

## REVIEW

# Mechanisms of resistance to EGFR tyrosine kinase inhibitors



Lihua Huang, Liwu Fu\*

State Key Laboratory of Oncology in Southern China, Cancer Center, Sun Yat-Sen University, Guangzhou 510060, China

Received 19 March 2015; received in revised form 12 June 2015; accepted 13 June 2015

## KEY WORDS

EGFR;  
TKIs;  
Resistance;  
Mechanisms

**Abstract** Since the discovery that non-small cell lung cancer (NSCLC) is driven by epidermal growth factor receptor (*EGFR*) mutations, the EGFR tyrosine kinase inhibitors (EGFR-TKIs, *e.g.*, gefitinib and erlotinib) have been effectively used for clinical treatment. However, patients eventually develop drug resistance. Resistance to EGFR-TKIs is inevitable due to various mechanisms, such as the secondary mutation (T790M), activation of alternative pathways (*c-Met*, HGF, AXL), aberrance of the downstream pathways (*K-RAS* mutations, loss of *PTEN*), impairment of the EGFR-TKIs-mediated apoptosis pathway (*BCL2*-like 11/*BIM* deletion polymorphism), histologic transformation, ATP binding cassette (ABC) transporter effusion, *etc.* Here we review and summarize the known resistant mechanisms to EGFR-TKIs and provide potential targets for development of new therapeutic strategies.

© 2015 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Abbreviations:** ABC, ATP binding cassette; ABCB1, ATP binding cassette, sub-family B, member 1; ABCC1, ATP binding cassette, sub-family C, member 1; ABCC10, ATP binding cassette, sub-family C, member 10; ABCG2, ATP binding cassette, sub-family G, member 2; AKT, protein kinase B; ALK, anaplastic lymphoma kinase; AXL, Anxelekt; BCL-2, B-cell CLL/lymphoma-2; BCL2L11/BIM, BCL2-like 11; BH3, BCL2-homology domain 3; BRAF, v-RAF murine sarcoma viral oncogene homolog B1; CML, chronic myelogenous leukemia; CRKL, Crk-like protein; EGFR, epidermal growth factor receptor; EGFR-TKIs, epidermal growth factor receptor tyrosine kinase inhibitors; EGFRvIII, EGFR variant III; EML4, echinoderm microtubule-associated protein-like 4; EMT, epithelial mesenchymal transition; ERK1/2, extracellular signal-regulated kinases; FGFs, fibroblast growth factors; FGFRs, fibroblast growth factor receptors; GAS6, growth-arrest-specific protein 6; HER, human epidermal receptor; HGF, hepatocyte growth factor; IGF, insulin growth factor; IGF-BPs, IGF-binding proteins; IGF-1R, IGF-1 receptor; IL, interleukin; IL-6R, IL-6 receptor; JAK, janus kinase; MAPK, mitogen-activated protein kinase; MEK, mitogen-activated protein kinase; NSCLC, non-small cell lung cancer; PDGFs, platelet-derived growth factors; PDGFRs, platelet-derived growth factor receptors; PI3K, phosphatidylinositol-3-kinase; PIK3CA, phosphatidylinositol-4,5-bisphosphate 3-kinase, catalytic subunit alpha; PTEN, phosphatase and tensin homolog; RAF, rapidly accelerated fibrosarcoma; RAS, rat sarcoma; RTK, tyrosine kinase receptor; SF, scatter factor; SOCS3, suppressor of cytokine signaling 3; STAT, signal transducers and activators of transcription; TKs, tyrosine kinases; TKIs, tyrosine kinase inhibitors; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor

\*Corresponding author. Tel.: +86 20 87343163; fax: +86 20 87343170.

E-mail address: [fulw@mail.sysu.edu.cn](mailto:fulw@mail.sysu.edu.cn) (Liwu Fu).

Peer review under responsibility of Institute of Materia Medica, Chinese Academy of Medical Sciences and Chinese Pharmaceutical Association.

<http://dx.doi.org/10.1016/j.apsb.2015.07.001>

2211-3835 © 2015 Chinese Pharmaceutical Association and Institute of Materia Medica, Chinese Academy of Medical Sciences. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. EGFR signal pathway and cancers

EGFR, also known as ERBB1 and HER1, is a transmembrane tyrosine kinase receptor (RTK). EGFR is a member of the human epidermal receptor (HER) family and a crucial component of cell signal pathways. Binding with ligands (EGF and TGF- $\alpha$ ) leads to conformational changes in EGFR and homodimerization or heterodimerization with other HER family members. There is subsequent autophosphorylation of the cytoplasmic tyrosine kinase (TK) domain with the help of adapter proteins (*e.g.*, SHC and GRB-2), which triggers downstream signaling. There are three main downstream pathways: (1) rat sarcoma (RAS)/rapidly accelerated fibrosarcoma (RAF)/mitogen-activated protein kinase (MAPK) pathway; (2) phosphatidylinositol-3-kinase (PI3K)/protein kinase B (AKT) pathway and (3) janus kinase (JAK)/signal transducers and activators of transcription (STAT) pathway, which stimulates mitosis, leading to cell proliferation and inhibition of apoptosis<sup>1</sup>. These pathways are crucial in normal cell growth (Fig. 1).

EGFR also serves as a stimulus for cancer growth. *EGFR* gene mutations and protein overexpression, both of which activate downstream pathways, are associated with cancers, especially lung cancer. The importance of EGFR to lung cancers supports the concept of ‘oncogene addiction’. Tyrosine kinase inhibitors (TKIs) have been used to treat the cancer harboring *EGFR* mutations or aberrant activation of EGFR. TKIs can inhibit the EGFR TK domain reversibly through competitive binding with ATP<sup>2</sup>. TKIs also lead

to tumor cell death through BCL2-like 11 (BIM)-mediated apoptosis. However, patients with *EGFR*-activating mutations benefit from treatment with EGFR-TKIs (*e.g.*, gefitinb and erlotinib) for less than approximately 1 year, after which drug resistance develops.

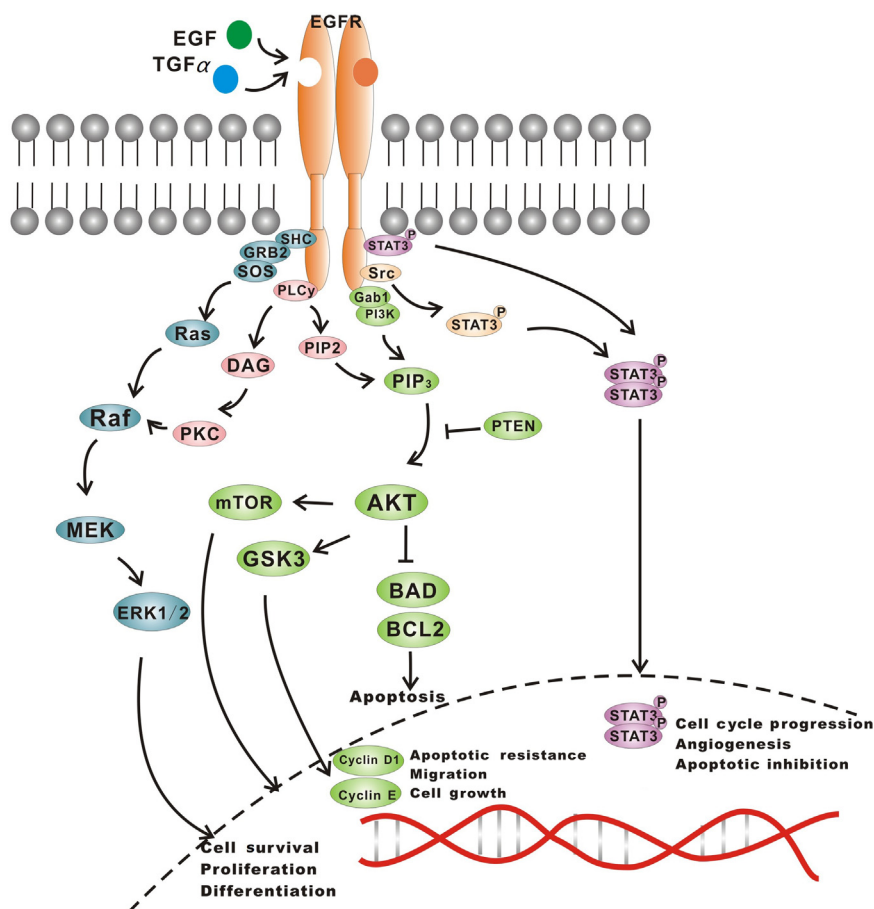
The etiology of EGFR-TKI resistance is complex. According to the cell signal transduction pathway, the etiology of EGFR-TKI resistance can be divided into subsequent categories.

## 2. EGFR mutations induce drug resistance, especially the secondary mutation T790M

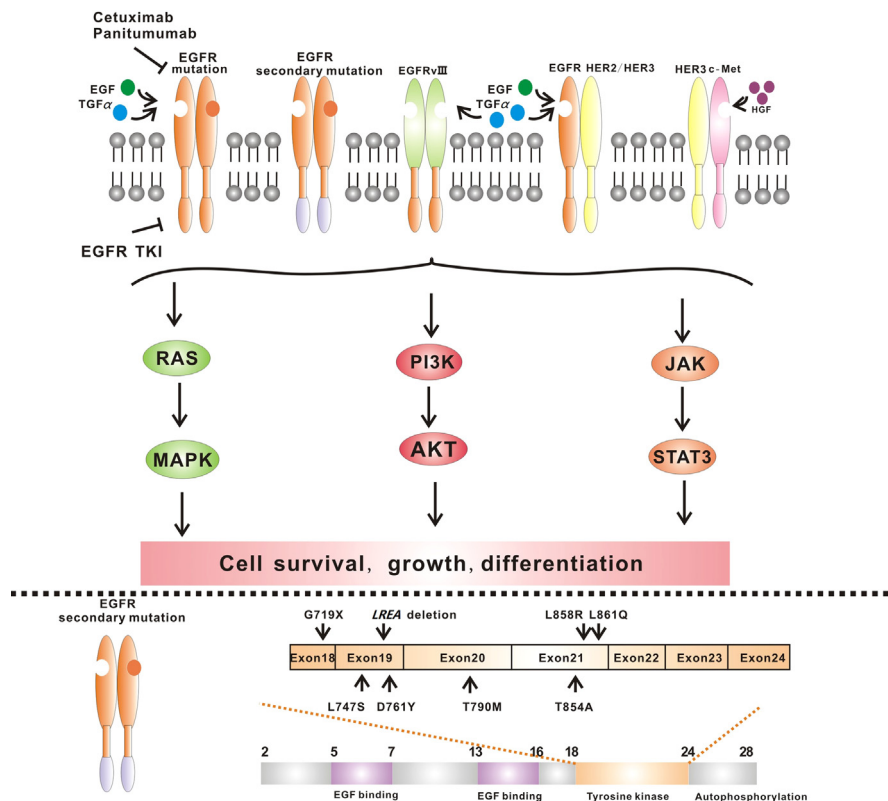
The *EGFR* gene, located in the 7p12–14 region in the short arm of chromosome 7, consists of 28 exons. The tyrosine kinase function is encoded by exons 18–24. Currently, more than 90% of the known *EGFR* mutations reside in exons 19–21 (Fig. 2). The rate of mutation in exon 19 is the highest, accounting for more than 60% of overall mutations<sup>3</sup>.

### 2.1. Secondary mutation—T790M

A secondary mutation of the *EGFR* gene reported in 2005 conferred acquired resistance to EGFR-TKIs<sup>4</sup>. This mutation (located in exon 20) results in the substitution of methionine for threonine at position 790 (T790M) in the kinase domain. Threonine 790 has been designated as a “gatekeeper” residue, important for regulating inhibitor specificity in



**Figure 1** EGFR and its signal pathway. There is subsequent autophosphorylation of the cytoplasmic tyrosine kinase domain, which, with the aid of adapter proteins (*e.g.*, SHC and GRB-2), triggers downstream signaling. The principal pathways included: (1) RAS/RAF/MEK, (2) PI3K/AKT and (3) JAK/STAT pathways.



**Figure 2** Aberration of HER families. Members of HER families get involved in the resistance to EGFR-TKIs. The secondary mutations of *EGFR*, *EGFR-vIII*, the overexpression of *HER2* or mutations of *HER2* contribute to the resistance in the presence of EGFR-TKIs. Compared to the other HER proteins, there are currently no mutational alterations known to confer oncogenic activities to *HER3*. In most cases, *HER3* phosphorylation is driven by one of HER family kinase partners, like *HER1* and *HER2*. What's more, resistance can also occur through amplification of the proto-oncogene *c-Met* and the *c-Met*-mediated phosphorylation of *HER3*. *HER3* serves as a key activator of downstream *PI3K/AKT* and *MEK/MAPK* signal pathways through dimerization with other HER family proteins or other molecules.

the ATP binding pocket. The T790M mutation enhances affinity of the ATP binding pocket for ATP, thus successfully competing with the TKIs, thereby conferring resistance<sup>5</sup>. Currently, two theories can explain the production of the second mutations: subcloning and induced mutation/acquisition<sup>6</sup>. Although the second mutation rarely occurs prior to treatment, it is found in approximately half of EGFR TKIs-treated patients. Experiments have identified a proportion of TKI-naive tumors that carry T790M, and these resistant clones may be selected after exposure to TKIs<sup>7,8</sup>. The T790M mutation can coexist with other mutations, like L858R and D761Y. The T790M mutation also possesses enhanced phosphorylating activity, especially in combination with the L858R mutation. The combination leads to lung cancer cell survival, indicating that the T790M mutant is actually an oncogene<sup>9</sup>. Furthermore, cyclin D1 and Hsp90 may contribute to resistance in cancer cells harboring the T790M mutant by inhibiting the degradation of EGFR and maintaining the conformation of mutant EGFR<sup>10</sup>. Recently, the Hsp90 inhibitor ganetespib has been shown to enhance the anti-tumor effect of TKIs<sup>11</sup>.

## 2.2. Other secondary resistance mutations: L747S, D761Y and T854A

The non-T790M secondary mutations mainly include D761Y, L747S and T854A<sup>12-14</sup>. They reduce the sensitivity of mutant EGFR to EGFR-TKIs, but the resistance mechanism remains unknown. A possible explanation may be that these secondary

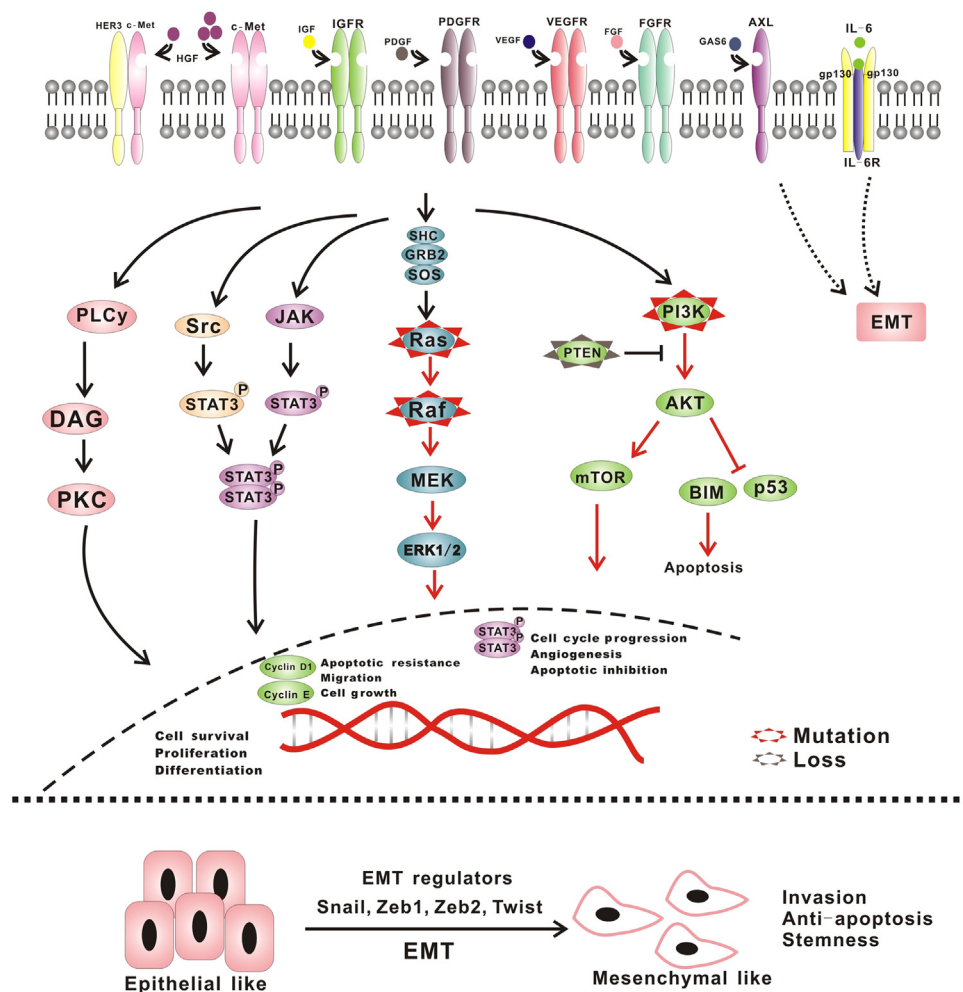
resistance mutations modify the conformation of EGFR and the combination between EGFR and TKIs. In addition, they may affect gefitinib-induced apoptosis and inhibit BIM up-regulation. Recently, another new insertion mutation on exon 20 of *EGFR* has been reported (Pro772\_His773insGlnCysPro)<sup>15,16</sup>. It was found in an individual who never smoked. The patient had previously been treated with cisplatin and gemcitabine, followed by carboplatin and pemetrexed. Finally, the patient developed resistance to erlotinib. Additional mutations still remain to be discovered. However, according to the reports of *EGFR*-mutated TKI-resistant patients, the frequency of non-T790M secondary mutations is low.

## 3. The aberrated activation of the bypass pathways induce resistance

The synchronous activation of redundant kinases also can induce resistance *via* activation of bypass pathways (Figs. 2 and 3). EGFR-TKI treatment of patients harboring such a change is not effective.

### 3.1. The aberrance of other members of HER family

The HER family is comprised of EGFR, HER2, HER3 and HER4. These receptors have a cytoplasmic TK domain which can be activated by ligand binding, followed by dimerization. Although



**Figure 3** Synchronous activation of redundant kinases and abnormality of the downstream pathway.

HER2 appears to have no intrinsic ligand-binding capability, it can interact reversibly with ligand-activated EGFR or HER3 to form active heterodimers which activate downstream signals to govern cell proliferation and migration. Overexpression of HER2 and mutations of *HER2* are involved in resistance of EGFR-TKIs<sup>17</sup>. Therefore, HER2 is a useful target for treatment. A covalent, irreversible inhibitor of HER2, afatinib, can overcome the resistance of the patient harboring HER2 overexpression or *HER2* gene mutations<sup>18</sup>. Compared to the other HER proteins, there are currently no mutational alterations known to confer oncogenic activities to HER3, but HER3 also takes part in resistance of EGFR-TKIs. HER3 phosphorylation is driven by one of the other HER family kinase partners, like HER1 and HER2, or through amplification of the proto-oncogene *c-Met*<sup>19</sup>. HER3 serves as a key activator of downstream PI3K/AKT and MEK/mitogen-activated protein kinase (MAPK) pathways and contributes to survival of the most tumor cells. However, researchers have found that a heregulin–EGFR–HER3 autocrine signaling axis mediated acquired lapatinib resistance in *HER2*<sup>+</sup> breast cancer models which no longer depended on HER2–HER3–PI3K signal pathway<sup>20</sup>. What's more, overexpression of HER3 in COLM-5 cells can lead to significant resistance to gefitinib *in vitro* and *in vivo*<sup>21</sup>. All of these studies highlight the central role of HER3 in cancers. Targeting of HER3 receptor with a monoclonal antibody, such as

MM-121 or MM-111 is an effective strategy currently under preclinical study and clinical evaluation<sup>22</sup>.

### 3.2. Amplification of *c-Met*

*c-Met* is a transmembrane RTK. Binding with its ligand, hepatocyte growth factor (HGF) triggers receptor dimerization and phosphorylation, leading to conformational changes of *c-Met* that activates the TK domain and activates a wide range of different cellular signal pathways, including those involved in proliferation, motility, migration and invasion. Although *c-Met* is important in the control of tissue homeostasis under normal physiological conditions, it has also been found to be aberrantly activated in human cancers *via* gene mutation, amplification or protein overexpression. In 2007, the second most common primary resistance to EGFR-TKIs involved amplification of the *c-Met* oncogene in HCC827 NSCLC cells after exposure to gefitinib. In *c-Met*-resistant patients, *c-Met* amplified clones existed prior to EGFR-TKIs therapy and were selected out by treatment<sup>23</sup>. The resistant mechanism of EGFR-TKIs may involve HER3-PI3K/AKT signaling by maintaining HER3 phosphorylation in the presence of gefitinib, which is independent of EGFR kinase activity<sup>19</sup>. Besides, a high *MET* gene copy number leads to shorter survival



in patients with NSCLC. Clinical trials have demonstrated that concurrent inhibition of EGFR and c-Met can overcome resistance of EGFR-TKIs and improve patient outcomes.

### 3.3. Overexpression of HGF

HGF is the ligand for c-Met. HGF acts as a pleiotropic factor and cytokine, promoting cell proliferation, survival, motility, scattering, differentiation and morphogenesis.

Overexpression of HGF is another EGFR-TKIs resistance mechanism and it may be more common among patients with mutations who had no response<sup>24</sup>. One study showed that HGF induces resistance by activating c-Met which restores phosphorylation of downstream MAPK/extracellular signal-regulated kinases (ERK1/2) and PI3K/AKT pathways<sup>25</sup>. Interestingly, although amplification of *c-Met* activates downstream pathways by activating ERBB3, HGF induces downstream pathways through c-Met; this activation is independent of ERBB3 or EGFR. The resistance induced by HGF not only appears in the NSCLC, but also exists in breast cancers. Blockade of EGFR and the downstream pathways can overcome HGF-mediated resistance.

### 3.4. The abnormality of insulin growth factor receptor (IGFR)

As early as 2002, it had been suggested that IGF-1 receptor (IGF-1R) signaling through PI3K may represent a novel and potentially important mechanism of resistance to anti-EGFR therapy. In 2008, researchers found that the loss of expression of IGF-binding proteins (IGFBPs) in tumor cells treated with EGFR-TKIs increased the activation of IGF-1R signaling which, in turn, mediates resistance of EGFR-TKIs<sup>26,27</sup>. Inhibition of IGF-1R signaling disrupted the association of IRS-1 with PI3K and restored the ability of gefitinib to down-regulate PI3K/AKT signaling and to inhibit cell growth. Concomitant inhibition of both EGFR and IGF1R was required to abort PI3K signaling, and treatment of the resistant cells with an IGF1R inhibitor restored their sensitivity to EGFR TKIs. Another study found that activation of IGF-1R can alter phosphorylation state and subcellular localization of p27, which can promote cell proliferation and cell motility<sup>28</sup>. Thus, IGF-1R and p27 can be used to be a biomarker of cell cycle arrest and response to therapy.

### 3.5. The abnormality molecules of multiple angiogenic pathways

Angiogenesis is an essential step in tumor growth and metastasis. Impaired vascularity and hypoxia can lead to increased metastasis and treatment resistance. Thus, targeting multiple angiogenic pathways may not only improve antitumor activity but also reduce the risk of resistance<sup>29</sup>. Important targets for the development of novel antiangiogenic therapies include vascular endothelial growth factors (VEGFs), fibroblast growth factors (FGFs), platelet-derived growth factors (PDGFs), and their receptors.

#### 3.5.1. The VEGFs and their receptors

The VEGF family and receptors are important regulators of angiogenesis and vascular permeability. Overexpression of vascular endothelial growth factor receptor (VEGFR) 1 in tumor cells leads to cell survival and invasion. It also reduces the inhibition by EGFR inhibitors and reduces sensitivity to gefitinib. Activation of the EGFR signal can increase the expression of VEGF. Researchers have demonstrated that a VEGF/VEGFR2 feed-forward loop in

NSCLC cells expressing VEGFR2, which leads to a signal amplification and a boost in VEGF secretion, is required for establishment of fully angiogenic tumors *in vivo*. This VEGF/VEGFR2 signaling cascade *via* VEGFR2/PI3K/mTOR induces an mTOR-dependent regulation of VEGF secretion<sup>30</sup>. And VEGF secretion is induced by the upregulation of HIF-1 $\alpha$ . Thus, the VEGF/VEGFR system is associated with resistance of anti-EGFR drugs through activation of downstream signal pathways *via* EGFR-independent mechanisms<sup>31</sup>. The therapeutic use of agents able to inhibit both EGFR and VEGFR may help to efficiently inhibit the activation of bypass pathways and overcome EGFR inhibitor resistance.

#### 3.5.2. The fibroblast growth factors and their receptors

The FGFs and the receptors (FGFRs) are involved in multiple cellular functions. During embryonic development, FGFs play a part in morphogenesis. In adults, FGFs are involved in wound healing and tissue repair as well as regulating the nervous system. FGFs also participate in tumor angiogenesis. FGFR autocrine signaling has been implicated in NSCLC cell lines. In 2010, researchers found EGFR-TKIs (gefitinib) increased the expression of FGFR2 and FGFR3. Importantly, FGFR2 and FGFR3 are capable of mediating FGF2 and FGF7 stimulated ERK activation, leading to cell survival and invasion and reducing the sensitivity to EGFR-TKIs<sup>32</sup>. FGFR activation is an escape mechanism in human lung cancer cells resistance to afatinib, that may compensate for the loss of EGFR-driven signaling pathway<sup>33</sup>. Treatment of NSCLC patients with combinations of EGFR and FGFR specific inhibitors (*e.g.*, PD173074) or FGF antibodies may be potential strategy to enhance efficacy of single EGFR-TKIs.

#### 3.5.3. The platelet-derived growth factors and their receptors (PDGFRs)

PDGFs are isolated from human platelets and promote angiogenesis. As angiogenesis factors, PDGFs/PDGFRs are closely related to tumor development. PDGFs/PDGFRs can induce tumor cell proliferation and migration, and inhibit apoptosis. In 2013, Akhavan et al.<sup>34</sup> found that a physiologic RTK switch to the PDGFR $\beta$  was required to maintain the growth of EGFR variant III (EGFRvIII)/EGFR-activated in response to EGFR-TKIs. Combination of EGFR and PDGFR inhibitors may overcome resistance of EGFR-TKIs.

### 3.6. EGFRvIII

*EGFRvIII* is a tumor-specific mutation that results from in-frame deletion of 801 base pairs spanning exons 2–7 of the coding sequence. This deletion removes 267 amino acids from the extracellular domain, creating a junction site between exons 1 and 8 and a new glycine residue. EGFRvIII has expressed in many kinds of tumors.

EGFRvIII confers enhanced tumorigenicity through multiple mechanisms and pathways. EGFRvIII expression is associated with the activation of downstream PI3K/AKT/mTOR pathway and increases proliferation and cell cycle progression mediated by a decrease in the level of p27<sup>KIP1</sup>. EGFRvIII also has been shown to activate the NF- $\kappa$ B pathway and regulate expression of IL-8<sup>35</sup>. Furthermore, EGFRvIII induces angiopoietin-like 4 expression through the ERK/c-Myc pathway and promotes tumor angiogenesis in malignant gliomas<sup>36</sup>. Cells harboring EGFRvIII have an enhanced capacity for dysregulated growth, survival, invasion and angiogenesis. EGFRvIII is an attractive

target and predictor in cancer immunotherapy because it is not expressed in normal tissue.

### 3.7. Overexpression of or overactivated Anexelekto (AXL)

AXL belongs to the Tyro/Axl/Mer (TAM) family of RTK. Growth-arrest-specific protein 6 (GAS6) is a ligand for AXL. Activation of AXL increases cell migration, aggregation and growth through multiple downstream pathways. AXL was overexpressed in *EGFR*-mutant NSCLC tumor xenografts with acquired resistance to erlotinib<sup>37</sup>. The researchers suggested that the AXL upregulation may activate AKT, MAPK or NF- $\kappa$ B signaling to promote resistance to erlotinib, perhaps in association with epithelial mesenchymal transition (EMT) process *via* inducing Slug expression<sup>38</sup>. Inhibitors of AXL (MP-470 or XL-880) enhance erlotinib sensitivity particularly in the context of NSCLC with acquired resistance to EGFR-targeting TKIs. Therefore, the inhibitors of AXL or GAS6 may overcome the AXL-mediated resistance of EGFR-TKIs.

### 3.8. Excess secretion of interleukin-6 (IL-6)

IL-6 is a cytokine which plays an important role in many chronic inflammatory diseases. Binding to the membrane-bound IL-6 receptor (IL-6R) causes the recruitment of two gp130 co-receptor molecules and the activation of intracellular signaling cascades *via* gp130. However, some reports suggest that IL-6 expression is involved in the regulation of tumor growth and metastatic spread, including lung cancers. One study demonstrated that activation of the IL-6R/JAK1/STAT3 pathway induced *de novo* resistance to irreversible EGFR-TKIs in NSCLC harboring T790M<sup>39</sup>. Afatinib activated IL-6R/JAK1/STAT3 signaling *via* autocrine IL-6 production<sup>40</sup>. Inhibition of the IL-6R/JAK1/STAT3 signal pathway can reverse the resistance. JAK1 activates STAT3 activity which is stopped rapidly by the major negative regulator suppressor of cytokine signaling 3 (SOCS3) in normal cells. However, association of the IL-6 receptor with the EGFR can activate STAT3 in presence of the inhibition of SOCS3 to JAK1 and JAK2<sup>41</sup>. Besides, paracrine or autocrine stimulation of the TGF- $\beta$  axis also increases the secretion of IL-6 and promotes resistance<sup>42</sup>.

### 3.9. Amplification of Crk-like protein (CRKL)

CRKL is a member of adapter proteins that participates in signal transduction in response to both extracellular and intracellular stimuli, such as growth factors, cytokines and the oncogenic BCR-ABL fusion protein. Oncogenic CRKL activates the SOS1/RAS/RAF/ERK and SRC/C3G/RAP1 pathways. Amplification of the *CRKL* gene was observed in 1/11 lung cancer patients with *EGFR* mutations who acquired resistance to EGFR-TKIs<sup>43</sup>. Amplification of *CRKL* in *EGFR*-mutant cells induces resistance to gefitinib by activating ERK and AKT signaling<sup>44</sup>. What's more, overexpression of CRKL promotes cell invasion *via* upregulating MMP9 expression and activating ERK pathway<sup>45</sup>. Therefore, CRKL can be a potential target in the treatment of NSCLC.

### 3.10. Overexpression and activation of integrin $\beta$ 1

Beta1 subunit of integrin is an adhesion molecule, sharing common downstream signaling elements with EGFR, such as the PI3K/AKT and ERK1/2 pathways. Overexpression of integrin

$\beta$ 1 induces gefitinib resistance, accompanied by increases in cell adhesion and migration<sup>46</sup>. Subsequently, researchers found that  $\beta$ 1 integrin activated an alternative survival pathway in breast cancer cells harboring resistance to lapatinib, which led to activation of  $\beta$ 1 integrin's downstream kinases, FAK and SRC. Inhibition of  $\beta$ 1 integrin by AIIB2 can enhance the sensitivity to EGFR-TKIs<sup>47</sup>. Further study found that ligand-dependent activation of integrin  $\beta$ 1 could induce EGFR-TKIs resistance through activating c-Met and downstream pathways<sup>48</sup>. Kanda et al.<sup>49</sup> identified the integrin  $\beta$ 1/Src/AKT signal pathway as a key mediator of acquired resistance to EGFR-targeted anticancer drugs. Researchers have also found that integrin mediates a stem-like phenotype and confers resistance to EGFR-targeted therapy through enhancing downstream coupling to a KRAS/RalB/NF- $\kappa$ B pathway<sup>50</sup>. Therefore, integrin  $\beta$ 1 and the pathway molecules provide potential agents for overcoming the resistance mediated by integrin  $\beta$ 1.

## 4. Abnormal downstream pathways induce drug resistance

The abnormal downstream pathways also can result in the resistance to EGFR-TKIs, even though the cells have not harbored any other mutations. The mutation of *K-RAS*, the loss of phosphatase and tensin homolog (*PTEN*), the mutations of *PIK3CA* and *BRAF* are the main points (Fig. 3).

### 4.1. K-RAS mutation

The RAS proteins include K-RAS, N-RAS and H-RAS. The RAS proteins are GTPases that are molecular switches for a variety of critical cellular activities and their function is tightly and temporally regulated in normal cells. Oncogenic mutations of *RAS* genes, which create constitutively-active RAS proteins, can result in uncontrolled proliferation or survival in tumor cells. In 2005, Pao et al.<sup>51</sup> showed that mutations in *K-RAS* are associated with primary resistance to single-agent gefitinib or erlotinib. If the mutations occur at codons 12, 13, 59, 61, 63, 116, 117, 119 or 146, its structure is altered by binding sites for guanine, affecting normal function. The effects of these mutations can be translated either in a reduction of the activity of oncoprotein GTPases, blocking it into the active form bound to GTP, or in decreased binding affinity and increasing the change in GDP by GTP attachment<sup>52</sup>. In total, 80% of *K-RAS* mutations occur in codon 12, and other mutations are mainly located in codons 13 and 61. *RAS* gene alteration is poor prognostic factors for survival of patients with NSCLC. Additionally, several studies have clearly demonstrated that RAS uses additional effectors to promote tumorigenesis<sup>53</sup>. Very little is known about the relevant mechanisms.

### 4.2. Loss of PTEN

*PTEN* dephosphorylates PI-(3, 4, 5)-triphosphate, which mediates activation of AKT, thereby negatively regulating the PI3K/AKT/mTOR pathway and leading to G1 cell cycle arrest and apoptosis. In addition, *PTEN* inhibits cell migration and spreading through regulation of focal adhesion kinase as well as regulates p53 protein levels and activity.

*PTEN* deleted on chromosome 10 is a tumor suppressor gene on chromosome 10q23.3 and encodes a 403 amino acid dual-specificity lipid and protein phosphatase. The loss of *PTEN* has only been investigated in a small number of NSCLC cases. However, *PTEN* loss contributes to erlotinib resistance in *EGFR* mutant lung cancer by activation of AKT and EGFR<sup>54,55</sup>. The

absence of PTEN protein expression is an independent prognostic marker in early stage resected lung adenocarcinoma. Inhibition of the PI3K/AKT/mTOR signal pathway can be an effective strategy to NSCLC harboring the *EGFR* activating mutations that acquires resistance to both TKIs and radiotherapy due to *PTEN* loss<sup>56</sup>.

#### 4.3. Mutations of *BRAF* (*v-RAF* murine sarcoma viral oncogene homolog B1)

*BRAF*, another component of the *EGFR*/RAS/RAF signal transduction pathway, encodes a RAS-regulated kinase that mediates cell growth, differentiation, apoptosis and malignant transformation. Mutations of *BRAF* (G469A, V600E and V599E) were found in several tumors, including malignant melanoma and colorectal cancer. Thus, *BRAF* mutations also induce drug resistance<sup>57</sup>. Activating *BRAF* mutations, especially the common mutant V599E, induces constitutive activation of the signal transduction pathway, providing a potent pro-mitogenic force that drives malignant transformation. The *BRAF* V599E mutant shows greatly increased activity in the RAF/MEK/ERK pathway both *in vitro* and *in vivo*. Here, *BRAF* provides a new target for the treatment to overcome the mutant *BRAF*-mediated resistance.

#### 4.4. A downstream mutation in *PIK3CA*

The *PIK3CA* gene, encoding a catalytic subunit of the PI3K, is mutated or amplified in various neoplasias, including lung cancer. Infrequently, a downstream mutation in *PIK3CA* has been identified as a mechanism of resistance<sup>58</sup>. Recently, a study

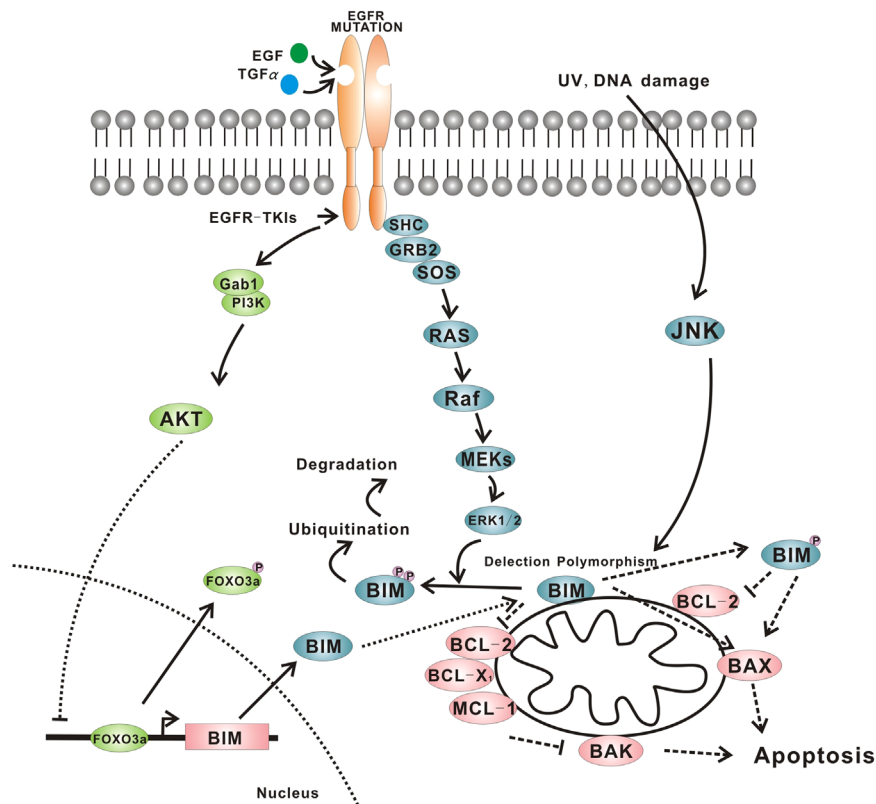
demonstrated that *PIK3CA* mutations frequently coexist with *EGFR* or *K-RAS* mutations<sup>59</sup>. Patients with single *PIK3CA* mutation in NSCLC have poor prognosis. The role of mutant *PIK3CA* in oncogenic signaling requires further investigation, including development of novel targets for therapeutic intervention in cancers harboring *PIK3CA* mutations.

#### 4.5. The aberrant expression of *NF1*

Neurofibromin, the RAS GTPase-activating protein, is encoded by *NF1* gene. A recent study demonstrated that reduced expression of *NF1* was associated with erlotinib resistance due to a failure to fully inhibit RAS/RAF/MEK/ERK pathway<sup>60</sup>. Combination therapy with *EGFR* and MEK inhibitors may restore sensitivity to *EGFR*-TKIs.

### 5. Impairment of a pathway that is essential for *EGFR*-TKIs-mediated apoptosis: a common intrinsic deletion polymorphism in the gene encoding *BIM*

*BIM* is a pro-apoptotic member of the B-cell CLL/lymphoma-2 (*BCL-2*) family, and plays a critical role in inducing cell apoptosis and tumor metastasis. Consequently, *BIM* has become the focus of intense interest as a potential target for cancer chemotherapy. Its upregulation is required for apoptosis induced by *EGFR* and *EGFR*-TKIs in tumors harboring *EGFR* mutations. The polymorphism switched *BIM* splicing from exon 4 to exon 3, which resulted in expression of *BIM* isoforms lacking the pro-apoptotic *BCL2*-homology domain 3 (*BH3*)<sup>61</sup> (Fig. 4). Although the polymorphism was sufficient to confer intrinsic TKI resistance in



**Figure 4** Apoptosis pathway mediated by *BIM*.



chronic myelogenous leukemia (CML) and EGFR NSCLC cell lines, this resistance could be overcome with BH3-mimetic drugs. Recently, researchers have shown that the PP2A activator FTY720 could induce apoptosis of CML cells *via* dual activation of BIM and BID and overcome various types of resistance of TKIs<sup>62</sup>.

### 6. Histologic transformation

#### 6.1. The EMT

EMT is a physiological process during embryogenesis that appears to be reinstated in adult tissues undergoing wound healing and tissue regeneration, or under certain pathological conditions, such as fibrosis and cancer. EMT is characterized by the combined loss of epithelial cell junction proteins, such as E-cadherin, and the gain of mesenchymal markers, such as vimentin and N-cadherin<sup>63</sup>. In the EMT process, epithelial cells lose their features, gain properties of mesenchymal, and become motile and invasive (Fig. 3).

In 2005, a transition to a mesenchymal phenotype was noted among patients treated with EGFR-TKIs. EMT may be induced by the activation of AXL *via* the PI3K/AKT pathway. One study also showed that loss of E-cadherin can activate EGFR–MEK/ERK/ZEB1/MMP2 axis, which is responsible for promoting invasion in NSCLC<sup>64</sup>. Tumor cells undergoing EMT are also known to increase the secretion of specific factors, including cytokines, chemokines and growth factors, which could play an important role in tumor progression. Well-established signals include those initiated by

TGF- $\beta$ , FGF, EGF and HGF, all of which have been shown to promote EMT in various tumor cell models. Other possible pathways or factors that have been reported to be associated with EMT include IL-8, IL-6, Notch-1, SOX9, FoxO4, SRC and CRIPTO-1<sup>65–69</sup>. Co-targeting the relative molecules of these pathways and EGFR can reverse the resistance mediated by EMT<sup>70</sup>. However, the specific mechanism of EMT still remains unknown.

#### 6.2. Small cell transformation

Conversion to small-cell carcinoma has been seen at the time of development of resistance. Tumor cells retained the original EGFR mutations but developed a histopathologic small-cell phenotype, which may benefit from a standard chemotherapy for small cell lung cancer<sup>71</sup>. Interestingly, after a period of conventional cytotoxic treatment, susceptibility to TKIs may redevelop. Research into this phenomenon is still insufficient.

### 7. ATP binding cassette (ABC) effusion

The ABC transporters are transmembrane proteins that involved in transporting biologically important substrates across the cellular membranes, such as amino acids, cholesterol, hydrophobic drugs and antibiotic. Overexpression of ABC transporters can reduce drug uptake, increase drug efflux and lead to low drug density in the cytoplasm, which will come to a lower drug efficacy and finally acquire drug resistance. When EGFR-TKIs (lapatinib)

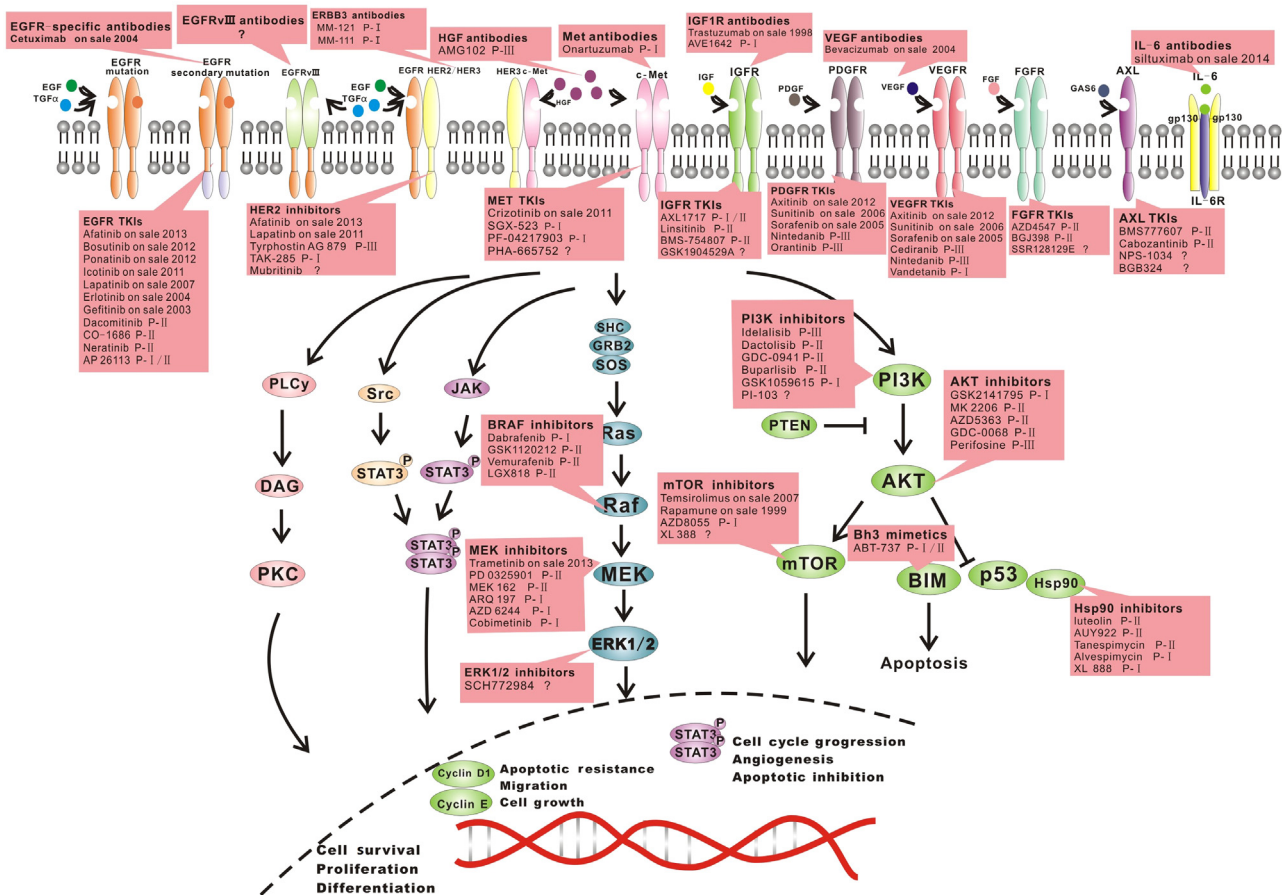


Figure 5 The potential targets for the relative treatment to overcome the resistance to EGFR-TKIs.



binds to the ATP binding cassette, sub-family B, member 1 (ABCB1) and ATP binding cassette, sub-family G, member 2 (ABCG2), substrate-binding sites with high affinity induced over-expression of ABC transporters<sup>72</sup>. Other ABC transporters, such as ATP binding cassette, sub-family C, member 1 (ABCC1) and ABCC10 also participate in drug resistance. Given the ABC transporter influence on TKI actions, ABC transporter inhibitors may reverse the resistance. In addition, researchers<sup>73</sup> found that GW583340 and GW2974, EGFR and HER-2 inhibitors can reverse ABCG2- and ABCB1-mediated drug resistance.

### 8. Echinoderm microtubule-associated protein-like 4-the anaplastic lymphoma kinase (*EML4-ALK*) fusion gene and the *ALK* secondary mutation

The *EML4-ALK* fusion oncogene was identified as a novel genetic alteration in NSCLC<sup>74</sup>. Patients harboring *ALK*

rearrangements tend to be non-smokers or light smokers, have a history of adenocarcinoma, and tend to be younger in age<sup>75</sup>. The *EML4-ALK* fusion gene was present at a high frequency in Chinese NSCLC patients, particularly in those with adenocarcinomas lacking *EGFR/K-RAS* mutations<sup>76</sup>. Recently, crizotinib was identified as a potent inhibitor of *ALK* and *MET* tyrosine kinases<sup>77</sup>. Crizotinib was well tolerated and resulted in important tumor shrinkage in NSCLC *EML4-ALK* positive patients. However, some patients have resistance to crizotinib and other *EML4-ALK* inhibitors are in development. Researchers<sup>78,79</sup> found that the naive NSCLC patients with *ALK* rearrangements also had concurrent *EGFR* activating mutations. The resistance mechanisms to *ALK* TKIs may be mediated by both *ALK* secondary mutation and a bypass signaling pathway such as *EGFR*. These mechanisms can occur independently, or in the same cancer. Combination therapy with *EGFR*-TKIs and *ALK* inhibitors can improve the anti-tumor effect.

**Table 1** Therapeutic strategies and clinical trials to overcome resistance of *EGFR*-TKIs, "-" stands for no drugs for therapy.

Resistant mechanism	Strategy	Clinical research	Ref.
<b><i>EGFR</i> mutation</b>			
T790M	EGFR-TKIs combined/+antibodies	Afatinib+cexitumab	80
	T790M-specific inhibitors	CO-1686/AZD9291	81,82
	c-Met inhibitors+PI3K inhibitors	GDC0973+GDC0941	83
	HSP90 inhibitors	Luteolin/ganetespib	11,84
	EGFR-TKIs+MEK inhibitors	Afatinib+ARQ 197	85
	Glycolysis inhibition+EGFR-TKIs	Afatinib+AUY922	86
<b>Bypass pathway</b>			
HER family abnormality	HER inhibitors+EGFR-TKIs	Afatinib/dacomitinib	18,87
<i>c-Met</i> amplification	EGFR-TKIs+c-Met inhibitors	Erlotinib+crizotinib	88
		Dacomitinib+crizotinib	
HGF overexpression	EGFR-TKIs+PI3K inhibitors	Gefitinib+PI-103	89
	Triple inhibition of <i>EGFR/Met/VEGF</i>	-	90
IGFR abnormality	IGFR inhibitors+EGFR-TKIs	AG1024+gefitinib	-
EGFRvIII	EGFRvIII antibodies	-	-
VEGF/VEGFR abnormality	EGFR-TKIs+VEGF inhibitors	ZD6474	91
	MEK inhibitors+VEGF inhibitors	ZD6474+PD0325901	30
PDGF/PDGFR abnormality	EGFR-TKIs+PDGF inhibitors	-	-
FGF/FGFR abnormality	EGFR-TKIs+FGF inhibitors	-	-
IL-6 abnormality	IL-6 antibodies	Siltuximab	92
AXL abnormality	AXL inhibitors	NPS-1034	93
<i>CRKL</i> amplification	Unknown	Unknown	-
Integrin beta1 overexpression	Unknown	Unknown	-
<b>Downstream pathway</b>			
<i>K-RAS</i> mutations	PI3K inhibitors+MEK inhibitors	GDC-0941+AZD6244	94
<i>BRAF</i> mutations	<i>BRAF</i> inhibitors+MEK inhibitors	Dabrafenib+trametinib	95
Loss of <i>PTEN</i>	mTOR inhibitors/AKT inhibitors	-	-
<i>PIK3CA</i> mutation	EGFR-TKIs+PI3K inhibitors	Gefitinib+BKM120	96
Low expression of NF1	Unknown	Unknown	-
<b>Apoptosis pathway</b>			
<i>BIM</i> BH3 deletion	EGFR-TKIs+PP2A activator	Erlotinib+FTY720 gefitinib+FTY720	62
<b>Histologic transformation</b>			
EMT	EGFR-TKIs+MEK1/2 inhibitors	-	-
SCLC transformation	Platinum+VP16/EGFR-TKIs	-	-
<b>ABC effusion</b>			
	EGFR-TKIs+HER-2 inhibitors	GW583340/GW2974	73
<b>Unknown mechanism</b>			
	EGFR-TKIs combined	Afatinib+cexitumab	80
	EGFR-TKIs+glycolysis inhibitors	Erlotinib+AUY922	

## 9. Therapeutic strategies to overcome resistance

The existence of various resistance mechanisms is clear, and it is wise to identify the specific mechanisms in a patient so that a suitable, effective strategy can be chosen. The next generation of EGFR-TKIs and specific antibodies of the relative molecules are under development. MicroRNAs will soon be employed in treating resistance to EGFR-TKIs in cancers. Clinical trials are engaged in examining the potential targets less susceptible to resistance. The relative clinical trials or therapeutic strategies are listed in the table (Table 1<sup>11,18,30,62,73,80–96</sup> and Fig. 5). In practice, combined treatments or therapies with multiple targets may show more powerful efficacy, and the multi-targeting drugs may show superior efficacy.

## 10. Conclusions

Increasing evidence shows that the primary or acquired resistance to first- or second-generation EGFR-TKIs explains why patients who initially benefited from these treatments later do not. Though some of the mechanisms of resistance have been identified, much additional information is needed to understand and overcome resistance to these agents.

## Acknowledgment

The work was supported by the National Natural Science Foundation of China (No. 81473233).

## References

- Ciardiello F, Tortora G. EGFR antagonists in cancer treatment. *N Engl J Med* 2008;**358**:1160–74.
- Bethune G, Bethune D, Ridgway N, Xu Z. Epidermal growth factor receptor (EGFR) in lung cancer: an overview and update. *J Thorac Dis* 2010;**2**:48–51.
- Yu HA, Arcila ME, Rekhtman N, Sima CS, Zakowski MF, Pao W, et al. Analysis of tumor specimens at the time of acquired resistance to EGFR-TKI therapy in 155 patients with EGFR-mutant lung cancers. *Clin Cancer Res* 2013;**19**:2240–7.
- Kobayashi S, Boggon TJ, Dayaram T, Jänne PA, Koehler O, Meyerson M, et al. EGFR mutation and resistance of non-small-cell lung cancer to gefitinib. *N Engl J Med* 2005;**352**:786–92.
- Yun CH, Mengwasser KE, Toms AV, Woo MS, Greulich H, Wong KK, et al. The T790M mutation in EGFR kinase causes drug resistance by increasing the affinity for ATP. *Proc Natl Acad Sci U S A* 2008;**105**:2070–5.
- Ma C, Wei S, Song Y. T790M and acquired resistance of EGFR TKI: a literature review of clinical reports. *J Thorac Dis* 2011;**3**:10–8.
- Fujita Y, Suda K, Kimura H, Matsumoto K, Arao T, Nagai T, et al. Highly sensitive detection of EGFR T790M mutation using colony hybridization predicts favorable prognosis of patients with lung cancer harboring activating EGFR mutation. *J Thorac Oncol* 2012;**7**:1640–4.
- Su KY, Chen HY, Li KC, Kuo ML, Yang JC, Chan WK, et al. Pretreatment epidermal growth factor receptor (EGFR) T790M mutation predicts shorter EGFR tyrosine kinase inhibitor response duration in patients with non-small-cell lung cancer. *J Clin Oncol* 2012;**30**:433–40.
- Suda K, Onozato R, Yatabe Y, Mitsudomi T. EGFR T790M mutation: a double role in lung cancer cell survival? *J Thorac Oncol* 2009;**4**:1–4.
- Hong YS, Jang WJ, Chun KS, Jeong CH. Hsp90 inhibition by WK88-1 potently suppresses the growth of gefitinib-resistant H1975 cells harboring the T790M mutation in EGFR. *Oncol Rep* 2014;**31**:2619–24.
- Smith DL, Acquaviva J, Sequeira M, Jimenez JP, Zhang C, Sang J, et al. The HSP90 inhibitor ganetespib potentiates the antitumor activity of EGFR tyrosine kinase inhibition in mutant and wild-type non-small cell lung cancer. *Target Oncol* 2015;**10**:235–45.
- Costa DB, Halmos B, Kumar A, Schumer ST, Huberman MS, Boggon TJ, et al. BIM mediates EGFR tyrosine kinase inhibitor-induced apoptosis in lung cancers with oncogenic EGFR mutations. *PLoS Med* 2007;**4**:1669–79.
- Toyooka S, Date H, Uchida A, Kiura K, Takata M. The epidermal growth factor receptor D761Y mutation and effect of tyrosine kinase inhibitor. *Clin Cancer Res* 2007;**13**:3431.
- Bean J, Riey GJ, Balak M, Marks JL, Ladanyi M, Miller VA, et al. Acquired resistance to epidermal growth factor receptor kinase inhibitors associated with a novel T854A mutation in a patient with EGFR-mutant lung adenocarcinoma. *Clin Cancer Res* 2008;**14**:7519–25.
- Woo HS, Ahn HK, Lee HY, Park I, Kim YS, Hong J, et al. Epidermal growth factor receptor (EGFR) exon 20 mutations in non-small-cell lung cancer and resistance to EGFR-tyrosine kinase inhibitors. *Invest New Drugs* 2014;**32**:1311–5.
- Khan NA, Mirshahidi S, Mirshahidi HR. A novel insertion mutation on exon 20 of epidermal growth factor receptor, conferring resistance to erlotinib. *Case Rep Oncol* 2014;**7**:491–6.
- Landi L, Cappuzzo F. HER2 and lung cancer. *Expert Rev Anticancer Ther* 2013;**13**:1219–28.
- Nishida Y, Kuwata T, Nitta H, Dennis E, Aizawa M, Kinoshita T, et al. A novel gene–protein assay for evaluating HER2 status in gastric cancer: simultaneous analyses of HER2 protein overexpression and gene amplification reveal intratumoral heterogeneity. *Gastric Cancer* 2014 Jun 11 <http://dx.doi.org/10.1007/s10120-014-0394-7>.
- Engelman JA, Zejnullahu K, Mitsudomi T, Song Y, Hyland C, Park JO, et al. MET amplification leads to gefitinib resistance in lung cancer by activating ERBB3 signaling. *Science* 2007;**316**:1039–43.
- Xia W, Petricoin 3rd EF, Zhao S, Liu L, Osada T, Cheng Q, et al. An heregulin–EGFR–HER3 autocrine signaling axis can mediate acquired lapatinib resistance in HER2<sup>+</sup> breast cancer models. *Breast Cancer Res* 2013;**15**:R85.
- Nakata S, Tanaka H, Ito Y, Hara M, Fujita M, Kondo E, et al. Deficient HER3 expression in poorly-differentiated colorectal cancer cells enhances gefitinib sensitivity. *Int J Oncol* 2014;**45**:1583–93.
- Schoeberl B, Faber AC, Li D, Liang MC, Crosby K, Onsum M, et al. An ErbB3 antibody, MM-121, is active in cancers with ligand-dependent activation. *Cancer Res* 2010;**70**:2485–94.
- Kubo T, Yamamoto H, Lockwood WW, Valencia I, Soh J, Peyton M, et al. MET gene amplification or EGFR mutation activate MET in lung cancers untreated with EGFR tyrosine kinase inhibitors. *Int J Cancer* 2009;**124**:1778–84.
- Yano S, Wang W, Li Q, Matsumoto K, Sakurama H, Nakamura T, et al. Hepatocyte growth factor induces gefitinib resistance of lung adenocarcinoma with epidermal growth factor receptor-activating mutations. *Cancer Res* 2008;**68**:9479–87.
- Mueller KL, Madden JM, Zoratti GL, Kuperwasser C, List K, Boerner JL. Fibroblast-secreted hepatocyte growth factor mediates epidermal growth factor receptor tyrosine kinase inhibitor resistance in triple-negative breast cancers through paracrine activation of Met. *Breast Cancer Res* 2012;**14**:R104.
- Guix M, Faber AC, Wang SE, Olivares MG, Song Y, Qu S, et al. Acquired resistance to EGFR tyrosine kinase inhibitors in cancer cells is mediated by loss of IGF-binding proteins. *J Clin Invest* 2008;**118**:2609–19.
- Peled N, Wynnes MW, Ikeda N, Ohira T, Yoshida K, Qian J. Insulin-like growth factor-1 receptor (IGF-1R) as a biomarker for resistance to the tyrosine kinase inhibitor gefitinib in non-small cell lung cancer. *Cell Oncol* 2013;**36**:277–88.
- Jameson MJ, Taniguchi LE, VanKoeveering KK, Stuart MM, Francom CR, Mendez RE, et al. Activation of the insulin-like growth factor-1 receptor alters p27 regulation by the epidermal growth factor receptor in oral squamous carcinoma cells. *J Oral Pathol Med* 2013;**42**:332–8.

29. Ballas MS, Chachoua A. Rationale for targeting VEGF, FGF, and PDGF for the treatment of NSCLC. *Onco Targets Ther* 2011;**4**:43–58.
30. Chatterjee S, Heukamp LC, Siobal M, Schöttle J, Wieczorek C, Peifer M, et al. Tumor VEGF:VEGFR2 autocrine feed-forward loop triggers angiogenesis in lung cancer. *J Clin Invest* 2013;**123**:1732–40.
31. Bianco R, Rosa R, Damiano V, Daniele G, Gelardi T, Garofalo S, et al. Vascular endothelial growth factor receptor-1 contributes to resistance to anti-epidermal growth factor receptor drugs in human cancer cells. *Clin Cancer Res* 2008;**14**:5069–80.
32. Ware KE, Marshall ME, Heasley LR, Marek L, Hinz TK, Hercule P, et al. Rapidly acquired resistance to EGFR tyrosine kinase inhibitors in NSCLC cell lines through de-repression of FGFR2 and FGFR3 expression. *PLoS One* 2010;**5**:e14117.
33. Azuma K, Kawahara A, Sonoda K, Nakashima K, Tashiro K, Watari K, et al. FGFR1 activation is an escape mechanism in human lung cancer cells resistant to afatinib, a pan-EGFR family kinase inhibitor. *Oncotarget* 2014;**5**:5908–19.
34. Akhavan D, Pourzia AL, Nourian AA, Williams KJ, Nathanson D, Babic I, et al. De-repression of PDGFR $\beta$  transcription promotes acquired resistance to EGFR tyrosine kinase inhibitors in glioblastoma patients. *Cancer Discov* 2013;**3**:534–47.
35. Bonavia R, Inda MM, Vandenberg S, Cheng SY, Nagane M, Hadwiger P, et al. EGFRVIII promotes glioma angiogenesis and growth through the NF- $\kappa$ B, interleukin-8 pathway. *Oncogene* 2012;**31**:4054–66.
36. Katanasaka Y, Koderia Y, Kitamura Y, Morimoto T, Tamura T, Koizumi F. Epidermal growth factor receptor variant type III markedly accelerates angiogenesis and tumor growth via inducing c-myc mediated angiopoietin-like 4 expression in malignant glioma. *Mol Cancer* 2013;**12**:31.
37. Zhang Z, Lee JC, Lin L, Olivas V, Au V, LaFramboise T, et al. Activation of the AXL kinase causes resistance to EGFR-targeted therapy in lung cancer. *Nat Genet* 2012;**44**:852–60.
38. Lee Y, Lee M, Kim S. GAS6 induces cancer cell migration and epithelial–mesenchymal transition through upregulation of MAPK and Slug. *Biochem Biophys Res Commun* 2013;**434**:8–14.
39. Kim SM, Kwon OJ, Hong YK, Kim JH, Solca F, Ha SJ, et al. Activation of IL-6R/JAK1/STAT3 signaling induces *de novo* resistance to irreversible EGFR inhibitors in non-small cell lung cancer with T790M resistance mutation. *Mol Cancer Ther* 2012;**11**:2254–64.
40. Ishiguro Y, Ishiguro H, Miyamoto H. Epidermal growth factor receptor tyrosine kinase inhibition up-regulates interleukin-6 in cancer cells and induces subsequent development of interstitial pneumonia. *Oncotarget* 2013;**4**:550–9.
41. Wang Y, van Boxel-Dezaire AH, Cheon H, Yang J, Stark GR. STAT3 activation in response to IL-6 is prolonged by the binding of IL-6 receptor to EGF receptor. *Proc Natl Acad Sci U S A* 2013;**110**:16975–80.
42. Yao Z, Fenoglio S, Gao DC, Camiolo M, Stiles B, Lindsted T, et al. TGF- $\beta$  IL-6 axis mediates selective and adaptive mechanisms of resistance to molecular targeted therapy in lung cancer. *Proc Natl Acad Sci U S A* 2010;**107**:15535–40.
43. Suda K, Mizuuchi H, Murakami I, Uramoto H, Tanaka F, Sato K, et al. CRKL amplification is rare as a mechanism for acquired resistance to kinase inhibitors in lung cancers with epidermal growth factor receptor mutation. *Lung Cancer* 2014;**85**:147–51.
44. Cheung HW, Du J, Boehm JS, He F, Weir BA, Wang X, et al. Amplification of CRKL induces transformation and epidermal growth factor receptor inhibitor resistance in human non-small cell lung cancers. *Cancer Discov* 2011;**1**:608–25.
45. Lin F, Xie CY, Li QC, Dong QZ, Wang EH, Wang Y. CRKL promotes lung cancer cell invasion through ERK–MMP9 pathway. *Mol Carcinog* 2015;**54** Suppl 1:E35–44.
46. Ju L, Zhou C, Li W, Yan L. Integrin beta1 over-expression associates with resistance to tyrosine kinase inhibitor gefitinib in non-small cell lung cancer. *J Cell Biochem* 2010;**111**:1565–74.
47. Huang C, Park CC, Hilsenbeck SG, Ward R, Rimawi MF, Wang YC, et al.  $\beta$ 1 Integrin mediates an alternative survival pathway in breast cancer cells resistant to lapatinib. *Breast Cancer Res* 2011;**13**:R84.
48. Ju L, Zhou C. Association of integrin beta1 and c-MET in mediating EGFR TKI gefitinib resistance in non-small cell lung cancer. *Cancer Cell Int* 2013;**13**:15.
49. Kanda R, Kawahara A, Watari K, Murakami Y, Sonoda K, Maeda M, et al. Erlotinib resistance in lung cancer cells mediated by integrin  $\beta$ 1/ Src/Akt-driven bypass signaling. *Cancer Res* 2013;**73**:6243–53.
50. Brown WS, Wendt MK. Integrin-mediated resistance to epidermal growth factor receptor-targeted therapy: an inflammatory situation. *Breast Cancer Res* 2014;**16**:448.
51. Pao W, Wang TY, Riely GJ, Miller VA, Pan Q, Ladanyi M, et al. KRAS mutations and primary resistance of lung adenocarcinomas to gefitinib or erlotinib. *PLoS Med* 2005;**2**:e17.
52. de Mello RA, Marques DS, Medeiros R, Araújo AM. Epidermal growth factor receptor and K-RAS in non-small cell lung cancer-molecular pathways involved and targeted therapies. *World J Clin Oncol* 2011;**2**:367–76.
53. Repasky GA, Chenette EJ, Der CJ. Renewing the conspiracy theory debate: does RAF function alone to mediate RAS oncogenesis? *Trends Cell Biol* 2004;**14**:639–47.
54. Bidkhorji G, Moeini A, Masoudi-Nejad A. Modeling of tumor progression in NSCLC and intrinsic resistance to TKI in loss of PTEN expression. *PLoS One* 2012;**7**:e48004.
55. Sos ML, Koker M, Weir BA, Heynck S, Rabinovsky R, Zander T, et al. PTEN loss contributes to erlotinib resistance in EGFR-mutant lung cancer by activation of Akt and EGFR. *Cancer Res* 2009;**69**:3256–61.
56. Kim EJ, Jeong JH, Bae S, Kang S, Kim CH, Lim YB. mTOR inhibitors radiosensitize PTEN-deficient non-small-cell lung cancer cells harboring an EGFR activating mutation by inducing autophagy. *J Cell Biochem* 2013;**114**:1248–56.
57. Ohashi K, Sequist LV, Arcila ME, Moran T, Chmielecki J, Lin YL, et al. Lung cancers with acquired resistance to EGFR inhibitors occasionally harbor BRAF gene mutations but lack mutations in KRAS, NRAS, or MEK1. *Proc Natl Acad Sci U S A* 2012;**109**:E2127–33.
58. Whyte DB, Holbeck SL. Correlation of PIK3CA mutations with gene expression and drug sensitivity in NCI-60 cell lines. *Biochem Biophys Res Commun* 2006;**340**:469–75.
59. Wang L, Hu H, Pan Y, Wang R, Li Y, Shen L, et al. PIK3CA mutations frequently coexist with EGFR/KRAS mutations in non-small cell lung cancer and suggest poor prognosis in EGFR/KRAS wildtype subgroup. *PLoS One* 2014;**9**:e88291.
60. de Bruin EC, Cowell C, Warne PH, Jiang M, Saunders RE, Melnick MA, et al. Reduced NF1 expression confers resistance to EGFR inhibition in lung cancer. *Cancer Discov* 2014;**4**:606–19.
61. Ng KP, Hillmer AM, Chuah CT, Juan WC, Ko TK, Teo AS, et al. A common BIM deletion polymorphism mediates intrinsic resistance and inferior responses to tyrosine kinase inhibitors in cancer. *Nat Med* 2012;**18**:521–8.
62. Kiyota M, Kuroda J, Yamamoto-Sugitani M, Shimura Y, Nakayama R, Nagoshi H, et al. FTY720 induces apoptosis of chronic myelogenous leukemia cells via dual activation of BIM and BID and overcomes various types of resistance to tyrosine kinase inhibitors. *Apoptosis* 2013;**18**:1437–46.
63. Thiery JP. Epithelial–mesenchymal transitions in tumour progression. *Nat Rev Cancer* 2002;**2**:442–54.
64. Bae GY, Choi SJ, Lee JS, Jo J, Lee J, Kim J, et al. Loss of E-cadherin activates EGFR–MEK/ERK signaling, which promotes invasion via the ZEB1/MMP2 axis in non-small cell lung cancer. *Oncotarget* 2013;**4**:2512–22.
65. Palena C, Hamilton DH, Fernando RI. Influence of IL-8 on the epithelial–mesenchymal transition and the tumor microenvironment. *Future Oncol* 2012;**8**:713–22.
66. Xie M, He CS, Wei SH, Zhang L. Notch-1 contributes to epidermal growth factor receptor tyrosine kinase inhibitor acquired resistance in

- non-small cell lung cancer *in vitro* and *in vivo*. *Eur J Cancer* 2013;**49**:3559–72.
67. Capaccione KM, Hong X, Morgan KM, Liu W, Bishop JM, Liu L, et al. Sox9 mediates Notch1-induced mesenchymal features in lung adenocarcinoma. *Oncotarget* 2014;**5**:3636–50.
68. Xu MM, Mao GX, Liu J, Li JC, Huang H, Liu YF, et al. Low expression of the *FoxO4* gene may contribute to the phenomenon of EMT in non-small cell lung cancer. *Asian Pac J Cancer Prev* 2014;**15**:4013–8.
69. Park KS, Raffeld M, Moon YW, Xi L, Bianco C, Pham T, et al. CRIPTO1 expression in *EGFR*-mutant NSCLC elicits intrinsic *EGFR*-inhibitor resistance. *J Clin Invest* 2014;**124**:3003–15.
70. Wilson C, Nicholes K, Bustos D, Lin E, Song Q, Stephan JP, et al. Overcoming EMT-associated resistance to anti-cancer drugs via *Src/FAK* pathway inhibition. *Oncotarget* 2014;**5**:7328–41.
71. Alam N, Gustafson KS, Ladanyi M, Zakowski MF, Kapoor A, Truskinovsky AM, et al. Small-cell carcinoma with an epidermal growth factor receptor mutation in a never-smoker with gefitinib-responsive adenocarcinoma of the lung. *Clin Lung Cancer* 2010;**11**:E1–4.
72. Shi Z, Tiwari AK, Shukla S, Robey RW, Singh S, Kim IW, et al. Sildenafil reverses ABCB1- and ABCG2-mediated chemotherapeutic drug resistance. *Cancer Res* 2011;**71**:3029–41.
73. Sodani K, Tiwari AK, Singh S, Patel A, Xiao ZJ, Chen JJ, et al. GW583340 and GW2974, human *EGFR* and *HER-2* inhibitors, reverse ABCG2- and ABCB1-mediated drug resistance. *Biochem Pharmacol* 2012;**83**:1613–22.
74. Soda M, Choi YL, Enomoto M, Takada S, Yamashita Y, Ishikawa S, et al. Identification of the transforming *EML4-ALK* fusion gene in non-small-cell lung cancer. *Nature* 2007;**448**:561–6.
75. Kwak EL, Bang YJ, Camidge DR, Shaw AT, Solomon B, Maki RG, et al. Anaplastic lymphoma kinase inhibition in non-small-cell lung cancer. *N Engl J Med* 2010;**363**:1693–703.
76. Zhang X, Zhang S, Yang X, Yang J, Zhou Q, Yin L, et al. Fusion of *EML4* and *ALK* is associated with development of lung adenocarcinomas lacking *EGFR* and *KRAS* mutations and is correlated with *ALK* expression. *Mol Cancer* 2010;**9**:188.
77. Pearson R, Kolesar JM. Targeted therapy for NSCLC: *ALK* inhibition. *J Oncol Pharm Pract* 2012;**18**:271–4.
78. Yamada T, Takeuchi S, Nakade J, Kita K, Nakagawa T, Nanjo S, et al. Paracrine receptor activation by microenvironment triggers bypass survival signals and *ALK* inhibitor resistance in *EML4-ALK* lung cancer cells. *Clin Cancer Res* 2012;**18**:3592–602.
79. Sasaki T, Koivunen J, Ogino A, Yanagita M, Nikiforov S, Zheng W, et al. A novel *ALK* secondary mutation and *EGFR* signaling cause resistance to *ALK* kinase inhibitors. *Cancer Res* 2011;**71**:6051–60.
80. Perez-Torres M, Guix M, Gonzalez A, Arteaga CL. Epidermal growth factor receptor (*EGFR*) antibody down-regulates mutant receptors and inhibits tumors expressing *EGFR* mutations. *J Biol Chem* 2006;**281**:40183–92.
81. Walter AO, Sjin RT, Haringsma HJ, Ohashi K, Sun J, Lee K, et al. Discovery of a mutant-selective covalent inhibitor of *EGFR* that overcomes T790M-mediated resistance in NSCLC. *Cancer Discov* 2013;**3**:1404–15.
82. Cross DA, Ashton SE, Ghiorghiu S, Eberlein C, Nebhan CA, Spitzler PJ, et al. AZD9291, an irreversible *EGFR* TKI, overcomes T790M-mediated resistance to *EGFR* inhibitors in lung cancer. *Cancer Discov* 2014;**4**:1046–61.
83. Hoeflich KP, Merchant M, Orr C, Chan J, Den Otter D, Berry L, et al. Intermittent administration of MEK inhibitor GDC-0973 plus PI3K inhibitor GDC-0941 triggers robust apoptosis and tumor growth inhibition. *Cancer Res* 2012;**72**:210–9.
84. Hong Z, Cao X, Li N, Zhang Y, Lan L, Zhou Y, et al. Luteolin is effective in the non-small cell lung cancer model with L858R/T790M *EGF* receptor mutation and erlotinib resistance. *Br J Pharmacol* 2014;**171**:2842–53.
85. Qu G, Liu C, Sun B, Zhou C, Zhang Z, Wang P. Combination of BIBW2992 and ARQ 197 is effective against erlotinib-resistant human lung cancer cells with the *EGFR* T790M mutation. *Oncol Rep* 2014;**32**:341–7.
86. Kim SM, Yun MR, Hong YK, Solca F, Kim JH, Kim HJ, et al. Glycolysis inhibition sensitizes non-small cell lung cancer with T790M mutation to irreversible *EGFR* inhibitors via translational suppression of Mcl-1 by AMPK activation. *Mol Cancer Ther* 2013;**12**:2145–56.
87. Engelman JA, Zejnullahu K, Gale CM, Lifshits E, Gonzales AJ, Shimamura T, et al. PF00299804, an irreversible pan-ERBB inhibitor, is effective in lung cancer models with *EGFR* and *ERBB2* mutations that are resistant to gefitinib. *Cancer Res* 2007;**67**:11924–32.
88. Nanjo S, Yamada T, Nishihara H, Takeuchi S, Sano T, Nakagawa T, et al. Ability of the Met kinase inhibitor crizotinib and new generation *EGFR* inhibitors to overcome resistance to *EGFR* inhibitors. *PLoS One* 2013;**8**:e84700.
89. Graves EE, Maity A, Le QT. The tumor microenvironment in non-small-cell lung cancer. *Semin Radiat Oncol* 2010;**20**:156–63.
90. Nakade J, Takeuchi S, Nakagawa T, Ishikawa D, Sano T, Nanjo S, et al. Triple inhibition of *EGFR*, Met, and VEGF suppresses regrowth of HGF-triggered, erlotinib-resistant lung cancer harboring an *EGFR* mutation. *J Thorac Oncol* 2014;**9**:775–83.
91. Sarkar S, Rajput S, Tripathi AK, Mandal M. Targeted therapy against *EGFR* and VEGFR using ZD6474 enhances the therapeutic potential of UV-B phototherapy in breast cancer cells. *Mol Cancer* 2013;**12**:122.
92. Song L, Smith MA, Doshi P, Sasser K, Fulp W, Altiock S, et al. Antitumor efficacy of the anti-interleukin-6 (IL-6) antibody siltuximab in mouse xenograft models of lung cancer. *J Thorac Oncol* 2014;**9**:974–82.
93. Rho JK, Choi YJ, Kim SY, Kim TW, Choi EK, Yoon SJ, et al. MET and AXL inhibitor NPS-1034 exerts efficacy against lung cancer cells resistant to *EGFR* kinase inhibitors because of MET or AXL activation. *Cancer Res* 2014;**74**:253–62.
94. Hofmann I, Weiss A, Elain G, Schwaederle M, Sterker D, Romanet V, et al. *K-RAS* mutant pancreatic tumors show higher sensitivity to MEK than to PI3K inhibition *in vivo*. *PLoS One* 2012;**7**:e44146.
95. Rutkowski P, Blank C. Dabrafenib for the treatment of BRAF V600-positive melanoma: a safety evaluation. *Expert Opin Drug Saf* 2014;**13**:1249–58.
96. Ren H, Zhao L, Li Y, Yue P, Deng X, Owonikoko TK, et al. The PI3 kinase inhibitor NVP-BKM120 induces GSK3/FBXW7-dependent Mcl-1 degradation, contributing to induction of apoptosis and enhancement of TRAIL-induced apoptosis. *Cancer Lett* 2013;**338**:229–38.