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Inflammatory Biomarkers in Osteoarthritis

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Summary

Osteoarthritis (OA) is highly prevalent and a leading cause of disability worldwide. Despite the global burden of OA, diagnostic tests and treatments for the molecular or early subclinical stages are still not available for clinical use. In recent years, there has been a large shift in the understanding of OA as a “wear and tear” disease to an inflammatory disease. This has been demonstrated through various studies using MRI, ultrasound, histochemistry, and biomarkers. It would of great value to be able to readily identify subclinical and/or sub-acute inflammation, particularly in such a way as to be appropriate for a clinical setting. Here we review several types of biomarkers associated with OA in human studies that point to a role of inflammation in OA.

Keywords

Inflammation; biomarker; osteoarthritis

Introduction

Inflammatory osteoarthritis (OA) is a debilitating and highly prevalent disease, but is often sub-clinical. There is an increasing body of evidence that inflammatory and destructive responses of the synovium play a major role in OA(1). Moreover, the role of inflammation in the illness of OA has been recognized through the association of joint effusion with joint pain(2). It is still unclear to what extent inflammation is an initiator versus an outcome of the joint destructive process(3). Of particular interest is the emerging evidence that the degree to which the immune and wound healing responses can be activated in part controls the predisposition of an individual to chronic diseases, among them OA. Despite the global burden of OA, diagnostic tests and treatments for the molecular or early subclinical stages are still not available for clinical use. It would of great value to be able to readily identify subclinical and/or sub-acute inflammation, particularly in such a way as to be appropriate

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for a clinical setting. Here we review several types of biomarkers associated with OA in human studies that point to a role of inflammation in OA.

Genetic and Genomic Markers

Increasingly, enrichment strategies are being used in clinical trials to select a study population in which detection of a drug effect is more likely than it would be in an unselected population (prognostic enrichment), or to increase the likelihood of predicting a response (predictive enrichment). In December 2012, the US Food and Drug Administration (FDA) published a draft guidance describing enrichment strategies to support approval of drugs and biological products(4). To date, labeling of more than 100 approved drugs contain information on companion diagnostics, all in the form of genomic biomarkers(5). Genomic selection to predict a response has primarily been used in hematology/oncology and only 4% of the companion diagnostics are currently in the field of rheumatology(6). However, a genomic strategy to identify a subset (4%) of patients with cystic fibrosis likely to respond to ivacaftor (via specific mutations of the CFTR gene) led to the recent FDA approval of this drug(7). In addition, the FDA has recently approved several new drugs and drug combinations(8–10) for the treatment of specific hepatitis C genotypes on the basis of companion genomic tests. A number of genetic/genomic markers are emerging in OA that relate to inflammatory phenotypes that might be similarly exploited in the future for enriching for specific patient subsets for trials and eventually selecting patients for specific clinical treatments.

For instance, several studies suggest that a proportion of the genetic susceptibility to OA may be encoded by variations in innate cytokine activity(11–15). Interestingly, it is possible that our access to antibiotics has shifted the role of the innate immune system from being protective against infectious diseases to intensifying age-related chronic diseases(16, 17). The fact that this may be the case has recently been demonstrated by the emerging story related to the forkhead-box class O (FOXO) genes, which have been found to mediate the inflammatory, apoptosis, and barrier function responses of keratinocytes exposed to bacteria(18). In particular, FOXO1 promotes wound healing by regulating the expression of transforming growth factor-beta (TGF- β) and playing a protective role against oxidative stress (19). FOXO1 and FOXO3 were also found to be expressed in human cartilage, and exposure of pro-inflammatory cytokines suppressed the activity of FOXO1 in chondrocytes(20). In addition, chondrocytes with reduced expression of FOXO transcription factors were more susceptible to apoptosis when exposed to oxidative stress(21). The injury of articular cartilage induces changes in genes associated with cell signaling, response to injury, and wound healing(22). Recently, the pro-apoptotic gene, PUMA, has been shown to be activated by the c-Jun N-terminal kinase (JNK)/c-Jun pathway in the regulation of chondrocyte apoptosis in cartilage of patients with OA(23). Moreover, the Wnt signaling pathway, which is involved in both cartilage and bone tissue homeostasis(24), was found to be up-regulated while its inhibitor (FRZB) was down-regulated in cartilage from joints with moderate to severe OA damage(22). Wnt signaling and DNA repair pathways have also been identified to be significantly correlated with articular cartilage healing(25).

A well-validated gene expression method has identified higher levels of interleukin (IL)-1 β expression in peripheral blood leukocytes (PBLs) in patients with knee OA in association with increased pain and greater risk of radiographic progression of OA(26). The expression of four microRNAs (miRNAs) (miR-146a, miR-155, miR-181a, and miR-223), which have been reported to be expressed in immune cells and are responsible for regulating inflammation, were significantly higher in the peripheral blood mononuclear cells (PBMCs) of patients with OA compared to the PBMCs of healthy controls(27). In another study, plasma concentrations of miR-132 were significantly higher in healthy controls compared to patients with either OA or rheumatoid arthritis (RA)(28). In addition, microarray analysis of blood samples from healthy controls and patients with OA identified six genes significantly down-regulated in mild OA (heat shock 90kDa protein 1, alpha; inhibitor of kappa light polypeptide gene enhancer in B-cells, kinase complex-associated protein; IL-13 receptor, alpha 1; laminin, gamma 1; CXCL4; and tumor necrosis factor-inducible gene (TSG)-6)(29), many of which play an important role in inflammatory cascades. Given the role that genomic companion testing has had in personalized treatments, these genetic markers may pave the path for the development of novel therapeutic targets to treat a subset of patients with OA.

Macrophages

The human body swiftly reacts in response to acute foreign intrusions or injury in a very complex and organized manner via the innate immune system(30). However, low-level inflammatory stimulation may also affect the body and cause a state of chronic inflammation. OA is increasingly recognized as a process involving low grade inflammation, often subclinical (31), that is predictive of articular chondropathy and OA progression (32–35).

A critical aspect of joint inflammation in OA involves synovial macrophages and macrophage-produced cytokines that drive aggrecanases, matrix metalloproteinases, and other destructive and inflammatory responses(36). These critical mediators of joint destruction may be used as tools to assess disease activity and severity. We demonstrated that the profile of cytokines in the OA joint is consistent with macrophage-mediated inflammasome activation(37). Interestingly, the synovial fluid (SF) concentration of uric acid in these OA knees was strongly associated with the concentration of several inflammatory cytokines. Uric acid is the canonical trigger of the macrophage-mediated inflammasome activity in gout(38) and is able to induce cytokine production by macrophages(39). Additional supporting evidence for a potential role of uric acid in OA has been demonstrated by the effect monosodium urate crystals have on decreasing chondrocyte viability and function in gout(40).

Targeting activated macrophages as a treatment for OA has been previously proposed(41–43). Similarly, the released proteins and cytokines of activated macrophages have great potential to become diagnostic biomarkers for OA. Our recent etarfolatide imaging study, which targets activated but not resting macrophages(44), demonstrated the preferential localization of activated macrophages to painful and more severely affected osteoarthritic knees(33). The macrophage markers CD14 and CD163 measured in the blood sera/plasma

and SF were associated with etarfolatide uptake, radiographic OA severity and progression, and joint symptoms(34). YKL-40, a member of the chitinase family, is a glycoprotein that is secreted mostly from macrophages(45, 46) and chondrocytes(47) and has been found to be elevated in the SF of patients with rheumatoid arthritis (RA) and OA(48). Moreover, the median value of SF YKL-40 was greater in patients with moderate/severe arthritis when compared to those with none/mild arthritis(48). Taken together, these results support the involvement of macrophages in OA with activation of the innate immune response in OA pathology and progression.

Complement

As part of the innate immune system, the complement system, as inferred in its name, complements and assists antibodies and phagocytic cells with the clearance of foreign objects(49). The complement system can be activated by several different pathways, including by many components of the cartilage extracellular matrix (reviewed by Hsueh et al.(50)). One proteomic study identified several complement proteins that were differentially expressed between healthy and OA knee SF samples(51). In another proteomic study, complement components were found to be modulated in sera of patients with moderate and severe OA(52). In particular, concentrations of C3a and C5b-9 have been found to be significantly higher in the SF from patients with early-stage OA in comparison to those from healthy individuals(53). In addition, expression of transcripts encoding the complement effectors C7, C4A, factor B, C9, and C5 was significantly higher, while factor H, C4-binding protein, C1 inhibitor, and clusterin, which are complement inhibitors, were lower in synovial membranes of patients with OA in comparison to those from healthy individuals(53). Interestingly, cartilage oligomeric matrix protein (COMP), which is elevated in the sera(54–56) and SF(57, 58) of patients with OA, can also activate the complement system (59, 60). The involvement of complement activation in OA is consistent with its involvement in several other chronic inflammatory diseases, including RA(61, 62), atherosclerosis(63, 64), Alzheimer's(65–67), and age-related macular degeneration(68, 69).

C-Reactive Protein

C reactive protein (CRP) is a central component of the innate immune inflammatory response; by binding to the cell surface of dead or dying cells and some bacteria it leads to the activation of the complement system(70). The synthesis of CRP is mediated by factors released by macrophages and adipocytes. CRP also leads to the promotion of pro-inflammatory cytokines, which increases the inflammatory response(71). CRP was elevated in patients with acute heart failure(72), and has been associated with the risk of developing coronary artery disease(73), head and neck squamous cell carcinoma mortality(74), and inflammatory bowel disease(75). Several studies have also sought to establish a connection between levels of serum CRP and OA(76). A recent meta-analysis of 32 studies revealed statistically significant differences in serum CRP between patients with OA and healthy controls. The study also revealed that CRP was significantly associated with pain and decreased physical function, but not radiographic OA (77). In studies that have adjusted for body mass, CRP has been shown to be independently associated with OA(78), while other studies have not proven CRP to be independently associated with OA(79, 80). The meta-

analysis study by Jin and colleagues(77) also compared studies that accounted for BMI from those that did not and concluded that the association of CRP with OA was independent of BMI.

Cytokines

Cytokines are small proteins released from a wide array of cells, including immune cells (macrophages, B lymphocytes, T lymphocytes, mast cells, natural killer cells, epithelial cells, endothelial cells, dendritic cells, fibroblasts, and stromal cells)(81, 82). Various cells utilize cytokines, chemokines (chemotactic cytokines), and adipokines (cytokines released by adipose tissue) as part of the inflammatory response to regulate cell signaling and interactions within the cell itself and between other cells(83, 84). Cytokines can have either pro-inflammatory or anti-inflammatory properties and often work concordantly to maintain cell homeostasis(85). We have shown that the concentrations of key SF cytokines (IL-1 α , IL-18 and tumor necrosis factor (TNF)- α) were associated with the level of OA severity; baseline IL-18 also predicted OA progression(37). Serum levels of IL-6 and TNF- α have also been associated with the prevalence of joint space narrowing and prediction of knee cartilage loss(86). Serum TNF- α levels have effectively been used to monitor the efficacy of various OA treatments in rabbits(87). However, in a multi-center, randomized, double blind, placebo-controlled trial, injection of anti-TNF- α agents did not show any improvement in patients with unmanageable hand OA(88). Soluble TNF- α receptor levels were decreased in the SF and plasma of patients with OA in comparison with healthy controls, with SF levels being negatively correlated with joint symptoms(89) evaluated by the Western Ontario and McMaster University Osteoarthritis Index (WOMAC) questionnaire. Recently, the activity of the protein encoded by TSG-6, which is an anti-inflammatory protein that is upregulated in response to the pro-inflammatory cytokines TNF and IL-1(90), and was previously detected in patients with arthritis(91), has been shown to be associated with progression of OA after 3-year follow up in a study of 91 patients(92). In a small pilot study, Vangness et al. (93) showed that higher levels of SF IL-2, IL-5, and monocyte chemoattractant protein (MCP)-1 were associated with greater degrees of OA severity. SF levels of IL-6, IL-13, and macrophage inflammatory protein (MIP)-1 β were elevated in patients with end-stage knee OA while granulocyte colony-stimulating factor (G-CSF) levels were lower when compared to SF levels of healthy controls(94).

From a prospective population-based study of 908 healthy middle-aged British women, individuals with higher age and baseline body mass index (BMI) and increased serum IL-6 after 5-year follow-up were more likely to be diagnosed with radiographic knee OA at their 10-year follow-up(95). Given the association of obesity with OA(96, 97), it is of interest that patients with knee OA have increased levels of IL-6 and soluble IL-6 receptor released from the infrapatellar fat pad of the knee compared with subcutaneous adipose tissue(98). Weight loss and increased physical activity in obese elderly patients with OA decreased levels of soluble TNF α Receptor 1 and increased physical function(99). Exercise has also been shown to increase both intraarticular and peri-synovial concentrations of anti-inflammatory IL-10 in women with knee OA(100).

Chemokines

Chemokines are a class of small protein cytokines that act as chemoattractants to guide cells to migrate to a specific location, and in the context of inflammation, toward the site of injury or pathogenic invasion(82). Interferon gamma inducible protein 10, also known as CXCL-10, is a chemokine that has been shown to be inversely associated with radiographic knee OA in both plasma and SF(62). Serum fractalkine, or CX3CL1, has also been shown to be significantly elevated in patients with knee OA in comparison to the sera from healthy individuals, while both serum and SF concentrations of the chemokine were associated with KL scores(101). SF CXCL12, but not serum CXCL12, was associated with radiographic severity of OA(102). A recent study has demonstrated the feasibility of using serum and SF MCP-1, also known as chemokine ligand 2 (CCL2), as a biomarker for self-reported pain and physical disability in patients with knee OA(103). To further support the role of macrophages in OA-related inflammation, MCP-1 has been shown to increase the recruitment of macrophages into adipose tissue(104) and atherosclerotic lesions(105). SF levels of macrophage derived chemokine (MDC) and interferon gamma-induced protein (IP-10) were elevated in patients with end-stage knee OA, while eotaxin levels were lower when compared to SF levels of healthy controls(94).

Adipokines/Hormones

A number of adipose tissue-secreted hormones mediate inflammatory effects(106, 107) and cartilage catabolism(108–110) in OA. Plasma adipokines adiponectin, which is responsible for modulating metabolic processes, and leptin, a regulator of fat storage, have been shown to be positively associated with joint symptom burden, while the glucose modulating adipokine, adipisin, was lower in patients with hip and knee OA(111). Adipsin is also known as complement factor D and is responsible for activating the alternative pathway of the complement system(112). Serum adiponectin levels have also been associated with radiographic hand OA progression after 6-year follow-up(113). Leptin is believed to play an important role in the pathogenesis of OA via its stimulation of anabolic functions of chondrocytes in cartilage(114). Serum leptin levels were also associated with prevalent and incident knee OA in the Study of Women's Health Across the Nation (SWAN) cohort(115). Serum leptin measured at baseline has been associated with increased levels of bone formation biomarkers (osteocalcin and procollagen type I N-terminal propeptide (PINP)) after two-year follow-up(116). In addition, baseline soluble leptin receptor was associated with lower levels of procollagen type II N-terminal propeptide (PIIANP) and decreased cartilage volume(116). Visfatin (also known as nicotinamide phosphoribosyltransferase (Nampt)), which is an adipokine and enzyme involved in the metabolism of nicotinate and nicotinamide, has recently been suggested to be a therapeutic target for OA given its role in chondrocyte and osteoblast activation(117). As an inflammatory biomarker, levels of SF visfatin were significantly greater in patients with knee OA compared to control subjects(118). In addition, SF visfatin was positively correlated with the cartilage degradation markers collagen II (CTX-II) and aggrecan (AGG1 and AGG2) and was significantly elevated in more severe knee OA grades (Kellgren Lawrence 4 versus 3)(118). In middle-aged women, lower levels of serum estradiol, which is a hormone that has anti-

inflammatory properties(119), and the urinary estrogen metabolite 2-hydroxyestrone were associated with both the prevalence and incidence of knee OA(120).

Conclusions

Given the inverse association of OA susceptibility with the ability of cartilage to repair itself(121), the subset of patients with inflammatory OA phenotypes are likely the ones most at risk for progression and symptoms. Moreover, given the wealth of agents available for treating autoimmune arthritides characterized by flagrant inflammation, there is a high likelihood that some of these agents, in appropriate dosing schemes, could be employed to treat the subset of patients with inflammatory OA. This OA subgroup would therefore seem to be an ideal one for which to develop algorithms and tools for reliable identification such as the artificial neural network model recently developed by Heard and colleagues(122). The descriptions above briefly summarize associations of inflammatory components with OA that could in future be exploited to assist with identification of patients for the most appropriate therapy. This however, will require tests with known precision and predictive capability (sensitivity, specificity, positive and negative predictive value) that represents a long developmental pathway before any of the research findings described above could be translated into use in clinical trials or practice. Nevertheless, a biomarker qualification project in OA is ongoing(123) that could serve as a paradigm for the qualification of inflammatory markers in OA. Although a number of OA-related biomarkers emanating from joint tissue have been identified, only a minority have been qualified in the context of inflammation due to the limitation of phenotyping tools to identify the subclinical inflammation in OA(1). Nevertheless, it has been possible to identify numerous inflammatory molecules and pathways in association with OA. This underscores the generalizability of inflammation in OA, albeit often subacute or subclinical. As we learn more about the genetic control of inflammation and wound healing in OA we will undoubtedly discover key targets for therapy in addition to being able to better subset patients for trials and eventually, specific therapies.

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References

1. Sellam J, Berenbaum F. The role of synovitis in pathophysiology and clinical symptoms of osteoarthritis. *Nat Rev Rheumatol*. 2010; 6(11):625–635. [PubMed: 20924410]
2. Takahashi T, Nagai H, Seki H, Fukuda M. Relationship between joint effusion, joint pain, and protein levels in joint lavage fluid of patients with internal derangement and osteoarthritis of the temporomandibular joint. *J Oral Maxillofac Surg*. 1999; 57(10):1187–1193. discussion 1193–4. [PubMed: 10513864]
3. Orłowsky EW, Kraus VB. The Role of Innate Immunity in Osteoarthritis: When ?Our First Line of Defense Goes On the Offensive. *The Journal of Rheumatology*. 2015 In Press.
4. Guidance for Industry: Enrichment Strategies for Clinical Trials to Support Approval of Human Drugs and Biological Products. 2012.
5. Paving the Way for Personalized Medicine. U.S. Department of Health and Human Services Food and Drug Administration; 2013.

6. Toward Precision Medicine: Building a Knowledge Network for Biomedical Research and a New Taxonomy of Disease. Washington, D.C.: National Academies Press; 2011.
7. Breakthrough therapy targets defective protein. U.S. Food and Drug Administration; 2012. FDA approves Kalydeco to treat rare form of cystic fibrosis.
8. FDA News Release. U.S. Department of Health and Human Services Food and Drug Administration; 2013. FDA approves new treatment for hepatitis C virus.
9. FDA News Release. U.S. Department of Health and Human Services Food and Drug Administration; 2013. FDA approves Sovaldi for chronic hepatitis C.
10. FDA News Release. U.S. Department of Health and Human Services Food and Drug Administration; 2014. FDA approves first combination pill to treat hepatitis C.
11. Riyazi N, Slagboom E, de Craen AJ, Meulenbelt I, Houwing-Duistermaat JJ, Kroon HM, et al. Association of the risk of osteoarthritis with high innate production of interleukin-1beta and low innate production of interleukin-10 ex vivo, upon lipopolysaccharide stimulation. *Arthritis Rheum.* 2005; 52(5):1443–1450. [PubMed: 15880595]
12. Goekoop RJ, Kloppenburg M, Kroon HM, Frolich M, Huizinga TW, Westendorp RG, et al. Low innate production of interleukin-1beta and interleukin-6 is associated with the absence of osteoarthritis in old age. *Osteoarthritis Cartilage.* 2010; 18(7):942–947. [PubMed: 20417290]
13. Han L, Lee HS, Yoon JH, Choi WS, Park YG, Nam SW, et al. Association of IL-17A and IL-17F single nucleotide polymorphisms with susceptibility to osteoarthritis in a Korean population. *Gene.* 2014; 533(1):119–122. [PubMed: 24096234]
14. Hamalainen S, Solovieva S, Vehmas T, Leino-Arjas P, Hirvonen A. Variations in the TNFalpha gene and their interactions with the IL4R and IL10 genes in relation to hand osteoarthritis. *BMC Musculoskelet Disord.* 2014; 15:311. [PubMed: 25252624]
15. Yigit S, Inanir A, Tekcan A, Tural E, Ozturk GT, Kismali G, et al. Significant association of interleukin-4 gene intron 3 VNTR polymorphism with susceptibility to knee osteoarthritis. *Gene.* 2014; 537(1):6–9. [PubMed: 24406619]
16. Kraus, VB. Preclinical osteoarthritis. In: Hochberg, MC.; Silman, AJ.; Smolen, JS.; Weinblatt, ME.; Weisman, MH., editors. *Rheumatology.* 6th ed. Philadelphia, PA: Mosby Elsevier; 2014. p. 1498-1547.
17. Grice EA, Segre JA. Interaction of the microbiome with the innate immune response in chronic wounds. *Adv Exp Med Biol.* 2012; 946:55–68. [PubMed: 21948362]
18. Li S, Dong G, Moschidis A, Ortiz J, Benakanakere MR, Kinane DF, et al. P. gingivalis modulates keratinocytes through FOXO transcription factors. *PLoS One.* 2013; 8(11):e78541. [PubMed: 24265696]
19. Hameedaldeen A, Liu J, Batres A, Graves GS, Graves DT. FOXO1, TGF-beta regulation and wound healing. *Int J Mol Sci.* 2014; 15(9):16257–16269. [PubMed: 25226535]
20. Akasaki Y, Hasegawa A, Saito M, Asahara H, Iwamoto Y, Lotz MK. Dysregulated FOXO transcription factors in articular cartilage in aging and osteoarthritis. *Osteoarthritis Cartilage.* 2014; 22(1):162–170. [PubMed: 24269635]
21. Akasaki Y, Alvarez-Garcia O, Saito M, Carames B, Iwamoto Y, Lotz MK. FoxO Transcription Factors Support Oxidative Stress Resistance in Human Chondrocytes. *Arthritis Rheumatol.* 2014; 66(12):3349–3358. [PubMed: 25186470]
22. Dell'accio F, De Bari C, Eltawil NM, Vanhummelen P, Pitzalis C. Identification of the molecular response of articular cartilage to injury, by microarray screening: Wnt-16 expression and signaling after injury and in osteoarthritis. *Arthritis Rheum.* 2008; 58(5):1410–1421. [PubMed: 18438861]
23. Lu H, Hou G, Zhang Y, Dai Y, Zhao H. c-Jun transactivates Puma gene expression to promote osteoarthritis. *Mol Med Rep.* 2014; 9(5):1606–1612. [PubMed: 24566851]
24. Luyten FP, Tylzanowski P, Lories RJ. Wnt signaling and osteoarthritis. *Bone.* 2009; 44(4):522–527. [PubMed: 19136083]
25. Rai MF, Schmidt EJ, McAlinden A, Cheverud JM, Sandell LJ. Molecular insight into the association between cartilage regeneration and ear wound healing in genetic mouse models: targeting new genes in regeneration. *G3 (Bethesda).* 2013; 3(11):1881–1891. [PubMed: 24002865]
26. Attur M, Belitskaya-Levy I, Oh C, Krasnokutsky S, Greenberg J, Samuels J, et al. Increased interleukin-1beta gene expression in peripheral blood leukocytes is associated with increased pain

- and predicts risk for progression of symptomatic knee osteoarthritis. *Arthritis Rheum.* 2011; 63(7): 1908–1917. [PubMed: 21717421]
27. Okuhara A, Nakasa T, Shibuya H, Niimoto T, Adachi N, Deie M, et al. Changes in microRNA expression in peripheral mononuclear cells according to the progression of osteoarthritis. *Mod Rheumatol.* 2012; 22(3):446–457. [PubMed: 22006119]
 28. Murata K, Yoshitomi H, Tanida S, Ishikawa M, Nishitani K, Ito H, et al. Plasma and synovial fluid microRNAs as potential biomarkers of rheumatoid arthritis and osteoarthritis. *Arthritis Res Ther.* 2010; 12(3):R86. [PubMed: 20470394]
 29. Marshall KW, Zhang H, Yager TD, Nossova N, Dempsey A, Zheng R, et al. Blood-based biomarkers for detecting mild osteoarthritis in the human knee. *Osteoarthritis Cartilage.* 2005; 13(10):861–871. [PubMed: 16139532]
 30. Barton GM. A calculated response: control of inflammation by the innate immune system. *J Clin Invest.* 2008; 118(2):413–420. [PubMed: 18246191]
 31. Fernandez-Madrid F, Karvonen RL, Teitge RA, Miller PR, An T, Negendank WG. Synovial thickening detected by MR imaging in osteoarthritis of the knee confirmed by biopsy as synovitis. *Magn Reson Imaging.* 1995; 13(2):177–183. [PubMed: 7739358]
 32. Ayril X, Pickering EH, Woodworth TG, Mackillop N, Dougados M. Synovitis: a potential predictive factor of structural progression of medial tibiofemoral knee osteoarthritis -- results of a 1 year longitudinal arthroscopic study in 422 patients. *Osteoarthritis Cartilage.* 2005; 13(5):361–367. [PubMed: 15882559]
 33. Kraus VB, McDaniel G, Huebner JL, Stabler TV, C P, Coleman RE, et al. Direct in vivo evidence of activated macrophages in human osteoarthritis. *Osteoarthritis Cartilage.* 2013 Apr. 21(Supplement)
 34. Daghestani HN, Pieper CF, Kraus VB. Soluble Macrophage Biomarkers Indicate Inflammatory Phenotypes in Patients with Knee Osteoarthritis. *Arthritis & Rheumatology.* 2015 In Press.
 35. Roemer FW, Guermazi A, Felson DT, Niu J, Nevitt MC, Crema MD, et al. Presence of MRI-detected joint effusion and synovitis increases the risk of cartilage loss in knees without osteoarthritis at 30-month follow-up: the MOST study. *Ann Rheum Dis.* 2011; 70(10):1804–1809. [PubMed: 21791448]
 36. Bondeson J, Wainwright SD, Lauder S, Amos N, Hughes CE. The role of synovial macrophages and macrophage-produced cytokines in driving aggrecanases, matrix metalloproteinases, and other destructive and inflammatory responses in osteoarthritis. *Arthritis Res Ther.* 2006; 8(6):R187. [PubMed: 17177994]
 37. Denoble AE, Huffman KM, Stabler TV, Kelly SJ, Hershfield MS, McDaniel GE, et al. Uric acid is a danger signal of increasing risk for osteoarthritis through inflammasome activation. *Proc Natl Acad Sci U S A.* 2011; 108(5):2088–2093. [PubMed: 21245324]
 38. Martinon F, Petrilli V, Mayor A, Tardivel A, Tschopp J. Gout-associated uric acid crystals activate the NALP3 inflammasome. *Nature.* 2006; 440(7081):237–241. [PubMed: 16407889]
 39. Orłowski EW, Stabler TV, Montell E, Verges J, Kraus VB. Monosodium urate crystal induced macrophage inflammation is attenuated by chondroitin sulphate: pre-clinical model for gout prophylaxis? *BMC Musculoskelet Disord.* 2014; 15:318. [PubMed: 25261974]
 40. Chhana A, Callon KE, Pool B, Naot D, Gamble GD, Dray M, et al. The effects of monosodium urate monohydrate crystals on chondrocyte viability and function: implications for development of cartilage damage in gout. *J Rheumatol.* 2013; 40(12):2067–2074. [PubMed: 24187106]
 41. Bondeson J. Activated synovial macrophages as targets for osteoarthritis drug therapy. *Curr Drug Targets.* 2010; 11(5):576–585. [PubMed: 20199392]
 42. Tsuneyoshi Y, Tanaka M, Nagai T, Sunahara N, Matsuda T, Sonoda T, et al. Functional folate receptor beta-expressing macrophages in osteoarthritis synovium and their M1/M2 expression profiles. *Scand J Rheumatol.* 2012; 41(2):132–140. [PubMed: 22211358]
 43. Low PS, Henne WA, Doorneweerd DD. Discovery and development of folic-acid-based receptor targeting for imaging and therapy of cancer and inflammatory diseases. *Acc Chem Res.* 2008; 41(1):120–129. [PubMed: 17655275]

44. Xia W, Hilgenbrink AR, Matteson EL, Lockwood MB, Cheng JX, Low PS. A functional folate receptor is induced during macrophage activation and can be used to target drugs to activated macrophages. *Blood*. 2009; 113(2):438–446. [PubMed: 18952896]
45. Renkema GH, Boot RG, Au FL, Donker-Koopman WE, Strijland A, Muijsers AO, et al. Chitotriosidase, a chitinase, and the 39-kDa human cartilage glycoprotein, a chitin-binding lectin, are homologues of family 18 glycosyl hydrolases secreted by human macrophages. *Eur J Biochem*. 1998; 251(1–2):504–509. [PubMed: 9492324]
46. Nyirkos P, Golds EE. Human synovial cells secrete a 39 kDa protein similar to a bovine mammary protein expressed during the non-lactating period. *Biochem J*. 1990; 269(1):265–268. [PubMed: 2375755]
47. Hakala BE, White C, Recklies AD. Human cartilage gp-39, a major secretory product of articular chondrocytes and synovial cells, is a mammalian member of a chitinase protein family. *J Biol Chem*. 1993; 268(34):25803–25810. [PubMed: 8245017]
48. Volck B, Johansen JS, Stoltenberg M, Garbarsch C, Price PA, Ostergaard M, et al. Studies on YKL-40 in knee joints of patients with rheumatoid arthritis and osteoarthritis. Involvement of YKL-40 in the joint pathology. *Osteoarthritis Cartilage*. 2001; 9(3):203–214. [PubMed: 11300743]
49. Sarma JV, Ward PA. The complement system. *Cell Tissue Res*. 2011; 343(1):227–235. [PubMed: 20838815]
50. Hsueh MF, Onnerfjord P, Kraus VB. Biomarkers and proteomic analysis of osteoarthritis. *Matrix Biol*. 2014; 39:56–66. [PubMed: 25179675]
51. Gobezie R, Kho A, Krastins B, Sarracino DA, Thornhill TS, Chase M, et al. High abundance synovial fluid proteome: distinct profiles in health and osteoarthritis. *Arthritis Res Ther*. 2007; 9(2):R36. [PubMed: 17407561]
52. Fernandez-Puente P, Mateos J, Fernandez-Costa C, Oreiro N, Fernandez-Lopez C, Ruiz-Romero C, et al. Identification of a panel of novel serum osteoarthritis biomarkers. *J Proteome Res*. 2011; 10(11):5095–5101. [PubMed: 21973172]
53. Wang Q, Rozelle AL, Lepus CM, Scanzello CR, Song JJ, Larsen DM, et al. Identification of a central role for complement in osteoarthritis. *Nat Med*. 2011; 17(12):1674–1679. [PubMed: 22057346]
54. Clark AG, Jordan JM, Vilim V, Renner JB, Dragomir AD, Luta G, et al. Serum cartilage oligomeric matrix protein reflects osteoarthritis presence and severity: the Johnston County Osteoarthritis Project. *Arthritis Rheum*. 1999; 42(11):2356–2364. [PubMed: 10555031]
55. Verma P, Dalal K. Serum cartilage oligomeric matrix protein (COMP) in knee osteoarthritis: a novel diagnostic and prognostic biomarker. *J Orthop Res*. 2013; 31(7):999–1006. [PubMed: 23423905]
56. Fernandes FA, Pucinelli ML, da Silva NP, Feldman D. Serum cartilage oligomeric matrix protein (COMP) levels in knee osteoarthritis in a Brazilian population: clinical and radiological correlation. *Scand J Rheumatol*. 2007; 36(3):211–215. [PubMed: 17657676]
57. Neidhart M, Hauser N, Paulsson M, DiCesare PE, Michel BA, Hauselmann HJ. Small fragments of cartilage oligomeric matrix protein in synovial fluid and serum as markers for cartilage degradation. *Br J Rheumatol*. 1997; 36(11):1151–1160. [PubMed: 9402858]
58. Lohmander LS, Saxne T, Heinegard DK. Release of cartilage oligomeric matrix protein (COMP) into joint fluid after knee injury and in osteoarthritis. *Ann Rheum Dis*. 1994; 53(1):8–13. [PubMed: 8311563]
59. Otteby KE, Holmquist E, Saxne T, Heinegard D, Hesselstrand R, Blom AM. Cartilage oligomeric matrix protein-induced complement activation in systemic sclerosis. *Arthritis Res Ther*. 2013; 15(6):R215. [PubMed: 24330664]
60. Happonen KE, Saxne T, Aspberg A, Morgelin M, Heinegard D, Blom AM. Regulation of complement by cartilage oligomeric matrix protein allows for a novel molecular diagnostic principle in rheumatoid arthritis. *Arthritis Rheum*. 2010; 62(12):3574–3583. [PubMed: 20737467]
61. Molenaar ET, Voskuyl AE, Familian A, van Mierlo GJ, Dijkmans BA, Hack CE. Complement activation in patients with rheumatoid arthritis mediated in part by C-reactive protein. *Arthritis Rheum*. 2001; 44(5):997–1002. [PubMed: 11352263]

62. Nydegger UE, Zubler RH, Gabay R, Joliat G, Karagevrekis CH, Lambert PH, et al. Circulating complement breakdown products in patients with rheumatoid arthritis. Correlation between plasma C3d, circulating immune complexes, and clinical activity. *J Clin Invest.* 1977; 59(5):862–868. [PubMed: 853126]
63. Niculescu F, Rus H. The role of complement activation in atherosclerosis. *Immunol Res.* 2004; 30(1):73–80. [PubMed: 15258311]
64. Niculescu F, Rus HG, Vlaicu R. Activation of the human terminal complement pathway in atherosclerosis. *Clin Immunol Immunopathol.* 1987; 45(2):147–155. [PubMed: 2444373]
65. Aiyaz M, Lupton MK, Proitsi P, Powell JF, Lovestone S. Complement activation as a biomarker for Alzheimer's disease. *Immunobiology.* 2012; 217(2):204–215. [PubMed: 21856034]
66. Eikelenboom P, Hack CE, Rozemuller JM, Stam FC. Complement activation in amyloid plaques in Alzheimer's dementia. *Virchows Arch B Cell Pathol Incl Mol Pathol.* 1989; 56(4):259–262. [PubMed: 2565620]
67. Rogers J, Cooper NR, Webster S, Schultz J, McGeer PL, Styren SD, et al. Complement activation by beta-amyloid in Alzheimer disease. *Proc Natl Acad Sci U S A.* 1992; 89(21):10016–10020. [PubMed: 1438191]
68. Scholl HP, Charbel Issa P, Walier M, Janzer S, Pollok-Kopp B, Borncke F, et al. Systemic complement activation in age-related macular degeneration. *PLoS One.* 2008; 3(7):e2593. [PubMed: 18596911]
69. Johnson LV, Leitner WP, Staples MK, Anderson DH. Complement activation and inflammatory processes in Drusen formation and age related macular degeneration. *Exp Eye Res.* 2001; 73(6): 887–896. [PubMed: 11846519]
70. Du Clos TW, Mold C. Pentraxins (CRP, SAP) in the process of complement activation and clearance of apoptotic bodies through Fcγ receptors. *Curr Opin Organ Transplant.* 2011; 16(1):15–20. [PubMed: 21150611]
71. Du Clos TW. Function of C-reactive protein. *Ann Med.* 2000; 32(4):274–278. [PubMed: 10852144]
72. Kalogeropoulos AP, Tang WH, Hsu A, Felker GM, Hernandez AF, Troughton RW, et al. High-sensitivity C-reactive protein in acute heart failure: insights from the ASCEND-HF trial. *J Card Fail.* 2014; 20(5):319–326. [PubMed: 24530944]
73. Emerging Risk Factors C, Kaptoge S, Di Angelantonio E, Lowe G, Pepys MB, Thompson SG, et al. C-reactive protein concentration and risk of coronary heart disease, stroke, and mortality: an individual participant meta-analysis. *Lancet.* 2010; 375(9709):132–140. [PubMed: 20031199]
74. Andersson BA, Lewin F, Lundgren J, Nilsson M, Rutqvist LE, Lofgren S, et al. Plasma tumor necrosis factor-alpha and C-reactive protein as biomarker for survival in head and neck squamous cell carcinoma. *J Cancer Res Clin Oncol.* 2014; 140(3):515–519. [PubMed: 24481866]
75. Vermeire S, Van Assche G, Rutgeerts P. C-reactive protein as a marker for inflammatory bowel disease. *Inflamm Bowel Dis.* 2004; 10(5):661–665. [PubMed: 15472532]
76. Pearle AD, Scanzello CR, George S, Mandl LA, DiCarlo EF, Peterson M, et al. Elevated high-sensitivity C-reactive protein levels are associated with local inflammatory findings in patients with osteoarthritis. *Osteoarthritis Cartilage.* 2007; 15(5):516–523. [PubMed: 17157039]
77. Jin X, Beguerie JR, Zhang W, Blizzard L, Otahal P, Jones G, et al. Circulating C reactive protein in osteoarthritis: a systematic review and meta-analysis. *Ann Rheum Dis.* 2013
78. Sowers M, Jannausch M, Stein E, Jamadar D, Hochberg M, Lachance L. C-reactive protein as a biomarker of emergent osteoarthritis. *Osteoarthritis Cartilage.* 2002; 10(8):595–601. [PubMed: 12479380]
79. Kraus VB, Stabler TV, Luta G, Renner JB, Dragomir AD, Jordan JM. Interpretation of serum C-reactive protein (CRP) levels for cardiovascular disease risk is complicated by race, pulmonary disease, body mass index, gender, and osteoarthritis. *Osteoarthritis Cartilage.* 2007; 15(8):966–971. [PubMed: 17395501]
80. Kerkhof HJ, Bierma-Zeinstra SM, Castano-Betancourt MC, de Maat MP, Hofman A, Pols HA, et al. Serum C reactive protein levels and genetic variation in the CRP gene are not associated with the prevalence, incidence or progression of osteoarthritis independent of body mass index. *Ann Rheum Dis.* 2010; 69(11):1976–1982. [PubMed: 20511616]

81. Liles WC, Van Voorhis WC. Review: nomenclature and biologic significance of cytokines involved in inflammation and the host immune response. *J Infect Dis.* 1995; 172(6):1573–1580. [PubMed: 7594719]
82. Commins SP, Borish L, Steinke JW. Immunologic messenger molecules: cytokines, interferons, and chemokines. *J Allergy Clin Immunol.* 2010; 125 Suppl 2(2):S53–S72. [PubMed: 19932918]
83. Hanada T, Yoshimura A. Regulation of cytokine signaling and inflammation. *Cytokine Growth Factor Rev.* 2002; 13(4–5):413–421. [PubMed: 12220554]
84. Rot A, von Andrian UH. Chemokines in innate and adaptive host defense: basic chemokines grammar for immune cells. *Annu Rev Immunol.* 2004; 22:891–928. [PubMed: 15032599]
85. Seruga B, Zhang HB, Bernstein LJ, Tannock IF. Cytokines and their relationship to the symptoms and outcome of cancer. *Nature Reviews Cancer.* 2008; 8(11):887–899. [PubMed: 18846100]
86. Stannus O, Jones G, Cicuttini F, Parameswaran V, Quinn S, Burgess J, et al. Circulating levels of IL-6 and TNF-alpha are associated with knee radiographic osteoarthritis and knee cartilage loss in older adults. *Osteoarthritis Cartilage.* 2010; 18(11):1441–1447. [PubMed: 20816981]
87. Guo H, Luo Q, Zhang J, Lin H, Xia L, He C. Comparing different physical factors on serum TNF-alpha levels, chondrocyte apoptosis, caspase-3 and caspase-8 expression in osteoarthritis of the knee in rabbits. *Joint Bone Spine.* 2011; 78(6):604–610. [PubMed: 21397547]
88. Chevalier X, Ravaud P, Maheu E, Baron G, Rialland A, Vergnaud P, et al. Adalimumab in patients with hand osteoarthritis refractory to analgesics and NSAIDs: a randomised, multicentre, double-blind, placebo-controlled trial. *Ann Rheum Dis.* 2014
89. Simao AP, Almeida TM, Mendonca VA, Santos SA, Gomes WF, Coimbra CC, et al. Soluble TNF receptors are produced at sites of inflammation and are inversely associated with self-reported symptoms (WOMAC) in knee osteoarthritis. *Rheumatol Int.* 2014
90. Milner CM, Day AJ. TSG-6: a multifunctional protein associated with inflammation. *J Cell Sci.* 2003; 116(Pt 10):1863–1873. [PubMed: 12692188]
91. Wisniewski HG, Maier R, Lotz M, Lee S, Klampfer L, Lee TH, et al. TSG-6: a TNF-, IL-1-, and LPS-inducible secreted glycoprotein associated with arthritis. *J Immunol.* 1993; 151(11):6593–6601. [PubMed: 8245487]
92. Wisniewski HG, Colon E, Liublińska V, Karia RJ, Stabler TV, Attur M, et al. TSG-6 activity as a novel biomarker of progression in knee osteoarthritis. *Osteoarthritis Cartilage.* 2014; 22(2):235–241. [PubMed: 24333293]
93. Vangsness CT Jr, Burke WS, Narvy SJ, MacPhee RD, Fedenko AN. Human knee synovial fluid cytokines correlated with grade of knee osteoarthritis—a pilot study. *Bull NYU Hosp Jt Dis.* 2011; 69(2):122–127. [PubMed: 22035391]
94. Beekhuizen M, Gierman LM, van Spil WE, Van Osch GJ, Huizinga TW, Saris DB, et al. An explorative study comparing levels of soluble mediators in control and osteoarthritic synovial fluid. *Osteoarthritis Cartilage.* 2013; 21(7):918–922. [PubMed: 23598178]
95. Livshits G, Zhai G, Hart DJ, Kato BS, Wang H, Williams FM, et al. Interleukin-6 is a significant predictor of radiographic knee osteoarthritis: The Chingford Study. *Arthritis Rheum.* 2009; 60(7):2037–2045. [PubMed: 19565477]
96. King LK, March L, Anandacoomarasamy A. Obesity & osteoarthritis. *Indian J Med Res.* 2013; 138:185–193. [PubMed: 24056594]
97. Salih S, Sutton P. Obesity, knee osteoarthritis and knee arthroplasty: a review. *BMC Sports Sci Med Rehabil.* 2013; 5(1):25. [PubMed: 24304704]
98. Distel E, Cadoudal T, Durant S, Poignard A, Chevalier X, Benelli C. The infrapatellar fat pad in knee osteoarthritis: an important source of interleukin-6 and its soluble receptor. *Arthritis Rheum.* 2009; 60(11):3374–3377. [PubMed: 19877065]
99. Miller GD, Nicklas BJ, Loeser RF. Inflammatory biomarkers and physical function in older, obese adults with knee pain and self-reported osteoarthritis after intensive weight-loss therapy. *J Am Geriatr Soc.* 2008; 56(4):644–651. [PubMed: 18312558]
100. Helmark IC, Mikkelsen UR, Borglum J, Rothe A, Petersen MC, Andersen O, et al. Exercise increases interleukin-10 levels both intraarticularly and peri-synovially in patients with knee osteoarthritis: a randomized controlled trial. *Arthritis Res Ther.* 2010; 12(4):R126. [PubMed: 20594330]

101. Zou Y, Li Y, Lu L, Lin Y, Liang W, Su Z, et al. Correlation of fractalkine concentrations in serum and synovial fluid with the radiographic severity of knee osteoarthritis. *Ann Clin Biochem.* 2013; 50(Pt 6):571–575. [PubMed: 23869024]
102. Xu Q, Sun XC, Shang XP, Jiang HS. Association of CXCL12 levels in synovial fluid with the radiographic severity of knee osteoarthritis. *J Investig Med.* 2012; 60(6):898–901.
103. Li L, Jiang BE. Serum and synovial fluid chemokine ligand 2/monocyte chemoattractant protein 1 concentrations correlates with symptomatic severity in patients with knee osteoarthritis. *Ann Clin Biochem.* 2014
104. Kanda H, Tateya S, Tamori Y, Kotani K, Hiasa K, Kitazawa R, et al. MCP-1 contributes to macrophage infiltration into adipose tissue, insulin resistance, and hepatic steatosis in obesity. *J Clin Invest.* 2006; 116(6):1494–1505. [PubMed: 16691291]
105. Yla-Herttuala S, Lipton BA, Rosenfeld ME, Sarkioja T, Yoshimura T, Leonard EJ, et al. Expression of monocyte chemoattractant protein 1 in macrophage-rich areas of human and rabbit atherosclerotic lesions. *Proc Natl Acad Sci U S A.* 1991; 88(12):5252–5256. [PubMed: 2052604]
106. Al-Suhaimi EA, Shehzad A. Leptin, resistin and visfatin: the missing link between endocrine metabolic disorders and immunity. *Eur J Med Res.* 2013; 18:12. [PubMed: 23634778]
107. Toussiot E, Streit G, Wendling D. The contribution of adipose tissue and adipokines to inflammation in joint diseases. *Curr Med Chem.* 2007; 14(10):1095–1100. [PubMed: 17456023]
108. Kang EH, Lee YJ, Kim TK, Chang CB, Chung JH, Shin K, et al. Adiponectin is a potential catabolic mediator in osteoarthritis cartilage. *Arthritis Res Ther.* 2010; 12(6):R231. [PubMed: 21194467]
109. Hui W, Litherland GJ, Elias MS, Kitson GI, Cawston TE, Rowan AD, et al. Leptin produced by joint white adipose tissue induces cartilage degradation via upregulation and activation of matrix metalloproteinases. *Ann Rheum Dis.* 2012; 71(3):455–462. [PubMed: 22072016]
110. Hu PF, Chen WP, Tang JL, Bao JP, Wu LD. Apelin plays a catabolic role on articular cartilage: in vivo and in vitro studies. *Int J Mol Med.* 2010; 26(3):357–363. [PubMed: 20664951]
111. Perruccio AV, Mahomed NN, Chandran V, Gandhi R. Plasma adipokine levels and their association with overall burden of painful joints among individuals with hip and knee osteoarthritis. *J Rheumatol.* 2014; 41(2):334–337. [PubMed: 24334649]
112. White RT, Damm D, Hancock N, Rosen BS, Lowell BB, Usher P, et al. Human adiponin is identical to complement factor D and is expressed at high levels in adipose tissue. *J Biol Chem.* 1992; 267(13):9210–9213. [PubMed: 1374388]
113. Yusuf E, Ioan-Facsinay A, Bijsterbosch J, Klein-Wieringa I, Kwekkeboom J, Slagboom PE, et al. Association between leptin, adiponectin and resistin and long-term progression of hand osteoarthritis. *Ann Rheum Dis.* 2011; 70(7):1282–1284. [PubMed: 21470970]
114. Dumond H, Presle N, Terlain B, Mainard D, Loeuille D, Netter P, et al. Evidence for a key role of leptin in osteoarthritis. *Arthritis Rheum.* 2003; 48(11):3118–3129. [PubMed: 14613274]
115. Karvonen-Gutierrez CA, Harlow SD, Mancuso P, Jacobson J, Mendes de Leon CF, Nan B. Association of leptin levels with radiographic knee osteoarthritis among a cohort of midlife women. *Arthritis Care Res (Hoboken).* 2013; 65(6):936–944. [PubMed: 23281224]
116. Berry PA, Jones SW, Cicuttini FM, Wluka AE, Maciewicz RA. Temporal relationship between serum adipokines, biomarkers of bone and cartilage turnover, and cartilage volume loss in a population with clinical knee osteoarthritis. *Arthritis Rheum.* 2011; 63(3):700–707. [PubMed: 21305502]
117. Laiguillon MC, Houard X, Bougault C, Gosset M, Nourissat G, Sautet A, et al. Expression and function of visfatin (Nampt), an adipokine-enzyme involved in inflammatory pathways of osteoarthritis. *Arthritis Res Ther.* 2014; 16(1):R38. [PubMed: 24479481]
118. Duan Y, Hao D, Li M, Wu Z, Li D, Yang X, et al. Increased synovial fluid visfatin is positively linked to cartilage degradation biomarkers in osteoarthritis. *Rheumatol Int.* 2012; 32(4):985–990. [PubMed: 21246369]
119. Salem ML, Hossain MS, Nomoto K. Mediation of the immunomodulatory effect of beta-estradiol on inflammatory responses by inhibition of recruitment and activation of inflammatory cells and their gene expression of TNF-alpha and IFN-gamma. *Int Arch Allergy Immunol.* 2000; 121(3): 235–245. [PubMed: 10729783]

120. Sowers MR, McConnell D, Jannausch M, Buyuktur AG, Hochberg M, Jamadar DA. Estradiol and its metabolites and their association with knee osteoarthritis. *Arthritis Rheum.* 2006; 54(8): 2481–2487. [PubMed: 16871545]
121. Hashimoto S, Rai MF, Janiszak KL, Cheverud JM, Sandell LJ. Cartilage and bone changes during development of post-traumatic osteoarthritis in selected LGXSM recombinant inbred mice. *Osteoarthritis Cartilage.* 2012; 20(6):562–571. [PubMed: 22361237]
122. Heard BJ, Rosvold JM, Fritzler MJ, El-Gabalawy H, Wiley JP, Krawetz RJ. A computational method to differentiate normal individuals, osteoarthritis and rheumatoid arthritis patients using serum biomarkers. *J R Soc Interface.* 2014; 11(97):20140428. [PubMed: 24920114]
123. Hunter DJ, Nevitt M, Losina E, Kraus V. Biomarkers for osteoarthritis: Current position and steps towards further validation. *Best Practice & Research in Clinical Rheumatology.* 2014; 28(1):61–71.