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Innate Immunity Fights Alzheimer's Disease

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Abstract

Alzheimer's disease (AD) is the most common age-related dementia. Pathognomonic accumulation of cerebral β -amyloid plaques likely results from imbalanced production and removal of amyloid- β (A β) peptides. In AD, innate immune cells lose their ability to restrict cerebral A β accumulation. At least in principle, mononuclear phagocytes can be enlisted to clear A β / β -amyloid from the brain. While the classical focus has been on dampening neuroinflammation in the context of AD, we hypothesize that rebalancing cerebral innate immunity by inhibiting actions of key anti-inflammatory cytokines returns the brain to a physiological state. Recent experiments demonstrating beneficial effects of blocking anti-inflammatory cytokine signaling in pre-clinical mouse models provide supportive evidence. This concept represents an important step toward innate immune targeted therapy to combat AD.

Keywords

amyloid; apolipoprotein E; immunotherapy; interleukin-10; microglia; phagocytosis

1. The central role of cerebral innate immunity in Alzheimer's disease

Alzheimer's disease (AD) currently affects more than 5 million Americans, and newly diagnosed cases are expected to reach 10–15 million in the U.S.A. by 2050 [1], highlighting the major impact of the disease on public health. AD is characterized by intracellular neurofibrillary tangles (comprised of misfolded tau protein), extracellular amyloid- β (A β) accumulation (consisting of β -pleated sheet conformers of insoluble A β peptides) and neuroinflammation, earmarked by reactive astrocytes and brain-resident monocytes (microglia) surrounding β -amyloid deposits [2]. While a number of preventative and therapeutic strategies are being pursued, an effective treatment for AD does not yet exist.

A widely-accepted theory of AD pathogenesis holds that imbalanced production versus clearance of A β peptides precipitates disease. In recent decades, the major therapeutic approach has been aimed at reducing cerebral A β production. Specifically, drugs have been designed to inhibit the β - and γ -secretases responsible for A β production from amyloid

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precursor protein (APP) endoproteolysis. However, the secretases cleave a variety of other substrates, and lack of specificity has been implicated in resultant adverse events [3,4]. An alternative strategy that is gaining momentum is targeting the other side of the equation: $A\beta$ clearance. This concept is rooted in the notion that failure of the innate immune system to clear $A\beta$, rather than overproduction of the peptides, is likely the etiologic culprit in sporadic AD [5]. Indeed, extensive microglial recruitment to plaques in human AD is accompanied by very little, if any, $A\beta$ phagocytosis [5,6], and occurs with increased production of pro-inflammatory cytokines that associate with cognitive decline [7]. A parsimonious conclusion from these findings is that innate immune cells (including both CNS-resident microglia and peripheral mononuclear phagocytes) lose their physiologic ability to restrict cerebral $A\beta$ accumulation and switch into a pathological state [2,7–10]. It remains unclear at what stage and to what degree a switch from 'good' to 'bad' microglia occurs, and whether this is reversible.

The most common form of the disease, sporadic or late-onset AD (LOAD), has a complex etiology that includes genetic, environmental, and lifestyle risk factors. Recent genome-wide association studies (GWAS) have identified a cluster of AD risk alleles belonging to core innate immune pathways [3,4,11]. A common phenotype linking these risk alleles is modulation of phagocytosis [12–15]. Consequently, regulation of inflammation and the cerebral innate immune response have become major areas of interest, both in terms of understanding AD etiology and for developing new therapeutic approaches. However, inconsistent results reported for non-steroidal anti-inflammatory drugs (NSAIDs) and AD prevention [16–19] imply that a more 'surgical' approach targeting select immune pathways is needed. However, refining therapeutic targets has been difficult. This is because activated microglia express a multitude of inflammatory cytokine and chemokine receptors in the context of AD-like pathology [20–24], and myriad cytokines and chemokines have been detected in AD patient brains and CSF [25–27].

While pro-inflammatory mediators have garnered the most attention in this regard, the cardinal anti-inflammatory regulators transforming growth factor-beta 1 (TGF-β1) [28] and interleukin-10 (IL-10) [25] are also elevated in human AD, raising a possible pathogenic role for these cytokines. Further, a functional polymorphism within the *IL10* gene has been linked to increased risk for LOAD in some [29–32], but not all populations [33–35]. While an initial report suggested that the *IL10* AD risk allele was associated with reduced IL-10 expression in healthy control plasma [32], those authors did not relate *IL10* polymorphism to IL-10 activity in AD patients. This is important, because CNS IL-10 abundance is increased in neurological diseases; including stroke, multiple sclerosis, meningitis, and AD [36]. Further, we and others have shown that the IL-10 signaling pathway is abnormally elevated in AD patient sera and brains [25,37]. The earlier conclusion that IL-10 down-regulation is a risk factor for AD is at odds with recent integrative genomic evidence showing increased IL-10 signaling in AD brains [38,39]. Clearly, further work is necessary to understand the *IL10* allele-AD risk relationship.

Contrary to the notion that all forms of inflammation are deleterious in the context of AD, awareness is being raised to the concept that blocking immunosuppressive pathways can be beneficial. In this regard, we demonstrated that TGF- β -Smad 2/3 signaling inhibition in

central and peripheral IL-10 abundance [40–42], prompting investigation into the putative contribution of IL-10 signaling in the context of AD. Indeed, new studies show that inhibiting IL-10/STAT3 signaling dramatically mitigates Alzheimer-like pathology [37], while brain overexpression of *Il10* produces complementary effects [43]. This has led us to theorize that 'rebalancing' activation of the innate immune system, as opposed to shutting it off completely, represents a novel AD therapeutic approach.

2. Interleukin-10 signaling suppresses cerebral β-amyloid clearance

IL-10 is a prototypical anti-inflammatory cytokine that is produced by and regulates activation of T cells, dendritic cells, peripheral macrophages, and CNS resident microglia [44,45]. Signaling is elicited by binding of IL-10 to its cognate receptor (IL-10R), which triggers phosphorylation of Janus kinase 1 (JAK1) that, in turn, phosphorylates signal transducer and activator of transcription 3 (STAT3). STAT3 homodimerizes and translocates to the nucleus, where it transactivates genes including suppressor of cytokine signaling 3 (SOCS3). SOCS3 is then phosphorylated by Src family kinases and interacts with receptors for inflammatory cytokines; targeting them for ubiquitin-mediated degradation. Activation of the IL-10 pathway referees essential functions of monocytes including phagocytosis, cytokine production, expression of costimulators, and antigen presentation. The *modus operandi* of this cytokine is to suppress overly exuberant inflammatory responses by blocking the action of pro-inflammatory cytokines [45,46].

In the central nervous system, IL-10 expression is increased in response to major neurological diseases including stroke, multiple sclerosis, meningitis, and AD [47]. Increased IL-10 abundance in AD patients' sera and brains has been reported [25,48], and we recently showed that all canonical IL-10 pathway signaling elements (*i.e.*, IL-10R1 and downstream JAK1, phospho-JAK1 (pJAK1), STAT3, phospho-STAT3 and SOCS3) are overly abundant in AD hippocampi compared to age-matched, non-demented controls [37]. These results led us to hypothesize that elevated IL-10 signaling in the AD brain blocks physiological innate immune recognition, phagocytosis, and clearance of cerebral β -amyloid.

To test this hypothesis, effects of IL-10 modulation were independently examined in three different mouse models (TgCRND8, Tg2576 and APP/PS1) exhibiting cortical and hippocampal Aβ plaques, gliosis, synaptic deficits, and associated cognitive impairment. Cerebral *Il10* overexpression using adeno-associated virus in TgCRND8 and Tg2576 mice exacerbated Aβ plaque number and size [43], while deletion of *Il10* reduced the severity of cerebral amyloid angiopathy and amyloidosis in APP/PS1 mice [37]. In neither experimental scenario did changes in IL-10 abundance affect APP metabolism or Aβ production; suggesting that IL-10-dependent alteration of cerebral amyloid deposition is owed to Aβ clearance [37,43]. Further support came from *in vitro* evidence showing that the IL-10/STAT3 pathway modulates microglial Aβ phagocytosis. Specifically, recombinant IL-10 treatment of primary microglial cultures promoted nuclear translocation of downstream STAT3 [37] and decreased phagocytosis of Aβ40 [43] and Aβ42 microaggregates [37].

However, IL-10 had no effect on A β 40 or A β 42 astrocytic uptake [43]. Importantly, genetic ablation of *Il10* or *Stat3* in microglial cultures increased A β 42 uptake into phagolysosomes [37].

Essential validation of these observations came from *in vivo* experiments, where *Il10* deficient APP/PS1 mice presented increased astrocytosis and reactive microglia [37]. Specifically, β -amyloid plaques, which appeared more diffuse than typical dense-cored plaques, were associated with greater numbers of monocytes compared with *Il10* sufficient APP/PS1 mice [37]. Cell number was not the only monocyte phenotype that *Il10* deficiency modified; intriguingly, mononuclear phagocytes infiltrating A β deposits expressed CD68 (a cell surface, phagolysosomal and endosomal marker) [49]. These striking results suggest that *Il10* deficient monocytes are prone to penetrate β -amyloid plaques and to phagocytose A β deposits [37]. Finally, cutting-edge quantitative three-dimensional *in silico* modeling (q3DISM) allowed us to quantitate A β phagocytosis *in vivo* (Figure 1). Results from this line of investigation confirm that *Il10* deficient monocytes are more efficient A β phagocytes versus *Il10* sufficient cells. Altogether, these data suggest that 1) endogenous overproduction of IL-10 causes defective innate immune A β phagocytosis leading to plaque build-up, and 2) inhibition of the IL-10/STAT3 pathway returns the system to homeostasis; thereby enabling cerebral A β clearance.

3. Beneficial effects of blocking IL-10/STAT3 signaling in the context of Alzheimer's disease

A key question is whether the effects of IL-10 modulation are restricted to cerebral amyloidosis or more broadly impact Alzheimer-like pathology. The first hint of this came from *Il10* overexpression, which decreased synaptic protein abundance and worsened cognitive impairment [43]; whereas *Il10* deletion led to stabilization of synaptic health and mitigated behavioral deficits [37]. These important findings indicate that the specific innate immune activation profile is of utmost importance for determining damaging vs. beneficial responses [10,41,50–52]. If left unchecked–or if overly repressed–cerebral innate immunity can perpetrate neuronal damage (directly or via $A\beta$); but if controlled, the system can be harnessed to clear the brain of damaging $A\beta$ species without coming at the cost of bystander neuronal injury.

The notion that monocyte activation states are clearly divided between a pro-inflammatory M1 phenotype and a pro-phagocytic M2 phenotype has become obsolete [9,10,50]. Rather, microglia can assume a continuum of activation states characterized by expression of intersecting markers [10,53]. This concept was corroborated by analysis of *Il10* dependent inflammatory genes via RNAseq [37] or NanoString [43]. In transgenic mouse models of cerebral amyloidosis, genes that were altered by *Il10* overexpression or *Il10* deficiency were further interrogated for immune-related function(s) [37,43]. Gene pathways were largely overlapping between these two opposing manipulations, and were responsible for: phagocytosis, chemoattraction/chemokine signaling, innate immune regulation (*e.g.*, toll-like receptor signaling), inflammation and the acute phase response (*e.g.*, LXR/RXR pathway), A β interaction, dendritic cell maturation and T cell activation, the protein complement system, communication between innate and adaptive immune cells, and IL-10

signaling itself (Figure 2). In both reports, mononuclear phagocyte activation state was neither absolutely M1 nor M2; rather, unique activation phenotypes fell within the continuum between M1 and M2 extremes [10,37,43]. Importantly, *Il10* deficient APP/PS1 animals exhibited polarization of microglial activation away from a traditional M2 state but had increased phagocytic capacity, confirming–yet again–how the M1/M2 dichotomy is not a useful classification system [10,37,50,52].

4. Intersection between IL-10 and apolipoprotein E pathways

Of the differentially expressed genes identified by large-scale expression profiling in *II10* targeted cerebral amyloidosis mouse models, one hit stood out: apolipoprotein E (*Apoe*) [37,43]. This is because the ε 4 allele of *APOE* is the strongest genetic susceptibility factor for human AD. Strikingly, *II10* overexpression increases cerebral *Apoe* mRNA abundance [43], while *II10* deficiency reduces *Apoe* expression [37] in mouse models. These findings are particularly interesting, because human ApoE isoform-specifically regulates A β metabolism, aggregation, deposition and clearance [54,55], and cerebral vasotonus [56]. Knocking in human *APOE4* into mouse models of cerebral amyloidosis exacerbates microglial inflammation and A β deposition [57], while *APOE2* promotes clearance of preexisting deposits [58].

Remarkably, cerebral *ll10* overexpression increases murine APOE association with insoluble amyloid and promotes direct ApoE- A β interaction that inhibits phagocytosis of the complex [43]. Additionally, treatment of microglial cultures with A β bound to human ApoE isoforms reveals isoform-specific reduction of A β phagocytosis (E4 > E3 > E2) [37]. These results are particularly significant in light of previous studies showing ApoE isoformdependent enhanced A β oligomerization and fibril stabilization [59–62], and A β clearance by glial cells can be negatively affected by ApoE-A β association [63]. Strikingly, this result mirrors the well-established relative risk of ApoE for human AD (E4 > E3 > E2) [64]. Importantly, *ll10* deficiency rescues impaired A β phagocytosis by ApoE3 [37]. Altogether, these results provide the basis for mechanistic intersection between IL-10 signaling and the greatest genetic risk factor for LOAD: *APOE*.

5. Conclusions

In this Opinion, we have illustrated the complex interplay between anti-inflammatory factors and AD pathoetiology. Specifically, evidence is presented that innate immunity is pathologically repressed by anti-inflammatory factors in the AD brain; obviating β -amyloid plaque phagocytosis and clearance. Specifically, elevated IL-10 signaling in the context of cerebral amyloidosis promotes a pathological form of cerebral innate immunity that licenses ApoE-A β binding and reduces A β clearance by mononuclear phagocytes. Therefore, IL-10 acts via dual mechanisms to promote cerebral amyloidosis–on the one hand, by retarding A β phagocytosis, and on the other, by promoting ApoE-A β co-deposition (Figure 3).

Modulating IL-10/STAT3 signaling has proven effective in other therapeutic settings [65,66], and rebalancing cerebral innate immunity to promote beneficial neuroinflammation may be more efficacious than anti-inflammatory therapy for AD. However, several key issues need to be explored before moving forward and enlisting the innate immune system to

clear A β as a therapeutic modality (see Outstanding Questions). First and foremost, whether activated monocytes in AD patients' brains arise from infiltrating peripheral macrophages or expansion of resident microglia remains unknown. As discussed above, in late stages of AD (*i.e.*, after amyloidosis is established) microglia are inefficient A β phagocytes [5,7]. However, several studies have shown that peripheral macrophages infiltrating the brain in mouse models of cerebral amyloidosis home to amyloid plaques and effectively clear A β [40,67,68]. To therapeutically target innate immunity in AD, it is therefore crucial to delineate the relative contribution(s) of central vs. peripheral mononuclear cells. Paninhibition of IL-10 signaling would not be judicious, because IL-10 is critically important to militate against peripheral inflammatory diseases (e.g., rheumathoid arthritis, psoriasis, and Crohn's disease). Yet, it deserves mentioning that IL-10-based therapeutics have already proven effective in aging populations [69]. The observation that IL-10 can modulate ApoE-A β interaction is original, and has major implications for therapeutic strategies. A deeper understanding of the intersecting points between IL-10 and ApoE in the context of AD is needed. It is important to note that genetic blockade of both major anti-inflammatory pathways: TGF-β-Smad 2/3 [40] and IL-10/STAT3 [37], results in an innate immune shift towards cerebral amyloid clearance. This exciting new research focus on blocking antiinflammatory pathways is eventually expected to lead to safe and effective AD treatment.

Outstanding Questions

- Do activated monocytes in AD patients' brains arise from infiltrating peripheral macrophages or expansion of resident microglia? Is it possible to specifically target peripheral monocytes or CNS microglia in LOAD?
- Are hematogenous monocytes more efficient Aβ phagocytes than brain-resident microglia? If so, what is the molecular mechanism for this important difference?
- What is the mechanism underlying increased LOAD risk incurred by the polymorphic *IL10* allele? Does the *IL10* polymorphism affect IL-10 signaling independently of its action on reducing IL-10 expression?
- Does IL-10 retard monocyte clearance of Aβ/β-amyloid, and therefore contribute to amyloid deposition in AD? What inflammatory genes does IL-10 signaling directly alter to influence monocyte Aβ phagocytosis and clearance?
- Can we determine more specific therapeutic targets that are downstream of the IL-10 pathway? At what stage would blocking IL-10 signaling be most effective against LOAD?
- How does IL-10 modulate ApoE-Aβ interaction, and does association of ApoE with lipids influence formation of this complex? Do lipids impact ApoE isoform-specific alteration of Aβ phagocytosis?
- Does *Il10* deletion mitigate Alzheimer-like pathology in *Apoe*-deficient mice, or in chimeric mice expressing human ApoE2, ApoE3, or ApoE4? Answering this important question could pave the way to understand the specific role(s) of

ApoE isoforms in *II10* deficient mouse models of cerebral amyloidosis, and in the human disease.

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References

- 1. Brookmeyer R, et al. National estimates of the prevalence of Alzheimer's disease in the United States. Alzheimers Dement. 2011; 7:61–73. [PubMed: 21255744]
- 2. Heneka MT, et al. Innate immunity in Alzheimer's disease. Nature Immunol. 2015; 16:229–236. [PubMed: 25689443]
- Yan R, Vassar R. Targeting the β secretase BACE1 for Alzheimer's disease therapy. Lancet Neurol. 2014; 13:319–329. [PubMed: 24556009]
- Golde TE, et al. γ-Secretase inhibitors and modulators. Biochim Biophys Acta. 2013; 1828:2898– 2907. [PubMed: 23791707]
- 5. Mawuenyega KG, et al. Decreased clearance of CNS beta-amyloid in Alzheimer's disease. Science. 2010; 330:1774. [PubMed: 21148344]
- 6. Hickman SE, et al. Microglial dysfunction and defective beta-amyloid clearance pathways in aging Alzheimer's disease mice. J Neurosci. 2008; 28:8354–8360. [PubMed: 18701698]
- 7. Johnston H, et al. Assessing the contribution of inflammation in models of Alzheimer's disease. Biochem Soc Trans. 2011; 39:886–890. [PubMed: 21787318]
- Orre M, et al. Isolation of glia from Alzheimer's mice reveals inflammation and dysfunction. Neurobiol. 2014; 35:2746–2760.
- 9. Heneka MT, et al. Neuroinflammation in Alzheimer's disease. Lancet Neurol. 2015; 14:388–405. [PubMed: 25792098]
- Town T, et al. The microglial "activation" continuum: from innate to adaptive responses. J Neuroinflammation. 2005; 2:24. [PubMed: 16259628]
- 11. Gjoneska E, et al. Conserved epigenomic signals in mice and humans reveal immune basis of Alzheimer's disease. Nature. 2015; 518:365–369. [PubMed: 25693568]
- Reitz C, Mayeux R. Alzheimer disease: epidemiology, diagnostic criteria, risk factors and biomarkers. Biochem Pharmacol. 2014; 88:640–651. [PubMed: 24398425]
- 13. Hazrati LN, et al. Genetic association of CR1 with Alzheimer's disease: a tentative disease mechanism. Neurobiol Aging. 2012; 33:2949.e5–2949.e12. [PubMed: 22819390]
- Griciuc A, et al. Alzheimer's Disease Risk Gene CD33 Inhibits Microglial Uptake of Amyloid Beta. Neuron. 2013; 78:631–43. [PubMed: 23623698]
- Hardy J, et al. Pathways to Alzheimer's disease. J Intern Med. 2014; 275:296–303. [PubMed: 24749173]
- 16. Jaturapatporn D, et al. Aspirin, steroidal and non-steroidal anti-inflammatory drugs for the treatment of Alzheimer's disease. Cochrane Database Syst Rev. 2012; 2:CD006378. [PubMed: 22336816]
- Szekely CA, Zandi PP. Non-steroidal anti-inflammatory drugs and Alzheimer's disease: the epidemiological evidence. CNS Neurol Disord Drug Targets. 2010; 9:132–139. [PubMed: 20205647]

- Leoutsakos JMS, et al. Effects of non-steroidal anti-inflammatory drug treatments on cognitive decline vary by phase of pre-clinical Alzheimer disease: findings from the randomized controlled Alzheimer's Disease Anti-inflammatory Prevention Trial. Int J Geriatr Psychiatry. 2012; 27:364– 374. [PubMed: 21560159]
- 20. Heneka MT, et al. NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. Nature. 2013; 493:674–678. [PubMed: 23254930]
- Fuhrmann M, et al. Microglial Cx3cr1 knockout prevents neuron loss in a mouse model of Alzheimer's disease. Nat Neurosci. 2010; 13:411–413. [PubMed: 20305648]
- 22. Berg vom J, et al. Inhibition of IL12/IL23 signaling reduces Alzheimer's disease–like pathology and cognitive decline. Nat Med. 2012; 18:1812–1819. [PubMed: 23178247]
- Lee S, et al. CX3CR1 deficiency alters microglial activation and reduces beta-amyloid deposition in two Alzheimer's disease mouse models. Am J Pathol. 2010; 177:2549–2562. [PubMed: 20864679]
- 24. Lee S, et al. Opposing effects of membrane-anchored CX3CL1 on amyloid and tau pathologies via the p38 MAPK pathway. J Neurosci. 2014; 34:12538–12546. [PubMed: 25209291]
- 25. Gezen-Ak D, et al. BDNF, TNFα, HSP90, CFH, and IL-10 serum levels in patients with early or late onset Alzheimer's disease or mild cognitive impairment. J Alzheimers Dis. 2013; 37:185–195. [PubMed: 23948885]
- 26. Akiyama H, et al. Inflammation and Alzheimer's disease. Neurobiol Aging. 2000; 21:383–421. [PubMed: 10858586]
- 27. Weeraratna AT, et al. Alterations in immunological and neurological gene expression patterns in Alzheimer's disease tissues. Exp Cell Res. 2007; 313:450–461. [PubMed: 17188679]
- Wyss-Coray T, et al. Amyloidogenic role of cytokine TGF-beta1 in transgenic mice and in Alzheimer's disease. Nature. 1997; 389:603–606. [PubMed: 9335500]
- Vural P, et al. The combinations of TNFalpha-308 and IL-6 -174 or IL-10 -1082 genes polymorphisms suggest an association with susceptibility to sporadic late-onset Alzheimer's disease. Acta Neurol Scand. 2009; 120:396–401. [PubMed: 19744138]
- Arosio B, et al. Interleukin-10 and interleukin-6 gene polymorphisms as risk factors for Alzheimer's disease. Neurobiol Aging. 2004; 25:1009–1015. [PubMed: 15212825]
- Lio D, et al. Interleukin-10 promoter polymorphism in sporadic Alzheimer's disease. Genes Immun. 2003; 4:234–238. [PubMed: 12700599]
- 32. Ma SL, et al. The association between promoter polymorphism of the interleukin-10 gene and Alzheimer's disease. Neurobiol Aging. 2005; 26:1005–1010. [PubMed: 15748779]
- 33. Depboylu C, et al. Lack of association of interleukin-10 promoter region polymorphisms with Alzheimer's disease. Neurosci Lett. 2003; 342:132–134. [PubMed: 12727335]
- 34. Ramos EM, et al. Tumor necrosis factor alpha and interleukin 10 promoter region polymorphisms and risk of late-onset Alzheimer disease. Arch Neurol. 2006; 63:1165–1169. [PubMed: 16908746]
- 35. Scassellati C, et al. Promoter haplotypes of interleukin-10 gene and sporadic Alzheimer's disease. Neurosci Lett. 2004; 356:119–122. [PubMed: 14746878]
- 36. Strle K, et al. Interleukin-10 in the brain. Crit Rev Immunol. 2001; 21:427–449. [PubMed: 11942558]
- Guillot-Sestier MV, et al. Il10 deficiency rebalances innate immunity to mitigate Alzheimer-like pathology. Neuron. 2015; 85:534–548. [PubMed: 25619654]
- 38. Zhang B, et al. Integrated systems approach identifies genetic nodes and networks in late-onset Alzheimer's disease. Cell. 2013; 153:707–720. [PubMed: 23622250]
- 39. Li X, et al. Integrated genomic approaches identify major pathways and upstream regulators in late onset Alzheimer's disease. Sci Rep. 2015; 5:12393. [PubMed: 26202100]
- 40. Town T, et al. Blocking TGF-beta-Smad2/3 innate immune signaling mitigates Alzheimer-like pathology. Nat Med. 2008; 14:681–687. [PubMed: 18516051]
- 41. Gate D, et al. Macrophages in Alzheimer's disease: the blood-borne identity. J Neural Transm. 2010; 117:961–970. [PubMed: 20517700]

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- 43. Chakrabarty P, et al. IL-10 alters immunoproteostasis in APP mice, increasing plaque burden and worsening cognitive behavior. Neuron. 2015; 85:519–533. [PubMed: 25619653]
- Williams LM, et al. Interleukin-10 suppression of myeloid cell activation--a continuing puzzle. Immunology. 2004; 113:281–292. [PubMed: 15500614]
- 45. Murray PJ. Understanding and exploiting the endogenous interleukin-10/STAT3-mediated antiinflammatory response. Curr Opin Pharmacol. 2006; 6:379–386. [PubMed: 16713356]
- 46. Banchereau J, et al. From IL-2 to IL-37: the expanding spectrum of anti-inflammatory cytokines. Nat Immunol. 2012; 13:925–931. [PubMed: 22990890]
- 47. Doty KR, et al. The role of the immune system in neurodegenerative disorders: Adaptive or maladaptive? Brain research. 201410.1016/j.brainres.2014.09.008
- Loewenbrueck KF, et al. Th1 responses to beta-amyloid in young humans convert to regulatory IL-10 responses in Down syndrome and Alzheimer's disease. Neurobiol Aging. 2010; 31:1732– 1742. [PubMed: 19058879]
- 49. Holness CL, Simmons DL. Molecular cloning of CD68, a human macrophage marker related to lysosomal glycoproteins. Blood. 1993; 81:1607–1613. [PubMed: 7680921]
- Guillot-Sestier MV, Town T. Innate immunity in Alzheimer's disease: a complex affair. CNS Neurol Disord Drug Targets. 2013; 12:593–607. [PubMed: 23574177]
- 51. Rezai-Zadeh K, et al. CNS infiltration of peripheral immune cells: D-Day for neurodegenerative disease? J Neuroimmune Pharmacol. 2009; 4:462–475. [PubMed: 19669892]
- 52. Weitz TM, Town T. Microglia in Alzheimer's Disease: It's All About Context. Int J Alzheimers Dis. 2012; 2012:314185. [PubMed: 22779026]
- Lee DC, et al. Review: experimental manipulations of microglia in mouse models of Alzheimer's pathology: activation reduces amyloid but hastens tau pathology. Neuropathol Appl Neurobiol. 2013; 39:69–85. [PubMed: 23171029]
- 54. Kanekiyo T, et al. ApoE and Aβ in Alzheimer's disease: accidental encounters or partners? Neuron. 2014; 81:740–754. [PubMed: 24559670]
- 55. Tai LM, et al. Levels of soluble apolipoprotein E/amyloid-β (Aβ) complex are reduced and oligomeric Aβ increased with APOE4 and Alzheimer disease in a transgenic mouse model and human samples. J Biol Chem. 2013; 288:5914–5926. [PubMed: 23293020]
- 56. Paris D, et al. Isoform-specific vasoconstriction induced by apolipoprotein E and modulation of this effect by Alzheimer's beta-amyloid peptide. Neurosci Lett. 1998; 256:73–76. [PubMed: 9853706]
- 57. Rodriguez GA, et al. Human APOE4 increases microglia reactivity at Aβ plaques in a mouse model of Aβ deposition. J Neuroinflammation. 2014; 11:111. [PubMed: 24948358]
- 58. Hudry E, et al. Gene transfer of human Apoe isoforms results in differential modulation of amyloid deposition and neurotoxicity in mouse brain. Sci Transl Med. 2013; 5:212ra161.
- 59. Hashimoto T, et al. Apolipoprotein E, especially apolipoprotein E4, increases the oligomerization of amyloid β peptide. J Neurosci. 2012; 32:15181–15192. [PubMed: 23100439]
- 60. Garai K, et al. The binding of apolipoprotein E to oligomers and fibrils of amyloid-β alters the kinetics of amyloid aggregation. Biochemistry. 2014; 53:6323–6331. [PubMed: 25207746]
- 61. Cerf E, et al. High ability of apolipoprotein E4 to stabilize amyloid-β peptide oligomers, the pathological entities responsible for Alzheimer's disease. FASEB J. 2011; 25:1585–1595. [PubMed: 21266538]
- 62. Pankiewicz JE, et al. Blocking the apoE/Aβ interaction ameliorates Aβ-related pathology in APOE ε2 and ε4 targeted replacement Alzheimer model mice. Acta Neuropathol Commun. 2014; 2:75. [PubMed: 24972680]
- 63. Mulder SD, et al. Apolipoproteins E and J interfere with amyloid-beta uptake by primary human astrocytes and microglia in vitro. Glia. 2014; 62:493–503. [PubMed: 24446231]
- Yu JT, et al. Apolipoprotein E in Alzheimer's disease: an update. Annu Rev Neurosci. 2014; 37:79–100. [PubMed: 24821312]

- Ni G, et al. Manipulating IL-10 signalling blockade for better immunotherapy. Cell Immunol. 2015; 293:126–129. [PubMed: 25596475]
- 66. Munoz J, et al. STAT3 inhibitors: finding a home in lymphoma and leukemia. Oncologist. 2014; 19:536–544. [PubMed: 24705981]
- 67. Simard AR, et al. Bone marrow-derived microglia play a critical role in restricting senile plaque formation in Alzheimer's disease. Neuron. 2006; 49:489–502. [PubMed: 16476660]
- 68. Simard AR, Rivest S. Neuroprotective properties of the innate immune system and bone marrow stem cells in Alzheimer's disease. Mol Psychiatry. 2006; 11:327–335. [PubMed: 16491130]
- 69. Kwilasz AJ, et al. The therapeutic potential of interleukin-10 in neuroimmune diseases. Neuropharmacology. 201410.1016/j.neuropharm.2014.10.020

Trends Box

- Recent GWAS have identified a cluster of AD risk alleles belonging to core innate immune pathways that modulate phagocytosis.
- Functional polymorphism within the *ll10* gene has been linked to increased risk for LOAD in certain populations, and IL-10 signaling is abnormally elevated in AD patient sera and brains.
- Inhibiting IL-10/STAT3 signaling dramatically mitigates Alzheimer-like pathology, while brain overexpression of *Il10* aggravates Aβ deposition in mouse models of cerebral amyloidosis.
- Elevated IL-10 signaling reduces Aβ clearance by mononuclear phagocytes and licenses ApoE-Aβ binding.
- IL-10/STAT3 pathway blockade enhances microglial Aβ phagocytic activity and decreases ApoE expression, thereby mitigating ApoE-Aβ binding that retards Aβ phagocytosis.
- Blocking anti-inflammatory mediators represents a promising future treatment approach for AD.



Figure 1. Methodology for quantitative 3D *in silico* modeling (q3DISM) of A β phagocytosis This multi-stage q3DISM technique is depicted. This cutting-edge technology allows for true 3D quantitation of A β phagocytosis *in vivo* by mononuclear phagocytes.



Figure 2. $I\!I\!0$ dependent neuroinflammatory gene profiles in mouse models of cerebral amyloidosis

The diagram shows the broad categories of major gene groups affected by experimental targeting of *Il10* (overexpression or genetic ablation) according to transcriptomics published in [32–33].



Figure 3. Proposed model for the impact of IL-10 on A β clearance by mononuclear phagocytes The top panel depicts effects of supra-physiologic IL-10; present in AD brains and in mouse models of cerebral amyloidosis. IL-10/STAT3 pathway activation reduces monocyte A β phagocytosis. Concomitantly, ApoE expression is increased and clusters in A β deposits, retarding A β clearance. The lower panel illustrates effects of *II10* ablation in mouse models of cerebral amyloidosis. Microglial phagocytic capacity is enhanced and ApoE expression, reduced, enabling A β clearance by mononuclear phagocytes.