Original Article

Low MiR-149 expression is associated with unfavorable prognosis and enhanced Akt/mTOR signaling in glioma

Liang Xue, Yi Wang, Shuyuan Yue, Jianning Zhang

Department of Neurosurgery, Tianjin Medical University General Hospital, Tianjin 300052, P. R. China Received July 1, 2015; Accepted August 20, 2015; Epub September 1, 2015; Published September 15, 2015

Abstract: microRNAs (miRs) play critical roles in the progression of glioma. Previous *in vitro* studies have described the anti-tumor role of miR-149 in cancer cells including glioma. In this study, we aimed to investigate whether miR-149 is associated with the prognosis of glioma patients. A total of 163 glioma patients who underwent tumor resection were included in our follow-up study. We found that the miR-149 expression was significantly lower in tumor tissues compared with that in normal tissues (P<0.05). Kaplan-Meier and analysis showed that the miR-149 expression status was significantly associated with the survival duration (logrank test, P<0.001), and multivariate Cox regression revealed that patients with low miR-149 expression were exposed to a 1.825 fold higher death risk (HR=1.825, 95% Cl=1.031-3.229, P=0.039) compared with those with high miR-149 expression. Further study showed that Akt/mTOR signaling was hyperactive in low miR-149 expressing tissues. Our study thus demonstrates that miR-149 expression in glioma tissues is critically associated with the prognosis of patients, suggesting its potential clinical significance.

Keywords: Glioma, miR-149, prognosis, Akt, mTOR

Introduction

Glioma, as the one of the most common malignancies in central nervous system, is characterized by high invasiveness, early recurrence and poor prognosis [1]. Despite the recently achieved advances in cancer diagnosis and treatment, the dismal survivalrate of high grade glioma still represents one of the major challenges in clinical practice. The unmet demand for the early prediction of prognosis, which might guide the therapeutic strategy for glioma, has underlined the importance of developing new diagnostic and prognostic approaches.

In recent years, microRNAs, a class of non-coding short single RNA strands, have emerged as key regulators of carcinogenesis and tumor progression [2-5]. Multiple microRNAs have exhibited their potency in governing the critical biological processes of tumor cells such as proliferation, apoptosis and differentiation by post-transcriptionally repressing their target genes [6]. The biology of glioma is also controlled by a microRNA network without exception [5, 7-9], a large spectrum of microRNAs in glioma has

been extensively investigated; for example, miR-10 and miR-16 have been shown to control glioma cell migration and invasion through regulating the epithelial-mesenchymal transition process [10, 11], and several miRs have been shown to be associated with the prognosis of glioma [12-14]. Among the miRs that play pivotal roles in the pathophysiology of glioma, miR-149 is believed to act as a versatile tumor-suppressionmiRand is downregulatedin several cancer types [15-17]. Recent studies have addressed its anti-proliferation role by targeting FOXM1 [15]; and importantly, it has been reported to inhibit the invasiveness of glioma cell through blockade of AKT1 signaling [18]. Although these studies have implied its potential values in the diagnosis and treatment of glioma, the clinical evidence of this microRNA is still lacking and remain to be identified; more importantly, elucidating its distinct expression profilein glioma patients may broaden our knowledge on the potentialrole of microRNAs in the emerging field of biomarker screen.

In the present study, we launched a follow-up study on 113 patients to examine whether mir-

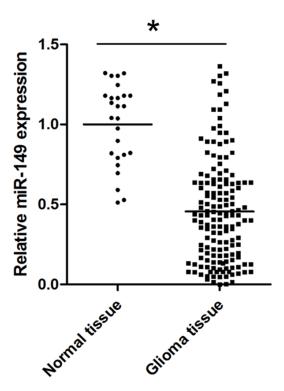


Figure 1. The differential expression of miR-149 in normal brain tissues and glioma tissues *P<0.05 n=26 for nomal tissue and n=163 for glioma tissue.

149 is associated with the clinical outcome of glioma patients. Our pilot data revealed its prognostic function in glioma patients.

Patients, materials and methods

Study subjects

We enrolled 163 glioma patients who underwent tumor resection from January 2005 to December 2012 in Tianjin Medical University General Hospital. Among these patients, 88 patients (mean age: 56.6±8.7 years) were male and 75 (mean age: 55.2±8.4 years) were female. Fresh tumor specimens were stored in -80°C. All the patients' diagnoses were confirmed based on the histopathological examination. The histological grade was determined according to the criteria formulated by the World Health Organization in 2007. The patients were followed-up every 3 months, and the follow-up was ended on December 31st 2014. 26 normal brain tissue samples were collected from the surgical waste from other surgeries and used as controls. We obtained a writing consent from each patient, and this study was approved by the Ethics Committee of Tianjin Medical University General Hospital.

RNA isolation and miR-149 quantification

The RNA was isolated using Trizol Reagent (Invitrogen) according to the instructions provided by the manufacturer. To assess the levels of miR-149 in each group, we utilized a TaqMan based quantitative PCR method. The TaqMan PCR assay kits were purchased from Applied Biosystems. The expression of U6 was used as an internal control to adjust miR-149 expression. And the relative expression of miR-149 was determined by the $2^{-\Delta \Delta Ct}$ method.

Western blot

Tissues were homogenized with RIPA lysis buffer and were subsequently subjected to electrophoresis, followed by transferring onto a NC membrane. Primary antibodies were all purchased from Cell Signaling Technology Inc. (Beverly, MA, USA). The bands were visualized by an ECL detection kit (P0018, Beyotime, Shanghai China). The band density was determined by ImageJ software (version 1.44, NIH).

Statistical analysis

The data in this study were analyzed by SPSS 19.0 software package, Wilcoxon-Mann-Whitney test was used to compare the difference of miR-149 expression between glioma tumor samples and normal brain samples. The patient population was dichotomized according to miR-149 expression level. Patients who had a 0.5-fold or above miR-149 expression than the mean expression of control group were categorized as high miR-149 expression, otherwise categorized as low miR-149 expression. Chi square test was performed to examine the association between miR-149 expression status and clinical-pathological characteristics. Survival analysis was performed by Kaplan-Meier estimator, comparison of the overall survival between two groups was performed by logrank test. Multivariate Cox regression analysis was also used to determine the hazard ratio of each covariate. A two tailed probability less than 0.05 was considered statistically significant.

Results

MiR-149 is down-regulated in gliomatissue

We first compared the miR-149 expression profile in normal brain tissues and glioma tissues.

Table 1. Association between miR-149 expression and clinical characteristics

Clinical characteristics	N	miR-149 expression	
		Low	High
Sex			
Male	88	57	31
Female	75	46	29
Age (years)			
≤60	106	68	38
>60	57	35	22
Tumor location			
Parenchyma	104	68	36
Ventricle	59	39	20
Tumor size			
≤3 cm	84	52	32
>3 cm	79	51	28
WHO grade*			
I+II	80	36	44
III+IV	83	67	16
Karnofsky score*			
≤80	94	88	6
>80	69	15	54
Resection range			
Total resection	93	55	38
Local resection	70	48	22
Adjuvant therapy			
Chemotherapy only	100	63	37
Radiotherapy and chemotherapy	63	40	23
Recurrence time*			
≤3 months	97	68	29
>3 months	66	33	33
Survival duration*			
<15 months	81	71	10
≥15 months	82	32	50

^{*}P<0.05, performed by χ^2 test.

As shown in **Figure 1**, miR-149 level was found to be significantly higher in normal brain tissues than that in glioma tissues (P<0.05).

Association between miR-149 expression and clinical characteristics

To examine the possible association between miR-149 expression and clinical characteristics, we first divided the patients into high-expression group and low-expression group based one the criteria described above. 103 patients were identified as low expression, and 60 patients were identified as high expression. We studied the association between miR-149

expression and general clinical characteristics in the study cohort. As presented in **Table 1**, miR-149 expression was not associated with age, sex, tumor location, tumor size, resection range and adjuvant therapy. By contrast, association between miR-149 expression and WHO grade, Karnofsky score, recurrence time or survival duration was observed.

MiR-149 correlates with the survival of glioma patients

To investigate whether miR-149 expression is correlated with the survival of glioma patients, we performed Kaplan-Meier survival analysis. All the cases were followed-up after surgery, and 7 patients lost follow-up due to the changes of their telephone number. As presented in Figure 2, patients with high expression of miR-149 exhibited greater overall survival than those with low miR-149 expression. Moreover, multivariate Cox regression analysis showed that the adjusted hazard ratio of miR-149 low expression was 1.825 (Table 2). Taken together, these data suggested that low expression of miR-149 is associated with a more aggressive type of glioma.

MiR-149 is associated with Akt/mTOR signaling in glioma patients

Previous *in vitro* study has demonstrated that miR-149 targets Akt. To test the potential role of miR-149 in regulating Akt/mTOR signaling in the clinical setting, we assayed the protein level of the Akt/mTOR axis. As shown in **Figure 3A**, both the phosphorylated and total AKT

level were lower in high miR-149 expressing tissues, consequently, mTOR, the downstream effector of AKT, was less phosphorylated. Correlation analysis confirmed the association between miR-149 expression and mTOR signaling, which suggested that the upregulatedAkt/mTOR might mediate the unfavorable effect of low miR-149 expression on prognosis of glioma patients (Figure 3B).

Discussion

In the current follow-up study, we evaluated the prognostic value of miR-149 in glioma patients. Our data showed that the differential expres-

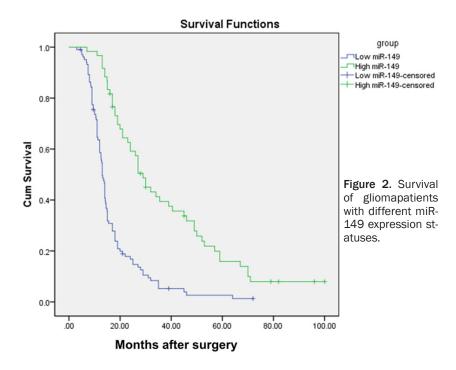


Table 2. Multivariate Cox regression analysis of risk factors associated with overall survival of glioma patients

Clinical characteristics	Adjusted HR	95% CI	Р
Age (>60)	1.369	0.958-1.956	0.085
Sex (Male)	1.096	0.783-1.535	0.593
Tumor size (>3 cm)	0.779	0.541-1.121	0.179
WHO grade (III+IV)	1.475	1.010-2.154	0.044
Karnofsky score (<80)	1.899	1.093-3.267	0.023
Resection range (≤3 cm)	1.050	0.743-1.484	0.780
Adjuvant therapy (Chemotherapy only)	1.255	0.845-1.863	0.261
miR-149 expression (Low)	1.825	1.031-3.229	0.039

sion of miR-149 in glioma tissues and normal tissues, Kaplan-Meier survival analysis and Cox regression analysis together showed that patients with low expression of miR-149 have a worse clinical outcome, which confirmed our hypothesis that miR-149 might play a favorable role in the survival of glioma patients. Therefore, our study demonstrates that miR-149 expression might be an independent indicator of prognosis of patients with glioma.

MicroRNAs have been recognized as indispensable regulators of normal cell function. Aberrant microRNA expression has been found in several diseases including glioma [19]. Several microRNAs have been implicated in the multi-

facetregulatory network of the pathogenesis of this cancer type. For example, a recent study shows that miR-218 functions as a tumor suppressor by affecting a series of critical biological processes of glioma including cell invasion, migration, proliferation and the maintenance of cancer cell stemness [20]. Particularly, growing numbers of studies have revealed their potential clinical values, circulating miR-128 has been identified as a diagnostic marker [21], and miR-218 has also been reported to show prognostic significance [22]. Although these studies revealed the pivotal role of microRNA in glioma biology, whether other microRNAs are also implicated in this issue is still to be explored. Moreover, given the broad effects of microRNAs are exhibiting, it is reasonable to speculate that a large number of previously

uncharacterized microRNAs may show macro effects in glioma patients.

Accumulating evidences have unraveled the role of miR-149 in cancer development, it is believed that this microRNA probably acts as a nodal point to control the apoptotic program. To date, a number of target genes have been validated such as PUMA and FOXM1 [15, 23]. Although most of the studies on miR-149 was conducted in other cancers with few studies in the setting of central nervous system, a recent in vitro study suggesting blockade of AKT1 by miR-149 in glioma cell has implicated its possible involvement in clinical samples [18]. Intriguingly, She X et al. described enhanced

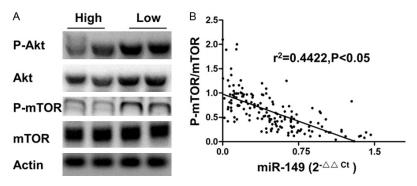


Figure 3. A. The representative western blot image of Akt/mTOR axis in low and high miR-149 expressing glioma tissues. B. Correlation between mTOR phosphorylation and miR-149 expression.

chemo sensitivity against temozolomide in miR-149 overexpressed glioma cell [24]. Our study, which deciphered the association between low miR-149 expression and poor prognosis, corroborates these in vitro findings. Although the overall miR-149 expression status of the principal cohort is low in our study, it is interesting to find out that a small proportion of these patients still presented high level of miR-149, which is comparable to that in normal tissues. Recent study by Ding et al. highlighted the crucial role of polymorphism within pre-miR-149 in its maturation, in their study, lower levels of miR-149 production was observed in rs71428439 pre-miR-149 expressing HEK293 cells [23]. More importantly, several studies also reported the association between miR-149 polymorphisms and cancer susceptibility or prognosis [25-27]. Therefore, it is conceivable that pre-miR-149 polymorphism at least partly account for its lowered expression level in glioma patients. Whether miR-149 polymorphisms are associated with glioma progression is expected to be an interesting topic awaiting to be uncovered.

AKT/mTOR signaling plays a central role in gliomacell proliferation. Considering that AKT1 has already been identified as the target of miR-149 in glioma cells, our finding could be expected to reinforce these *in vitro*mechanistic studies. Importantly, the association between miR-149 and AKT/mTOR signaling in the clinical sample may represent the possible mechanism that drives the unfavorable clinical outcome of low miR-149 expression. However, it should be noted that miR-149 was also described to be related with myocardium infarction [28, 29], and functional tests revealed its anti-apoptotic

action in myocardium [23], which is contrary to the reports in cancers. This discrepancy may be explained by the reprogrammed signaling network in cancer cells. Due to the several mutations occurring in proapoptotic genes in cancer cells, the overall effect of miR-149 may be deathprone, making it as a potential tumor suppressor in glioma.

The current prognostic model is largely dependent upon the WHO histological grading and immunohistochemistry analysis. However, histological examinations are often subjective and call for rich clinical diagnostic experience of pathologists, and the false positive of immunohistochemistry also limits its diagnostic power. The application of microRNA in the clinical practice would be expected to meet the needs of clinical practitioner. MicroRNA detection in the paraffin section by the RNA in situ method might represents a more reliable diagnostic approach. Moreover, as several circulating microRNAs are emerging as the biomarkers for cancer diagnosis, investigating whether circulating miR-149 level correlates with its expression in tumor site should represent a more convenient alternative to assess the prognosis of patients. Identifying miR-149 as a prognostic marker enables the quantitative evaluation of the death risk at early diagnosis, which might be helpful to tailoring individualized treatment. To go further, concerning that we have identified low miR-149 expression is associated with poorer prognosis, it might be an effective therapeutic approach to complement miR-149 in low expression patients.

In summary, although our study population is relatively small, we demonstrate the aberrant miR-149 expression in glioma patients for the first time; more importantly, the expression status is significantly associated with the survival duration of patients. In the future, large scale studies and more detailed experimental studies are still needed to confirm the prognostic significance and the mechanism of miR-149 in glioma.

Disclosure of conflict of interest

None.

Address correspondence to: Liang Xue, Department of Neurosurgery, Tianjin Medical University General Hospital, 154 Anshan Road, Heping District, Tianjin 300052, P. R China. E-mail: xueliang32@126.com

References

- [1] Ahmed R, Oborski MJ, Hwang M, Lieberman FS and Mountz JM. Malignant gliomas: current perspectives in diagnosis, treatment, and early response assessment using advanced quantitative imaging methods. Cancer Manag Res 2014; 6: 149-170.
- [2] Chan B, Manley J, Lee J and Singh SR. The emerging roles of microRNAs in cancer metabolism. Cancer Lett 2015; 356: 301-308.
- [3] Weis SM and Cheresh DA. Tumor angiogenesis: molecular pathways and therapeutic targets. Nat Med 2011; 17: 1359-1370.
- [4] Olson P, Lu J, Zhang H, Shai A, Chun MG, Wang Y, Libutti SK, Nakakura EK, Golub TR and Hanahan D. MicroRNA dynamics in the stages of tumorigenesis correlate with hallmark capabilities of cancer. Genes Dev 2009; 23: 2152-2165
- [5] Low SY, Ho YK, Too HP, Yap CT and Ng WH. MicroRNA as potential modulators in chemore-sistant high-grade gliomas. J Clin Neurosci 2014; 21: 395-400.
- [6] Zhang B, Pan X, Cobb GP and Anderson TA. microRNAs as oncogenes and tumor suppressors. Dev Biol 2007; 302: 1-12.
- [7] Katsushima K and Kondo Y. Non-coding RNAs as epigenetic regulator of glioma stem-like cell differentiation. Front Genet 2014; 5: 14.
- [8] Zhao B, Bian EB, Li J and Li J. New advances of microRNAs in glioma stem cells, with special emphasis on aberrant methylation of microR-NAs. J Cell Physiol 2014; 229: 1141-1147.
- [9] Palumbo S, Miracco C, Pirtoli L and Comincini S. Emerging roles of microRNA in modulating cell-death processes in malignant glioma. J Cell Physiol 2014; 229: 277-286.
- [10] Yan Y, Wang Q, Yan XL, Zhang Y, Li W, Tang F, Li X and Yang P. miR-10a controls glioma migration and invasion through regulating epithelialmesenchymal transition via EphA8. FEBS Lett 2015; 589: 756-765.
- [11] Wang Q, Li X, Zhu Y and Yang P. MicroRNA-16 suppresses epithelial-mesenchymal transition-related gene expression in human glioma. Mol Med Rep 2014; 10: 3310-3314.
- [12] Sun G, Yan S, Shi L, Wan Z, Jiang N, Li M and Guo J. Decreased Expression of miR-15b in Human Gliomas Is Associated with Poor Prognosis. Cancer Biother Radiopharm 2015; 30: 169-73.

- [13] Ji Y, Wei Y, Wang J, Gong K, Zhang Y and Zuo H. Correlation of microRNA-10b upregulation and poor prognosis in human gliomas. Tumour Biol 2015; 36: 6249-6254.
- [14] Lai NS, Wu DG, Fang XG, Lin YC, Chen SS, Li ZB and Xu SS. Serum microRNA-210 as a potential noninvasive biomarker for the diagnosis and prognosis of glioma. Br J Cancer 2015; 112: 1241-1246.
- [15] Xu K, Liu X, Mao X, Xue L, Wang R, Chen L and Chu X. MicroRNA-149 suppresses colorectal cancer cell migration and invasion by directly targeting forkhead box transcription factor FOXM1. Cell Physiol Biochem 2015; 35: 499-515
- [16] He DX, Gu XT, Li YR, Jiang L, Jin J and Ma X. Methylation-regulated miR-149 modulates chemoresistance by targeting GlcNAc Ndeacetylase/N-sulfotransferase-1 in human breast cancer. FEBS J 2014; 281: 4718-4730.
- [17] Perez-Rivas LG, Jerez JM, Carmona R, de Luque V, Vicioso L, Claros MG, Viguera E, Pajares B, Sanchez A, Ribelles N, Alba E and Lozano J. A microRNA signature associated with early recurrence in breast cancer. PLoS One 2014; 9: e91884.
- [18] Pan SJ, Zhan SK, Pei BG, Sun QF, Bian LG and Sun BM. MicroRNA-149 inhibits proliferation and invasion of glioma cells via blockade of AKT1 signaling. Int J Immunopathol Pharmacol 2012; 25: 871-881.
- [19] Speranza MC, Frattini V, Pisati F, Kapetis D, Porrati P, Eoli M, Pellegatta S and Finocchiaro G. NEDD9, a novel target of miR-145, increases the invasiveness of glioblastoma. Oncotarget 2012; 3: 723-734.
- [20] Tu Y, Gao X, Li G, Fu H, Cui D, Liu H, Jin W and Zhang Y. MicroRNA-218 inhibits glioma invasion, migration, proliferation, and cancer stemlike cell self-renewal by targeting the polycomb group gene Bmi1. Cancer Res 2013; 73: 6046-6055.
- [21] Sun J, Liao K, Wu X, Huang J, Zhang S and Lu X. Serum microRNA-128 as a biomarker for diagnosis of glioma. Int J Clin Exp Med 2015; 8: 456-463.
- [22] Cheng MW, Wang LL and Hu GY. Expression of microRNA-218 and its Clinicopathological and Prognostic Significance in Human Glioma Cases. Asian Pac J Cancer Prev 2015; 16: 1839-1843.
- [23] Ding SL, Wang JX, Jiao JQ, Tu X, Wang Q, Liu F, Li Q, Gao J, Zhou QY, Gu DF and Li PF. A premicroRNA-149 (miR-149) genetic variation affects miR-149 maturation and its ability to regulate the Puma protein in apoptosis. J Biol Chem 2013; 288: 26865-26877.
- [24] She X, Yu Z, Cui Y, Lei Q, Wang Z, Xu G, Xiang J, Wu M and Li G. miR-128 and miR-149 enhance the chemosensitivity of temozolomide by Rap1B-mediated cytoskeletal remodeling in glioblastoma. Oncol Rep 2014; 32: 957-964.

miR-149 predicts glioma prognosis

- [25] Wei WJ, Lu ZW, Li DS, Wang Y, Zhu YX, Wang ZY, Wu Y, Wang YL and Ji QH. Association of the miR-149 Rs2292832 polymorphism with papillary thyroid cancer risk and clinicopathologic characteristics in a Chinese population. Int J Mol Sci 2014; 15: 20968-20981.
- [26] Xia L, Ren Y, Fang X, Yin Z, Li X, Wu W, Guan P and Zhou B. Prognostic role of common microRNA polymorphisms in cancers: evidence from a meta-analysis. PLoS One 2014; 9: e106799.
- [27] Wang Z, Wei M, Ren Y, Liu H, Wang M, Shi K and Jiang H. miR149 rs71428439 polymorphism and risk of clear cell renal cell carcinoma: a case-control study. Tumour Biol 2014; 35: 12127-12130.
- [28] van Rooij E, Sutherland LB, Thatcher JE, Di-Maio JM, Naseem RH, Marshall WS, Hill JA and Olson EN. Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. Proc Natl Acad Sci U S A 2008; 105: 13027-13032.
- [29] Liu Z, Yang D, Xie P, Ren G, Sun G, Zeng X and Sun X. MiR-106b and MiR-15b modulate apoptosis and angiogenesis in myocardial infarction. Cell Physiol Biochem 2012; 29: 851-862.