

Diagnostic and treatment strategies in mucopolysaccharidosis VI

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Abstract: Mucopolysaccharidosis VI (MPS VI) is a very rare autosomal recessive disorder caused by mutations in the *ARSB* gene, which lead to deficient activity of the lysosomal enzyme ASB. This enzyme is important for the breakdown of the glycosaminoglycans (GAGs) dermatan sulfate and chondroitin sulfate, which accumulate in body tissues and organs of MPS VI patients. The storage of GAGs (especially dermatan sulfate) causes bone dysplasia, joint restriction, organomegaly, heart disease, and corneal clouding, among several other problems, and reduced life span. Despite the fact that most cases are severe, there is a spectrum of severity and some cases are so attenuated that diagnosis is made late in life. Although the analysis of urinary GAGs and/or the measurement of enzyme activity in dried blood spots are useful screening methods, the diagnosis is based in the demonstration of the enzyme deficiency in leucocytes or fibroblasts, and/or in the identification of pathogenic mutations in the *ARSB* gene. Specific treatment with enzyme replacement has been available since 2005. It is safe and effective, bringing measurable benefits and increased survival to patients. As several evidences indicate that early initiation of therapy may lead to a better outcome, newborn screening is being considered for this condition, and it is already in place in selected areas where the incidence of MPS VI is increased. However, as enzyme replacement therapy is not curative, associated therapies should be considered, and research on innovative therapies continues. The management of affected patients by a multidisciplinary team with experience in MPS diseases is highly recommended.

Keywords: mucopolysaccharidosis VI, Maroteaux–Lamy syndrome, dermatan sulfate, arylsulfatase b, enzyme replacement therapy, lysosomal storage diseases

Introduction

Mucopolysaccharidosis VI (MPS VI), or Maroteaux–Lamy syndrome (OMIM 253200) is a rare genetic disease first described in 1963 by the French doctors Pierre Maroteaux and Maurice Lamy.¹ The disease is inherited as an autosomal recessive trait and is caused by mutations in the *ARSB* gene, that encodes the lysosomal enzyme E.C.3.1.6.12 or ASB.²

The enzyme deficiency leads to the accumulation of the glycosaminoglycan (GAG) dermatan sulfate (DS) into the lysosome, leading to a variety of signs and symptoms in multiple organs and systems.³ The disorder is clinically heterogeneous. The majority of patients are severely affected (rapidly advancing form). In a minority of patients the disease is more attenuated (slowly-progressing form).⁴ International studies report that the incidence of MPS VI ranges from one in 43,261 births in a Turkish population in Germany to one in 1,505,160 births in Sweden.⁵ Table 1 shows the epidemiological studies performed in different countries to date.^{4,6-17} Some populations seem to have increased frequencies of mutations in the gene *ARSB*. The prevalence identified in a

Table 1 Frequency of MPS (all types) and MPS VI reported in the literature

Country/ Region	Frequency of MPS (all types) ^a	Frequency of MPS VI ^a	Reference
The Netherlands	4.5	0.15	Poorthuis et al ⁶
Australia	4.4	0.4	Meikle et al ⁷
Western Australia	3.4	0.31	Nelson et al ⁸
Northern Portugal	4.8	0.42	Pinto et al ⁹
Germany	3.53	0.23	Baehner et al ¹⁰
Sweden	1.75	0.07	Malm et al ¹¹
Norway	3.08	0.07	Malm et al ¹¹
Denmark	1.77	0.05	Malm et al ¹¹
Tunisia	2.29	0.35	Ben Turkia et al ¹²
Taiwan	2.04	0.14	Lin et al ¹³
Czech Republic	3.72	0.05	Poupetová et al ¹⁴
Saudi Arabia	NA	8.0	Moammar et al ¹⁵
Estonia	4.05	0.27	Krabbi et al ¹⁶
Brazil (Monte Santo, a small city in the Northeast)	NA	25.0	Costa- Motta et al ⁴
Poland	1.81	0.0132	Jurecka et al ¹⁷

Note: ^aCases per 100,000 live births.

Abbreviations: MPS VI, mucopolysaccharidosis VI; NA, not available.

Turkish population living in Germany, for example, was higher than the non-Turkish population.¹⁰ It has also been reported that in Brazil,¹⁸ and in Portugal,⁹ from which many people migrated to Brazil, the frequency of MPS VI is proportionally increased considering the usual distribution of MPS types. In Brazil, for example, in the city of Monte Santo, state of Bahia, the prevalence of MPS VI is estimated as 1:5,000, probably due to a combination of founder effect and endogamy, with a single mutation (p.H178L) present in homozygosis in all cases. A pilot program of newborn screening for MPS VI was implemented in this region, based on measurement of the enzyme activity and the detection of the common mutation.⁴

Clinical features

The clinical presentation of MPS VI is quite variable regarding the age of onset of symptoms and the disease progression.

The severe form of MPS VI, or rapidly progressive, is characterized by the onset of symptoms between 2 and 3 years of age, restricted joint mobility in infancy, delayed puberty, cervical spine compression, and respiratory failure. Most

patients die of cardiac decompensation in the second or third decade of life.² Although they have no cognitive deficit related directly to the disease, patients may present difficulties due to hearing impairment, visual, and physical limitations. Physical characteristics include joint restriction, pectus carinatum, kyphoscoliosis, macrocephaly, hepatosplenomegaly, umbilical and inguinal hernia, beyond the typical facial features such as low nasal bridge, hirsutism, gingival hypertrophy, dental malocclusion, macroglossia, and corneal opacification (which can lead to glaucoma and papilledema)^{19,20} (Figure 1). Patients may progress to respiratory failure, heart disease, severe joint involvement, and spinal cord compression (SCC) requiring surgical intervention.^{3,21} Cholestasis is rare, but has been reported.²²

The mild form, or slowly progressive, is characterized by attenuated symptoms, due to lower levels of DS.²³ The patients can develop skeletal complications such as short stature, carpal tunnel syndrome, Dupuytren's contracture, and hip dysplasia. In addition, adult patients may have valve disease, obstructive sleep apnea, and pulmonary complications²⁴ (Figure 2).

Unfortunately, delays to or missed diagnosis are common especially related to patients with mild form since they may present without a typical coarse facies and with less pronounced skeletal disease.

It is believed that the greater the residual activity of ASB, the more attenuated the symptoms,²⁵ but this correlation was not confirmed by all groups.²⁶ The correlation with urinary excretion of GAGs is another factor to be considered for the prognosis. A study with 121 patients with untreated MPS VI was performed and a classification into two groups was suggested. Patients with urinary GAG excretion greater than 200 ug/mg creatinine had more severe disease characterized by short stature, low weight, greater difficulty in performing the 6-minute walk test, greater impairment of lung function, and joint restriction. They were characterized as patients with the rapidly progressive form. Patients over 20 years of age rarely showed GAG excretion greater than 100 ug/mg creatinine, suggesting that levels below 100 ug/mg creatinine predict a greater survival.

They were classified as slowly progressive patients. These cases tend to have higher height and levels of GAGs below 100 ug/mg creatinine.²³ Although this classification is widespread and frequently used, there are several descriptions of patients with intermediate phenotypes.²⁷⁻²⁹ In general, MPS VI patients have a chronic progressive course and multisystem especially the musculoskeletal system, cardiopulmonary system, cornea, skin, liver, spleen, brain, and meninges.³



Figure 1 Male patient (14 years old) with the rapidly advancing form of MPS VI.
Abbreviation: MPS VI, mucopolysaccharidosis VI.



Figure 2 Female patient (11 years old) with the slowly advancing form of MPS VI.
Abbreviation: MPS VI, mucopolysaccharidosis VI.

Biochemical diagnosis

In general, the first step in diagnosis of MPS is the assessment of urinary GAGs excretion. GAGs are unbranched polysaccharide chains containing repeated disaccharides with uronic acid, and galactose or hexosamine. Except for hyaluronic acid, sulfate ions are also associated with these substances. Chondroitin sulfate (CS), heparan sulfate (HS), DS, and keratan sulfate are the main GAGs present in body tissues. ASB deficiency in MPS VI causes an important accumulation in tissues of DS and CS, and these compounds are excreted in high concentrations in the urine.²

Using only qualitative screening tests (such as Berry spot test) is not recommended anymore (due to the high proportion of false results) and was replaced by more refined and quantitative colorimetric methods.³⁰ The urinary GAGs may be analyzed quantitatively (to measure the total amount of GAGs) and/or qualitatively (to identify the specific GAG species which are elevated). Performing both analyses simultaneously is strongly recommended, as it reduces the risk of missing an MPS diagnosis. Since quantitative analysis is non-specific, qualitative urinary GAGs analysis is needed to identify the exact GAG types which are elevated, guiding the subsequent investigation needed to identify the specific MPS type. It is important to note that urinary GAGs analyses, quantitative or qualitative, are screening methods and its results should never be considered diagnostic for a specific MPS type. Quantitative GAGs analysis is also important because it allows monitoring a patient's response to specific treatment.^{31–33}

Quantitative analysis is usually performed by colorimetric analysis of urine combined with a cationic blue dye. The most used is the dimethylmethylene blue based method. Urinary GAGs concentration varies with age, and newborns have higher excretion than adults, therefore there are different reference ranges according to the age of the patients.³⁴

To identify the distinct GAGs excreted in urine, a qualitative pattern can be obtained by thin-layer chromatography (TLC) and/or electrophoresis (ELP). TLC and ELP are qualitative techniques, and a careful interpretation of the results by an experienced technician is required. Generally, these methods provide good results, especially when ELP is used. However, some ambiguous results or even false positives may also occur due to subjective interpretation of the different patterns.³²

In MPS VI, the GAGs which are increased are DS and CS. However, DS is also excreted in excess in MPS I, II, and VII, in conjunction with HS (and also with CS, in the case of MPS VII) (Figure 3). Although differentiation of MPS VI from the other MPS disorders causing excess DS excretion

seems straightforward, there are several complicating factors. It is not uncommon to observe a strong signal of DS with a faint signal of HS in patients with MPS I or II. Therefore, even if DS is detected, other MPS types should not be immediately ruled out. Conversely, it has been noted that trace amounts of HS may be present in urine of patients with MPS VI or even in the urine of young children (<3 years of age). It is not possible to firmly differentiate MPS I, II, VI, and VII using qualitative urinary GAGs analysis.³¹ Advantages of urinary GAGs analysis include non-invasive sample collection, relative ease of performance, and availability at many laboratories, including non-specialized ones. Urine collected in the morning is preferred, as it is usually more concentrated, but not mandatory. However, diluted samples should be avoided, as it can lead to false or ambiguous results. In normal neonates the concentration of GAGs may be increased, hindering the detection of MPS patients.³¹ In an attempt to overcome the difficulties of transportation of liquid urine samples, a new method which uses dried-urine filter paper samples for reliable mucopolysacchariduria detection has been described.³⁵

Quantitative urinary GAGs excretion can be in normal range in some MPS patients, especially in adults with MPS III and MPS IV. So, a negative result in the assay of urinary GAGs should be considered with caution, and the investigation should be continued if clinical suspicion of MPS persists.^{31,32,36}

It is expected that the measurement of GAGs by tandem mass spectrometry would progressively replace colorimetric quantification and TLC/ELP qualitative evaluations,³⁷ but

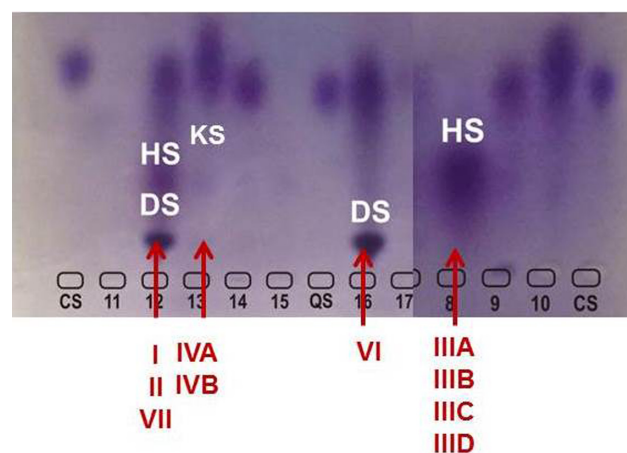


Figure 3 Electrophoresis of urinary GAGs.

Notes: The pattern observed in MPS VI, with presence of marked amounts of DS is indicated. The patterns observed in other MPS types are also indicated (with increased amounts of DS + HS, HS only or KS only).

Abbreviations: MPS, mucopolysaccharidosis; GAGs, glycosaminoglycans; DS, dermatan sulfate; HS, heparan sulfate; KS, keratan sulfate.

technology may take some time to be incorporated in the routine evaluation due to the high cost of equipment.

A positive result in the evaluation of urinary GAGs should be followed by more specific methods until a conclusion is reached. The usual following step is the measurement of ASB enzyme activity to demonstrate its deficiency and complete the biochemical diagnosis. This can be done using different types of samples, such as dried blood spots (DBS), leukocytes (including lymphocytes), or fibroblasts. Leukocytes (isolated from whole blood) and fibroblasts are considered as the gold standard samples for the measurement of enzyme activity. The enzyme analysis using DBS is a good option, particularly for regions of the world where collecting and/or shipping whole blood is not practical, but should be considered as a screening method. For ASB analysis the activity can be measured using the natural substrate, but in routine practice artificial substrates are used, such as the chromogenic p-nitrocatechol sulfate, or the fluorogenic 4-methylumbelliferyl sulfate. It is always recommended to assay another lysosomal enzyme in the same sample used for ASB analysis to assess the quality of the sample. If the reference enzyme is normal and ASB activity is low, another sulfatase (eg, iduronate-2-sulfatase) should be measured to rule out multiple sulfatase deficiency, a different lysosomal disease in which low levels of all sulfatasases are found, including ASB.

It should be pointed out that no clear correlation could be drawn from the level of residual enzyme activity and clinical phenotype, making the prediction of clinical course difficult to be estimated on the basis of the enzyme assay. Although the biochemical tests (urinary GAGs analyses and enzyme assays) are sufficient to confirm the diagnosis of MPS VI, whenever possible the genetic analysis of the *ARSB* gene should be performed for the characterization of the molecular defect.³²

For the biochemical prenatal diagnosis of MPS VI the sample of choice is amniocytes, because there is the presence of high arylsulfatase C activity in chorionic villi which could interfere with the result if this material is used.³¹

Mutation identification

The gene responsible for MPS VI (*ARSB*) was identified and its sequence cloned and described in 1990.³⁸ It is mapped at chromosome 5q13-14 and has 2.8 kb organized in eight exons interrupted by seven introns.³⁹ As genetic abnormalities began to be described,²⁵ there was a growing attempt to correlate the genotypes with phenotypes, which has been disappointing most of the time, as a consequence of the broad molecular

heterogeneity present in this gene, including disease causing mutations and several polymorphisms.

To date, more than 164 mutations have been described in the ASB gene, most of them being point mutations.⁴⁰ Also, several polymorphisms have been identified and expression studies have shown that one of them, p.V358M, could reduce the residual activity of ARSB, depending on the concomitant disease causing mutation present. Most mutations are private, and those patients presenting both alleles with mutation predicting synthesis of a non-functional protein such as nonsense, small insertions and/or deletions tend to show a more severe phenotype.²⁵

Generally, protocols for ARSB molecular analysis starts with genomic DNA extraction from several sources such as total blood, leucocytes, saliva, DBS, and others, followed by polymerase chain reaction (PCR) amplification⁴¹ of the coding region of the gene, including exons 1–8 and adjacent intronic regions. The amplified product is then analyzed by Sanger sequencing⁴² in order to identify the alteration which leads to the enzyme defect. When the mutation is identified, it is prudent to confirm it through cDNA analysis obtained from mRNA. This is particularly interesting for mutations within the intron/exon boundaries which can cause the use of alternative splicing sites and consequently deletion or insertion of several nucleotides within the coding sequence.

There is a group of mutations that can be present in more than one unrelated patient (Table 2).⁴⁰ For those, it is possible to apply a strategy for detection of known recurrent mutations. Nevertheless, this procedure depends on the population studied, since the mutation profile can vary in distinct ethnic groups. A common mutation was described (p.H178L) in a small community in the Northeast of Brazil which was present in homozygosis in all patients tested.

Table 2 Recurrent mutations observed in specific populations worldwide

Mutation	Nucleotide	Exon/region	Reference
p.L72R	g.215T>G	I	Petry et al ⁹⁰
p.R152W	g.454C>T	II	Voskoboeva et al ⁹¹
g.427delG	g.427delG	II	Karageorgos et al ⁹²
p.G144R	g.430G>A	II	Isbrandt et al ⁹³
p.H178L	g.533A>T	III	Karageorgos et al ⁹²
p.Y210C	g.629A>G	III	Litjens et al ⁹⁴
p.R315Q	g.944G>A	V	Villani et al ⁹⁵
IVS5-1g>c	g.1143-1G>C	Intron V	Karageorgos et al ⁹²
IVS5-8t>g	g.1143-8T>G	Intron V	Petry et al ⁹⁰
p.H393P	g.1178A>C	VII	Litjens et al ⁹⁴
p.F399L	g.1197C>G	VII	Yang et al ⁹⁶
p.P531R	g.1592C>G	VIII	Villani et al ⁹⁵
g.1533del23	g.1533del23	VIII	Petry et al ⁹⁰

In this same sample a haplotype analysis was performed using intragenic polymorphisms and the same haplotype was identified amongst the individuals tested indicating a founder effect.⁴³

The observation of a high carrier frequency for the p.R152W mutation in the Lithuanian population may indicate a possible founder effect in this region.⁴⁴ Although its high prevalence in a series consisting of patients from Russia, Kazakhstan, and Central and Eastern Europe suggests that p.R152W may be of Slavic, not Lithuanian origin.⁴⁵

After the identification of both mutations in the index case, they can be tested in the parents to confirm inheritance and rule out the presence of two “in cis” mutations, as has already been reported for other MPS.⁴⁶ It is also possible to analyze its presence in other family members for identification of carriers or other affected individuals, using several methodologies according to the lab expertise and equipment available. Simple methods such as restriction fragment length polymorphisms can be used for mutations which lay within an enzyme restriction site which allows the digestion of the PCR product with specific restriction enzymes. This digested product can then be analyzed by ELP in agarose gel and visualization under ultra violet light using double strand DNA intercalants such as ethidium bromide, SYBR Safe, and gel red.

When the mutation found is not within an enzyme restriction site, it is possible to use a technique based on hybridization with a fluorescent labeled probe which is either complementary to the normal or to the mutated sequence. The presence of a positive reaction with one or both probes indicates the homozygous or heterozygous genotype. This method is known as real time PCR and can be very useful for identification of recurrent and frequent mutations in a given community,⁴ including neonatal screening as it allows for a throughput analysis with quick and sensible results.

Nowadays, modern techniques allow for the identification of a broad number of sequencing reactions in a single assay, allowing for a complete analysis of the whole gene (or even several genes). This technology is based on next generation sequencing (NGS) and can be used as panels of certain genes or even whole exome and genome sequencing and it is becoming an extremely advantageous technique due to its quickness and robustness. Although costs for this kind of analysis are still high, there is a decreasing cost trend as already shown with other novel technologies. NGS can even be useful for diagnosis of patients without a definitive biochemical diagnosis.

Nevertheless, even promising technologies have some pitfalls. For a specific group of mutations such as complete gene deletions including contiguous genes and gene inversions/rearrangements, this strategy would fail to identify the disease causing mutations. In the first example, sequencing or even NGS would not allow for deletion breakpoints mapping and other techniques such as comparative genomic hybridization (CGH) array would be necessary as ancillary techniques.⁴⁷

If the molecular defect is known in the family, prenatal diagnosis is possible through mutation analysis using several sources of biological material such as chorionic villi, amniocytes, or umbilical cord blood and can be performed concomitantly with the enzyme assay or independently.

DNA sequencing is the primary clinical technique for identifying mutations in human disease, but sequencing often does not detect intragenic or whole-gene deletions/duplications. CGH using oligonucleotide arrays has been implemented in cytogenetic and molecular diagnostic laboratories as a robust, rapid, and sensitive assay for detecting targeted gene deletions.⁴⁷ Targeted oligonucleotides array can be designed to detect both whole-gene deletions and small intragenic deletions in metabolic disorders and lysosomal storage genes.^{48,49} This technology may be particularly useful as complementary diagnostic test to the gene sequencing analyses in groups where intragenic deletions, duplications, and rearrangements are causative. Furthermore, genomic positions of the breakpoints cannot be determined by traditional methods. In contrast, the target CGH array is a rapid, highly sensitive, and accurate method that can be useful for detecting single- and multi-exon deletions and duplications in a large set of metabolic and lysosomal disorder genes simultaneously.⁴⁹ As gene CGH array is a recent technology being used in the metabolic and lysosomal disorder diagnostics and not yet a standard test, the intragenic deletions/duplications detected need to be confirmed. Methods such as breakpoints PCR, Southern blot, real time PCR, or confirmation with dye swap array CGH can be considered to determine the exact breakpoints.

Treatment

Until recently, support therapy and bone marrow transplantation therapies were the only options available for MPS VI. Supportive therapy was focused on nutrition, occupational and physiotherapy, in addition to symptomatic treatment of complications such as the use of oxygen with positive pressure during sleep, tonsillectomy, adenoidectomy, tracheostomy, medical or surgical treatment for heart failure, cervical

decompression, corneal transplantation, and ventricular shunt, among others.³

Hematopoietic stem cell transplantation

Bone marrow or hematopoietic cell transplantation has been used for some patients since 1984⁵⁰ with reports of favorable outcome several years after transplantation with.^{51,52} The rationale is that the hematopoietic system of the receptor is reconstituted with stem cells from an immune-compatible healthy donor, in order to establish a long-term enzyme source.^{53,54} Although there is an increase in the enzyme ASB and decrease of GAG excretion, skeletal changes already established do not improve, even in cases of successful transplantation. The use of this form of treatment is limited by the high rate of mortality and morbidity related to the procedure, such as graft versus host disease and because of the difficulty of finding donors.⁵⁵

Enzyme replacement therapy (ERT)

ERT is a specific form of therapy for MPS VI and is safer than hematopoietic cell transplantation. ERT is carried out by a recombinant form of ASB (rhASB) synthesized by genetic engineering from ovarian cells of the Chinese hamster by BioMarin Pharmaceutical Laboratory, Novato, CA, USA.^{56,57} Galsulfase (Naglazyme®) is administered intravenously in order to provide an active form of the enzyme that lacks or is insufficient in patients with MPS VI. In cats treated from birth, the enzyme prevented or slowed the progression of the skeletal related disease^{57,58} but was ineffective regarding the GAG accumulation in corneal keratocytes and cartilage.^{58,59} Some studies reported immune response to the recombinant enzyme, which led to adverse effects such as hypersensitivity reactions and anaphylaxis. The antibody response can inactivate or degrade the enzyme, affecting its efficacy.^{60,61}

A Phase III study was conducted using a weekly dose of 1 mg/kg in 39 patients, for 24 weeks, confirming the Phase II study results, with improvement in overall resistance measured by the 12-minute walk test and stair climbing, reduced GAG excretion in the urine, and good tolerability.^{62,63} Of the 54 patients who participated in clinical studies to its end, 53 developed specific antibodies to galsulfase without a direct correlation with the excretion of GAGs in the urine.³ A Phase IV study was performed using two different doses, 1 mg/kg and 2 mg/kg, weekly, in patients with MPS VI between 3.3 and 12.7 months, for at least 52 weeks. From this study it was demonstrated that the medication was well tolerated at both doses and no serious adverse events were related to medication. The main measure of improvement

was a reduction of approximately 70% of urinary GAG excretion.⁶⁴ Harmatz et al⁶⁵ conducted an observational study with 56 patients receiving ERT during Phase I/II, Phase II, Phase III, and extension clinical trials, and reported that the improvement in lung function is a factor that may be contributing to the improved resistance observed in these patients and it was not related to age. Decker et al⁶⁶ demonstrated that the growth rate in the prepubertal period was higher in patients receiving long-term ERT and that the increase was even greater in patients under the age of 16 years.

Recently, a study with 34 MPS VI patients under 5 years of age showed that ERT with galsulfase is safe and effective in this population⁶⁷ and has a similar profile to that reported in clinical trials with older patients.^{62,68} The results of this study also showed a significant negative correlation between the ERT and levels of urinary GAGs. In addition, patients maintained or increased the growth rate after treatment, and from the nine patients who underwent sleep studies at the beginning of this study, one showed improvement in sleep apnea after 56 months of ERT.⁶⁷

Clinical trials have shown that ERT with galsulfase has been well tolerated by patients with MPS VI.⁶⁹ However, there are a few reports of adverse reactions.^{70,71}

Di Natale et al⁷² conducted a study with four patients with the severe form of MPS VI who were on ERT with galsulfase to analyze genes that could be potential biomarkers of response to therapy. The results of this study showed that the expression of TNF- α , a cytokine produced by cells such as macrophages, chondrocytes, and synoviocytes significantly reduced after treatment, suggesting TNF- α as a potential biomarker of response to treatment.

A major complication: SCC

SCC is a frequent complication of MPS VI and may lead to important morbidity. Surgery for spinal decompression usually results in a significant neurological improvement^{73,74} and must be performed in experienced and specialized MPS treatment centers. ERT cannot cross the blood–brain barrier and has no access to the central nervous system including spinal cord by intravenous infusion. Horovitz et al⁷⁵ described six MPS VI children with SCC who presented increased joint mobility after ERT. As ERT increases neck mobility and reduces stiffness by GAG clearance from paravertebral structures, patients can experience an increased range of vertebral motion. In MPS VI animal models, intrathecal (IT) ERT prevents and removes dural storage⁷⁶ with no adverse reactions. Furthermore, a pioneer report of IT ERT to treat SCC in a single case of MPS VI has been described in the

literature.⁷⁷ The patient had pachymeningitis cervicalis hypertrophica and received IT infusions of rhASB administered monthly by lumbar puncture. Despite significant urodynamic and neurological improvement, the patient had worsened walking capacity. The authors concluded that IT ERT can be used as an alternative approach for MPS VI patients with pachymeningitis cervicalis hypertrophica, as long as neck stability is previously obtained, but further studies are ongoing to support these findings.

Perspectives

Based on the limitations observed with the current available treatments, new approaches have been tested for MPS VI. In that scenario, mainly the rat and cat animal models have been particularly useful for studies of disease pathogenesis and effectiveness of new treatments.^{78,79}

Using MPS VI rats, and based on previous studies in other animal models that suggested an important inflammatory component on the pathogenesis of this disease,⁸⁰ Eliyahu et al⁸¹ demonstrated that, when an anti-TNF- α drug is used in combination with ERT, benefits in organs such as the trachea and even the bones can be achieved. Further studies using pentosan polysulfate, a US Food and Drug Administration approved medication with anti-inflammatory effects have shown reduction in eye and nasal secretions and improvement in bone and tracheal deformities as well.⁸² Currently, Phase I clinical trials are under development, and anti-inflammatory therapy for MPS VI seems to be a

promising approach at least to alleviate some of the disease symptoms.

Gene therapy has also been tested in animal models of MPS VI. Different types of viral vectors have shown promising results in delivering a correct version of the *ARSB* gene and achieving high levels of enzyme activity. A study with an adeno-associated virus 2/8 suggests that this approach could be used as good therapy for MPS VI. A study in cats has shown improvement in visceral organs, joint mobility, skeletal abnormalities, and survival.⁸³ A second study used a retroviral vector in neonatal MPS VI cats. This approach was able to produce sustained high ASB serum levels for 8 years, with improvements in visceral organs, increased bone length as well as improvements in joints, heart valves, and aorta, which are known to be organs not easily corrected by ERT.⁸⁴ Major problems with gene therapy approaches are still the pre-existing immunity against adeno-associated viruses and the risk of insertional oncogenesis.^{83,84}

One point that several reports seem to agree on is that treatment is more effective when started earlier in life.⁸⁵ Newborn screening may play a role in the early detection of cases, with high-throughput methods based on tandem mass spectrometry already in development.⁸⁶ While no formal massive screening for MPS VI is in place so far, an interesting approach is being used in an area with high incidence of MPS VI. In this selected area, a community-based neonatal screening will enable early diagnosis and more effective treatments. Also, it opens up the possibility of more effective genetic

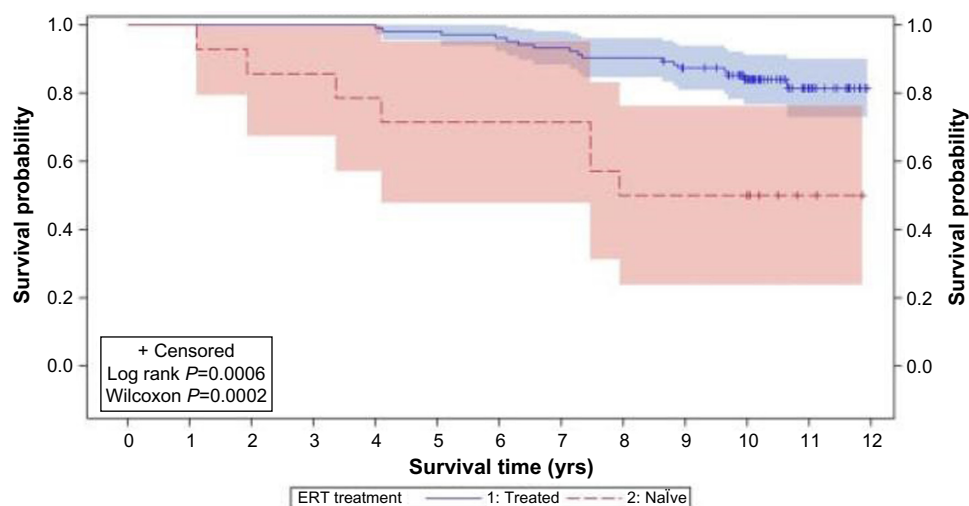


Figure 4 Product-limit survival estimates of MPS VI patients who received ERT and who were treatment naïve.

Notes: Product-limit survival estimates with number of subjects at risk and 95% confidence intervals. Kaplan–Meier survival curves of ERT (n=103; blue solid line) and naïve (n=14; red dashed line) patient groups. The 95% confidence intervals are indicated by blue (ERT) or red (naïve) shaded boxes. The calculated P-values were: Log rank $P=0.0006$, Wilcoxon $P=0.0002$. Reproduced from Giugliani R, Lampe C, Guffon N, et al. Natural history and galsulfase treatment in mucopolysaccharidosis VI (MPS VI, Maroteaux-Lamy syndrome) – 10-year follow-up of patients who previously participated in an MPS VI Survey Study. *Am J Med Genet A*. 2014;164A(8):1953–1964, with permission from Johny Wiley and Sons. © 2014 Wiley Periodicals, Inc.⁸⁸

Abbreviations: MPS VI, mucopolysaccharidosis VI; ERT, enzyme replacement therapy; yrs, years.

counseling and development of comprehensive prevention programs for these families.^{4,87}

Final remarks

Ten years after a specific ERT for MPS VI was approved, it still remains a challenging disease. Despite the clear benefits of ERT in terms of reduced morbidity and even mortality⁸⁸ (Figure 4), the affected patients still present significant disease manifestations. Early start of treatment, as already elegantly demonstrated in a sibling study⁸⁹ should bring additional benefits. Present methods for biochemical and genetic diagnosis are accurate, but depend on clinical suspicion, which could be delayed. The detection of affected babies by newborn screening is being considered as a potential diagnostic tool, and pilot screening programs are already in place in selected areas. Additional treatment options (such as pentosan polysulfate) to be used jointly with ERT, or gene therapy, in combination with early diagnosis, will probably positively change the course of the disease in affected patients.

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The authors have no conflicts of interest to disclose.

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