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EDITORIAL

# Intimal pericytes as the second line of immune defence in atherosclerosis

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#### Abstract

Inflammation plays an essential role in the development of atherosclerosis. The initiation and growth of atherosclerotic plaques is accompanied by recruitment of inflammatory and precursor cells from the bloodstream and their differentiation towards pro-inflammatory phenotypes. This process is orchestrated by the production of a number of pro-inflammatory cytokines and chemokines. Human arterial intima consists of structurally distinct leaflets, with a proteoglycan-rich layer lying immediately below the endothelial lining. Recent studies reveal the important role of stellate pericyte-like cells (intimal pericytes) populating the proteoglycan-rich layer in the development of atherosclerosis. During the pathologic process, intimal pericytes may participate in the recruitment of inflammatory cells by producing signalling molecules and play a role in the antigen presentation. Intimal pericytes are also involved in lipid accumulation and the formation of foam cells. This review focuses on the role of pericytelike cells in the development of atherosclerotic lesions.

Key words: Atherosclerosis; Arteries; Intima; Immuneinflammatory processes; Pericyte-like cells

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**Core tip:** Intimal stellate cells, expressing smooth muscle  $\alpha$ -actin that co-express antigen 3G5 antigen (known to be specific for pericytes), reside in the intima of human large arteries. There cells have been defined as "pericytes-like satellite cells" or "intimal pericytes". Because of the peculiarities of the distribution of these cells and because of the current lack of our knowledge about the spectrum of the expression of other pericyte-associated markers in the arterial wall, it is reasonable to avoiding at this time to identify smooth muscle  $\alpha$ -actin(+)/3G5 antigen(+) stellate-shaped cells as true pericytes. The review highlights the importance of intimal pericytes in atherosclerosis.

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### INTRODUCTION

Cardiovascular disorders (CVDs) are currently the main cause of mortality in developed countries, and atherosclerosis is the major contributor to the CVD risk<sup>[1]</sup>. The disease affects large arteries causing lipid deposition in the arterial wall, local inflammatory process and the formation of plaques. Atherosclerotic plaques progression leads to luminal narrowing of blood vessels, which by itself can result in the ischemia of corresponding organs. However, most dangerous is the acute thrombus formation on the surface of a ruptured atherosclerotic plaque. Thrombosis can lead to rapid and life-threatening events, such as acute coronary syndrome and stroke<sup>[2]</sup>. Early stages of atherosclerosis development are mostly clinically silent, and the first manifestations of the disease are often fatal. At the same time, subclinical atherosclerosis is a prevalent condition in the modern society, especially among the ageing population. It has been reported recently that up to 5% of women and 12% of men over the age of 80 suffer from asymptomatic atherosclerotic carotid artery stenosis<sup>[3]</sup>. Other studies demonstrated the high rate of asymptomatic atherosclerosis incidence in young and middle-aged people, with up to 100% of apparently healthy individuals having atherosclerotic lesions at various stages of progression<sup>[4,5]</sup>. Along with the wellknown benefits of life style correction, the development of more specific therapies could help decreasing the risk of atherosclerosis progression and consequent CVD development. It is therefore important to improve our understanding of the pathological mechanisms that trigger atherosclerotic lesion development in the arterial wall.

The intimal barrier of human arterial wall consists of the endothelium and the subendothelial net-like tissue layer formed by pericyte-like cells that have recently been characterized as true pericytes<sup>[6,7]</sup>. Whereas the endothelium serves as the first line of defence against the development of atherosclerotic lesion, the pericytecontaining subendothelial layer can be regarded as the second defence line essential for normal functioning and protection of the arterial wall. Apart from pericytes, this layer contains other cell types, including smooth muscle cells, macrophages, lymphocytes, mast cells, dendritic cells, and other cell types. Pericyte-containing subendothelial layer is a characteristic component of both arterial and venous intima<sup>[6]</sup>. Its role in the development of human pathology remained unknown until recently. However, the accumulating evidence demonstrated its importance in many severe vascular disorders, such as saphenous vein graft disease, thrombosis and atherosclerosis. Macrovascular pericytes are capable of rapid proliferation and participate in immune reactions. Moreover, these cells have been identified as a source of coagulation-inducing tissue factor<sup>[7]</sup>. In this review we will summarize the current knowledge on the structure and function of the subendothelial leaflet of human macrovascular intima and its role in the pathogenesis of atherosclerosis.

## STRUCTURE OF NORMAL AND ATHEROSCLEROTIC ARTERIAL INTIMA

Aortic intima is the part of the blood vessel wall located between the internal elastic lamina and the lumen with endothelial lining, and consists of two distinct layers (Figure 1)<sup>[8,9]</sup>. Muscular-elastic layer represents the external part of the intima, and is separated from the internal intimal layer by the limiting membrane. This layer is formed by longitudinally oriented elongated cells and elastic fibres. The internal intimal layer, also known as proteoglycan-rich or connective tissue layer<sup>[10,11]</sup>, contains randomly-oriented connective tissue fibres and morphologically heterogeneous cell population<sup>[12]</sup>. The intimal layers differ from each other by the composition of glycosaminoglycans and fibres. Muscular-elastic layer is rich in elastin fibres, and proteoglycan-rich layer - in collagen and reticulin<sup>[13]</sup>. Moreover, collagen composition and structure are different between the two layers: longitudinally oriented collagens I and III are common in the muscular-elastic layer, while collagens  ${\rm I\!V}$  and  ${\rm V}$ are detected primarily in the proteoglycan layer, where they form the endothelial basal membrane.

The development of atherosclerotic lesion has been carefully studied by our group on human aorta samples from healthy subjects and atherosclerotic patients<sup>[8-13]</sup>. Atherosclerotic process is tightly associated with thickening of the arterial intima. Careful analysis of the intima thickness along the arteries affected by atherosclerosis in comparison to normal tissues revealed that muscularelastic layer remained unaltered in the fatty streak area and became only slightly thicker in the atherosclerotic plaque area<sup>[8-13]</sup>. By contrast, the proteoglycan layer was 2-fold thicker in the fatty streak and 4-fold - in

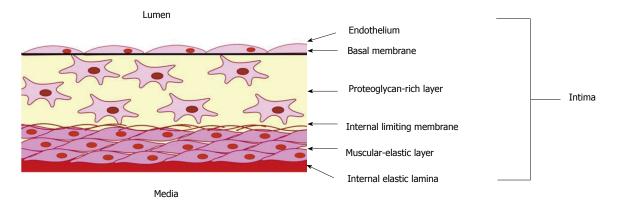


Figure 1 Schema showing the organization of the intima of the arterial wall. The proteoglycan-rich layer which contains a heterogeneous population of cells, including macrovascular pericytes, is located just below the endothelial monolayer. Intimal pericytes form a network of cells interconnected through gap junctions. The muscular-elastic layer, formed by elongated contractile smooth muscular cells, lies below the proteoglycan-rich layer.

the atherosclerotic lesion areas, and was the major contributor of the arterial stenosis. This thickened proteoglycan layer had increased collagen content. Moreover, longitudinal orientation of collagen fibres was altered, and lipid droplets were present between the interstitial collagen fibres<sup>[8-13]</sup>. Atherosclerotic plaques were characterized by the accumulation of all collagen types, especially in the fibrous cap area. Collagens IV and V formed a thick layer in the endothelial basal membrane area and surrounded the subendothelial cells. At the same time, no prominent alterations of collagen structure and composition were present in the muscular-elastic layer of the intima<sup>[8-13]</sup>.

Changes of lipid content accompany the development of atherosclerotic plaques in the aortic intima. In the normal tissue, extracellular lipid droplets visualised by Oil Red O staining were located along the elastic membrane. In the fatty streaks, both extra- and intracellular droplets were present, mostly in the proximity of the lumen. In atherosclerotic plaques, lipid deposition could be observed in the proteoglycan layer, and total lipid content was 8-fold higher than in the unaffected tissue<sup>[6]</sup>. Lipid content of muscular-elastic layer was increased to a lesser extent, being 4.4-fold higher than control. Cellular composition of the proteoglycanrich layer of atherosclerotic intima was also altered in comparison with the healthy tissue<sup>[6]</sup>. Total cell count assessed by alcohol-alkaline dissociation of the fixed tissue, was 1.5 and 2-fold higher in the fatty streak and atherosclerotic plaque respectively<sup>[6]</sup>.

Taken together, these observations clearly indicate that the subendothelial proteoglycan-rich layer of the arterial intima undergoes the most prominent alterations in course of the atherosclerotic plaque development, including thickening, lipid deposition and the accumulation of collagen fibres with disturbed orientation<sup>[6,8-13]</sup>.

## CELLULAR COMPOSITION OF MACROVASCULAR INTIMA

Human macrovascular intima is made of a heterogeneous

cell population. Immediately below the endothelial lining and in close contact with it, there is a threedimensional net formed by pericyte-like cells. Early studies have identified pericytes as mostly microvascular cells that play an important role in angiogenesis and vessel branching and are essential for maintaining the normal structure of the capillary wall and the endothelial barrier function, including the maintenance the bloodbrain barrier<sup>[14,15]</sup>. Pericytes were also demonstrated to contribute to embryonic development of the aorta<sup>[16]</sup>. It has become evident, however, that pericyte-like cells are also present in the walls of large blood vessels<sup>[17]</sup>. Apart from vasa vasorum microvessels<sup>[18]</sup>, pericyte-like stellate cells were found in the intima of large arteries<sup>[6,7,19-21]</sup>.

Accurate identification of pericytes, as well as their characterisation in in vitro studies, remains challenging, since these cells have flexible phenotype. The expression of numerous pericyte marker proteins is highly dynamic and depends on the surrounding milieu. Moreover, many marker proteins are shared between pericytes and vascular smooth muscle cells (VSMCs)<sup>[14]</sup>. One of such markers is the  $\alpha$ -smooth muscle actin ( $\alpha$ SMA), which is normally expressed in VSMCs, but is also common in pericytes. However, its expression in the latter cell type can vary depending on the surrounding milieu<sup>[22]</sup>. For instance, pericytes from non-contractile capillaries can be  $\alpha$ SMA-negative<sup>[23]</sup>. The expression of  $\alpha$ SMA in pericytes can also be regulated by cell maturity<sup>[23]</sup> or stimulation by endothelin-1<sup>[24]</sup>. These considerations have to be taken into account while analyzing the cell population of macrovesicular intima. In the intima of large human arteries, the majority of resident cells were found to be  $\alpha$ SMA-positive, especially in the muscularelastic layer, where this population reached two thirds of the total amount of cells<sup>[25]</sup>. In the proteoglycan layer, the percentage of aSMA-positive cells was much lower. These cells were different from typical VSMCs, were less densely packed and had a characteristic branched morphology, with long processes forming intracellular contacts. Moreover, some aSMA-positive intimal cells expressed marker proteins that were unusual for muscular cells,

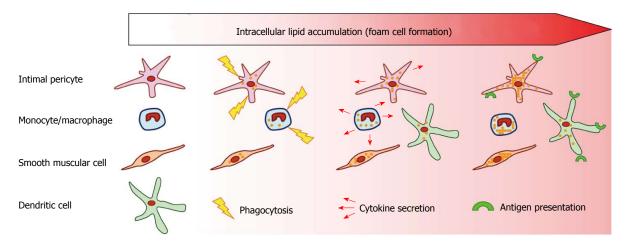


Figure 2 Scheme showing the roles of arterial wall cells in atherogenesis. Several types of arterial wall cells participate in lipid accumulation and foam cell formation. Macrophages and intimal pericytes accumulate lipids through phagocytosis and participate in the local inflammatory process secreting pro-inflammatory cytokines. Dendritic cells, together with macrophages and intimal pericytes express antigen-presenting complexes, further promoting the inflammatory process.

such as CD68, which is considered a macrophagespecific antigen<sup>[26]</sup>. Interestingly, the proportion of  $\alpha$ SMA+CD68+ cells increased in atherosclerotic lesions, as well as in primary cell culture exposed to atherogenic modified low-density lipoprotein (LDL). Apparently, the acquisition of the "macrophage"-specific marker reflects the engagement of cells into phagocytic activity<sup>[27,28]</sup>. The large percentage of  $\alpha$ SMA-negative cells in the arterial intima can partly be explained by the loss of contractile apparatus, since these cells acquire other functions than maintaining the vascular tone, such as metabolic homeostasis and nutrition or participation in the immune response in the blood vessel wall<sup>[29]</sup>.

Therefore, accurate identification of pericytes in the intimal layer should not be based on  $\alpha$ SMA expression exclusively, and has to include the analysis of other pericyte markers expression and morphological data. Commonly used pericyte markers include plateletderived growth factor receptor  $\beta$ , CD146, aminopeptidase A and N (CD13), endoglin, neuron-glial 2, non-muscle myosin, desmin, vimentin and nestin<sup>[14,30]</sup>. However, most of these markers are shared with VSMCs and/or dependent on pericyte maturity or activation state, and therefore should be used in combination to avoid falsepositive results. It has been demonstrated that some antigens expressed in resident intimal cells are not present in VSMCs. Antigen 3G5 is an O-sialoganglioside specific for microvascular pericytes<sup>[31]</sup>. It was found in some intimal resident cells, but not in the tunica media<sup>[26,32]</sup>. In humans, 3G5-positive pericytes form the net-like subendothelial tissue layer in normal human arterial intima and account for approximately one third of intimal cells (Figure 2)<sup>[26]</sup>. Another pericyte antigen detected in the macrovascular intima is 2A7, also known as melanoma chondroitin sulphate proteoglycan<sup>[26,33]</sup>. The expression of this antigen is typical for activated pericytes in the active angiogenesis phase. The expression of these pericyte marker antigens alters in atherogenesis. As

demonstrated on primary cultures of subendothelial cells exposed to atherogenic modified LDL, intracellular lipid deposition results in the reduction of 3G5-positive cell fraction, whereas 2A7-expressing fraction increases<sup>[33]</sup>.

Studies of the primary cell culture obtained from human macrovascular intima helped revealing the mechanisms of pericyte acquisition of characteristic stellate shape. Pericyte arborisation could be induced by increasing the intracellular cAMP concentration<sup>[33]</sup>. Interestingly, cells originating from the proteoglycan layer were more susceptible of arborisation, with up to 100% of them being capable of acquiring the stellate shape, whereas in population originating from the muscular-elastic layer it could be induced in only 50% of cells. The arborisation was associated with re-distribution of connexion 43 (Cx43), which is responsible for gap junction formation. Stellate cells formed large Cx43-positive sites located at the ends of cellular processes, where intracellular contacts were formed. In vivo, pericytes form stable contacts with the endothelial cells, which could also be reproduced in coculture of the two cell types<sup>[7]</sup>.

Apart from pericytes, other cell types populating the arterial intima have been described. These include cells of hematogenous lineage - macrophages<sup>[34-36]</sup>, lymphocytes<sup>[37,38]</sup>, mast cells<sup>[39]</sup> and dendritic cells. Cells of hematogenous lineage are located exclusively in the subendothelial proteoglycan layer of intima and are enriched in atherosclerotic plaques, where they can account for up to 20% of total cell population.

## ROLE OF PERICYTE-LIKE CELLS (INTIMAL PERICYTES) IN THE PATHOGENESIS OF ATHEROSCLEROSIS

According to current consensus, pericytes are pluripotent cells that can serve as progenitors for other cell types of



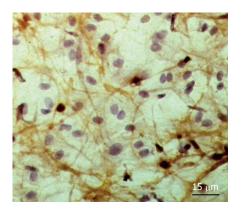


Figure 3 Cellular network formed by 3G5+ cells located under the luminal endothelium in undiseased intima. This cellular network was identified by means of the application of peroxidase-anti-peroxidase immunohistochemical analysis (brown colour of reaction product) in en face preparation of a tissue specimen of the human aorta. Counterstain with haematoxylin.

mesenchymal origin, including smooth muscle cells<sup>[40]</sup>, fibroblasts, osteoblasts  $^{[41,42]}$ , chondrocytes  $^{[40]}$  and adipocytes<sup>[43]</sup>. Pericytes are actively involved in various conditions associated with impaired microcirculation, such as diabetes, inflammation, wound healing and tumor growth. Converging evidence indicates pericyte implication in the pathogenesis of atherosclerosis. In presence of proinflammatory microenvironment and atherogenic modified LDL typical for early stages of atherosclerosis, intimal pericytes can accumulate lipids and become activated. The development of atherosclerotic plaque is associated with the increase of total cell count and the percentage of pericytes indicative of their involvement into the pathological process<sup>[21]</sup>. In fatty streaks, these cells actively accumulate lipids, which leads to the increase of cell size, acquisition of irregular shape, loss of Cx43-mediated cell contacts<sup>[44]</sup> and disturbance of the net-like subendothelial tissue<sup>[45]</sup>. Pericytes may not only proliferate and store lipids contributing to the plaque growth<sup>[46,47]</sup>, but also promote thrombogenesis being a source of tissue factor<sup>[17]</sup>. Figure 3 depicts the impact of different arterial cells, including intimal pericytes, in the atherosclerotic process.

Microvascular pericytes are important for atherosclerosis progression, as they participate in neovascularization of the atherosclerotic plaques<sup>[48]</sup>. The recruitment of pericytes to growing neovessels could potentially be mediated by T-cadherin signalling. Tcadherin is an unusual member of the cadherin family, which is up-regulated in atherosclerosis<sup>[49]</sup>. It might play a role during LDL-induced pericyte activation through ERK1/2-nuclear factor  $\kappa$ B signalling pathway<sup>[50]</sup>. It has been demonstrated that pericytes could also be recruited to neovessels in atherosclerotic plaques through hepatocyte growth factor signalling triggering c-Met-PI3K/Akt pathway in pericytes<sup>[51]</sup>.

Participation in vascular wall remodelling and calcification is another possible contribution of pericytes to the pathological process in atherosclerosis. Activation of Wnt/ $\beta$ -catenin signalling stimulates chondrogenic

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differentiation of pericytes<sup>[52]</sup>, which can be enhanced by transforming growth factor- $\beta 3^{[53]}$ . The latter is abundantly produced by macrophages, foam cells and VSMCs in the atherosclerotic plaque<sup>[54]</sup>. Intimal pericytes were shown to express vascular calcification-associated factor. Moreover, pro-inflammatory environment in the plaque may promote osteogenic differentiation of the resident vascular cells. These observations indicate the potential involvement of pericytes in ectopic vascular calcification associated with atherosclerosis<sup>[55,56]</sup>.

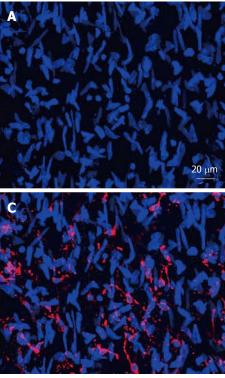
Taken together, the presented observations strongly support the hypothesis that macrovascular pericytes play an important role in all stages of the atherosclerotic lesion development, which makes them an attractive potential target for therapy development<sup>[7,57]</sup>. Interestingly, pericytes may also take part in the innate immunity reactions, which will be discussed in the next section.

## PERICYTE-LIKE CELLS IN INNATE

The development of atherosclerotic lesion is tightly associated with local inflammatory process. At the initial stages of atherosclerosis, formation of focal intimal thickening is accompanied by the increase of intimal cell count<sup>[58-60]</sup>. Such increase can be explained by the enhanced proliferation of intimal cells, recruitment of circulating hematologous cells or a combination of these processes. Cell proliferation has been demonstrated to be important for atherosclerotic process<sup>[61]</sup>, although the differences in cell composition depending on different types of large arteries remained unknown. Detailed study of cell count and proliferation was performed on 29 post-mortem specimens of human carotid and coronary arteries with different stages of atherosclerotic plaque development: diffuse intimal thickening (grossly normal tissue), initial lesions, fatty streaks, lipofibrous plaques and fibrous plaques<sup>[62]</sup>. Proliferating cells in S phase were identified by proliferating cell nuclear antigen (PCNA) staining. The authors report the increased total cell count in atherosclerotic lesions and more frequent macrophages and lymphocytes in comparison with normal tissue. Number of proliferating cells was also increased in atherosclerotic plaques as compared to normal intima. Maximal total cell count and maximal amount of proliferating cells were detected in lipofibrous plaques in both types of arteries. Interestingly, the proliferative index (determined as the percentage of PCNA-positive cells in the total subpopulation) hematogenous cells in atherosclerotic lesions was lower than in the unaffected intima. It is therefore likely that the increase of hematogenous cells in atherosclerotic intima occurs due to immune cell recruitment from the bloodstream and not from the local proliferation<sup>[63-65]</sup>. By contrast, proliferative index of resident cells was higher in atherosclerotic lesions than in unaffected tissue, reaching its maximum in lipofibrous plaques<sup>[62]</sup>.

It is well documented that modified atherogenic LDL

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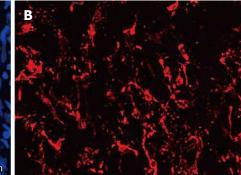


Figure 4 Immunofluorescent images representing the merge of 45 optic "confocal" sections of the intima obtained with interval of 1 µm between consecutive sections (A and B). A: Nuclei were visualized by staining with ethidium bromide; B: Distribution and networks formed by HLA-DR+ cells, detected with the use of anti-HLA-DR antibody conjugated with Alexa 633; C: A merge of the images shown in (A) and (B). Human aorta specimen. Reproduced from Bobryshev *et al*<sup>74</sup>, with permission from Elsevier. HLA: Human leukocyte antigen.

not only causes lipid deposition in the arterial intima, but also initiates pro-inflammatory conditions, stimulating both adaptive and innate immunity<sup>[66,67]</sup>. During the atherosclerotic plaque formation, inflammatory cells are migrating into the lesion zone, where they release proinflammatory cytokines and chemokines, inducing proliferation of the resident intimal cells<sup>[65,68]</sup>. This model may explain the observed divergence of inflammatory and resident cells proliferative index dynamics in atherosclerotic lesions. Study of immune-inflammatory processes in diffuse intimal thickening demonstrated the link between lipid deposition in the arterial wall and the immune-inflammatory cell content<sup>[69]</sup>. Lipid deposition in the proteoglycan-rich layer of intima positively correlated with the expression of major histocompatibility complex class  ${\rm I\hspace{-1.5pt}I}$  (MHC  ${\rm I\hspace{-1.5pt}I}$  ) molecule  $\mathsf{HLA}\text{-}\mathsf{DR}^{\scriptscriptstyle[70]}$  and the amount of immune-inflammatory cells in that area. These findings demonstrate that lipid accumulation at the early stages of atherosclerotic lesion development is accompanied by local immune activation. HLA-DR is a marker of antigen presenting cells (APCs)<sup>[70,71]</sup> that include macrophages, B cells and dendritic cells. The latter were described as professional APCs, as they play the most prominent role in antigen presentation and initiation of the immune response. Apart from these cell types of hematogenous origin, some non-hematogenous cells can serve as APCs. For instance, epithelial cells of the thymus are involved in antigen presentation<sup>[71]</sup>. Human endothelial cells are capable of T cell activation and basally express both MHCI and MHC  ${\rm II}$ , although the expression is less prominent in larger arteries<sup>[72]</sup>. Moreover, fibroblasts and endothelial cells can become activated to express MHC II under certain conditions both *in vitro* and *in vivo*<sup>[70,73]</sup>.

Immunofluorescent analysis of HLA-DR expression in the aortic intima revealed a large population of interconnected HLA-DR-positive cells that formed a netlike tissue (Figure 4)<sup>[74]</sup>. In some segments of the intima up to 15% of total cell population was expressing HLA-DR<sup>[74]</sup>. Morphologically, these cells could be identified as pericytes, with their typical net-forming intercellular contacts.

These results indicate that antigen presentation in the arterial wall can be performed by a variety of intimal cell types, including macrovascular pericytes. Importantly, microscopic study has revealed some HLA-DR-positive cells that also contained apo-B in the perinuclear space, possibly representing the early events of lipid accumulation in the intimal cells<sup>[74]</sup>. It is therefore possible that pericytes play a double role in atherogenesis, participating both in lipid accumulation and in local immune reaction.

Immunohistochemistry demonstrated the expression of tumor necrosis factor (TNF)- $\alpha$  and CCL18 cytokines in both normal and atherosclerotic human aorta. These two cytokines were expressed by distinct populations of cells, with CCL18 mostly located in the subendothelial layer, and TNF- $\alpha$  - in deeper layers of the unaffected intima. The expression pattern was altered in atherosclerotic plaques, with both TNF- $\alpha$  and CCL18 upregulated compared to grossly normal arterial intima. Apart from macrophages, other cytokine-producing cells types were detected, including stellate pericyte-like cells.

The immune functions of pericytes remain an attractive topic for future investigation<sup>[75]</sup>. Microvascular placental pericytes were demonstrated to interact with CD4<sup>+</sup> T cells in co-culture. Pericytes stimulated the expression of CD25 and CD69 in resting allogenic CD4<sup>+</sup> T cells, indicative of MHC recognition. This, however, did not lead to the induction of cytokine production and proliferation of T cells<sup>[76]</sup>. The possibility that macrovascular pericytes have immune-modulating properties remains to be explored. The three-dimensional network of subendothelial cells participating in immune reactions may play important protective functions in the blood vessel wall. As demonstrated on the biopsy material from arteries, unaffected by atherosclerosis or with different stages of atherosclerotic lesion development, changes in this network occur early in the pathological process.

## OTHER CELL TYPES PARTICIPATING IN IMMUNE REACTIONS IN THE SUBENDOTHELIAL INTIMA

Cells of hematogenous origin actively participate in immune reactions in the blood vessel wall. The development of atherosclerotic lesion is accompanied by infiltration and population of the lesion site by lymphocytes and macrophages. In human atherosclerotic lesions, the population of T cells consists mostly of the effector memory T cells. Substantial part of them are CD4<sup>+</sup> T helper cells<sup>[77]</sup>. CD8<sup>+</sup> cytotoxic T lymphocytes are also present in atherosclerotic lesions. T cells are especially numerous in so called vulnerable plaques that are susceptible to rupture and thrombosis initiation<sup>[78]</sup>. B cells have also been demonstrated to participate in atherogenesis<sup>[79,80]</sup>. However, these cells are mostly found in the adventitia of the affected vessels, and not in the atherosclerotic plaques<sup>[77,81]</sup>.

Pro-inflammatory stimuli in blood vessels affected by atherosclerosis attract circulating monocytes that rapidly migrate to the atherosclerotic lesion site and differentiate where, predominantly to macrophages. This polarization is driven by stimulation of specific receptors, including C-C chemokine receptor type 2 and CX3C chemokine receptor 1<sup>[82,83]</sup>. Both of these chemokines are implicated in the atherosclerotic plaque progression<sup>[84]</sup>. Macrophages play a prominent role in the inflammatory process associated with the initiation and progression of atherosclerotic lesions. Macrophages are involved in the uptake of atherogenic modified LDL and form typical foam cells. Toll-like receptor (TLR) signalling in macrophages plays important role in lipid accumulation through regulation of scavenger receptor expression and suppression of cholesterol efflux<sup>[85,86]</sup>. Macrophages also produce a number of inflammatory cytokines and contribute to vascular remodelling.

In atherosclerotic lesions, two distinct macrophage subpopulations can be found: M1 (classical) and M2 (alternative). Pro-inflammatory M1 phenotype can be induced in response to bacterial infection and pro-inflammatory cytokines, such as interferon (IFN)- $\gamma$  and TNF- $\alpha^{[87]}$ . They produce a spectrum of cytokines and chemokines stimulating the inflammatory res-

ponse, as well as nitric oxide and reactive oxygen species. M2 macrophages have anti-inflammatory properties, producing interleukin (IL)-10. In addition, M2 subpopulation of macrophages is heterogeneous: M2a macrophages are induced in response to IL-4 and IL-13 and participate in tissue remodelling, M2b cells are involved in immunoregulation, and M2c, induced by IL-10 and transforming growth factor- $\beta$ , are involved in clearance of apoptotic cells<sup>[88,89]</sup>. All three subtypes of M2 macrophages can be found in human atherosclerotic lesions. Human-specific M4 macrophages can be induced by platelet-derived chemokine CXCL4 and are characterized by resistance to excessive lipid uptake. This makes them susceptible to rapid transformation into foam cells and indicates their potential role in atherosclerotic lesion development<sup>[90]</sup>. Specific populations of macrophages can be found in hemorrhagic atherosclerotic plaques. In humans, they represent M(Hb), HA-mac and Mhem subsets. HA-mac and M(Hb) macrophages are involved in hemoglobine clearance<sup>[91]</sup>. All three subpopulations possess antiinflammatory properties and can play a protective role in atherosclerosis.

Dendritic cells are also frequent in the subendothelial layer of intima, just below the liminal endothelium, where they contribute up to 10% of total cell population<sup>[74,92,93]</sup>. In the subendothelial space, dendritic cells form cellular networks by means of their long interconnected processes<sup>[74,92,93]</sup>. Dendritic cells play an important role in the immune response, producing a spectrum of cytokines and surface co-stimulatory molecules. They can be differentiated from monocytes at the periphery through stimulation with granulocyte-macrophage colony-stimulating factor (GM-CSF) and TLR4 ligands<sup>[94]</sup>. These monocyte-derived dendritic cells are involved in antigen presentation and cross-presentation<sup>[95]</sup>. It has been demonstrated that monocyte-derived DCs from patients with atherosclerosis had increased sensitivity to GM-CSF and IL-4 as compared to healthy subjects<sup>[96]</sup>. Pro-inflammatory microenvironment of the atherosclerotic plaque accounts for preferential differentiation of monocytes and dendritic cell precursors towards pro-inflammatory dendritic cells that play an important role in the inflammatory process and stimulate differentiation of the naïve T cells to Th1 and Th17 phenotypes<sup>[97]</sup>. Dendritic cells also produce pro-inflammatory IFN- $\alpha$  and a number of chemokines that induce the migration of inflammatory cells to the lesion site<sup>[98]</sup>. In summary, pro-inflammatory dendritic cells play a prominent role in the development of atherosclerotic plaque. Populations of atheroprotective tolerogenic and anti-inflammatory dendritic cells could also be found in atherosclerotic plaques. In Apo-Edeficient mice, tolerogenic dendritic cells generated in response to immunogenic bacterial peptides were demonstrated to decrease inflammation and contribute to the plaque decrease through induction of IL-10producing Tregs<sup>[99]</sup>. It is possible that employing the

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pathways of tolerogenic dendritic cell induction can be beneficial for treatment of atherosclerosis.

#### CONCLUSION

The subendothelial layer of human arterial intima contains a cellular network made of pluripotent pericytes that form multiple intracellular contacts through gap junctions. Together with the professional immune cells, such as macrophages and dendritic cells, pericytes may execute functions, typical for innate immune cell types: participate in LDL trapping by phagocytosis, produce pro- and anti-inflammatory cytokines and other factors and present antigens (Figure 2). Whereas the aortic endothelium plays a role as the first line of defence in atherosclerosis development, the subendothelial pericytes can be regarded as the second defence line and should be considered for the development therapeutic approaches for atherosclerosis treatment.

#### REFERENCES

- Mozaffarian D, Benjamin EJ, Go AS, Arnett DK, Blaha MJ, Cushman M, de Ferranti S, Després JP, Fullerton HJ, Howard VJ, Huffman MD, Judd SE, Kissela BM, Lackland DT, Lichtman JH, Lisabeth LD, Liu S, Mackey RH, Matchar DB, McGuire DK, Mohler ER, Moy CS, Muntner P, Mussolino ME, Nasir K, Neumar RW, Nichol G, Palaniappan L, Pandey DK, Reeves MJ, Rodriguez CJ, Sorlie PD, Stein J, Towfighi A, Turan TN, Virani SS, Willey JZ, Woo D, Yeh RW, Turner MB; American Heart Association Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics--2015 update: a report from the American Heart Association. *Circulation* 2015; **131**: e29-322 [PMID: 25520374 DOI: 10.1161/CIR.000000000000152]
- 2 Hellings WE, Peeters W, Moll FL, Pasterkamp G. From vulnerable plaque to vulnerable patient: the search for biomarkers of plaque destabilization. *Trends Cardiovasc Med* 2007; 17: 162-171 [PMID: 17574124 DOI: 10.1016/j.tcm.2007.03.006]
- 3 de Weerd M, Greving JP, Hedblad B, Lorenz MW, Mathiesen EB, O'Leary DH, Rosvall M, Sitzer M, Buskens E, Bots ML. Prevalence of asymptomatic carotid artery stenosis in the general population: an individual participant data meta-analysis. *Stroke* 2010; **41**: 1294-1297 [PMID: 20431077 DOI: 10.1161/STROKEAHA.110.581058]
- 4 McGill HC, Herderick EE, McMahan CA, Zieske AW, Malcolm GT, Tracy RE, Strong JP. Atherosclerosis in youth. *Minerva Pediatr* 2002; 54: 437-447 [PMID: 12244281]
- 5 Tuzcu EM, Kapadia SR, Tutar E, Ziada KM, Hobbs RE, McCarthy PM, Young JB, Nissen SE. High prevalence of coronary atherosclerosis in asymptomatic teenagers and young adults: evidence from intravascular ultrasound. *Circulation* 2001; 103: 2705-2710 [PMID: 11390341 DOI: 10.1161/01.CIR.103.22.2705]
- 6 Andreeva ER, Pugach IM, Gordon D, Orekhov AN. Continuous subendothelial network formed by pericyte-like cells in human vascular bed. *Tissue Cell* 1998; 30: 127-135 [PMID: 9569686 DOI: 10.1016/S0040-8166(98)80014-1]
- 7 Juchem G, Weiss DR, Gansera B, Kemkes BM, Mueller-Hoecker J, Nees S. Pericytes in the macrovascular intima: possible physiological and pathogenetic impact. *Am J Physiol Heart Circ Physiol* 2010; **298**: H754-H770 [PMID: 20023125 DOI: 10.1152/ ajpheart.00343.2009]
- 8 Gross L, Epstein EZ, Kugel MA. Histology of the Coronary Arteries and their Branches in the Human Heart. *Am J Pathol* 1934; 10: 253-274.7 [PMID: 19970139]
- 9 Movat HZ, More RH, Haust MD. The diffuse intimal thickening of the human aorta with aging. *Am J Pathol* 1958; 34: 1023-1031

[PMID: 13583094]

- 10 Geer JC, Haust MD. Smooth muscle cells in atherosclerosis. Monogr Atheroscler 1972; 2: 1-140 [PMID: 4600684]
- Haust MD. Recent concepts on the pathogenesis of atherosclerosis. CMAJ 1989; 140: 929 [PMID: 2702530]
- 12 Stary HC. Evolution and progression of atherosclerotic lesions in coronary arteries of children and young adults. *Arteriosclerosis* 1989; 9: 119-132 [PMID: 2912430]
- 13 Velican D, Velican C. Histochemical study on the glycosaminoglycans (acid mucopolysaccharides) of the human coronary arteries. *Acta Histochem* 1977; **59**: 190-200 [PMID: 412386 DOI: 10.1016/ S0065-1281(77)80039-1]
- 14 van Dijk CG, Nieuweboer FE, Pei JY, Xu YJ, Burgisser P, van Mulligen E, el Azzouzi H, Duncker DJ, Verhaar MC, Cheng C. The complex mural cell: pericyte function in health and disease. *Int J Cardiol* 2015; 190: 75-89 [PMID: 25918055 DOI: 10.1016/ j.ijcard.2015.03.258]
- 15 Hall AP. Review of the pericyte during angiogenesis and its role in cancer and diabetic retinopathy. *Toxicol Pathol* 2006; 34: 763-775 [PMID: 17162534]
- 16 Nicosia RF. The aortic ring model of angiogenesis: a quarter century of search and discovery. *J Cell Mol Med* 2009; 13: 4113-4136 [PMID: 19725916 DOI: 10.1111/j.1582-4934.2009.00891.x]
- 17 Orekhov AN, Bobryshev YV, Chistiakov DA. The complexity of cell composition of the intima of large arteries: focus on pericyte-like cells. *Cardiovasc Res* 2014; 103: 438-451 [PMID: 25016615 DOI: 10.1093/cvr/cvu168]
- 18 Campagnolo P, Cesselli D, Al Haj Zen A, Beltrami AP, Kränkel N, Katare R, Angelini G, Emanueli C, Madeddu P. Human adult vena saphena contains perivascular progenitor cells endowed with clonogenic and proangiogenic potential. *Circulation* 2010; **121**: 1735-1745 [PMID: 20368523 DOI: 10.1161/CIRCULATIONAHA.109.899252]
- 19 Rekhter MD, Andreeva ER, Andrianova IV, Mironov AA, Orekhov AN. Stellate cells of aortic intima: I. Human and rabbit. *Tissue Cell* 1992; 24: 689-696 [PMID: 1440588]
- 20 Krushinsky AV, Orekhov AN, Smirnov VN. Stellate cells in the intima of human aorta. Application of alkaline dissociation method in the analysis of the vessel wall cellular content. *Acta Anat* (Basel) 1983; 117: 266-269 [PMID: 6359799 DOI: 10.1159/000145797]
- 21 Orekhov AN, Karpova II, Tertov VV, Rudchenko SA, Andreeva ER, Krushinsky AV, Smirnov VN. Cellular composition of atherosclerotic and uninvolved human aortic subendothelial intima. Light-microscopic study of dissociated aortic cells. *Am J Pathol* 1984; 115: 17-24 [PMID: 6711678]
- 22 Dore-Duffy P, Wang S, Mehedi A, Katyshev V, Cleary K, Tapper A, Reynolds C, Ding Y, Zhan P, Rafols J, Kreipke CW. Pericyte-mediated vasoconstriction underlies TBI-induced hypoperfusion. *Neurol Res* 2011; 33: 176-186 [PMID: 21801592 DOI: 10.1179/016 164111X12881719352372]
- 23 Boado RJ, Pardridge WM. Differential expression of alpha-actin mRNA and immunoreactive protein in brain microvascular pericytes and smooth muscle cells. *J Neurosci Res* 1994; **39**: 430-435 [PMID: 7884822 DOI: 10.1002/jnr.490390410]
- Hughes S, Gardiner T, Hu P, Baxter L, Rosinova E, Chan-Ling T. Altered pericyte-endothelial relations in the rat retina during aging: implications for vessel stability. *Neurobiol Aging* 2006; 27: 1838-1847 [PMID: 16387390 DOI: 10.1016/j.neurobiolaging.2005. 10.021]
- 25 Glukhova MA, Kabakov AE, Frid MG, Ornatsky OI, Belkin AM, Mukhin DN, Orekhov AN, Koteliansky VE, Smirnov VN. Modulation of human aorta smooth muscle cell phenotype: a study of muscle-specific variants of vinculin, caldesmon, and actin expression. *Proc Natl Acad Sci USA* 1988; **85**: 9542-9546 [PMID: 3143999 DOI: 10.1073/pnas.85.24.9542]
- 26 Andreeva ER, Pugach IM, Orekhov AN. Subendothelial smooth muscle cells of human aorta express macrophage antigen in situ and in vitro. *Atherosclerosis* 1997; 135: 19-27 [PMID: 9395269 DOI: 10.1016/S0021-9150(97)00136-6]
- 27 **Greaves DR**, Gordon S. Macrophage-specific gene expression: current paradigms and future challenges. *Int J Hematol* 2002; **76**: 6-15



[PMID: 12138897 DOI: 10.1007/BF02982713]

- 28 Song L, Lee C, Schindler C. Deletion of the murine scavenger receptor CD68. *J Lipid Res* 2011; **52**: 1542-1550 [PMID: 21572087 DOI: 10.1194/jlr.M015412]
- 29 Davies PF. Vascular cell interactions with special reference to the pathogenesis of atherosclerosis. *Lab Invest* 1986; 55: 5-24 [PMID: 3014215]
- 30 Armulik A, Abramsson A, Betsholtz C. Endothelial/pericyte interactions. *Circ Res* 2005; 97: 512-523 [PMID: 16166562 DOI: 10.1161/01.RES.0000182903.16652.d7]
- 31 Nayak RC, Berman AB, George KL, Eisenbarth GS, King GL. A monoclonal antibody (3G5)-defined ganglioside antigen is expressed on the cell surface of microvascular pericytes. *J Exp Med* 1988; 167: 1003-1015 [PMID: 3351433 DOI: 10.1084/jem.167.3.1003]
- 32 Boström K, Watson KE, Horn S, Wortham C, Herman IM, Demer LL. Bone morphogenetic protein expression in human atherosclerotic lesions. *J Clin Invest* 1993; 91: 1800-1809 [PMID: 8473518 DOI: 10.1172/JCI116391]
- 33 Andreeva ER, Rekhter MD, Romanov YuA GM, Antonov AS, Mironov AA, Orekhov AN. Stellate cells of aortic intima: II. Arborization of intimal cells in culture. *Tissue Cell* 1992; 24: 697-704 [PMID: 1332216 DOI: 10.1016/0040-8166(92)90040-E]
- 34 Aqel NM, Ball RY, Waldmann H, Mitchinson MJ. Identification of macrophages and smooth muscle cells in human atherosclerosis using monoclonal antibodies. *J Pathol* 1985; 146: 197-204 [PMID: 3897495 DOI: 10.1002/path.1711460306]
- 35 Munro JM, van der Walt JD, Munro CS, Chalmers JA, Cox EL. An immunohistochemical analysis of human aortic fatty streaks. *Hum Pathol* 1987; 18: 375-380 [PMID: 3549534 DOI: 10.1016/ S0046-8177(87)80168-5]
- 36 Roessner A, Herrera A, Höning HJ, Vollmer E, Zwadlo G, Schürmann R, Sorg C, Grundmann E. Identification of macrophages and smooth muscle cells with monoclonal antibodies in the human atherosclerotic plaque. *Virchows Arch A Pathol Anat Histopathol* 1987; 412: 169-174 [PMID: 3122417 DOI: 10.1007/BF00716190]
- 37 van der Wal AC, Das PK, Bentz van de Berg D, van der Loos CM, Becker AE. Atherosclerotic lesions in humans. In situ immunophenotypic analysis suggesting an immune mediated response. *Lab Invest* 1989; 61: 166-170 [PMID: 2787872]
- 38 Hansson GK, Holm J, Jonasson L. Detection of activated T lymphocytes in the human atherosclerotic plaque. *Am J Pathol* 1989; 135: 169-175 [PMID: 2505620]
- 39 Kaartinen M, Penttilä A, Kovanen PT. Mast cells of two types differing in neutral protease composition in the human aortic intima. Demonstration of tryptase- and tryptase/chymase-containing mast cells in normal intimas, fatty streaks, and the shoulder region of atheromas. *Arterioscler Thromb* 1994; 14: 966-972 [PMID: 7515278 DOI: 10.1161/01.ATV.14.6.966]
- 40 Sims DE. Recent advances in pericyte biology--implications for health and disease. *Can J Cardiol* 1991; 7: 431-443 [PMID: 1768982]
- 41 Diaz-Flores L, Gutierrez R, Lopez-Alonso A, Gonzalez R, Varela H. Pericytes as a supplementary source of osteoblasts in periosteal osteogenesis. *Clin Orthop Relat Res* 1992; (275): 280-286 [PMID: 1735226 DOI: 10.1097/00003086-199202000-00042]
- 42 Brighton CT, Lorich DG, Kupcha R, Reilly TM, Jones AR, Woodbury RA. The pericyte as a possible osteoblast progenitor cell. *Clin Orthop Relat Res* 1992; (275): 287-299 [PMID: 1735227 DOI: 10.1097/00003086-199202000-00043]
- 43 Iyama K, Ohzono K, Usuku G. Electron microscopical studies on the genesis of white adipocytes: differentiation of immature pericytes into adipocytes in transplanted preadipose tissue. *Virchows Arch B Cell Pathol Incl Mol Pathol* 1979; **31**: 143-155 [PMID: 42211 DOI: 10.1007/BF02889932]
- 44 Andreeva ER, Serebryakov VN, Orekhov AN. Gap junctional communication in primary culture of cells derived from human aortic intima. *Tissue Cell* 1995; 27: 591-597 [PMID: 7491628 DOI: 10.1016/S0040-8166(05)80069-2]
- 45 Andreeva ER, Orekhov AN, Smirnov VN. Quantitative estimation of lipid-laden cells in atherosclerotic lesions of the human aorta.

*Acta Anat* (Basel) 1991; **141**: 316-323 [PMID: 1660667 DOI: 10.1159/000147142]

- 46 Orekhov AN, Andreeva ER, Krushinsky AV, Novikov ID, Tertov VV, Nestaiko GV, Khashimov KhA, Repin VS, Smirnov VN. Intimal cells and atherosclerosis. Relationship between the number of intimal cells and major manifestations of atherosclerosis in the human aorta. *Am J Pathol* 1986; 125: 402-415 [PMID: 3789095]
- 47 Rekhter MD, Andreeva ER, Mironov AA, Orekhov AN. Threedimensional cytoarchitecture of normal and atherosclerotic intima of human aorta. *Am J Pathol* 1991; 138: 569-580 [PMID: 2000936]
- 48 Moreno PR, Purushothaman M, Purushothaman KR. Plaque neovascularization: defense mechanisms, betrayal, or a war in progress. *Ann N Y Acad Sci* 2012; **1254**: 7-17 [PMID: 22548565 DOI: 10.1111/j.1749-6632.2012.06497.x]
- 49 Ivanov D, Philippova M, Antropova J, Gubaeva F, Iljinskaya O, Tararak E, Bochkov V, Erne P, Resink T, Tkachuk V. Expression of cell adhesion molecule T-cadherin in the human vasculature. *Histochem Cell Biol* 2001; 115: 231-242 [PMID: 11326751]
- 50 Takeuchi T, Ohtsuki Y. Recent progress in T-cadherin (CDH13, H-cadherin) research. *Histol Histopathol* 2001; 16: 1287-1293 [PMID: 11642747]
- 51 Liu Y, Wilkinson FL, Kirton JP, Jeziorska M, Iizasa H, Sai Y, Nakashima E, Heagerty AM, Canfield AE, Alexander MY. Hepatocyte growth factor and c-Met expression in pericytes: implications for atherosclerotic plaque development. *J Pathol* 2007; 212: 12-19 [PMID: 17405187 DOI: 10.1002/path.2155]
- 52 Kirton JP, Crofts NJ, George SJ, Brennan K, Canfield AE. Wnt/beta-catenin signaling stimulates chondrogenic and inhibits adipogenic differentiation of pericytes: potential relevance to vascular disease? *Circ Res* 2007; **101**: 581-589 [PMID: 17673669 DOI: 10.1161/CIRCRESAHA.107.156372]
- 53 Farrington-Rock C, Crofts NJ, Doherty MJ, Ashton BA, Griffin-Jones C, Canfield AE. Chondrogenic and adipogenic potential of microvascular pericytes. *Circulation* 2004; 110: 2226-2232 [PMID: 15466630 DOI: 10.1161/01.CIR.0000144457.55518.E5]
- 54 Bobik A, Agrotis A, Kanellakis P, Dilley R, Krushinsky A, Smirnov V, Tararak E, Condron M, Kostolias G. Distinct patterns of transforming growth factor-beta isoform and receptor expression in human atherosclerotic lesions. Colocalization implicates TGF-beta in fibrofatty lesion development. *Circulation* 1999; **99**: 2883-2891 [PMID: 10359732 DOI: 10.1161/01.CIR.99.22.2883]
- 55 Wilkinson FL, Liu Y, Rucka AK, Jeziorska M, Hoyland JA, Heagerty AM, Canfield AE, Alexander MY. Contribution of VCAF-positive cells to neovascularization and calcification in atherosclerotic plaque development. *J Pathol* 2007; **211**: 362-369 [PMID: 17154367 DOI: 10.1002/path.2114]
- 56 Collett GD, Canfield AE. Angiogenesis and pericytes in the initiation of ectopic calcification. *Circ Res* 2005; 96: 930-938 [PMID: 15890980 DOI: 10.1161/01.RES.0000163634.51301.0d]
- Kelly-Goss MR, Sweat RS, Stapor PC, Peirce SM, Murfee WL. Targeting pericytes for angiogenic therapies. *Microcirculation* 2014; 21: 345-357 [PMID: 24267154 DOI: 10.1111/micc.12107]
- 58 Andreeva ER, Pugach IM, Orekhov AN. Collagen-synthesizing cells in initial and advanced atherosclerotic lesions of human aorta. *Atherosclerosis* 1997; 130: 133-142 [PMID: 9126657 DOI: 10.1016/ S0021-9150(96)06056-X]
- 59 Orekhov AN, Andreeva ER, Mikhailova IA, Gordon D. Cell proliferation in normal and atherosclerotic human aorta: proliferative splash in lipid-rich lesions. *Atherosclerosis* 1998; **139**: 41-48 [PMID: 9699890 DOI: 10.1016/S0021-9150(98)00044-6]
- 60 Babaev VR, Bobryshev YV, Sukhova GK, Kasantseva IA. Monocyte/macrophage accumulation and smooth muscle cell phenotypes in early atherosclerotic lesions of human aorta. *Atherosclerosis* 1993; 100: 237-248 [PMID: 8357356 DOI: 10.1016/ 0021-9150(93)90210-L]
- 61 Ross R. Atherosclerosis--an inflammatory disease. N Engl J Med 1999; 340: 115-126 [PMID: 9887164 DOI: 10.1056/ NEJM199901143400207]
- 62 **Orekhov AN**, Andreeva ER, Andrianova IV, Bobryshev YV. Peculiarities of cell composition and cell proliferation in different

type atherosclerotic lesions in carotid and coronary arteries. *Atherosclerosis* 2010; **212**: 436-443 [PMID: 20692661 DOI: 10.1016/j.atherosclerosis.2010.07.009]

- 63 Galkina E, Ley K. Leukocyte influx in atherosclerosis. *Curr Drug Targets* 2007; 8: 1239-1248 [PMID: 18220701 DOI: 10.2174/13894 5007783220650]
- 64 Bobryshev YV. Monocyte recruitment and foam cell formation in atherosclerosis. *Micron* 2006; 37: 208-222 [PMID: 16360317 DOI: 10.1016/j.micron.2005.10.007]
- 65 Galkina E, Ley K. Immune and inflammatory mechanisms of atherosclerosis (\*). *Annu Rev Immunol* 2009; 27: 165-197 [PMID: 19302038 DOI: 10.1146/annurev.immunol.021908.132620]
- 66 Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. N Engl J Med 2005; 352: 1685-1695 [PMID: 15843671 DOI: 10.1056/NEJMra043430]
- 67 Hartvigsen K, Chou MY, Hansen LF, Shaw PX, Tsimikas S, Binder CJ, Witztum JL. The role of innate immunity in atherogenesis. J Lipid Res 2009; 50 Suppl: S388-S393 [PMID: 19106070 DOI: 10.1194/jlr.R800100-JLR200]
- 68 Shimada K. Immune system and atherosclerotic disease: heterogeneity of leukocyte subsets participating in the pathogenesis of atherosclerosis. *Circ J* 2009; 73: 994-1001 [PMID: 19430164 DOI: 10.1253/circj.CJ-09-0277]
- 69 Bobryshev YV, Andreeva ER, Mikhailova IA, Andrianova IV, Moisenovich MM, Khapchaev S, Agapov II, Sobenin IA, Lusta KA, Orekhov AN. Correlation between lipid deposition, immuneinflammatory cell content and MHC class II expression in diffuse intimal thickening of the human aorta. *Atherosclerosis* 2011; 219: 1711-183 [PMID: 21831373 DOI: 10.1016/j.atherosclerosis.2011.07. 016]
- 70 Handunnetthi L, Ramagopalan SV, Ebers GC, Knight JC. Regulation of major histocompatibility complex class II gene expression, genetic variation and disease. *Genes Immun* 2010; 11: 99-112 [PMID: 19890353 DOI: 10.1038/gene.2009.83]
- 71 Geissmann F, Manz MG, Jung S, Sieweke MH, Merad M, Ley K. Development of monocytes, macrophages, and dendritic cells. *Science* 2010; **327**: 656-661 [PMID: 20133564 DOI: 10.1126/ science.1178331]
- 72 von Willebrand E, Lautenschlager I, Inkinen K, Lehto VP, Virtanen I, Häyry P. Distribution of the major histocompatibility complex antigens in human and rat kidney. *Kidney Int* 1985; 27: 616-621 [PMID: 3892132]
- 73 Nielsen M, Lund O, Buus S, Lundegaard C. MHC class II epitope predictive algorithms. *Immunology* 2010; 130: 319-328 [PMID: 20408898 DOI: 10.1111/j.1365-2567.2010.03268.x]
- 74 Bobryshev YV, Moisenovich MM, Pustovalova OL, Agapov II, Orekhov AN. Widespread distribution of HLA-DR-expressing cells in macroscopically undiseased intima of the human aorta: a possible role in surveillance and maintenance of vascular homeostasis. *Immunobiology* 2012; 217: 558-568 [PMID: 21601938 DOI: 10.1016/j.imbio.2011.03.014]
- 75 Pober JS, Tellides G. Participation of blood vessel cells in human adaptive immune responses. *Trends Immunol* 2012; 33: 49-57 [PMID: 22030237 DOI: 10.1016/j.it.2011.09.006]
- 76 Maier CL, Pober JS. Human placental pericytes poorly stimulate and actively regulate allogeneic CD4 T cell responses. *Arterioscler Thromb Vasc Biol* 2011; 31: 183-189 [PMID: 21051666 DOI: 10.1161/ATVBAHA.110.217117]
- 77 Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol* 2011; 12: 204-212 [PMID: 21321594 DOI: 10.1038/ ni.2001]
- 78 Arbab-Zadeh A, Nakano M, Virmani R, Fuster V. Acute coronary events. *Circulation* 2012; 125: 1147-1156 [PMID: 22392862 DOI: 10.1161/CIRCULATIONAHA.111.047431]
- 79 Caligiuri G, Paulsson G, Nicoletti A, Maseri A, Hansson GK. Evidence for antigen-driven T-cell response in unstable angina. *Circulation* 2000; 102: 1114-1119 [PMID: 10973839]
- 80 Huan T, Zhang B, Wang Z, Joehanes R, Zhu J, Johnson AD, Ying S, Munson PJ, Raghavachari N, Wang R, Liu P, Courchesne P, Hwang SJ, Assimes TL, McPherson R, Samani NJ, Schunkert H, Meng Q,

Suver C, O'Donnell CJ, Derry J, Yang X, Levy D. A systems biology framework identifies molecular underpinnings of coronary heart disease. *Arterioscler Thromb Vasc Biol* 2013; **33**: 1427-1434 [PMID: 23539213 DOI: 10.1161/ATVBAHA.112.300112]

- 81 Canducci F, Saita D, Foglieni C, Piscopiello MR, Chiesa R, Colombo A, Cianflone D, Maseri A, Clementi M, Burioni R. Crossreacting antibacterial auto-antibodies are produced within coronary atherosclerotic plaques of acute coronary syndrome patients. *PLoS One* 2012; 7: e42283 [PMID: 22879930 DOI: 10.1371/journal. pone.0042283]
- 82 Lauvau G, Chorro L, Spaulding E, Soudja SM. Inflammatory monocyte effector mechanisms. *Cell Immunol* 2014; 291: 32-40 [PMID: 25205002 DOI: 10.1016/j.cellimm.2014.07.007]
- 83 Fong AM, Robinson LA, Steeber DA, Tedder TF, Yoshie O, Imai T, Patel DD. Fractalkine and CX3CR1 mediate a novel mechanism of leukocyte capture, firm adhesion, and activation under physiologic flow. *J Exp Med* 1998; 188: 1413-1419 [PMID: 9782118 DOI: 10.1084/jem.188.8.1413]
- 84 Tacke F, Alvarez D, Kaplan TJ, Jakubzick C, Spanbroek R, Llodra J, Garin A, Liu J, Mack M, van Rooijen N, Lira SA, Habenicht AJ, Randolph GJ. Monocyte subsets differentially employ CCR2, CCR5, and CX3CR1 to accumulate within atherosclerotic plaques. *J Clin Invest* 2007; 117: 185-194 [PMID: 17200718]
- 85 Cole JE, Georgiou E, Monaco C. The expression and functions of toll-like receptors in atherosclerosis. *Mediators Inflamm* 2010; 2010: 393946 [PMID: 20652007 DOI: 10.1155/2010/393946]
- 86 Seneviratne AN, Sivagurunathan B, Monaco C. Toll-like receptors and macrophage activation in atherosclerosis. *Clin Chim Acta* 2012; 413: 3-14 [PMID: 21884686 DOI: 10.1016/j.cca.2011.08.021]
- 87 Zhang S, Kim CC, Batra S, McKerrow JH, Loke P. Delineation of diverse macrophage activation programs in response to intracellular parasites and cytokines. *PLoS Negl Trop Dis* 2010; 4: e648 [PMID: 20361029 DOI: 10.1371/journal.pntd.0000648]
- 88 Martinez FO, Sica A, Mantovani A, Locati M. Macrophage activation and polarization. *Front Biosci* 2008; 13: 453-461 [PMID: 17981560]
- 89 Zizzo G, Hilliard BA, Monestier M, Cohen PL. Efficient clearance of early apoptotic cells by human macrophages requires M2c polarization and MerTK induction. *J Immunol* 2012; 189: 3508-3520 [PMID: 22942426]
- 90 Gleissner CA. Macrophage Phenotype Modulation by CXCL4 in Atherosclerosis. *Front Physiol* 2012; 3: 1 [PMID: 22275902 DOI: 10.3389/fphys.2012.00001]
- 91 Chinetti-Gbaguidi G, Colin S, Staels B. Macrophage subsets in atherosclerosis. *Nat Rev Cardiol* 2015; 12: 10-17 [PMID: 25367649 DOI: 10.1038/nrcardio.2014.173]
- 92 Bobryshev YV, Lord RS. Ultrastructural recognition of cells with dendritic cell morphology in human aortic intima. Contacting interactions of Vascular Dendritic Cells in athero-resistant and athero-prone areas of the normal aorta. *Arch Histol Cytol* 1995; 58: 307-322 [PMID: 8527238 DOI: 10.1679/aohc.58.307]
- 93 Millonig G, Niederegger H, Rabl W, Hochleitner BW, Hoefer D, Romani N, Wick G. Network of vascular-associated dendritic cells in intima of healthy young individuals. *Arterioscler Thromb Vasc Biol* 2001; 21: 503-508 [PMID: 11304464 DOI: 10.1161/01. ATV.21.4.503]
- 94 Randolph GJ, Ochando J, Partida-Sánchez S. Migration of dendritic cell subsets and their precursors. *Annu Rev Immunol* 2008; 26: 293-316 [PMID: 18045026 DOI: 10.1146/annurev. immunol.26.021607.090254]
- 95 Legein B, Temmerman L, Biessen EA, Lutgens E. Inflammation and immune system interactions in atherosclerosis. *Cell Mol Life Sci* 2013; **70**: 3847-3869 [PMID: 23430000 DOI: 10.1007/ s00018-013-1289-1]
- 96 Dopheide JF, Sester U, Schlitt A, Horstick G, Rupprecht HJ, Münzel T, Blankenberg S. Monocyte-derived dendritic cells of patients with coronary artery disease show an increased expression of costimulatory molecules CD40, CD80 and CD86 in vitro. *Coron Artery Dis* 2007; 18: 523-531 [PMID: 17925605]
- 97 Ait-Oufella H, Sage AP, Mallat Z, Tedgui A. Adaptive (T and B

cells) immunity and control by dendritic cells in atherosclerosis. *Circ Res* 2014; **114**: 1640-1660 [PMID: 24812352 DOI: 10.1161/ CIRCRESAHA.114.302761]

98 Sozzani S, Vermi W, Del Prete A, Facchetti F. Trafficking properties of plasmacytoid dendritic cells in health and disease. *Trends Immunol* 2010; 31: 270-277 [PMID: 20579936 DOI: 10.1016/j.it.2010.05.004] Ivanova EA et al. Intimal pericytes and atherosclerosis

- 99 Ovchinnikova OA, Berge N, Kang C, Urien C, Ketelhuth DF, Pottier J, Drouet L, Hansson GK, Marchal G, Bäck M, Schwartz-Cornil I, Lagranderie M. Mycobacterium bovis BCG killed by extended freeze-drying induces an immunoregulatory profile and protects against atherosclerosis. *J Intern Med* 2014; 275: 49-58 [PMID: 23962000 DOI: 10.1111/joim.12127]
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