

Efficiency to Discovery Transgenic Loci in GM Rice Using Next Generation Sequencing Whole Genome Re-sequencing

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Molecular characterization technology in genetically modified organisms, in addition to how transgenic biotechnologies are developed now require full transparency to assess the risk to living modified and non-modified organisms. Next generation sequencing (NGS) methodology is suggested as an effective means in genome characterization and detection of transgenic insertion locations. In the present study, we applied NGS to insert transgenic loci, specifically the epidermal growth factor (EGF) in genetically modified rice cells. A total of 29.3 Gb (~72× coverage) was sequenced with a 2 × 150 bp paired end method by Illumina HiSeq2500, which was consecutively mapped to the rice genome and T-vector sequence. The compatible pairs of reads were successfully mapped to 10 loci on the rice chromosome and vector sequences were validated to the insertion location by polymerase chain reaction (PCR) amplification. The EGF transgenic site was confirmed only on chromosome 4 by PCR. Results of this study demonstrated the success of NGS data to characterize the rice genome. Bioinformatics analyses must be developed in association with NGS data to identify highly accurate transgenic sites.

Keywords: genetically modified organisms, next generation sequencing (NGS) T-DNA, rice, risk assessment

Introduction

Genetic engineering technology is widely used in the agricultural and plant biotechnology fields, ranging from the food and feed industries to bio-pharmaceuticals and cosmetics [1, 2]. The history of genetically modified (GM) technology began with the discovery of plasmid DNA, where the plasmid could be transferred from one cell to another genome [3]. Scientists subsequently applied the basic plasmid vector system principle and developed recombinant DNA technology to create genetically engineered organisms. Today, GM techniques have been applied to various research fields, including crop sciences, drug manufacturing, and animal husbandry.

The development of transgenic biotechnologies over the last 20 years has led to safety concerns regarding genetically modified organisms (GMOs), particularly in food crops and new pharmaceuticals, which are the most controversial issues. Safety concerns regarding GMOs have resulted in research, debates, and ongoing public unease. Therefore, the European Union (EU) and National Institutes of Health (NIH) in the United States proposed an authorization process in commercial GMO use; however, public apprehension for transgenic techniques remains uncertain and controversial [4-8].

Generally, molecular characterization and identification of GMOs are performed using Southern blots and polymerase chain reaction (PCR) based detection followed by conventional sequencing methods [7]. However, these appro-

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aches are limited to evaluate whether the host genome has unintended sequence substitutions and indels [9]. Moreover, if sufficient genomic information is not available for the chosen comparative model species, it is difficult to detect the correct transgenic insert site location or sequence contamination of vector DNA [9, 10].

Recent publications of GMO molecular characterizations reported the use of next generation sequencing (NGS) approaches as an effective means to detect the precise transgenic insert location [9, 11, 12]. High-throughput DNA sequencing technologies and bioinformatics can be coupled with NGS to offer new possibilities in drawing genetic maps with feasible costs. For these reasons, researchers have tested new approaches in the molecular characterization of GMOs using NGS technologies [9, 10, 12].

Here, we examined transgenic insertion sites using paired-end whole genome re-sequencing data following Yang *et al.* with modifications [9]. Human epidermal growth factor (EGF) was inserted into GM rice cells, which could produce EGF safely without endotoxin derived from bacteria and was used as material for this study. Deep sequencing was performed with the Illumina HiSeq2500 platforms (Illumina Inc., San Diego, CA, USA). In this pilot study, we demonstrated the potential of NGS for examination of transgenic insertion loci and discuss some technical bottlenecks of this new method.

Methods

GM rice samples

The GM rice event PJKS131-2 was transformed with the EGF inserted pJKS131 vector, produced by Natural Bio-Materials Inc. (Jeonju, Korea). Taxonomically, the event PJKS131-2 was derived from *Oryza sativa* L. cv. Dongjin. The T-vector was transformed with rice callus as described by Chan *et al.* [13]. Transgenic rice calli were incubated with

50 mg/L of hygromycin B antibiotic (A.G. Scientific Inc., San Diego, CA, USA) for selection. The GM rice callus samples were subjected to NGS and further validated by PCR amplification.

DNA extraction and whole genome shotgun library and sequencing

The calli of GM rice event PJKS131-2 were collected and stored at -80°C . Total genomic DNA was extracted using the CTAB method in liquid nitrogen. Genomic DNA quality was evaluated by 0.5% agarose gel electrophoresis. Following the quality check, genomic DNA was sheared with average 500 bp fragment sizes. Truseq DNA PCR free Library Preparation Kit (Illumina Inc.) was used to construct the DNA library according to the manufacturer's protocol. The quality of constructed DNA libraries was confirmed by the LabChip GX system (PerkinElmer, Waltham, MA, USA). DNA libraries were sequenced with 150-bp paired-end sequencing using Illumina HiSeq2500.

Transgenic insertion analysis

Initially, paired-end reads were filtered out by phred scores < 20 and duplicate sequences were removed. After filtration, DNA fragments were consecutively mapped against the rice reference genome (phytozome v9 [14]) and T-vector sequence (Supplementary Fig. 1). The transgene insertion types were classified by adaptation and modification of the analytical strategies reported in Yang *et al.* [9]. Fig. 1 shows the workflow applied in this method. Initially, all NGS reads were individually mapped to the rice reference genome and transgenic vector (types A and C in Yang *et al.* [9]). Subsequently, these NGS reads were eliminated to conduct the following analyses. NGS reads not classified as above were classified into the following two classes: one side of the NGS read matched the reference genome, (1) the other one matched to vector (type B in Yang *et al.* [9]); or (2) one

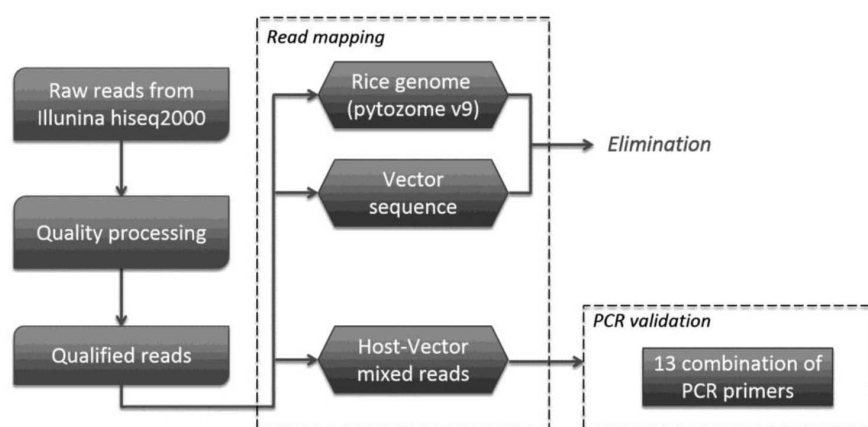
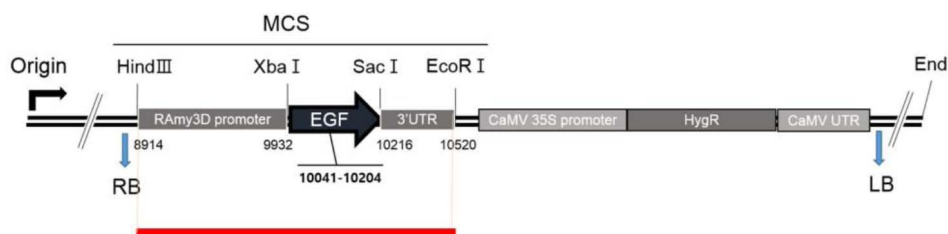


Fig. 1. Summary of the work-flow. PCR, polymerase chain reaction.

Table 1. Whole genome sequencing summary

Event	No. of reads	Total read length (bp)	Q30 (%)	GC ratio (%)
PJKS131-2	194,965,440	29,359,127,691 (72×)	71.56	41.58

**Fig. 2.** Schematic drawing of the transgenic vector genome. Red line represent the region of mapped reads. MCS, multiple cloning site; RB, right border; LB, left border.

side of the NGS read exhibited both elements from the rice reference and transgenic vector (types D and E in Yang *et al.* [9]).

Experimental validation of transgenic inserts

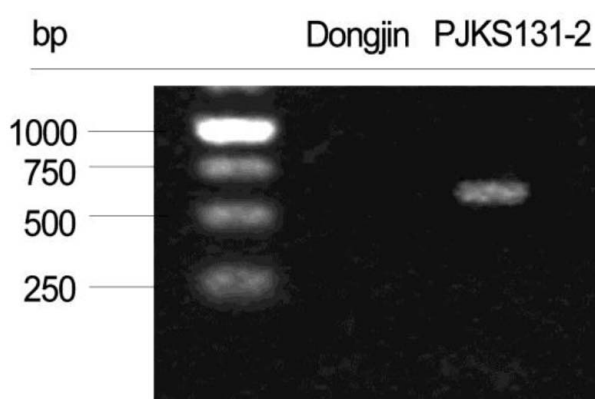
Each of the 13 combination primer sets was designed congruent with the transgenic insertion region orientation. PCR was conducted using DNA polymerase (Solgent Co., Daejeon, Korea) following the manufacturer's instructions. The reaction was performed under the following conditions: a pre-denaturation step at 95°C for 5 min; denaturation at 95°C for 60 s; 30 amplification cycles, including annealing at 60°C for 45 s, and elongation at 72°C for 120 s; and a final elongation at 72°C for 5 min.

Results

Whole genome re-sequencing and mapping to discover the transgenic position

The transgenic GM rice site, PJKS131-2, was detected by performing whole genome re-sequencing using callus tissue. Genomic DNA libraries were constructed with an average 500 bp and both ends were read with 150 bp paired-end sequencing methods. A total length of raw sequencing reads were 29.3 Gb (~194.9 million reads), which showed ~72× coverage in the total read length (Table 1). Following quality control processing, reads with average phred scores ≥ 30 were estimated at ~71.5% (Table 1).

The types of mapped reads were classified by alignment of all NGS reads to the rice reference genome and transgenic vector sequences. Fig. 2 shows construction of the pJKS131 transgenic vector. Reads were aligned on the cloning vector positions 8,500 bp to 10,500 bp, similar to transgenic insert locations. Detailed mapping strategies were described in the Methods. The transgene insertion site was identified by classifying reads where one end matched the host genome

**Fig. 3.** Polymerase chain reaction validation of transgenic site.

and the other end matched the vector sequences (i.e., types B, D, and E) mapped back to the rice chromosome and known vector sequences. Eleven pairs of reads were identified on rice chromosome including chromosome 4. The total mapped reads described above were compatible with the transgenic vector backbone sequences.

PCR validation of mapping prediction

Thirteen PCR primers designed based on mapping direction validated the mapping results of 10 transgenic insert candidates. PCR results confirmed the target *EGF* sequence was successfully inserted on rice chromosome 4 (Figs. 3 and 4). The remaining reads were concluded to be artifacts, because all matches were not detected with PCR.

Discussion

Recent developments in NGS methods and accompanying bioinformatics tools have paved the way for ongoing genomics research widely used in the agricultural biotechnology field. Consequently, several studies reported new

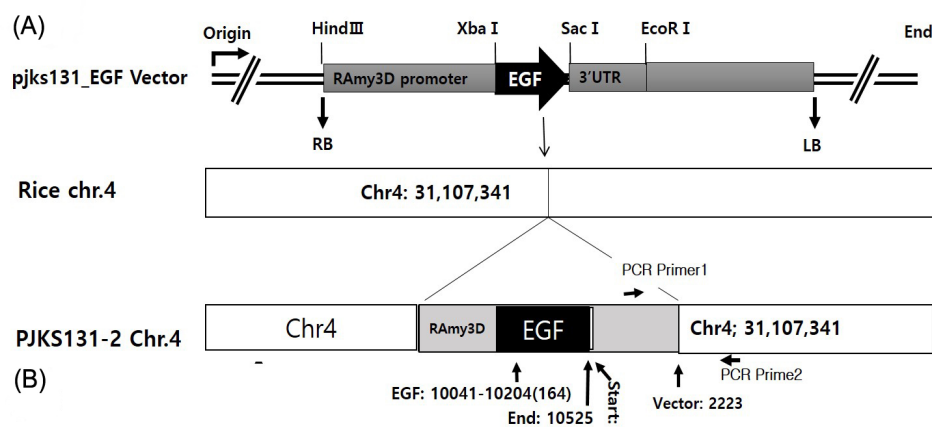


Fig. 4. Transgenic position of epidermal growth factor (*EGF*) locus on the rice chromosome 4 and polymerase chain reaction (PCR) test to identify T-DNA junction sequence. (A) The *EGF* is inserted on the position 31,104,341 of the chromosome 4. (B) The bold with underline is T-DNA sequence of the vector 2,026–2,223 bp and the next bases is rice transgenic locus chromosome 4 (31107341–31107690) in the fragment amplified by PCR test primer1 (5' TACCTGCA-TGCTGCGGTGAAG 3') and primer2 (5'AGGGCTGTGTAGAAGTACTCGC 3').

GAAGTACTCGCCGATAGTGGAAACCGACGCCAGCACTCGTCCGAGGGCAAAGAAATAGAGTAGATGCCGACCGGGATCTGTG
ATCGACAAGCTCGAGTTTCTCCATAATAATGTGTGAGTAGTCCAGATAAGGGAATTAGGGTTCCTATAGGGTTTCGCTCATGTGT
TGAGCATATAAGAAACCTTAGTATGTGATGCAAGTAAACAATTATGGATTCTACTTATGTTCTTTCTTTTCTGACTGGGTTGCAGG
 TATGAGAAGAGGCATTCCAACATTCCGGCTCACGTCTCCCATGCTTCGTTGCAAGGAAAGGTGACCATGTCATCATTGGCCAGTGCAGGT
 AAAACTTAACCTCCATACCTTAGTTTTGACTCCCTAGAAATCATCTTAAATAACAGAGGTGCTATTAGTTCAAACATTATCAGTTCACCCAGC
 TAGTACTATGAGATTTGGTTGCTGAAGTACAATTTCTGCATTTCAACAGGCCGCTGTCGAAAATGTGAGGTTCAACGCTCTGAAGTGCAT
 CCCAGCTGGATCCACCGCGGCAGCGCGGCAAGAAGGCCTTACCG

approaches in GM crop safety assessment using NGS platforms [10-12]. In our study, we investigated *EGF* inserted GM rice events using NGS technology and bioinformatics to test the potential uses of this new approach in molecular assessment of transgenic organisms.

Results were successful in differentiating NGS read types using *in silico* analyses from GM rice, PJKS131-2 and hypothetically, the outcome was acceptable in terms of read classification. However, as a validation step, we experienced unexpected problems. Consistent with mapping and aligning data, we considered all possible transgenic insertion directions on the rice chromosomes and designed PCR primers based on loci information. Among the primers, except for locus specific primers on chromosome 4, results showed all matches were mismatches, which was caused by computational errors derived from analogous sequences between the rice genome and the transgenic vector. Therefore, we concluded it is essential to develop more accurate algorithms based on the transformation vector.

In addition, it is important to note our experimental sample was collected from rice callus tissues, with *Agrobacterium* co-incubation and a plant cell suspension culture system. Transgenic plant cell suspension culture system exhibits several advantages, including a low microorganism risk and chemical contamination, simple cell culture methods, economical facilities, and stable productivity. However, it is difficult to obtain pure genomic DNA of the host plant without plasmid DNA mixing using the plant cell culture method. We eliminated NGS raw reads mapped only against vector DNA (type C), however if raw reads contained too many vector backbone sequences, problems in further

bioinformatics analyses would still occur. Further studies are required with appropriate controls of GM plants in cell culture environments.

In the present study, we completed a proof-of-concept experiment to examine the molecular characterization of a recombinant-protein produced GM rice event using NGS methods. New approaches have recently been reported to assess the development and release of GM crops, however these techniques are not popularized in the field of GM risk assessment. However, previous studies in other disciplines have successfully established NGS, but for practical reasons, it has not been easy to apply this new method for testing GMOs. NGS strategies largely depend on sample quality, amount of data, and subsequent bioinformatics analyses. Therefore, it is critical proper guidelines to discovery transgenic site by NGS data matched and PCR test in the GMOs established and required.

Supplementary material

Supplementary data including one figure can be found with this article online at <http://www.genominfo.org/src/sm/gni-13-81-s001.pdf>.

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SUPPLEMENTARY INFORMATION

Efficiency to Discovery Transgenic Loci in GM Rice Using Next Generation Sequencing Whole Genome Re-sequencing

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Korea

※ pJKS131 vector sequence

1060-2085 : HPTII

2376-2401 : LB T-DNA

8634-8661 : RB T-DNA

8907-9938 : RAmy3D promoter

9945-9965 : Hwang`s 5`UTR

9966-10040 : RAmy3D signal peptide

10041-10204 : EGF mature peptide

10211-10519 : RAmy3D 3`UTR

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      10      20      30      40      50      60
GTAATCATGG TCATAGCTGT TTCCTGTGTG AAATTGTTAT CCGCTCACAA TTCCACACAA

      70      80      90     100     110     120
CATACGAGCC GGAAGCATAA AGTGTAAGC CTGGGGTGCC TAATGAGTGA GCTAACTCAC

     130     140     150     160     170     180
ATTAATTGCG TTGCGCTCAC TGCCCGCTTT CCAGTCGGGA AACCTGTCGT GCCAGCTGCA

     190     200     210     220     230     240
TTAATGAATC GGCCAACGCG CGGGGAGAGG CGGTTTGCGT ATTGGCTAGA GCAGCTTGCC

     250     260     270     280     290     300
AACATGGTGG AGCACGACAC TCTCGTCTAC TCCAAGAATA TCAAAGATAC AGTCTCAGAA

     310     320     330     340     350     360
GACCAAAGGG CTATTGAGAC TTTTCAACAA AGGGTAATAT CGGGAAACCT CCTCGGATTC

     370     380     390     400     410     420
CATTGCCCAG CTATCTGTCA CTTTCATCAA AGGACAGTAG AAAAGGAAGG TGGCACCTAC

     430     440     450     460     470     480
AAATGCCATC ATTGCGATAA AGGAAAGGCT ATCGTTCAAG ATGCCTCTGC CGACAGTGGT

     490     500     510     520     530     540
CCCAAAGATG GACCCCCACC CACGAGGAGC ATCGTGGAAA AAGAAGACGT TCCAACCACG

     550     560     570     580     590     600
TCTTCAAAGC AAGTGGATTG ATGTGATAAC ATGGTGGAGC ACGACACTCT CGTCTACTCC

     610     620     630     640     650     660
AAGAATATCA AAGATACAGT CTCAGAAGAC CAAAGGGCTA TTGAGACTTT TCAACAAAGG

     670     680     690     700     710     720
GTAATATCGG GAAACCTCCT CGGATTCCAT TGCCAGCTA TCTGTCACTT CATCAAAGG

     730     740     750     760     770     780
ACAGTAGAAA AGGAAGGTGG CACCTACAAA TGCCATCATT GCGATAAAGG AAAGGCTATC
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GTTCAAGATG CCTCTGCCGA CAGTGGTCCC AAAGATGGAC CCCCACCCAC GAGGAGCATC

850 860 870 880 890 900
GTGGAAAAAG AAGACGTTCC AACCACGTCT TCAAAGCAAG TGGATTGATG TGATATCTCC

910 920 930 940 950 960
ACTGACGTAA GGGATGACGC ACAATCCAC TATCCTTCGC AAGACCTTCC TCTATATAAG

970 980 990 1000 1010 1020
GAAGTTCATT TCATTTGGAG AGGACACGCT GAAATCACCA GTCTCTCTCT ACAAATCTAT

1030 1040 1050 1060 1070 1080
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1090 1100 1110 1120 1130 1140
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1150 1160 1170 1180 1190 1200
CTCTCGGAGG GCGAAGAATC TCGTGCTTTC AGCTTCGATG TAGGAGGGCG TGGATATGTC

1210 1220 1230 1240 1250 1260
CTGCGGTAA ATAGCTGCGC CGATGGTTTC TACAAAGATC GTTATGTTTA TCGGCACTTT

1270 1280 1290 1300 1310 1320
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1390 1400 1410 1420 1430 1440
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1570 1580 1590 1600 1610 1620
GACACCGTCA GTGCGTCCGT CGCGCAGGCT CTCGATGAGC TGATGCTTTG GGCCGAGGAC

1630 1640 1650 1660 1670 1680
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1690 1700 1710 1720 1730 1740
AATGGCCGCA TAACAGCGGT CATTGACTGG AGCGAGGCGA TGTTCCGGGA TTCCAATAC

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1810 1820 1830 1840 1850 1860
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1930 1940 1950 1960 1970 1980
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1990 2000 2010 2020 2030 2040
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2110 2120 2130 2140 2150 2160
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2170 2180 2190 2200 2210 2220
GGGAATTAGG GTTCCTATAG GGTTTCGCTC ATGTGTTGAG CATATAAGAA ACCCTTAGTA

2230 2240 2250 2260 2270 2280
TGTATTTGTA TTTGTAAAAT ACTTCTATCA ATAAAATTC TAATTCCTAA AACCAAAATC

2290 2300 2310 2320 2330 2340
CAGTACTAAA ATCCAGATCC CCCGAATTAA TTCGGCGTTA ATTCAGTACA TTAAAAAGT

2350 2360 2370 2380 2390 2400
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4090 4100 4110 4120 4130 4140
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4150 4160 4170 4180 4190 4200
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4330 4340 4350 4360 4370 4380
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4390 4400 4410 4420 4430 4440
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4450 4460 4470 4480 4490 4500
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4510 4520 4530 4540 4550 4560
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4570 4580 4590 4600 4610 4620
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4630 4640 4650 4660 4670 4680
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4690 4700 4710 4720 4730 4740
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4750 4760 4770 4780 4790 4800
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4810 4820 4830 4840 4850 4860
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4870 4880 4890 4900 4910 4920
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4930 4940 4950 4960 4970 4980
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4990 5000 5010 5020 5030 5040
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5050 5060 5070 5080 5090 5100
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5170 5180 5190 5200 5210 5220
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5230 5240 5250 5260 5270 5280
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5290 5300 5310 5320 5330 5340
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5350 5360 5370 5380 5390 5400
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5470 5480 5490 5500 5510 5520
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5530 5540 5550 5560 5570 5580
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5650 5660 5670 5680 5690 5700
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5770 5780 5790 5800 5810 5820
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5830 5840 5850 5860 5870 5880
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5950 5960 5970 5980 5990 6000
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6370 6380 6390 6400 6410 6420
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6430 6440 6450 6460 6470 6480
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6490 6500 6510 6520 6530 6540
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6550 6560 6570 6580 6590 6600
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6670 6680 6690 6700 6710 6720
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6850 6860 6870 6880 6890 6900
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6970 6980 6990 7000 7010 7020
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7270 7280 7290 7300 7310 7320
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7330 7340 7350 7360 7370 7380
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7390 7400 7410 7420 7430 7440
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7450 7460 7470 7480 7490 7500
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8230 8240 8250 8260 8270 8280
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8290 8300 8310 8320 8330 8340
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8350 8360 8370 8380 8390 8400
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8470 8480 8490 8500 8510 8520
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8650 8660 8670 8680 8690 8700
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9370 9380 9390 9400 9410 9420
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9430 9440 9450 9460 9470 9480
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9490 9500 9510 9520 9530 9540
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9550 9560 9570 9580 9590 9600
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9670 9680 9690 9700 9710 9720
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9730 9740 9750 9760 9770 9780
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10270 10280 10290 10300 10310 10320
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10330 10340 10350 10360 10370 10380
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10450      10460      10470      10480      10490      10500
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10510      10520      10530      10540      10550      10560
CCGATCAGTG AGTTTTTATG AATTC..... .....
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Supplementary Fig. 1. pJKS131 vector sequence to transfer the *EGF* to the rice genome.