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A Multibiomarker-Based Outcome Risk Stratification Model for Adult Septic Shock

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Abstract

Objectives—Clinical trials in septic shock continue to fail due, in part, to inequitable and sometimes unknown distribution of baseline mortality risk between study arms. Investigators advocate that interventional trials in septic shock require effective outcome risk stratification. We derived and tested a multibiomarker-based approach to estimate mortality risk in adults with septic shock.

Design—Previous genome-wide expression studies identified 12 plasma proteins as candidates for biomarker-based risk stratification. The current analysis used banked plasma samples and clinical data from existing studies. Biomarkers were assayed in plasma samples obtained from 341 subjects with septic shock within 24 hours of admission to the ICU. Classification and regression tree analysis was used to generate a decision tree predicting 28-day mortality based on a combination of both biomarkers and clinical variables. The derived tree was first tested in an independent cohort of 331 subjects, then calibrated using all subjects (n = 672), and subsequently validated in another independent cohort (n = 209).

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The remaining authors have disclosed that they do not have any potential conflicts of interest.

Setting—Multiple ICUs in Canada, Finland, and the United States.

Subjects—Eight hundred eighty-one adults with septic shock or severe sepsis.

Intervention-None.

Measurements and Main Results—The derived decision tree included five candidate biomarkers, admission lactate concentration, age, and chronic disease burden. In the derivation cohort, sensitivity for mortality was 94% (95% CI, 87–97), specificity was 56% (50–63), positive predictive value was 50% (43–57), and negative predictive value was 95% (89–98). Performance was comparable in the test cohort. The calibrated decision tree had the following test characteristics in the validation cohort: sensitivity 85% (76–92), specificity 60% (51–69), positive predictive value 61% (52–70), and negative predictive value 85% (75–91).

Conclusions—We have derived, tested, calibrated, and validated a risk stratification tool and found that it reliably estimates the probability of mortality in adults with septic shock.

Keywords

biomarkers; decision tree; modeling; outcome; sepsis; stratification

Substantial resources are invested in trying to find new treatments for septic shock, but the vast majority of new treatments fail to demonstrate efficacy during phase 3 trials. There are many reasons for these failures, but one consistent reason is that baseline mortality risk varies widely in patients with septic shock (1). Consequently, current trial designs lead to the inclusion of patients with low mortality risk who are unlikely to benefit from novel therapies beyond standard care as well as patients having an extremely high mortality risk who may be beyond salvage by experimental therapy. This approach can dilute any potential beneficial effect of an experimental therapy for patients with a significant, but modifiable, mortality risk and consequently lead to a negative clinical trial.

Researchers advocate that interventional trials in sepsis must be conducted in the context of effective outcome risk stratification (1-3). A recent trial used a mortality risk scoring system to screen for patients with a high, but potentially modifiable, risk of mortality. The trial failed to demonstrate efficacy, in part, because the actual mortality in the placebo group was far lower than predicted by the scoring system (4). This highlights the importance of developing stratification tools for septic shock to better inform the conduct of clinical trials.

We recently derived and validated a biomarker-based model that reliably predicts 28-day mortality in pediatric septic shock (5). The plasma protein biomarkers were selected objectively based on extensive genome-wide expression studies and predictive modeling (6, 7). Furthermore, the biomarkers were measured from samples obtained during the first 24 hours of admission to the ICU, which is during the time when patients are typically evaluated for inclusion in clinical trials. We hypothesized that the biomarkers used in the pediatric study could also be used to estimate a mortality probability in adults with septic shock. In the current study, we extend our methodology to derive, test, and validate an analogous model in adults with septic shock.

METHODS

Overall Study Design

All analyses of plasma samples and clinical data are based on secondary use of existing data from previous studies, with approval of the respective institutional review boards.

Derivation Cohort

Derivation cohort study subjects (n = 341) were participants in the Vasopressin and Septic Shock Trial (VASST), a randomized, concealed, norepinephrine-controlled trial testing the efficacy of low-dose vasopressin versus norepinephrine in adults with septic shock (Current Controlled Trials number: ISRCTN9485869). The original VASST publication describes all protocol details (8).

Test Cohort

Test cohort study subjects (n = 331) were pooled from two sources. Two hundred and fortythree subjects were participants in a prospective, observational, multicenter cohort study of prevalence and outcome of severe sepsis and septic shock in Finland (FINNSEPSIS) (9). An additional 88 subjects were participants in a single center, observational study at St. Paul's Hospital in Vancouver, British Columbia (10).

Validation Cohort

Validation cohort study subjects (n = 209) were participants in the Molecular Epidemiology of Severe Sepsis in the Intensive Care Unit study, an ongoing cohort study at the Hospital of the University of Pennsylvania. Eligible patients with septic shock were enrolled in either the emergency department or the medical ICU, and patients or their proxies provided informed consent. Septic shock was defined using published criteria (11).

Candidate Stratification Biomarkers

The 12 candidate biomarkers (gene symbols) included C-C chemokine ligand 3 (CCL3), C-C chemokine ligand 4 (CCL4), neutrophil elastase 2 (ELA2), granzyme B (GZMB), heat shock protein 70 kDa 1B (HSPA1B), interleukin-1a (IL1A), interleukin-8 (IL8), lipocalin 2 (LCN2), lactotransferrin (LTF), matrix metallopeptidase 8 (MMP8), resistin (RETN), and thrombospondin 1 (THBS1). These biomarkers were selected from 117 gene probes previously shown to have predictive strength for poor outcomes in microarray-based studies involving children with septic shock (6, 7). Final biomarker selection was based on a priori criteria: 1) the gene product (i.e., protein) has biological and mechanistic plausibility regarding the host response to infection, immunity, and/or inflammation, and 2) the gene product is readily measured in the blood compartment.

All plasma samples were collected within the first 24 hours of presentation to the ICU. The plasma concentrations of the candidate biomarkers were measured using a multiplex magnetic bead platform (MILLIPLEX MAP, EMD Millipore Corporation, Billerica, MA) and a Luminex 100/200 System (Luminex Corporation, Austin, TX) according to the manufacturers' specifications. Technical assay performance data were previously published (5).

Additional Stratification Variables

We abstracted available data elements for consideration in the risk modeling that, based on existing literature, we hypothesized could be associated with poor outcomes: serum lactate concentration (mmol/L) at study entry, age, gender, and Acute Physiology and Chronic Health Evaluation (APACHE) II/III score. We also recorded the presence of the following comorbid conditions: New York Heart Association Class IV congestive heart failure, chronic obstructive pulmonary disease, requirement for chronic dialysis, chronic hepatic failure, hematologic or metastatic solid organ malignancy, and requirement for chronic steroids at study entry. We derived a binary "chronic disease" variable to indicate the presence of any one of these comorbidities.

Statistical Analysis

Initially, data are described using medians, interquartile ranges (IQRs), frequencies, and percentages. Comparisons between survivors and nonsurvivors used the Mann-Whitney *U* test, chi-square test, or Fisher exact test as appropriate. Descriptive statistics and comparisons used SigmaStat Software (Systat Software, San Jose, CA).

All-cause 28-day mortality is the primary outcome variable for the modeling procedures. To derive the decision tree, we employed a classification and regression tree (CART) approach (12, 13). The CART analysis procedure considered all 12 candidate biomarkers as well as other potential clinical predictor variables listed above. The procedure selects cut points and ordering of decision nodes that maximally discriminate between survivors and nonsurvivors. The tree was built using Salford Predictive Modeler v6.6 (Salford Systems, San Diego, CA). Performance of the tree is reported using diagnostic test statistics with 95% CIs computed using the VassarStats Website for Statistical Computation (14). Areas under the receiver operating characteristic (ROC) curves were compared using the method of Hanley and McNeil (15). The net reclassification improvement (NRI) was also used to estimate the incremental predictive ability of the biomarker-based model compared to using APACHE II scores alone (16). The NRI was computed using the R-package Hmisc.

RESULTS

Model Derivation

Table 1 provides the clinical and demographic data for the derivation cohort (n = 341), all of whom had septic shock. The 109 nonsurvivors (32.0%) were older, had a higher median APACHE II score, and a higher proportion had chronic disease at study entry, compared to the 232 survivors. The mean and median times to death in the derivation cohort nonsurvivors were 8.7 ± 7.9 (sp) and 6 days (IQR, 2–13 d), respectively.

Supplemental Table 1 (Supplemental Digital Content 1, http://links.lww.com/CCM/A829) shows the performance of the individual biomarkers. Figure 1 shows the derived decision tree. Maximum accuracy was achieved with five of the 12 candidate stratification biomarkers: CCL3, HSPA1B, IL8, GZMB, and CCL4. Serum lactate concentration at study entry, age, and presence of chronic disease further improved predictive accuracy. The individual comorbid conditions did not improve predictive capacity. There were six low-risk

terminal nodes (0.0–12.5% risk of death; terminal nodes 1, 3, 5, 7, 8, and 10) and six highrisk terminal nodes (34.4–84.2% risk of death; terminal nodes 2, 4, 6, 9, 11, and 12). Of the 138 subjects classified as low risk, 131 survived (94.9%) and 7 (5.1%) had died by 28 days. Of the 203 subjects classified as high risk, 102 (50.2%) had died by 28 days. Table 2 shows the diagnostic test characteristics of the decision tree in the derivation cohort.

Model Testing

The test cohort consisted of 331 subjects with septic shock (81.9%) or severe sepsis (18.1%), of whom 99 (29.9%) did not survive to 28 days. Table 1 provides the clinical and demographic data. Compared to the derivation cohort, the test cohort had a higher proportion of male subjects, a lower median APACHE II score, a lower proportion of subjects with chronic disease, and a lower proportion of subjects with septic shock. The mortality rate of the test cohort (29.9%) was not significantly different compared to the derivation cohort (32.0%). Within the test cohort, nonsurvivors had a higher median age, a lower proportion of subjects with either chronic disease or septic shock, compared to the survivors. The mean and median times to death in the test cohort nonsurvivors were 11.1 ± 8.0 and 10 days (IQR, 4–17 d), respectively, both of which were significantly greater compared to the derivation cohort.

Supplemental Figure 1 (Supplemental Digital Content 2, http://links.lww.com/CCM/A830) shows the classification of the test cohort subjects according to the decision tree. The algorithm from the derivation cohort (Fig. 1) was applied to the test cohort with no modifications. One hundred and twenty-six test cohort subjects were classified as low risk (terminal nodes 1, 3, 5, 7, 8, and 10), whereas 205 were classified as high risk (terminal nodes 2, 4, 6, 9, 11, and 12). Among the low-risk subjects, the mortality rate was 9.5%, whereas among the high- risk subjects, the mortality rate was 42.4%. Table 2 shows the diagnostic test characteristics of the decision tree in the test cohort. The model did not perform differently when applied against only the test cohort subjects with septic shock (n = 271, data not shown).

Comparison With APACHE II

We compared the performance of the biomarker-based model with that of APACHE II for all subjects in the derivation and test cohorts (n = 672). Figure 2 shows the ROC curves for the biomarker-based model and APACHE II. The area under the curve (AUC) for the biomarker-based model (0.784; 95% CI, 0.747–0.820) was superior to that of APACHE II (0.676; 95% CI, 0.632–0.721; p = 0.0001).

When adding the information from the biomarker-based model to the information in APACHE II, the NRI was 0.576 (95% CI, 0.341–0.812). The NRI is a measure of how much the accuracy of predicted outcomes is improved when adding information (16). The NRI ranges between -2 and +2. A score of -2 indicates that all true positives are reclassified as false negatives and all true negatives are reclassified as false positives, and no false classifications are reclassified as true classifications. Conversely, when the score is 2, adding the information correctly reclassifies every case. Our results demonstrate that the biomarker-

based model provides additional classification value beyond the information included in APACHE II.

Model Calibration

Assuming that a larger sample size could generate a more generalizable model, the decision tree was calibrated by combining all subjects in the derivation and test cohorts (n = 672) and repeating the CART analysis. The calibrated decision tree is shown in Figure 3. Notable changes included the addition of IL1A as a lower level decision leading to terminal nodes 7 and 8, and the replacement of a GZMB-based decision leading to terminal nodes 4 and 5 with an IL8-based decision. In addition, the decisions in the center of the tree based on lactate and chronic disease status changed their relative level positions. The calibrated tree contained six low-risk terminal nodes (2.7-17.4% risk of death; terminal nodes 1, 3, 4, 6, 8, and 10) and six high-risk terminal nodes (45.3-75.0% risk of death; terminal nodes 2, 5, 7, 9, 11, and 12). Of the 317 subjects classified as low risk, 291 survived (91.8%) and 26 (8.9%) had died by 28 days. Of the 355 subjects classified as high risk, 182 (51.3%) had died by 28 days. Table 2 shows the diagnostic test characteristics of the calibrated decision tree.

Validating the Calibrated Decision Tree

The calibrated decision tree was validated in a cohort of 209 subjects with septic shock, of whom 88 (42.1%) did not survive to 28 days. Table 1 provides the clinical and demographic data. Compared with the derivation cohort, the validation cohort had a higher mortality rate, a higher proportion of subjects with chronic disease, and a lower proportion of subjects with a surgical diagnosis. Within the validation cohort, nonsurvivors had a higher proportion of subjects with chronic disease, and a lower proportion of subjects with chronic disease, and a lower proportion of subjects with a surgical diagnosis, compared to the survivors. The mean and median times to death in the validation cohort nonsurvivors were 7.5 ± 6.9 and 5 days (IQR, 2–11 d), respectively.

Supplemental Figure 2 (Supplemental Digital Content 3, http://links.lww.com/CCM/A831) shows the classification of the validation cohort subjects according to the calibrated decision tree. The calibrated algorithm (Fig. 2) was applied to the validation cohort with no modifications. Eighty-six subjects were classified as low risk (terminal nodes 1, 3, 4, 6, 8, and 10), whereas 123 were classified as high risk (terminal nodes 2, 5, 7, 9, 11, and 12). Among the low-risk subjects, the mortality rate was 15.1%, whereas among the high-risk subjects, the mortality rate was 60.9%. Table 2 shows the performance of the calibrated decision tree in the validation cohort.

Since the validation cohort had APACHE III data available, we compared the performance of the calibrated model with that of APACHE III. In the validation cohort, the AUC for the calibrated model was 0.726 (95% CI, 0.660–0.792), whereas the AUC for APACHE III was 0.514 (95% CI, 0.434–0.595; p < 0.0001).

DISCUSSION

We have derived, tested, calibrated, and validated a risk stratification tool for adult septic shock that estimates the risk of 28-day mortality. A panel of biomarkers measured during the

initial presentation to the ICU with septic shock, as well as admission lactate concentrations, age, and chronic disease status, form the basis of the model. On testing and validation, we observe good performance of the model. For all subjects in the study (n = 881), the high-risk terminal nodes of the calibrated model identified a cohort with a mortality rate of 56%, whereas the low-risk terminal nodes identified a cohort with a mortality rate of 11%. This dichotomous interpretation demonstrates that the model can partition a heterogeneous cohort of patients into two broad groups having an approximately five-fold difference in mortality risk. A more comprehensive interpretation allows assigning risk based on the respective terminal nodes, and this partitions patients across a clinically relevant range of mortality probabilities. The negative predictive value and the negative likelihood ratio of the model indicate that it may be most reliable as a "rule-out tool."

A strength of our modeling is the initial approach to deriving the candidate stratification biomarkers. Using our extensive genome-wide expression databank, we identified 117 gene probes possibly associated with outcome in children with septic shock (6, 7, 17–24). From these, we selected the 12 biomarkers using a priori criteria.

The modeling process considered the candidate stratification biomarkers and clinical variables potentially associated with outcome. Interestingly, the biomarkers dominated the upper level decision rules, whereas the clinical variables contributed to either the lower level decision rules or not at all. This suggests that the biomarkers contribute significant and consistent stratification information that adds to stratification based on clinical findings; the NRI supports this assertion. The pediatric modeling procedures yielded similar results (5). In fact, the upper level decision rules of the pediatric and adult models consisted of the same three biomarkers (i.e., CCL3, HSPA1B, and IL8), albeit with different cutoff values. This suggests consistent utility for these three particular stratification biomarkers. We do note a limitation of our current study in that we did not consider other biomarkers having potential prognostic utility in adult populations (25), nor could we consider all possible clinical variables potentially associated with outcome due to reliance on secondary data sources.

The 2008 Surviving Sepsis Campaign International Guidelines recommend a serum lactate concentration greater than 4 mmol/L as a threshold indicator of tissue hypoperfusion warranting initiation of protocolized, quantitative resuscitation (26). The 2012 guidelines provide the same recommendation based on a reported mortality of 46% in septic patients with both hypotension and serum lactate concentration greater than 4 mmol/L (27, 28). Our calibrated decision tree indicates that serum lactate concentrations below these levels are associated with increased risk of mortality, which is consistent with two recent studies (29, 30).

The available data allowed a comparison of the biomarker- based model performance with both APACHE II and APACHE III, and the biomarker-based model outperformed APACHE II and III in these cohorts. We note that the Mortality Probability Model II at 24 hours (MPM II₂₄), customized for patients with sepsis, yielded an AUC of 0.826 in a previous study (31). MPM II₂₄ may perform better than our biomarker-based model, but MPM II₂₄ data were not recorded in the studies from which our data were pooled.

There are important differences between the cohorts included in this study. First, the initial derivation cohort was drawn from participants in an interventional clinical trial. They may represent a more selected population of patients than the subjects in the test and validation cohorts who were drawn from observational databases. Second, the cohorts were drawn from three different healthcare systems, each with their own healthcare practices, cultures, and demographics. Third, while all subjects in the derivation and validation cohorts met criteria for septic shock, the subjects in the test cohort met criteria for either septic shock (81.9%) or severe sepsis (19.1%). Fourth, the cohorts differed in age, gender, illness severity, time to death, surgical status, and chronic disease burden. All of these differences represent potential confounders, yet the models performed consistently well across cohorts.

We propose that the primary potential application of the biomarker-based model is to enhance patient selection for interventional clinical trials. Excluding the lowest risk patients who are likely to survive without experimental intervention reduces their exposure to risk of adverse effects from the new intervention. Furthermore, the approach may increase the likelihood of positive trial findings; excluding the highest risk patients unlikely to survive with any therapy removes patients who may be too sick to respond to treatment, which could enhance the potential for measurable risk reduction for new therapy among moderate or high-risk patients with modifiable outcomes. Application of the model in this manner would require the development of a rapid, multiassay platform and computer support to reliably apply the decision rule. Although neither of these currently exist, the technologies to develop them are routinely available.

In conclusion, we have derived, tested, and validated a multibiomarker-based risk model that estimates mortality probability in adults with septic shock. Favorable comparisons to existing scoring systems and good performance in the context of potentially profound confounding factors support the generalizability and utility of the model. We propose that the model may enhance clinical trial design. Another potential application of the model may include serving as a benchmarking metric for quality improvement efforts.

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Figure 1.

Classification tree from the derivation cohort (n = 341). The classification tree consists of 11 decision points and 22 daughter nodes. The classification tree includes five of the 12 candidate stratification biomarkers: C-C chemokine ligand 3 (CCL3), heat shock protein 70 kDa 1B (HSPA1B), interleukin-8 (IL8), granzyme B (GZMB), and C-C chemokine ligand 4 (CCL4). For consistency, the serum concentrations of all candidate stratification biomarkers are provided in pg/mL. The classification tree also includes serum lactate concentrations (mmol/L), age (yr), and the presence/absence of chronic disease as defined in the *Methods* section. The root node provides the total number of patients in the derivation cohort and the number of survivors and nonsurvivors with the respective rates. Each daughter node provides the respective rates. The numbers above daughter nodes designate terminal nodes. Terminal nodes 1, 3, 5, 7, 8, and 10 are considered high-risk terminal nodes.



Figure 2.

Comparison of receiver operating characteristic (ROC) curves for the biomarker-based model and Acute Physiology and Chronic Health Evaluation (APACHE) II. The ROC curves are calculated based on the respective mortality probabilities and 28-day all-cause mortality and are based on all subjects in the combined derivation and test cohorts (n = 672). The ROC curve for the biomarker-based model (*solid line*) yielded an area under the curve (AUC) of 0.784 (0.747–0.820), whereas the ROC curve for APACHE II (*dashed line*) yielded an AUC of 0.676 (0.632–0.721).

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Figure 3.

The calibrated classification tree based on the combination of all subjects in the derivation and test cohorts (calibration cohort, n = 672). The classification tree consists of 11 decision points and 22 daughter nodes. The classification tree includes six of the 12 candidate stratification biomarkers: C-C chemokine ligand 3 (CCL3), heat shock protein 70 kDa 1B (HSPA1B), interleukin-8 (IL8), granzyme B (GZMB), C-C chemokine ligand 4 (CCL4), and interleukin-1 α (IL1A). The classification tree also includes serum lactate concentrations (mmol/L), age (yr), and the presence/absence of chronic disease as defined in the *Methods* section. The conventions of the calibrated classification tree are the same as that described for Figure 1. Terminal nodes 1, 3, 4, 6, 8, and 10 are considered low-risk terminal nodes, whereas terminal nodes 2, 5, 7, 9, 11, and 12 are considered high-risk terminal nodes.

Table 1

Clinical and Demographic Data for the Derivation, Test, and Validation Cohorts

				Derivation (Cohort
Variable			ПА	Survivor	s Nonsurvivors
и			341	232	109
Median age (I	QR)		63 (51–73) 61 (48–72) 64 (54–76) ^a
Males, n (%)			201 (58.9)	142 (61.2)) 59 (54.1)
Median APA(CHE II (IQR) ^d		27 (22–32) 25 (20–30) 31 (24–35) ^a
Median APA0	CHE III (IQR)	p	NA	NA	NA
Number of pe	ople with chro	nic disease (%) f	165 (48.4)	96 (41.4)	69 (63.3) ^C
Mean days to	death \pm sD		NA	NA	8.7 ± 7.9
Median days t	to death (IQR)		NA	NA	6 (2–13)
Number of pe	ople with septi	ic shock (%)	341 (100)	232 (100)	109 (100)
Number of pe	ople with a sur	gical diagnosis (%).	78 (23)	56 (24)	22 (20)
	Test Cohort		•	alidation Coh	ort
ЧI	Survivors	Nonsurvivors	ЧΙ	Survivors	Nonsurvivors
331	232	66	209	121	88
61 (50–72)	58 (47–69)	69 (56–76) ^a	62 (51–71)	63 (50–72)	62 (53–71)
229 (69.2) b	164 (70.7)	65 (59.6) ^C	116 (55.5)	67 (55.3)	49 (55.7)
23 (18–29) e	22 (17–27)	27 (20–32) ^a	NA	NA	NA
NA	NA	NA	50 (37–79)	51 (37–76)	50 (38-82)
95 (28.7) ^b	56 (24.1)	39 (35.8) ^c	$132~(63.2)^{b}$	68 (56.2)	64 (74.4) ^C
NA	NA	$11.1\pm 8.0^{\textit{\theta}}$	NA	NA	7.5 ± 6.9
NA	NA	$10 \ (4-17)^{e}$	NA	NA	5 (2–11)
271 (81.9) ^b	182 (78.4)	89 (89.9) ^c	209 (100)	121 (100)	88 (100)
90 (27)	68 (29)	22 (22)	$(4)^{b}$	8 (7)	$0 (0)_{C}$
IQR = interquat	rtile range, AP	ACHE = Acute Phy	'siology and C	hronic Health	Evaluation, NA = no

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 $\frac{a}{p} < 0.05$ versus respective survivors; Mann-Whitney U test.

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b > 0.05 versus derivation cohort; chi-square test.

 $^{c}_{p}<0.05$ versus respective survivors; chi-square test.

^dThe derivation and test cohorts had APACHE II scores recorded, whereas the validation cohort had APACHE III scores recorded.

 $e \over p < 0.05$ versus derivation cohort; Mann-Whitney U test.

f The presence of at least one of the following at study entry: New York Heart Association Class 4 congestive heart failure, chronic obstructive pulmonary disease, requirement for chronic dialysis, chronic hepatic failure, hematologic or metastatic solid organ malignancy, or requirement for chronic steroids.

Table 2

Diagnostic Test Characteristics of the Decision Trees

Variable	Derivation Cohort	Test Cohort	Calibration Cohort (Combined Derivation and Test Cohorts)	Validation Cohort
No. of subjects	341	331	672	209
No. of true positives	102	87	182	75
No. of true negatives	131	114	291	73
No. of false positives	101	118	173	48
No. of false negatives	7	12	26	13
Sensitivity	94% (87–97) ^a	88% (79–93)	88% (82–92)	85% (76–92)
Specificity	56% (50-63)	49% (43–56)	63% (58–67)	60% (51–69)
Positive predictive value	50% (43-57)	42% (36–50)	51% (46–57)	61% (52–70)
Negative predictive value	95% (89–98)	90% (84–95)	92% (88–94)	85% (75–91)
Positive likelihood ratio	2.1 (1.8–2.5)	1.7 (1.5–2.0)	2.3 (2.1–2.7)	2.1 (1.7–2.7)
Negative likelihood ratio	0.1 (0.06–0.2)	0.2 (0.1–0.4)	0.2 (0.1–0.3)	0.2 (0.1–0.4)
Area under the curve	0.834 (0.792–0.875)	0.720 (0.661–0.780)	0.793 (0.758–0.823)	0.726 (0.660–0.792)

^aNumbers in parentheses represent 95% CIs.