

RESEARCH

Open Access

Effective cellular internalization of silica-coated CdSe quantum dots for high contrast cancer imaging and labelling applications

Muthunayagam Vibin¹, Ramachandran Vinayakan², Annie John³, Francis Boniface Fernandez³ and Annie Abraham^{1*}

* Correspondence: annieab2001@gmail.com

¹Department of Biochemistry, University of Kerala, Kariavattom campus, 695581 Thiruvananthapuram, Kerala, India
Full list of author information is available at the end of the article

Abstract

The possibility of developing novel contrast imaging agents for cancer cellular labelling and fluorescence imaging applications were explored using silica-coated cadmium selenide (CdSe) quantum dots (QDs). The time dependent cellular internalization efficiency study was carried out using Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES) and Confocal Laser Scanning Microscopy (cLSM) after exposing QDs to stem cells and cancer cells. The strong fluorescence from the cytoplasm confirmed that the QDs were efficiently internalized by the cells. The internalization maxima were observed at the fourth hour of incubation in both stem and cancer cells. Further, the in vitro fluorescence imaging as well as localization study of QDs were performed in various cells. Moreover, high contrast in vivo tumor imaging efficiency of silica-coated CdSe QDs was performed in ultrathin sections of tumor mice, and the results confirmed its effective role in cellular imaging and labelling in cancer and other diseases.

Keywords: Stem cells; Cellular imaging; Fluorescence; Internalization; Quantum dots; Ultrathin

Background

The development of highly sensitive and specific biological probes which lack the intrinsic limitations of organic dyes and fluorescent proteins is focus of many areas of research like molecular and cellular biology [1,2] and medical diagnostics [3-6]. The conventional organic fluorescent tags mainly suffer from low photo stability, poor quantum yield under biological conditions and interference from auto fluorescence. The exceptional photophysical properties of quantum dots (QDs), particularly photostability and emission as a function of size, make them superior to organic dyes for biological applications. These properties have opened new possibilities for advanced molecular and cellular imaging as well as for ultrasensitive bioassays and diagnostics [7-10]. Besides, QDs are better labelling agents in long-term imaging such as fluorescence marking of transport processes in cells and in tracking the path of single membrane-bound molecules [11,12].

Although imaging of fixed cells is useful and sufficient for many applications, live cell microscopy is ideal for visualizing cellular processes, which is considerably more

difficult. It has been shown that many cell types naturally engulf QDs through a non-specific uptake mechanism. This mechanism was used to track the migration of breast tumor cells on a substrate coated with red emitting QDs; the fluorescence within the cells were increased due to the uptake of QDs, leaving behind a dark path [13-15].

Many reports highlight the use of QDs as a fluorescent probe to visualize the biological processes both in vitro and in vivo [16-18]. With proper surface functionalization using peptides, proteins or antibodies, QDs shows specificity in targeting and imaging tumors in living subjects. The fluorescence imaging can easily be achieved by monitoring the stable and strong fluorescence from the QDs [19]; Law et al. [20]. These studies confirm that QDs have opened up a new avenue for investigating biomolecular processes inside the cells.

The inherent hydrophobicity of QDs can be overcome by various approaches; among these overcoating with silica is most preferred method [21-24]. Besides, the cytocompatibility of CdSe QDs on overcoating with silica has been well established by many groups [25-27]. This is based on the observation that the core constituents viz., cadmium and selenium ions are well encapsulated within the silica shell preventing surface oxidation, even in the biological media. Thus a silica shell plays dual role i.e., it makes QDs dispersible in aqueous media and it eliminates the toxicity. A detailed investigation of the cytocompatibility of silanised QDs was reported by our group [28]. By time and concentration dependent studies we observed that the internalized silica-coated CdSe QDs were non-toxic proving the cytocompatibility even at higher concentrations and longer incubation periods. We have also reported the non-specific cellular uptake and subcellular localization of silica-coated CdSe quantum dots [29].

Several research groups have described the use of QDs for sensitive bioassays and cellular imaging in vitro and in vivo [30-34]. But many aspects of this approach need to be further optimized particularly for in vivo applications. Our preliminary in vitro investigations proved the cytocompatibility and stability of silica-coated CdSe QDs under biological conditions [29]. The objective of this study was to demonstrate that the silica-coated CdSe QDs can be used as labelling agents for long-life cellular imaging, cancer cellular imaging using cLSM and fluorescence microscopy. We use Inductively Coupled Plasma-Optical Emission (ICP-OES) Spectroscopy to quantify the in vitro cellular internalization efficiency of silica-coated CdSe QDs in a couple of cell models, New Zealand rabbit adipose derived mesenchymal stem cells (RADMSCs) and Human cervical cancer cells (HeLa). Imaging using Confocal microscopy is also performed to obtain a visual image of the QDs internalised by cells after incubation for a time interval, assessed from the results of ICP-OES studies. Simple fluorescence imaging and localization of the silica-coated CdSe QDs studies using fluorescent microscopy after staining with 4,6-diamidino-2-phenylindole (DAPI) have also been carried out in RADMSCs and HeLa cells. The high cellular internalisation efficiency as well as the high contrast confocal images of tumor sections obtained indicates the role of silica-coated CdSe QDs in imaging and labelling of cancer cells and other diseases.

Methods

Chemicals

Chemicals for QD synthesis and silica overcoating such as trioctylphosphine oxide (TOPO), trioctylphosphine, cadmium oxide (CdO), selenium powder, dodecylamine,

igepal and aminopropyl silane (APS) were purchased from Sigma–Aldrich and tetradecylphosphonic acid (TDPA) from Alfa Aesar and used as such without further purification.

Synthesis and characterization of silica-coated CdSe QDs

QDs were synthesized by following a reported procedure [35,36]. Cadmium oxide (0.067 g, 0.52 mmol), dodecylamine (3.8 g, 20.72 mmol), trioctylphosphine oxide (2.7 g, 6.9 mmol) and tetradecylphosphonic acid (0.40 g, 1.44 mmol) were heated to 300°C under vacuum, until CdO dissolved completely to produce an optically clear solution. At this temperature, an injection mixture containing trioctylphosphine (5.2 mL) and Se (0.083 mmol) was introduced. After desired crystal growth, the reaction was arrested by reducing the reaction temperature down to ambient conditions. The QDs were purified by reprecipitation with methanol and redispersed in dry chloroform for silica overcoating.

For overcoating CdSe QDs with silica, we have modified the reported procedure [18, 24] as follows: A mixture of TOPO capped CdSe QDs in chloroform (400 µL) and APS (0.075 g, 0.36 mM) was vortexed for 30 min, in an inert atmosphere. This mixture was added to Igepal CO-520 (1.3 mL) in cyclohexane (10 mL) and stirred for 30 min under dry conditions. Ammonia solution (150 µL, 33 wt.%) was added drop wise and the stirring was continued for 1 day. The silica-coated QDs were purified by washing repeatedly with dry chloroform and redispersed in PBS buffer (pH 7.3). Dots were stored in the dark at room temperature and the emission stability was investigated as a function of time, pH and ionic strength of the medium.

The electronic absorption spectra were recorded on a Shimadzu UV-3101 scanning spectrophotometer; emission spectra were collected using SPEX-Fluorolog F112X spectrofluorimeter. For HRTEM studies, a drop of nanoparticle solution was placed on a carbon coated Cu grid and the solvent was allowed to evaporate. Specimens were imaged on a FEI Tecnai G2 S-TWIN 300 kV high resolution transmission electron microscope.

Materials and animals

For cellular imaging experiments, all chemicals and reagents used were purchased from Sigma–Aldrich, USA. The New Zealand rabbit adipose tissue-derived mesenchymal stem cells (RADMSCs) and Human cervical cancer cells (HeLa) were maintained at 37°C and 5% CO₂ in Dulbecco's Modified Eagle's Medium (DMEM) supplemented with 10% fetal bovine serum (FBS) and antibiotics.

The animal models used for the tumor imaging study (Normal male, Swiss albino mice of 6 weeks old, weighing 20–25 g) were from the animal house facility, Department of Biochemistry, University of Kerala. For maintaining the experimental animals, the institutional ethical guidelines were absolutely followed as per Committee for the Purpose of CPCSEA rules [Sanction No: IAEC-KU-9/06-07/BC-AA (8) (ii)], Government of India. All animal experiments were performed in triplicate.

Solid tumor development

The solid tumor model was established by subcutaneous injection of DLA cells (1×10^6 cells/animal) into the back right hind limb of mice [37]. The mice were subjected to

imaging studies when the tumor volume reached 200–450 mm³ (3–4 weeks after inoculation).

Percentage of cellular internalization of silica-coated CdSe QDs – ICP-OES

For silica-coated CdSe QDs internalization studies, HeLa and RADMSCs were seeded 1×10^6 cells/well in 6-well plates and incubated overnight to allow for cell attachment. After the overnight incubation, cell medium was removed and replaced with 1 mL of DMEM medium. Cells were serum starved for 30 min and fresh medium containing the silica-coated CdSe QDs was added. Cells were incubated for different time interval (0, 1, 2, 3, 4 and 5 h) in the presence of silica-coated CdSe QDs. At each time point, cell supernatant was removed and cells were washed three times with PBS. Samples were digested in 500 μ L of nitric acid for 30 min at 70°C and diluted in 3 mL of double distilled water. Quantum dots internalization was quantified using ICP-OES (Optima 5300 DV instrument, Perkin Elmer). All experiments were performed in triplicate.

Cellular internalization studies of silica-coated CdSe QDs – cLSM

To study the cellular internalization efficiency of the silica-coated CdSe QDs in vitro, RADMSCs and HeLa cells were cultured on coverslips and the medium was changed on every 2 days, till 85% cell confluence was achieved. Then the cells were incubated at 37°C with 5% CO₂ for 2 and 4 h after adding with silica-coated CdSe QD solution (100 nM) in PBS. After incubation, the coverslips were taken out, rinsed thrice at 37°C using pre-heated PBS, fixed with 3.7% paraformaldehyde and fluorescence images were observed in a confocal laser scanning microscope (cLSM). An argon laser (488 nm) was used for QD excitation under confocal fluorescence microscope (Carl Zeiss LSM 510 META). All experiments were performed in triplicate.

Cellular imaging and localization study using silica-coated CdSe QDs – Fluorescence microscopy

For cellular imaging experiment, stem cells (RADMSC) and cancer cells (HeLa) were used. RADMSCs and HeLa were cultured on coverslips, and the medium was changed every 2 days, until 85% cell confluence was achieved. Then the cells were incubated at 37°C with 5% CO₂ for 4 h after adding with silica-coated CdSe QD solution (100 nM in PBS). Then, silica-coated CdSe QD solution (100 nM) was added to the cells. After incubation, the coverslips were taken out, rinsed thrice at 37°C using pre-heated PBS, fixed with 3.7% paraformaldehyde, after staining with 4,6-diamidino-2-phenylindole (DAPI) and fluorescence cellular images were observed using a DM 6000 fluorescence microscope (Leica, Germany, 20x objective, equipped with DFC 300 FX digital camera).

In the case of fluorescence imaging of cancer cells - Daltons Lymphoma Ascites (DLA) cells, were procured from Amala Cancer Research Centre, Thrissur, India and maintained in the peritoneal cavity of mice. Approximately 2 weeks were taken for the development of tumor in the peritoneal cavity and matured cells were aspirated from the peritoneum, washed with PBS and seeded on coverslips. Then the cells were incubated at 37°C with 5% CO₂ for 2 and 4 h after adding with silica-coated CdSe QD solution (100 nM) in PBS. After incubation, the coverslips were taken out, rinsed thrice at 37°C using pre-heated PBS, fixed with 3.7% paraformaldehyde and fluorescence images were observed in a DM 6000 fluorescence microscope (Leica, Germany, 20x objective, equipped with DFC 300 FX digital camera).

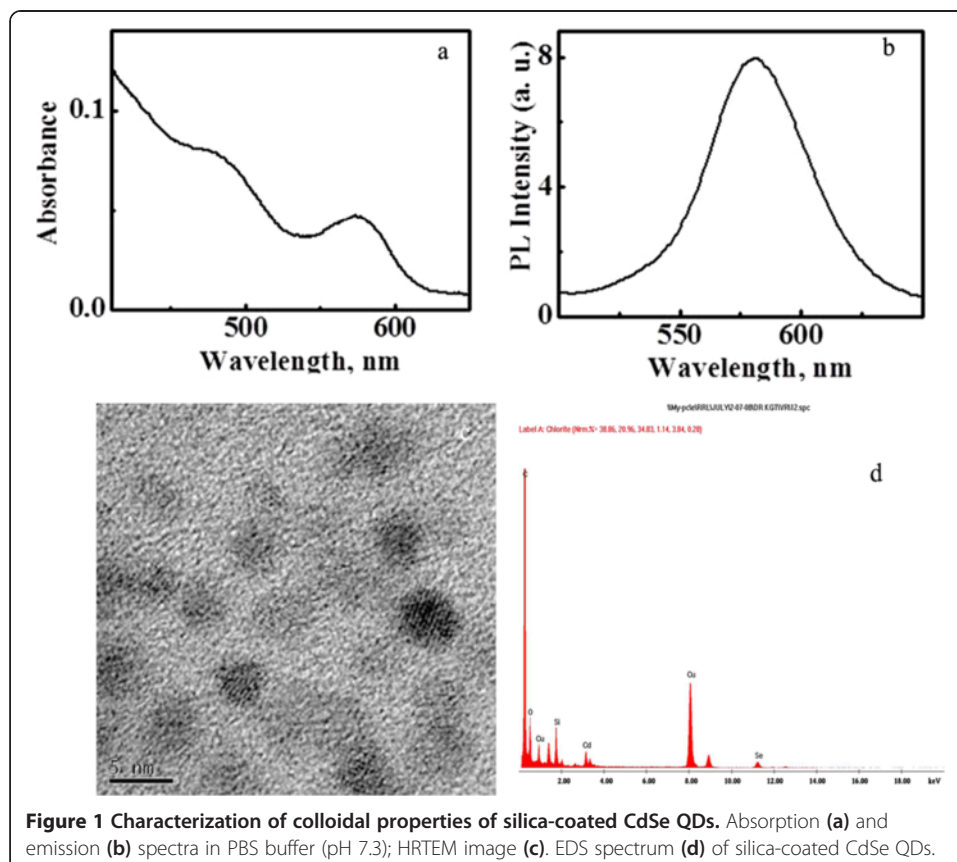
Fluorescence imaging of ultrathin sections of tumor- cLSM

Tumor mice were anesthetized by intra-abdominal injection of 3% pentobarbital sodium (45 mg/kg). QD and QD-Ab probe were (10 nM) injected at the dosage of 20 ml/kg into the tail vein. After the injection, the mice were put in a dark chamber for 4 h, and then tumor region were stripped from the mice. Then tumor tissues was fixed and sectioned by ultramicrotome (LKB; Bromma-2088-Ultratome[®]V, Sweden), to obtain 100 nm ultrathin sections of tumor. For fluorescence imaging, these ultrathin tissue sections were directly examined with a cLSM. The ultrathin sections were stained with toluidine blue and observed under a light microscope (Leica-DMIL, Germany).

Results and discussion

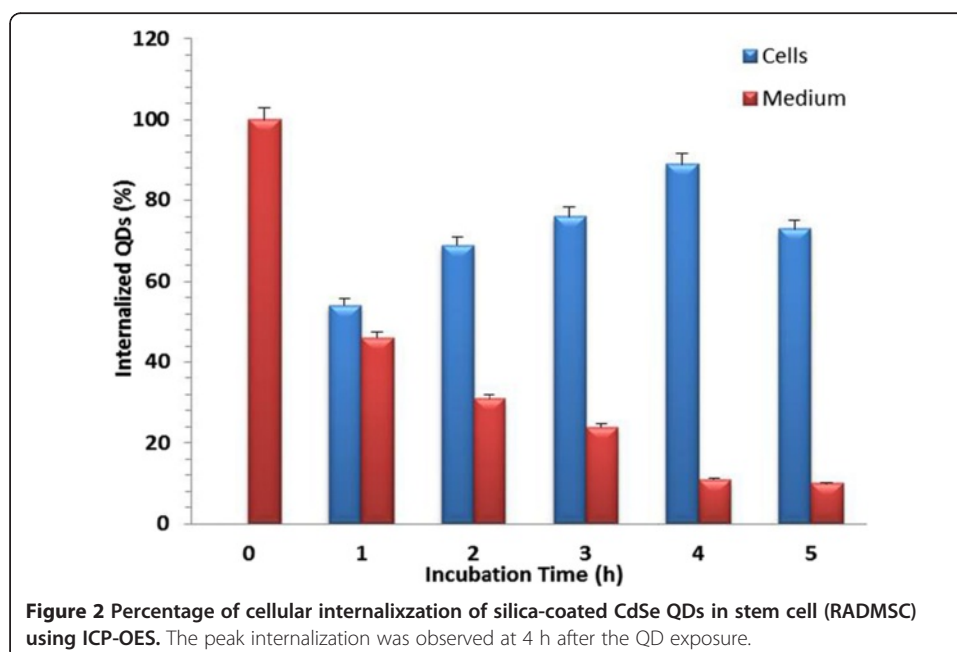
Triethylphosphine oxide capped CdSe QDs were synthesised and made water soluble by silanisation. The details of QDs synthesis, silanisation and the evaluation of colloidal properties of QDs have been published elsewhere in detail [29]. The silica-coated CdSe QDs obtained showed a greenish yellow emission in PBS buffer (pH 7.3). The size homogeneity was ensured using high resolution transmission electron microscope (HRTEM), and colloidal properties of QDs were depicted in Figure 1. The QDs were found to be readily soluble in aqueous medium, stable and luminescing under biological conditions.

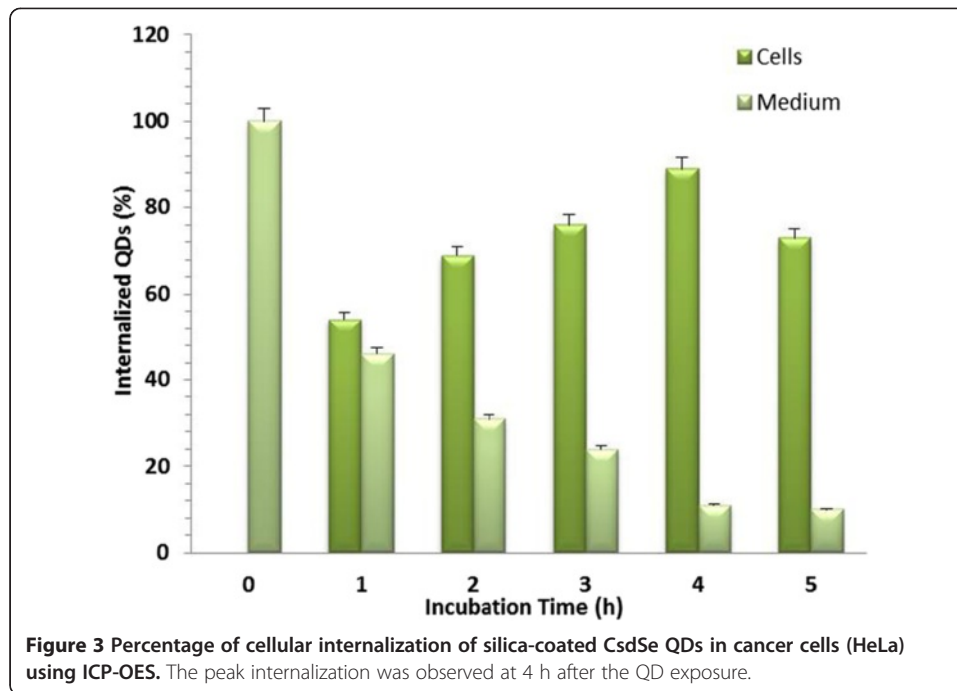
The cytocompatibility of cadmium chalcogenides based QDs are of primary attention when there biological applications are concerned. The toxicological effect is attributed to the leaching out of core materials, particularly cadmium ions into the cell media



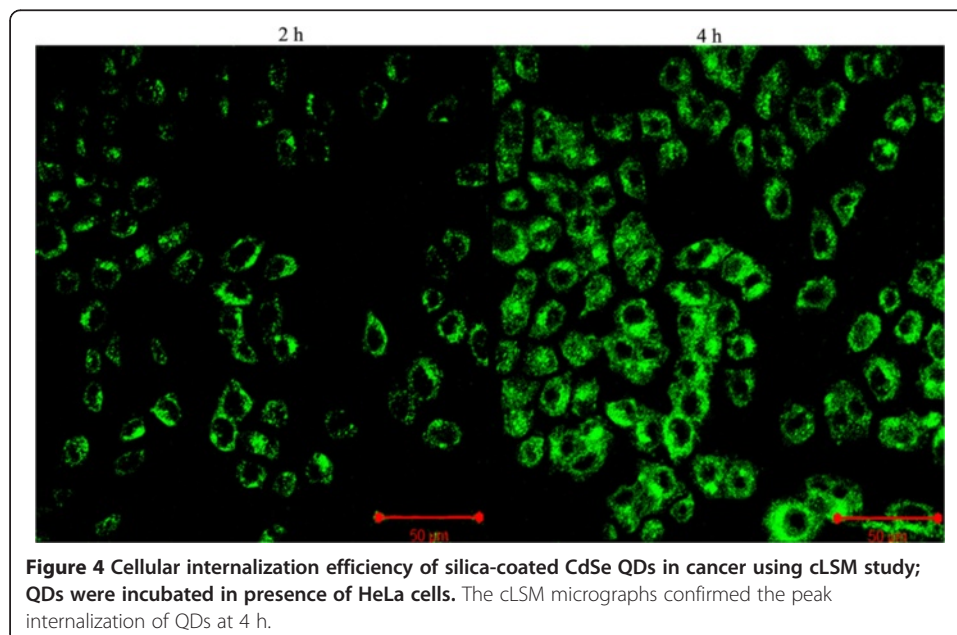
[38,39]. The non-toxicity of the QDs used in our studies is ensured by generating a CdSe/Silica core shell structure preventing any leakage of core materials. Also, the silica shell ensures aqueous solubility which is essential for biological studies and applications [28]. One of the emerging applications of QDs is cellular labelling and imaging. In the current experiment, we have used a couple of cell models-HeLa cells and RADMSCs, to monitor the in vitro cellular internalisation efficiency of silica-coated QDs as a function of time. The results obtained from the ICP-OES studies undoubtedly prove the high internalisation efficiency and cytocompatibility of silica-coated CdSe QDs. These observations confirm their potential applications in cellular imaging. This aspect was further confirmed by confocal laser scanning microscopy (cLSM) studies. For this, cells were incubated with QDs for 1, 2, 3, 4 and 5 h and then washed to remove any unbound QDs. Percentage of cellular internalization of silica-coated CdSe QDs were studied by using Inductively Coupled Plasma-Optical Emission Spectroscopy (ICP-OES). ICP-OES was performed utilizing an Optima 5300 DV instrument (Perkin Elmer). ICP-OES was used to measure the cadmium content to indicate the concentration of QDs in cells collected at predetermined time intervals (0–5 h). The ICP-OES instrument was initialized, optimized and standardized using the manufacturer's recommendations. Standardization was performed using multielement standards (Perkin Elmer Life and Analytical Sciences). The result obtained from the ICP-OES studies is shown in Figures 2 & 3, where the percentage of internalised QDs is plotted as a function of incubation time. Results showed that the maximum internalization observed at 4 h in both stem and cancer cells after the QD exposure. Overall internalization efficiency was found to be significant high in cancer cells when compared to stem cells (Figures 2 & 3).

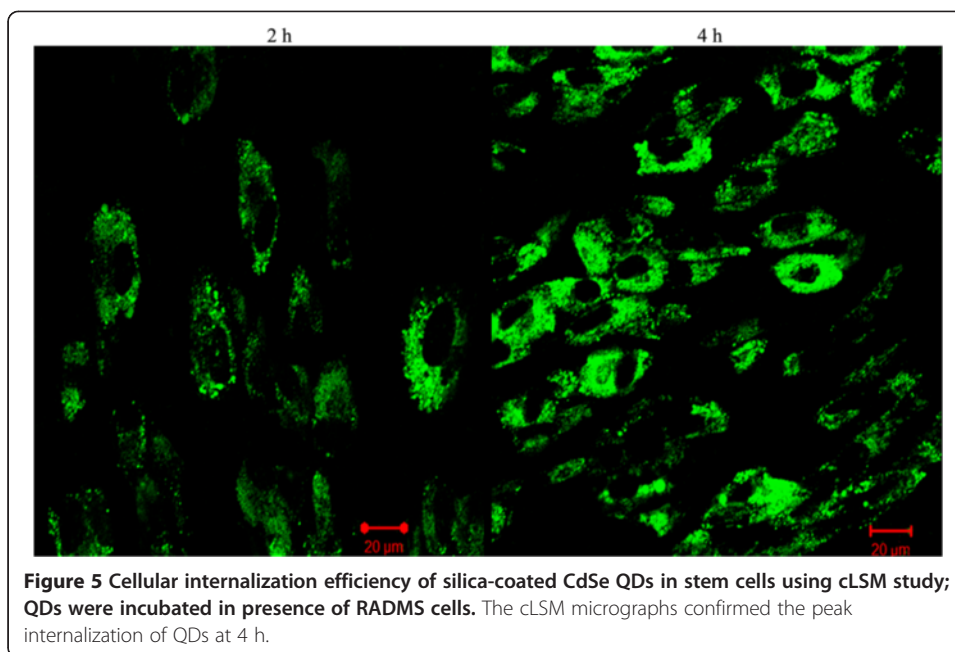
The cellular internalization efficiency of silica-coated CdSe QDs was monitored by confocal laser scanning microscopy (cLSM). For this, cells were incubated with QDs for the predetermined time intervals and then washed to remove any unbound QDs. Interestingly, we were able to observe a uniform fluorescence with high contrast in





most of the cells, when examined under cLSM (Figures 4 & 5). The peak internalization time (4 h) was again confirmed by the images obtained from cLSM analysis. cLSM imaging were performed in all time intervals (1, 2, 3, 4 and 5 h), and images for 2 and 4 h are showing in Figures 4 & 5. The fluorescence signal was seen in cytoplasm as well as in the vicinity of nucleus, in case of both cell models under study. These results confirmed the significant internalization of QDs into cytoplasmic vesicles. The fluorescence obtained within the cells is attributed to the trapping of QDs in the endocytic intracellular vesicles. These observations are in agreement with previous reports demonstrating the uptake of QD conjugates,





which was mediated by endocytosis [40]. Jaiswal and his co-workers also reported a similar mechanism of QDs uptake, and the fluorescence observed from cells was attributed to the internalization of QDs by endocytosis [13]. Thus internalized silica-coated CdSe QDs were non-toxic during this time span at concentrations needed for uptake study in these cells. Cytocompatibility of these fluorescent materials were reported earlier by our previous report [28], indicating that the silica layer makes the QD dispersible in aqueous media and also prevents the leakage of toxic cadmium and selenium ions. The silica-coated CdSe QDs were found to be efficiently internalized by cancer cells and stem cells *in vitro*, thereby highlighting their potential to be used as a non-toxic optical probe for biomedical diagnostics as well as cellular labelling applications. To the best of our knowledge, this is the first report of using silica-coated CdSe QDs for the cellular imaging, cancer imaging and internalization studies.

Further, we have applied these QDs for different cellular imaging applications and localization studies by growing the cancer cells (HeLa) and stem cells (RADMSC) for 4 h in the presence of QDs. Before the cellular imaging experiment, the cell viability was studied systematically using cell viability assays, and the cell proliferation assay data showed the absence of cell death (in comparison to control experiments), confirming that the core material is well protected inside the silica shell (data not shown) and these observations were published elsewhere [28]. In 2009, Han's group reported the CdSeS QD/SiO₂ nanoparticles showing good resistance to photobleaching and low cytotoxicity, which facilitates their use in live cell imaging [27]. Our cellular imaging results showed that silica-coated CdSe QDs were significantly internalized into cytoplasmic vesicles and produce the good fluorescence in and around the cells (both stem and cancer cells) (Figure 6). Generally, we found that the silica-coated CdSe QDs show much higher cellular internalization efficiency, possibly because of cytocompatibility of silica shell.

Interestingly, *in vivo* fluorescence based tumor imaging results showed better localisation of QDs in the tumor site (Figure 7). Evident from the results shown in Figure 7b

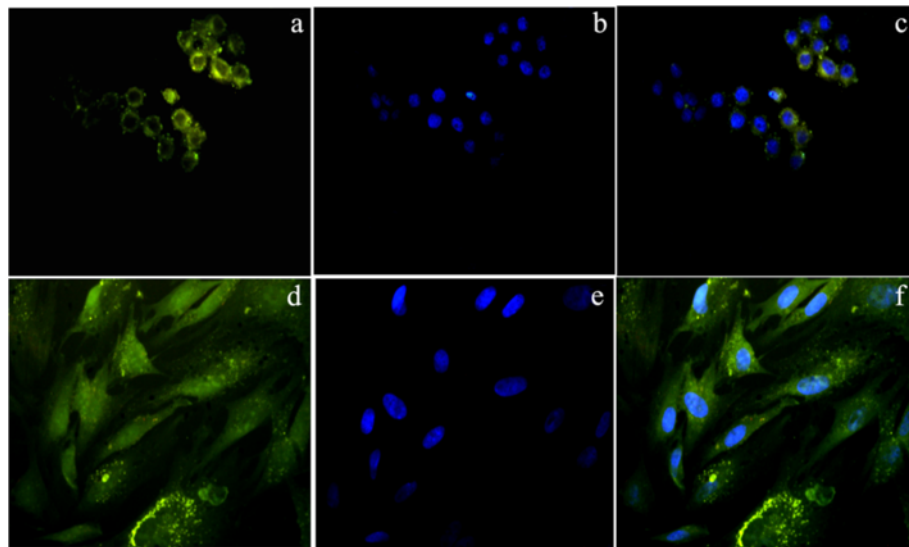


Figure 6 Cellular imaging-fluorescence microscopy: (a & d) HeLa and RADMSC cells showed the efficient cellular internalization of QDs in the cytoplasm and nearby nucleus [yellowish green fluorescence]; (b & e) Nucleus were counterstained blue with DAPI [blue fluorescence]; (c & f) Overlay of QDs and DAPI fluorescence images.

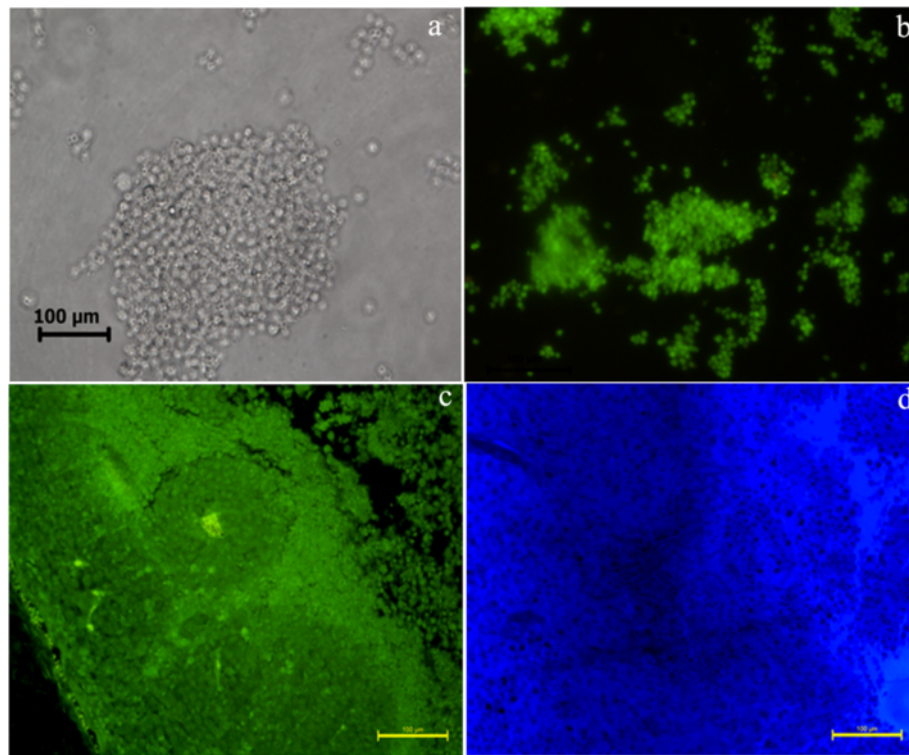


Figure 7 Fluorescence imaging of tumor sections using silica-coated CdSe QDs. Phase contrast microscopic image of DLA cells (a). Fluorescence microscope image of DLA cells (b). Fluorescence microscope image of ultra thin section of solid tumor (c). Light microscopic image of toluidine blue stained ultra-thin section of tumor (d).

& c, the high contrast fluorescence signal appeared in the DLA cells in vitro and in vivo (ultra-thin section), and Figure 7a shows morphology of DLA cells using phase contrast microscopy. Ultrathin sections of tumor tissue exhibited strong fluorescence which specifies the tumor-targeted imaging efficiency of QDs in living mice (Figure 7c). The light microscopic image of toluidine blue stained ultrathin tumor sections showing the presence of growing DLA cells in the tumor (Figure 7d). With these excellent properties and efficient internalization efficiency, silica-coated CdSe QDs might become a highly promising probe for biological applications. These results establish silica-coated CdSe QDs as an extremely useful tools for molecular imaging, cell tracking and labeling to study the cell division and every cellular events especially metastasis of cancer and other diseases.

Conclusions

The in vitro cellular internalization efficiency of silica-coated CdSe QDs is systematically followed in this report. Cytocompatibility assessment experiments based on various assays showed that the silanised QDs were non-toxic, aqueous soluble and showed stable fluorescence under biological conditions. The ICP-OES measurements in stem cells (RADMSC) and in cancer cells (HeLa) confirmed that silica-coated CdSe QDs have excellent effective internalization efficiency and the peak concentration was observed after 4 hours. In addition, the high contrast images obtained in confocal laser scanning microscopy (cLSM) from the in vitro cellular imaging study on stem cells and cancer cells and in vivo tumor imaging studies further confirmed that these QDs are highly useful for cellular imaging and labelling applications using their relatively stable fluorescence emission under biological conditions. Overall, this study implies that silica-coated CdSe QDs could be used as labelling and imaging agents for cancer cellular imaging and cell tracking applications for the study of cancer and other diseases.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

MV carried out the Solid tumor development, cellular internalization, Cellular imaging and localization study, Fluorescence imaging Article composing; RV carried out the Synthesis and characterization of silica-coated CdSe QDs; AJ carried out the tumor ultrathin sectioning study; FBF carried out the cLSM study; AA carried out the designing of experiments, interpretation of results. All authors read and approved the final manuscript.

Acknowledgement

This work was supported by the research grant from the Department of Biotechnology, Ministry of Science and Technology, Govt. of India, New Delhi (Order No. BT/PR9904/NNT/28/63/2007) to the corresponding author Prof. Annie Abraham. We thank Prof. K. George Thomas, Dean, Indian Institutes of Science Education and Research (IISER), Thiruvananthapuram, India for providing nanomaterials for the study. Thanks are also due to Dr. T.V. Anilkumar, SCTIMST, Thiruvananthapuram for confocal microscopy, and Dr. H. K. Varma for the ICP-OES analysis.

Author details

¹Department of Biochemistry, University of Kerala, Kariavattom campus, 695581 Thiruvananthapuram, Kerala, India.

²Photosciences and Photonics, National Institute for Interdisciplinary Science and Technology (CSIR), Thiruvananthapuram, Kerala, India. ³TEM Laboratory, Sree Chitra Tirunal Institute of Medical Sciences and Technology, Biomedical Technology Wing, Thiruvananthapuram, Kerala, India.

Received: 10 June 2013 Accepted: 29 April 2014

Published online: 27 June 2014

References

1. Fottner C, Mettler E, Goetz M, Schirmacher E, Anlauf M, Strand D, Schirmacher R, Klöppel G, Delaney P, Schreckenberger M, Galle PR, Neurath MF, Kiesslich R, Weber MM (2010) In Vivo molecular imaging of somatostatin receptors in pancreatic islet cells and neuroendocrine tumors by miniaturized confocal laser-scanning fluorescence microscopy. *Endocrinology* 151(5):2179–2188

2. Zhang J, Jia X, Lv XJ, Deng YL, Xie HY (2010) Fluorescent quantum dot-labeled aptamer bioprobes specifically targeting mouse liver cancer cells. *Talanta* 81(1–2):505–509
3. Davis SC, Samkoe KS, O'Hara JA, Gibbs-Strauss SL, Payne HL, Hoopes PJ, Paulsen KD, Pogue BW (2010) MRI-coupled fluorescence tomography quantifies EGFR activity in brain tumors. *Acad Radiol* 17(3):271–276
4. Do W, Hwang HY, Ko JH, Lee HK, Ryu SH, Song IC, Lee DS, Kim S (2010) A nucleolin-targeted multimodal nanoparticle imaging probe for tracking cancer cells using an aptamer. *J Nucl Med* 51(1):98–105
5. Zhang Q (2013) Imaging single synaptic vesicles in Mammalian central synapses with quantum dots. *Methods Mol Biol* 995:179–191
6. Madani SY, Shabani F, Dwek MV, Seifalian AM (2013) Conjugation of quantum dots on carbon nanotubes for medical diagnosis and treatment. *Int J Nanomedicine* 8:941–950
7. Gao XH, Nie SM (2003) Molecular profiling of single cells and tissue specimens with quantum dots. *Trends Biotechnol* 21(9):371–373
8. Jovin TM (2003) Quantum dots finally come of age. *Nat Biotechnol* 21(1):32–33
9. Deng D, Cao J, Qu L, Achilefu S, Gu Y (2013) Highly luminescent water-soluble quaternary Zn-Ag-In-S quantum dots for tumor cell-targeted imaging. *Phys Chem Chem Phys* 15(14):5078–5083
10. Lian S, Zhang P, Gong P, Hu D, Shi B, Zeng C, Cai L (2012) A universal quantum dots-aptamer probe for efficient cancer detection and targeted imaging. *J Nanosci Nanotechnol* 12(10):7703–7708
11. Lidke DS, Nagy P, Heintzmann R, Jovin DJA, Post JN, Grecco HE, Jares-Erijman EA, Jovin T (2004) Quantum dot ligands provide new insights into erbB/HER receptor-mediated signal transduction. *Nat Biotechnol* 22(2):198–203
12. Bonasio R, Carman CV, Kim E, Sage PT, Love KR, Mempel TR, Springer TA, von Andrian UH (2007) Specific and covalent labeling of a membrane protein with organic fluorochromes and quantum dots. *Proc Natl Acad Sci USA* 104(37):14753–14758
13. Jaiswal JK, Mattoussi H, Mauro JM, Simon SM (2003) Long-term multiple color imaging of live cells using quantum dot bioconjugates. *Nat Biotechnol* 21(1):47–51
14. Parak WJ, Boudreau R, Le Gros M, Gerion D, Zanchet D, Micheel CM, Williams SC, Alivisatos AP, Larabell C (2002) Cell motility and metastatic potential studies based on quantum dot imaging of phagokinetic tracks. *Adv Mater* 14(12):882–885
15. Hanaki K, Momo A, Oku T, Komoto A, Maenosono S, Yamaguchi Y, Yamamoto K (2003) Semiconductor quantum dot/albumin complex is a long-life and highly photostable endosome marker. *Biochem Biophys Res Commun* 302(3):496–501
16. Stroh M, Zimmer JP, Duda DG, Levchenko TS, Cohen KS, Brown EB, Scadden DT, Torchilin VP, Bawendi MG, Fukumura D, Jain RK (2005) Quantum dots spectrally distinguish multiple species within the tumor milieu in vivo. *Nat Med* 11:678–72
17. Cai WB, Hsu AR, Li ZB, Chen XY (2007) Are quantum dots ready for in vivo imaging in human subjects? *Nanoscale Res Lett* 2:265–281
18. Vibin M, Vinayakan R, John A, Raji V, Rejija CS, Abraham A (2011) Fluorescence imaging of stem cells, cancer cells and semi-thin sections of tissues using silica-coated CdSe quantum dots. *J Fluoresc* 21:1365–1370
19. Yong KT, Ding H, Roy I, Law WC, Bergey EJ, Maitra A, Prasad PN (2009) Imaging pancreatic cancer using bioconjugated InP quantum dots. *ACS Nano* 3(3):502–510
20. Law WC, Yong KT, Roy I, Ding H, Hu R, Zhao WW, Prasad PN (2009) Aqueous-phase synthesis of highly luminescent CdTe/ZnTe core/shell quantum dots optimized for targeted bioimaging. *Small* 5(11):1302–1310
21. Yang LL, Mao H, Wang YA, Cao ZH, Peng XH, Wang XX, Duan HW, Ni CC, Yuan QG, Adams G, Smith MQ, Wood WC, Gao XH, Nie SM (2009) Single chain epidermal growth factor receptor antibody conjugated nanoparticles for in vivo tumor targeting and imaging. *Small* 5(2):235–243
22. Pellegrino T, Kudera S, Liedl T, Javier AM, Manna L, Parak WJ (2005) On the development of colloidal nanoparticles towards multifunctional structures and their possible use for biological applications. *Small* 1(1):48–63
23. Darbandi M, Thomann R, Nann T (2005) Single Quantum dots in silica spheres by microemulsion synthesis. *Chem Mater* 17(23):5720–5725
24. Selvan ST, Patra PK, Ang CY, Ying JY (2007) Synthesis of silica-coated semiconductor and magnetic quantum dots and their use in the imaging of live cells. *Angew Chem Int Ed Engl* 46(14):2448–2452
25. Bakalova R, Zhelev Z, Aoki I, Ohba H, Imai Y, Kanno I (2006) Silica-shelled single quantum dot micelles as imaging probes with dual or multimodality. *Anal Chem* 78(16):5925–5932
26. Chen F, Gerion D (2004) Fluorescent CdSe/ZnS nanocrystal – peptide conjugates for long-term, nontoxic imaging and nuclear targeting in living cells. *Nano Lett* 4(10):1827–1832
27. Han R, Yu M, Zheng Q, Wang L, Hong Y, Sha Y (2009) A facile synthesis of small-sized, highly photoluminescent, and monodisperse CdSeS QD/SiO₂ (2) for live cell imaging. *Langmuir* 25(20):12250–12255
28. Vibin M, Vinayakan R, John A, Raji V, Rejija CS, Vinesh NS, Abraham A (2011) Cytotoxicity and fluorescence studies of silica-coated CdSe quantum dots for bioimaging applications. *J Nanopart Res* 13:2587–2596
29. Vibin M, Vinayakan R, John A, Raji V, Rejija CS, Abraham A (2011) Cellular uptake and subcellular localization of highly luminescent silica-coated CdSe quantum dots – In vitro and in vivo. *J Colloid Interface Sci* 357:366–371
30. Alivisatos AP (1996) Semiconductor clusters, nanocrystals, and quantum dots. *Science* 271:933–937
31. Kim S, Lim YT, Soltesz EG, De Grand AM, Lee J, Nakayama A, Parker JA, Mihaljevic T, Laurence RG, Dor DM, Cohn LH, Bawendi MG, Frangioni JV (2004) Near-infrared fluorescent type II quantum dots for sentinel lymph node mapping. *Nat Biotechnol* 22:93–97
32. Gao XH, Cui YY, Levenson RM, Chung LWK, Nie SM (2004) In vivo cancer targeting and imaging with semiconductor quantum dots. *Nat Biotechnol* 22(8):969–976
33. Pic E, Pons T, Bezdetsnaya L, Leroux A, Guillemin F, Dubertret B, Marcha F (2010) Fluorescence imaging and whole-body biodistribution of near-infrared-emitting quantum dots after subcutaneous injection for regional lymph node mapping in mice. *Mol Imaging Biol* 12(4):394–405
34. Kwon H, Lee J, Song R, Hwang SI, Lee J, Kim YH, Lee HJ (2013) In vitro and in vivo imaging of prostate cancer angiogenesis using anti-vascular endothelial growth factor receptor 2 antibody-conjugated quantum dot. *Korean J Radiol* 14(1):30–37

35. Peng XA, Peng XG (2001) Formation of high-quality CdTe, CdSe, and CdS nanocrystals using CdO as precursor. *J Am Chem Soc* 123(1):183–184
36. Vinayakan R, Shanmugapriya T, Nair PV, Ramamurthy P, Thomas K (2007) An approach for optimizing the shell thickness of core-shell quantum dots using photoinduced charge transfer. *J Phys Chem C* 111:10146–10149
37. Siveen KS, Kuttan G (2011) Immunomodulatory and antitumor activity of *Aerva lanata* ethanolic extract. *Immunopharmacol Immunotoxicol* 33:423–432
38. Derfus AM, Chan WCW, Bhatia SN (2004) Probing the cytotoxicity of semiconductor quantum dots. *Nano Lett* 4:11–18
39. Kirchner C, Liedl T, Kudera S, Pellegrino T, Javier AM, Gaub HE (2005) Cytotoxicity of colloidal CdSe and CdSe/ZnS nanoparticles. *Nano Lett* 5:331–338
40. Lim YT, Kim S, Nakayama A, Stott NE, Bawendi MG, Frangioni JV (2003) Selection of quantum dot wavelengths for biomedical assays and imaging. *Mol Imaging* 2(1):50–64

doi:10.1186/s12645-014-0001-y

Cite this article as: Vibin et al.: Effective cellular internalization of silica-coated CdSe quantum dots for high contrast cancer imaging and labelling applications. *Cancer Nanotechnology* 2014 5:1.

Submit your manuscript to a SpringerOpen[®] journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com
