease, and urinary MDA, in 73 asymptomatic middle-aged men who were not severely obese. We examined relationships between multiple measures of obesity, metabolic markers, and urinary MDA levels and evaluated associations between VFA and urinary MDA. In the visceral obesity group, γ -glutamyl transferase (GGT), uric acid, and urinary MDA levels were significantly higher than in the nonvisceral obesity group (P = 0.008, P = 0.002, and P = 0.018). Urinary MDA (P = 0.003) levels were only significantly positively correlated with VFA among measures of obesity. Urinary MDA, serum GGT, and serum CRP were significantly positively associated with VFA (P = 0.001, P = 0.0046, and P = 0.023, resp.), even after adjusting for BMI and WC.

1. Introduction

Obesity is associated with hypertension, dyslipidemia, metabolic syndrome, and type 2 diabetes [1, 2]. In particular, visceral abdominal fat accumulation is closely linked to insulin resistance and cardiovascular disease (CVD) [3, 4]. Obesity is usually defined by body mass index (BMI), waist circumference (WC), or body fat percentage; however, these measures are limited in their ability to distinguish visceral obesity, which plays a key role in the pathogenesis of cardiometabolic syndrome. Although their average BMI is low, Asians demonstrate higher fat percentages compared with

Caucasians of equivalent BMI and have higher prevalence of diabetes and CVD [5, 6]. Alanine aminotransferase (ALT) [7], γ -glutamyl transferase (GGT) [8], uric acid [9], and C-reactive protein (CRP) [10] are suggested to be metabolic markers associated with hepatic dysfunction, oxidative stress, and inflammation.

Oxidative stress refers to an imbalance between oxidants and antioxidants on a cellular or individual level [11]. Many studies found higher oxidative stress levels in obese than nonobese people [12–14], which may be due to several potential mechanisms including chronic inflammation [15–17], hyperglycemia [18], or impairment of antioxidant defense

¹Department of Laboratory Medicine and Molecular Genetics and Research Institute of Convergence of Biomedical Science and Technology, Pusan National University Yangsan Hospital, Yangsan 626-770, Republic of Korea

²Family Medicine Clinic and Research Institute of Convergence of Biomedical Science and Technology,

Pusan National University Yangsan Hospital, Yangsan 626-770, Republic of Korea ³Department of Family Medicine, Pusan National University Hospital, Busan 49241, Republic of Korea

⁴Research And Statistical Support, Research Institute of Convergence for Biomedical Science and Technology, Pusan National University Yangsan Hospital, Yangsan 626-770, Republic of Korea

systems [19, 20]. Malondialdehyde (MDA) has been recognized as an important indicator of lipid peroxidation that is generated as an end product from oxidative degradation of polyunsaturated fatty acids [21, 22]. Urinary MDA is especially useful as a noninvasive test for monitoring oxidative stress.

Studies that evaluate the correlation between various measures of obesity and metabolic parameters, focusing on visceral obesity, are scarce. Therefore, the present study was designed to investigate the associations between multiple anthropometric parameters of obesity, particularly those relating to visceral abdominal fat area, and various metabolic parameters in middle-aged men who were not severely obese. We were particularly interested in the association between visceral obesity and metabolic parameters, especially urinary MDA, after adjusting for BMI and WC.

2. Material and Methods

2.1. Study Subjects and Ethics. The study was approved by the Institutional Review Board at Pusan National University Yangsan Hospital, and informed written consent was obtained from all subjects before participating. Study subjects included 80 asymptomatic men between the ages of 40 and 60 years who visited the health promotion center in Pusan National University Yangsan Hospital. Subjects with a history of malignancy or a cardiovascular event and subjects receiving medication for acute diseases, such as myocardial infarction and angina pectoris, were excluded. Subjects with low body weight (BMI $< 18.5 \, \text{kg/m}^2$) or severe obesity (BMI $\ge 30.0 \, \text{kg/m}^2$) were also excluded. Seventy-three middle-aged men were enrolled in this study.

2.2. Measurements. Following an overnight fast, blood samples were obtained from an antecubital vein between 08:00 and 09:00 a.m. ALT, GGT, and uric acid concentrations were measured using the Hitachi 7600 Analyzer (Hitachi Co., Ltd., Tokyo, Japan) by an enzymatic colorimetric method. Low-density lipoprotein and high-density lipoprotein were measured with Toshiba TBA200FR using a direct measurement method and triglycerides (TG) were measured using lipase, GK, GPO, and POD with a glycerol blank. CRP was measured using a Behring BN II nephelometer (Dade Behring, Marburg, Germany).

Height and weight were measured down to the first decimal place, and BMI was calculated as body weight (kg) divided by the squared height (m). Using a tape measure, WC was measured from the half-way point between the lower line of the last rib and the upper line of the iliac crest when a subject exhaled and was measured down to the first decimal place. Each subject's sagittal abdominal diameter (SAD) was measured in a supine position with legs extended using a portable sliding-beam caliper (Holtain Ltd., Dyfed, Wales, UK). The vertical abdominal length was measured in a supine position by letting the subject slightly lift the midsection up and inserting the fixed lower arm of the portable sliding-beam caliper at the location of iliac crest of pelvis (space between L4 and L5), letting the subject inhale deeply and

slowly exhale, and lowering the upper arm of the portable sliding-beam caliper down until it touched the abdomen without pressing it. Measurements were taken to the nearest tenth of a cm [23].

Abdominal fat was assessed from computed tomography (CT) scans taken at the L4-L5 level. Abdominal fat was defined as the area corresponding to the pixel range from –190 to –30 Hounsfield units [24]. The visceral and subcutaneous abdominal adipose tissue areas were measured. The fat inside the peritoneum was considered visceral adipose tissue, and the fat between the dermis and muscle fascia was considered subcutaneous adipose tissue. Whole body fat composition was obtained using dual-energy X-ray absorptiometry (DXA) (Hologic Inc., Bedford, MA, USA).

Data on alcohol intake and smoking habits were obtained by interview. Subjects were divided into two groups by the amount of alcohol consumption: nondrinker 0–180 g/week and drinker >180 g/week. Smoking status was classified as nonsmoker or (former or current) smoker. Both diet and physical activity were assessed due to their possible effects on insulin sensitivity. Diet was monitored by using a semi-quantitative food frequency questionnaire [25] and physical activity was assessed using the International Physical Activity Questionnaire [26]. Physical activity levels are expressed in MET-minute.

2.3. Urine MDA. Urinary MDA was measured with high performance liquid chromatography (HPLC). For the quantification of urinary MDA by HPLC [27], 3.0 mL (1%) phosphoric (V) acid, 0.4 mL ultrapure water, and 0.6 mL of sample or standard (1-125 µM MDA) were added to a screw-capped test tube and mixed thoroughly. Then, 1.0 mL of 0.67% thiobarbituric acid was added to all tubes and kept in a 95°C water bath for 1 hour. After incubation, the tubes were placed in an ice bath. Then, the tubes were centrifuged for 15 minutes. The supernatant was filtered before being applied to a Zorbax Eclipse XDB-C8 (I.d. $4.6 \times$ $250 \,\mathrm{mm}$, $5 \,\mu\mathrm{m}$, Agilent). Measurements were made with a UV-Vis detector (Agilent 1100 series HPLC system, USA) at 532 nm. The mobile phase was 0.05 M potassium phosphate buffer (pH 6.8) with methanol (50:50, v/v). The flow rate was 0.5 mL/min. MDA and creatinine were analyzed in two voided specimens. Urinary MDA levels were expressed as μmol/g creatinine, averaged, and used for analysis.

2.4. Statistical Analysis. Descriptive data were expressed as the mean value (\pm SD) or number (%). The 73 subjects were divided into two groups: the visceral obesity (VO) group and nonvisceral obesity (non-VO) group. The visceral obesity group consisted of the subjects whose CT visceral fat area (CT-VFA) was ≥100 cm². The Shapiro-Wilk test was used to test the normality of the variables. To compare the means of two groups, we used the two-sample t-test or the Mann-Whitney U test for continuous variables depending on the normality of the variables. For the categorical variables, the Chi-square test was used to verify the group effect. Correlations between variables were tested using Spearman's correlation coefficients. Finally, multiple regression analysis

TABLE 1: General characteristics of subjects.

Characteristics	Value
Number of participants	73
Age (y; mean \pm SD)	51.2 ± 5.8
Height (cm; mean ± SD)	171.4 ± 5.3
Weight (kg; mean ± SD)	71.8 ± 7.7
BMI (kg/m^2 ; mean \pm SD)	24.4 ± 2.1
Waist circumference (cm; mean ± SD)	86.3 ± 6.7
Abdominal VFA (cm ² ; mean ± SD)	100.8 ± 35.2
Abdominal SFA (cm ² ; mean ± SD)	127.6 ± 50.2

BMI: body mass index; VFA: visceral fat area; SFA: subcutaneous fat area.

was performed to investigate the relationship among the variables considering GGT, UA, CRP, TG, and urinary MDA levels as dependent variables and abdominal VFA as the independent variable after adjusting for BMI and WC. The analysis was conducted using SPSS version 18.0 for Windows (SPSS Inc., Chicago, IL). Statistical significance was accepted for *P* values < 0.05.

3. Results

3.1. Clinical Characteristics of Subjects. The general characteristics of the study subjects are presented in Table 1. The overall average age was 51.2 years and mean BMI was 24.4 kg/m². Mean abdominal VFA was 100.8 cm².

3.2. Differences between the Characteristics of Patients in the Two Study Groups. No differences in age, smoking status, alcohol habits, physical activity, or dietary intake were observed between the VO group and non-VO group (Table 2).

The VO group had significantly higher BMI, WC, and SAD than the non-VO group (P=0.009, P=0.002, and P=0.003, resp.), and DXA-measured trunk fat and total fat percentages were also significantly higher in the VO group (P=0.005 and P=0.014, resp.). However, there were no differences in DXA-measured upper arm fat and lower leg fat percentages. CT-measured subcutaneous fat area was similar between the two groups (P=0.117). GGT and uric acid levels were higher in the VO group (P=0.008 and P=0.002, resp.), and TG levels were marginally higher in the VO group (P=0.063). Urinary MDA levels were significantly higher in the VO group (P=0.018).

3.3. Correlations between Obesity Measures, Metabolic Parameters, and Urinary MDA. BMI, WC, and DXA-measured total fat percentages were significantly positively correlated with ALT, GGT, CRP, and TG (Table 3), but not significantly correlated with uric acid or urinary MDA. However, urinary MDA (r=0.357, P=0.002) and uric acid (r=0.263, P=0.027) levels were only significantly positively correlated with VFA among the measures of obesity.

3.4. Multiple Regression Analysis of Abdominal VFA and Metabolic Parameter and Urinary MDA. In all subjects, the association between VFA and GGT ($\beta = 0.23$, P = 0.028)

and CRP ($\beta=0.278, P=0.01$), as well as urinary MDA ($\beta=0.362, P=0.001$), remained significant after adjusting for BMI (Table 4). These associations also remained after adjusting for BMI and WC (GGT: $\beta=0.207, P=0.046$; CRP: $\beta=0.245, P=0.023$; urinary MDA: $\beta=0.349, P<0.001$). On the other hand, the associations between VFA and uric acid or TG were not significant after adjusting for BMI and WC.

4. Discussion

In the present study, we investigated associations between abdominal VFA and common clinical metabolic biomarkers in middle-aged men without morbid obesity. In addition, we were particularly interested in the correlation between urinary MDA, a known oxidative stress marker, and abdominal visceral adiposity. We demonstrated that urinary MDA, GGT, and CRP were significantly positively associated with VFA, even after adjusting for BMI and WC.

Oxidative stress is considered a crucial factor because this is an early instigator of metabolic syndrome [12] and a contributor to the development of major obesity-related comorbidities such as CVD [13]. MDA is a biomarker derived from lipid peroxides and that is considered useful marker of oxidative marker [28]. Previously published data showed that participants with a high VFA (≥100 cm²) were more likely to have high plasma MDA levels, which is consistent with our findings, although our study examined urine MDA level [29]. We checked urinary MDA as oxidative stress marker because that is noninvasive test. In our data, urine MDA is a marker of oxidative stress in obese people, especially those with visceral obesity.

We wondered if previously known cardiometabolic biomarkers as well as urinary MDA levels have been connected with various measures of obesity and specifically those laboratory markers that reflect visceral obesity. We confirmed that urinary MDA levels were related to abdominal visceral fat area after adjusting for BMI and WC, which are typically used to evaluate clinical obesity. Urinary MDA levels can indicate inflamed adipose tissue. Further study is required to determine the clinical utility of urinary MDA levels.

Serum uric acid was higher in the VO group than the non-VO group and showed a positive correlation with VFA; however, after adjusting for BMI and WC, there was no association with VFA. On the other hand, GGT and CRP demonstrated a significant association with VFA even after adjusting for BMI and WC. Previous studies showed GGT was strongly associated with metabolic syndrome, which is a combined expression of metabolic disorders including abdominal obesity [8]. In several studies, CRP levels already showed positive and significant correlations with body fat mass and VFA measured by CT [30, 31] in men. These results suggested that CRP levels can reflect inflammation by visceral adiposity. Previously published data demonstrated a highly significant association between smoke exposure and MDA [32, 33]; however, in the present study, the levels of urinary MDA in smokers were not elevated compared with nonsmokers.

TABLE 2: Characteristics of patients in the two study groups.

	Nonvisceral obesity	Visceral obesity	P value
	(n = 38)	(n = 35)	1 value
Sociodemographic parameters			
Age (years)	50.3 ± 6.1	52.2 ± 5.5	0.185
Smoking status			0.162
Nonsmoker	24 (60.0)	16 (40.0)	
Smoker	14 (42.4)	19 (57.6)	
Alcohol consumer	30 (50.8)	33 (49.2)	0.770
Activity (METS/week)*	1602.3 ± 2000.8	907.1 ± 932.2	0.268
Dietary parameters			
Energy intake (Kcal/kg/day)	28.9 ± 5.0	28.0 ± 5.1	0.475
Protein intake (g/kg/day)	1.2 ± 0.3	1.2 ± 0.3	0.800
Fat intake (g/kg/day)	0.7 ± 0.2	0.7 ± 0.2	0.543
Carbohydrate (g/kg/day)	4.6 ± 0.7	4.3 ± 0.8	0.163
Anthropometric parameters			
Body mass index (kg/m²)	23.8 ± 1.8	25.1 ± 2.2	0.009
Waist circumference (cm)	83.9 ± 5.1	88.8 ± 7.4	0.002
Sagittal abdominal diameter (cm)	18.3 ± 1.4	19.8 ± 2.6	0.003
DXA-measured fat			
Upper arm fat (%)	24.8 ± 4.6	26.4 ± 4.5	0.139
Lower leg fat (%)	23.6 ± 4.0	25.1 ± 4.6	0.148
Trunk fat (%)	27.6 ± 3.9	30.6 ± 5.1	0.005
Total fat (%)	25.6 ± 3.4	27.8 ± 4.2	0.014
CT-measured abdominal fat area			
VFA (cm ²)*	74.9 ± 16.4	128.8 ± 27.9	<0.001
SFA (cm ²)*	120.2 ± 42.1	135.7 ± 57.3	0.117
Metabolic parameters			
ALT (IU/L)*	31.2 ± 17.0	35.1 ± 16.0	0.156
GGT (IU/L)*	46.2 ± 28.7	99.5 ± 99.5	0.008
Uric acid (IU/L)*	6.2 ± 1.0	7.1 ± 1.2	0.002
CRP (mg/dL)*	0.10 ± 0.07	0.18 ± 0.25	0.147
LDL-cholesterol (mg/dL)	138.1 ± 35.6	137.9 ± 32.9	0.981
HDL-cholesterol (mg/dL)*	53.2 ± 10.7	49.3 ± 10.9	0.154
TG (mg/dL)*	132.4 ± 72.0	173.2 ± 97.0	0.063
Oxidative stress parameter			
Urinary MDA* (μmol/g creatinine)	1.56 ± 0.85	2.08 ± 1.16	0.018

Data are expressed as the mean \pm SE or the number (%).

P value by 2-sample t-test or chi-square test.

 $VFA: visceral\ fat\ area; SFA: subcutaneous\ fat\ area; DXA:\ dual-energy\ X-ray\ absorptiometry;\ ALT:\ alanine\ aminotransferase;\ GGT:\ \gamma-glutamyl\ transferase;\ CRP:\ C-reactive\ protein;\ LDL:\ low-density\ lipoprotein;\ HDL:\ high-density\ lipoprotein;\ TG:\ triglyceride;\ MDA:\ malondialdehyde.$

One MET is roughly equivalent to 1 kcal/min for a person weighing 60 kg.

This study has several limitations, which include a relatively small sample size and a study population limited to relatively healthy middle-aged men. However, the use of this population limits the effects of various unmeasured confounding factors. Unfortunately, we did not measure blood MDA concentration and could not evaluate their correlation with urinary MDA and VFA. In addition, the study was conducted

using a cross-sectional design, and thus further studies are required. Nonetheless, we believe our findings are meaningful because this study represents a new attempt to evaluate multiple anthropometric parameters evaluating obesity including visceral abdominal fat area and various metabolic parameters. Furthermore, we suggest that urinary MDA levels may be useful as a marker of inflamed adipose tissue.

^{*} P value by Mann-Whitney U test.

	BMI	WC	Fat percent	VFA	SFA
A T /TD				· · · · · · · · · · · · · · · · · · ·	
ALT	0.394*	0.305^{*}	0.277^{*}	0.154	0.384*
GGT	$0.355^{^{T}}$	0.335^{\dagger}	0.380^{\dagger}	$\boldsymbol{0.389}^{\dagger}$	0.376^{\dagger}
Uric acid	0.112	0.168	0.105	0.263*	0.066
CRP	0.343^{\dagger}	0.346^{\dagger}	0.290*	0.326^{\dagger}	0.345^{\dagger}
TG	0.255*	0.274^*	0.257*	0.254^*	0.233*
Urinary MDA	0.081	0.031	-0.013	$\boldsymbol{0.357}^{\dagger}$	0.005

Table 3: Correlations between obesity indices, metabolic parameters, and urinary malondialdehyde levels.

ALT: alanine aminotransferase; GGT: γ -glutamyl transferase; CRP: C-reactive protein; TG: triglyceride; MDA: malondialdehyde; BMI: body mass index; VFA: visceral fat area; SFA: subcutaneous fat area; WC: waist circumference. P value by Spearman's correlation.

Table 4: Multiple regression analysis of associations between abdominal visceral fat area and metabolic parameters.

Abdominal VFA	Adjusted BMI			P value	Adjusted BMI and WC				P value	
	F	B	SE	β	r value	F	B	SE	β	r value
GGT	14.03	0.106	0.047	0.230	0.028*	10.70 [†]	0.095	0.047	0.207	0.046*
Uric acid	12.43^{\dagger}	5.703	3.240	0.185	0.083	10.23^{\dagger}	4.680	3.196	0.152	0.148
CRP	15.34^{\dagger}	53.703	20.231	0.278	0.010*	11.296^{\dagger}	47.434	20.396	0.245	0.023*
TG	11.17^{\dagger}	0.035	0.043	0.086	0.419	9.008^{\dagger}	0.028	0.042	0.069	0.507
Urinary MDA	19.37 [†]	12.321	3.301	0.362	<0.001 [†]	14.389 [†]	11.867	3.259	0.349	0.001

GGT: γ -glutamyl transferase; CRP: C-reactive protein; TG: triglyceride; MDA: malondialdehyde; BMI: body mass index; VFA: visceral fat area; WC: waist circumference.

5. Conclusion

Urinary MDA, serum GGT, and serum CRP were significantly positively correlated with VFA, even after adjusting for BMI and WC in middle-aged healthy men. Further study is needed to confirm the validity of urinary MDA as a marker of inflamed adipose tissue.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgment

This study was supported by a grant from DFI Co., Ltd.

References

- [1] J. B. Meigs, P. W. F. Wilson, C. S. Fox et al., "Body mass index, metabolic syndrome, and risk of type 2 diabetes or cardiovascular disease," *Journal of Clinical Endocrinology and Metabolism*, vol. 91, no. 8, pp. 2906–2912, 2006.
- [2] P. W. Wilson, R. B. D'Agostino, L. Sullivan, H. Parise, and W. B. Kannel, "Overweight and obesity as determinants of cardiovascular risk: the framingham experience," *Archives of Internal Medicine*, vol. 162, no. 16, p. 1867, 2002.
- [3] A. Hiuge-Shimizu, K. Kishida, T. Funahashi et al., "Absolute value of visceral fat area measured on computed tomography scans and obesity-related cardiovascular risk factors in large-scale Japanese general population (the VACATION-J study)," *Annals of Medicine*, vol. 44, no. 1, pp. 82–92, 2012.

- [4] K. Kishida, T. Funahashi, and I. Shimomura, "Clinical importance of assessment of type 2 diabetes mellitus with visceral obesity. A Japanese perspective," *Current Diabetes Reviews*, vol. 8, no. 2, pp. 84–91, 2012.
- [5] Y.-W. Park, D. B. Allison, S. B. Heymsfield, and D. Gallagher, "Larger amounts of visceral adipose tissue in Asian Americans," *Obesity Research*, vol. 9, no. 7, pp. 381–387, 2001.
- [6] C. M. Khoo, S. Sairazi, S. Taslim et al., "Ethnicity modifies the relationships of insulin resistance, inflammation, and adiponectin with obesity in a multiethnic Asian population," *Diabetes Care*, vol. 34, no. 5, pp. 1120–1126, 2011.
- [7] G. Marchesini, M. Brizi, G. Blanchi et al., "Nonalcoholic fatty liver disease: A feature of the metabolic syndrome," *Diabetes*, vol. 50, no. 8, pp. 1844–1850, 2001.
- [8] J. G. Lee, S. Lee, Y. J. Kim et al., "Multiple biomarkers and their relative contributions to identifying metabolic syndrome," *Clinica Chimica Acta*, vol. 408, no. 1-2, pp. 50–55, 2009.
- [9] A. Onat, H. Uyarel, G. Hergenc et al., "Serum uric acid is a determinant of metabolic syndrome in a population-based study," *American Journal of Hypertension*, vol. 19, no. 10, pp. 1055–1062, 2006.
- [10] M. K. Rutter, J. B. Meigs, L. M. Sullivan, R. B. D'Agostino Sr., and P. W. F. Wilson, "C-reactive protein, the metabolic syndrome, and prediction of cardiovascular events in the framingham offspring study," *Circulation*, vol. 110, no. 4, pp. 380–385, 2004.
- [11] H. Sies, "Biochemistry of oxidative stress," *Angewandte Chemie—International Edition in English*, vol. 25, no. 12, pp. 1058–1071, 1986.
- [12] S. Furukawa, T. Fujita, M. Shimabukuro et al., "Increased oxidative stress in obesity and its impact on metabolic syndrome," *Journal of Clinical Investigation*, vol. 114, no. 12, pp. 1752–1761, 2004.

^{*}P < 0.05, †P < 0.01.

^{*} P < 0.05, † P < 0.01.

[13] J. F. Keaney Jr., M. G. Larson, R. S. Vasan et al., "Obesity and systemic oxidative stress: clinical correlates of oxidative stress in the framingham study," *Arteriosclerosis, Thrombosis,* and Vascular Biology, vol. 23, no. 3, pp. 434–439, 2003.

- [14] S. O. Olusi, "Obesity is an independent risk factor for plasma lipid peroxidation and depletion of erythrocyte cytoprotectic enzymes in humans," *International Journal of Obesity*, vol. 26, no. 9, pp. 1159–1164, 2002.
- [15] J.-M. Fernández-Real, M. Broch, J. Vendrell, and W. Ricart, "Insulin resistance, inflammation, and serum fatty acid composition," *Diabetes Care*, vol. 26, no. 5, pp. 1362–1368, 2003.
- [16] I. Saito, K. Yonemasu, and F. Inami, "Association of body mass index, body fat, and weight gain with inflammation markers among rural residents in Japan," *Circulation Journal*, vol. 67, no. 4, pp. 323–329, 2003.
- [17] G. Davì, M. T. Guagnano, G. Ciabattoni et al., "Platelet activation in obese women: role of inflammation and oxidant stress," *Journal of the American Medical Association*, vol. 288, no. 16, pp. 2008–2014, 2002.
- [18] D. Aronson and E. J. Rayfield, "How hyperglycemia promotes atherosclerosis: molecular mechanisms," *Cardiovascular Diabetology*, vol. 1, article 1, 2002.
- [19] H. K. Vincent, S. K. Powers, A. J. Dirks, and P. J. Scarpace, "Mechanism for obesity-induced increase in myocardial lipid peroxidation," *International Journal of Obesity*, vol. 25, no. 3, pp. 378–388, 2001.
- [20] P. Wallström, E. Wirfält, P. H. Lahmann, B. Gullberg, L. Janzon, and G. Berglund, "Serum concentrations of β -carotene and α -tocopherol are associated with diet, smoking, and general and central adiposity," *American Journal of Clinical Nutrition*, vol. 73, no. 4, pp. 777–785, 2001.
- [21] A. A. Horton, S. Fairhurst, and J. S. Bus, "Lipid peroxidation and mechanisms of toxicity," *Critical Reviews in Toxicology*, vol. 18, no. 1, pp. 27–79, 1987.
- [22] E. Seljeskog, T. Hervig, and M. A. Mansoor, "A novel HPLC method for the measurement of thiobarbituric acid reactive substances (TBARS). A comparison with a commercially available kit," *Clinical Biochemistry*, vol. 39, no. 9, pp. 947–954, 2006.
- [23] U. Risérus, J. Ärnlöv, K. Brismar, B. Zethelius, L. Berglund, and B. Vessby, "Sagittal abdominal diameter is a strong anthropometric marker of insulin resistance and hyperproinsulinemia in obese men," *Diabetes Care*, vol. 27, no. 8, pp. 2041–2046, 2004.
- [24] B.-K. Lee and Y. Kim, "Iron deficiency is associated with increased levels of blood cadmium in the Korean general population: analysis of 2008-2009 Korean National Health and Nutrition Examination Survey data," *Environmental Research*, vol. 112, pp. 155–163, 2012.
- [25] S. Lee, H. K. Park, S. P. Son, C. W. Lee, I. J. Kim, and H. J. Kim, "Effects of oral magnesium supplementation on insulin sensitivity and blood pressure in normo-magnesemic nondiabetic overweight Korean adults," *Nutrition, Metabolism & Cardiovascular Diseases*, vol. 19, no. 11, pp. 781–788, 2009.
- [26] C. L. Craig, A. L. Marshall, M. Sjöström et al., "International physical activity questionnaire: 12-country reliability and validity," *Medicine and Science in Sports and Exercise*, vol. 35, no. 8, pp. 1381–1395, 2003.
- [27] R. Mendes, C. Cardoso, and C. Pestana, "Measurement of malondialdehyde in fish: a comparison study between HPLC methods and the traditional spectrophotometric test," *Food Chemistry*, vol. 112, no. 4, pp. 1038–1045, 2009.

[28] J. Lykkesfeldt, "Malondialdehyde as biomarker of oxidative damage to lipids caused by smoking," *Clinica Chimica Acta*, vol. 380, no. 1-2, pp. 50–58, 2007.

- [29] Y. Jang, O. Y. Kim, H. J. Ryu et al., "Visceral fat accumulation determines postprandial lipemic response, lipid peroxidation, DNA damage, and endothelial dysfunction in nonobese Korean men," *Journal of Lipid Research*, vol. 44, no. 12, pp. 2356–2364, 2003.
- [30] I. Lemieux, A. Pascot, D. Prud'homme et al., "Elevated C-reactive protein: another component of the atherothrombotic profile of abdominal obesity," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 21, no. 6, pp. 961–967, 2001.
- [31] A. Cartier, M. Côté, I. Lemieux et al., "Age-related differences in inflammatory markers in men: contribution of visceral adiposity," *Metabolism*, vol. 58, no. 10, pp. 1452–1458, 2009.
- [32] G. Block, M. Dietrich, E. P. Norkus et al., "Factors associated with oxidative stress in human populations," *American Journal of Epidemiology*, vol. 156, no. 3, pp. 274–285, 2002.
- [33] F. Ozguner, A. Koyu, and G. Cesur, "Active smoking causes oxidative stress and decreases blood melatonin levels," *Toxicology and Industrial Health*, vol. 21, no. 1-2, pp. 21–26, 2005.