Hindawi Publishing Corporation BioMed Research International Volume 2015, Article ID 623121, 12 pages http://dx.doi.org/10.1155/2015/623121

Research Article

Mining for Candidate Genes Related to Pancreatic Cancer Using Protein-Protein Interactions and a Shortest Path Approach

Fei Yuan, Yu-Hang Zhang, Sibao Wan, ShaoPeng Wang, and Xiang-Yin Kong

¹Institute of Health Sciences, Shanghai Institutes for Biological Sciences (SIBS), Chinese Academy of Sciences (CAS) & Shanghai Jiao Tong University School of Medicine (SJTUSM), Shanghai 200031, China ²College of Life Sciences, Shanghai University, Shanghai 200444, China

Correspondence should be addressed to Xiang-Yin Kong; xykong@sibs.ac.cn

Received 6 September 2015; Accepted 15 October 2015

Academic Editor: Jialiang Yang

Copyright © 2015 Fei Yuan et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Pancreatic cancer (PC) is a highly malignant tumor derived from pancreas tissue and is one of the leading causes of death from cancer. Its molecular mechanism has been partially revealed by validating its oncogenes and tumor suppressor genes; however, the available data remain insufficient for medical workers to design effective treatments. Large-scale identification of PC-related genes can promote studies on PC. In this study, we propose a computational method for mining new candidate PC-related genes. A large network was constructed using protein-protein interaction information, and a shortest path approach was applied to mine new candidate genes based on validated PC-related genes. In addition, a permutation test was adopted to further select key candidate genes. Finally, for all discovered candidate genes, the likelihood that the genes are novel PC-related genes is discussed based on their currently known functions.

1. Introduction

The pancreas is a significant versatile organ of the digestive system and endocrine system; it is significant because it assists in digestion and maintains hormonal balance via digestive enzymes and specific hormones, respectively. Pancreatic cancer (PC) originates from the pancreas, has a high rate of metastasis, and is a highly malignant tumor derived from pancreas tissue. Among various types of PC, pancreatic ductal adenocarcinoma (PDAC) accounts for more than 90% of all pancreatic tumors. PDAC is a malignancy with a poor prognosis, which is demonstrated through its one-year survival rate of approximately 18% for all stages of the disease [1]. In the Western world, pancreatic cancer is one of the top killers for human beings [2]. In 2012 alone, it resulted in 33000 deaths all over the world. In the Western world, pancreatic cancer is the fourth leading cause of death from cancer with a poor prognosis (5-year survival in less than 5% of cases according to most reports). Such a high fatality rate is attributed to the low rate of diagnosis at early age. Only a minority of patients can receive proper treatment with a 5year survival rate up to 22%. Therefore, it is significant and

crucial to study this severe disease. Similar to most types of tumors, PC is induced by both environmental and hereditary elements. Extrinsic factors such as age, gender, race, cigarette smoking, and obesity are all factors that may contribute to tumor initiation [3–5]. Further, certain chronic pancreas-associated diseases, such as diabetes mellitus and chronic pancreatitis, are also related to PC [6, 7].

Over the last decade, the genetic background for PC, especially pancreatic adenocarcinoma (PAC) that comprises most cases, has been revealed through validating a list of oncogenes and tumor suppressor genes for PC. Based on hereditary features, mutations in pancreatic adenocarcinoma can be divided into two clusters defined as common somatic mutations and germline mutations. A somatic mutation, KRAS, is regarded as the earliest and key mutation in nonfamilial PAC initiation, and it aids in maintaining invasion status and tumor progression [8]. In addition to tumor development, more mutated genes contribute to malignant phenotypes. The tumor suppressor genes p16/INK4A are significant somatic mutations and are downregulated in pancreatic adenocarcinoma [9, 10]. In many types of tumors, excessive activation of the TGF- β pathway is a mechanism of

tumor progression and invasion [11]. Another pair of tumor suppressor genes, SMAD/DPC4, is involved in PAC through regulating the TGF- β pathway and is critical to advanced tumors [12].

Further, heredofamilial pancreatic adenocarcinoma is associated with certain other significant genes with more complex mechanisms. As indicated by the available literature, most such genes participate in the DNA repair process, such as MSH1/2, PMS1/2, and BRCA1/2, which may participate in nonspecific tumor induction [13–15]. However, in several inherited familial conditions, specific known mutations have not been identified, which may hint at the complexity of carcinogenic mechanisms and the potential oncogenes as well as tumor suppressor genes [16].

To predict more potential tumor-related genes, we proposed a new method that considers protein interactions from STRING (Search Tool for the Retrieval of Interacting Genes/Proteins) [17] and mines potential PC-related genes. STRING is a database with massive amounts of information on physical and functional associations between different proteins. With an established score system, STRING enables a user to search and browse the protein interactions data as well as simultaneously quantify the statistic cooccurrence in the background [17]. Here, depending on the method and using a comprehensive analysis of the protein interaction network, we fully utilized the database containing reported PC-related genes and predicted potential genes involved in PC.

2. Materials and Methods

2.1. Materials. PC-related genes were retrieved from KEGG PATHWAY, which is one of the main databases in KEGG (Kyoto Encyclopedia of Genes and Genomes) [18]. By examining the pathway hsa05212, pancreatic cancer (http://www.genome.jp/dbget-bin/www_bget?hsa05212, accessed in December 2014), we obtained 65 PC-related genes, which comprise the gene set S. Detailed information on these genes is listed in Supplementary Material I available online at http://dx.doi.org/10.1155/2015/623121.

2.2. Method for Mining New Candidate Genes in a Protein-Protein Interaction Network. Protein-protein interaction (PPI) information is useful for investigating protein-related and gene-related problems [19-25]. Most methods for predicting protein attributes are based on the notion that two proteins that interact always share similar attributes [19-23]. Because PC-related genes must share common features related to PC, it is reasonable to use PPI information to identify whether a gene is related to PC. Here, we adopted the PPI information reported in STRING (version 9.1, http://www.string-db.org/) [17], which is a large online database that reports known and predicted protein interactions. The PPIs reported in STRING are derived from the following four sources: (1) genomic context, (2) highthroughput experiments, (3) (conserved) coexpression, and (4) previous knowledge, which imply that the physical and functional associations of the proteins were measured. To extract PPI information for humans, we downloaded the file "protein.links.v9.1.txt.gz" and selected protein interactions beginning with "9606.", which produced 2,425,314 human PPIs. Each PPI includes two proteins and one score, which measures the strength of the interaction in the range between 150 and 999. For the formulation, let us denote the score of an interaction between proteins p_1 and p_2 as $Q_i(p_1, p_2)$.

The aforementioned PPIs were used to construct a large network by taking proteins as nodes. Two nodes are adjacent if and only if the corresponding proteins comprise an interaction that is a member of the 2,425,314 PPIs. Furthermore, the interaction score should also be added to the constructed network. To generate compatibility between our network and a shortest path approach, each edge e was assigned a weight w(e) as follows: $w(e) = 1000 - Q_i(p_1, p_2)$, where p_1 and p_2 are corresponding proteins of the endpoints of e.

Two proteins that can interact with each other always share similar attributes [19-23]. Further, considering the interaction score, this notion can be generalized as follows: two proteins in an interaction with a high score are more likely to share similar attributes than those with a low score. Moreover, if we consider a series of proteins $p_1, p_2, ..., p_n$ such that two consecutive proteins can comprise an interaction with a high score, then these proteins may all share some common attributes. By mapping these proteins onto the constructed network, they may comprise a shortest path connecting p_1 and p_n , which is a path connecting p_1 and p_n such that the summation of the weights of edges on the path is minimum. Thus, we searched all shortest paths connecting any two PC-related genes. For two consecutive nodes in each of these paths, their corresponding proteins can comprise a PPI with a high score because they lie on a shortest path. As mentioned above, they can share similar functions. For a specific shortest path p_1, p_2, \dots, p_n , where p_1 and p_n are encoded by PC-related genes, p_2 shares similar functions with p_1 ; that is, p_2 may be encoded by an invalidated or known PC-related gene; p_3 shares similar functions with p_2 ; thus p_3 may also share similar functions with p_1 and may be encoded by an invalidated or known PC-related gene. This can be induced to p_4, p_5, \dots, p_{n-1} . Thus, we extracted genes in these shortest paths and excluded those that were members of 65 PC-related genes. These genes were referred to as shortest path genes and deemed to have special relationships with PC. The similar scheme has been applied to extract novel genes or chemicals related to other diseases or some biological processes [26-29]. In addition, to distinguish those genes, each shortest path gene was assigned a value, which is referred to as betweenness and defined as the number of shortest paths that contain the gene. The betweenness indicates the direct and indirect associations between shortest path genes and PC-related genes [30].

2.3. Further Selection. By executing the method mentioned in Section 2.2, certain shortest path genes can be extracted from the constructed network. However, certain such genes may be false positives. To exclude this type of gene, the following method was adopted.

Certain nodes in the constructed network are general hubs; their corresponding genes may always receive a high

betweenness value even if we randomly selected certain genes as PC-related genes, but the genes exhibit few relationships with PC. To exclude this type of gene among shortest path genes, we randomly produced 1,000 gene sets with the same size as *S* and compared the betweenness of the shortest path genes in these sets to *S*. The detailed procedures are described as follows.

- (I) Randomly produce 1,000 gene sets, such as S_1 , S_2, \ldots, S_{1000} , each of which with the same size as S, the set consisting of PC-related genes.
- (II) The method described in Section 2.2 was executed for 1,000 rounds. For the ith round, the PC-related genes in S were replaced with the genes in S_i , and the shortest paths connecting any pair of genes in S_i were searched, thereby counting the betweenness of each shortest path gene based on these shortest paths.
- (III) For each shortest path gene, there were 1,000 betweenness on randomly produced gene sets and one betweenness on *S*. After comparing them, we generated a measurement referred to as permutation FDR, which defines the proportion of randomly produced gene sets on which the betweenness was larger than that on *S*.
- (IV) To exclude shortest path genes with general hubs in the network and few relationships with PC, we excluded shortest path genes with permutation FDR values equal to or larger than 0.05. The remaining shortest path genes are referred to as candidate genes.

3. Results and Discussion

- 3.1. Shortest Path Genes. Based on the method described in Section 2.2, we searched all shortest paths connecting any pair of PC-related genes and produced 2,080 shortest paths (each pair of PC-related genes can be connected by a shortest path), which are provided in Supplementary Material II. A graph with these 2,080 shortest paths is shown in Figure 1. The detailed information of edges in this graph is provided in Supplementary Material III. These 2,080 paths involved 134 Ensembl gene IDs. We excluded 65 Ensembl IDs for PC-related genes, resulting in 69 shortest path genes, which are listed in Supplementary Material IV. In addition, we counted the betweenness of each shortest path gene, which is also provided in Supplementary Material IV.
- 3.2. Candidate Genes. As mentioned in Section 3.1, several shortest path genes were retrieved. According to the principle underlying the method in Section 2.2, these genes may have special relationships with PC. However, certain such genes may be false positives and have few relationships with PC. Thus, we performed a permutation test to control for this type of genes. After calculating the permutation FDR for each shortest path gene, which are listed in Supplementary Material IV, we discarded the genes with a permutation FDR greater than or equal to 0.05, thereby generating thirteen candidate genes, which are listed in Table 1.

3.3. Analysis of Significant Candidate Genes. Using our method, we predicted thirteen genes that may participate in PC. Based on the principle underlying our method, the candidate genes are specifically connected with PC-related genes. Moreover, such genes have all been reported as genes that are relevant to PC and may exhibit diverse functions in tumor initiation and invasion, especially in PC.

Among the candidate genes, three are inhibitory genes that may participate in PC through their respective mechanisms. The first candidate gene is NFKBIA (see row 1 of Table 1, with betweenness 73 and permutation FDR 0.002), which is expressed in pancreatic tissues [31]. NFKBIA encodes a member of the NF-kappa-B inhibitor family, which has been confirmed to further participate in interactions with REL dimers to inhibit the NF-kappa-B pathway in processes of inflammation immune response and tumorigenesis [32, 33]. Targeted by a specific microRNA, miR196a, NFKBIA has been proved to be associated with PC, especially in the metastasis process [31]. As we have mentioned above, in most tumors including pancreatic cancer, NF-kappa-B pathway has been widely reported to be overactivated and has a close connection with the patients' prognosis, indicating the underlying relationship between NFKBIA and the pancreatic cancer [34-37]. Mutations have been widely reported to contribute to specific functional alteration of crucial proteins including NFKBIA in diseases especially in various cancer subtypes like pancreatic cancer [38-41]. What is more, as one of the crucial inhibitory components of NF-kappa-B pathway, NFKBIA has been reported to be downregulated in cancer [42, 43]. The overexpression of NFKBIA has also been reported to be associated with a better prognosis of various treatment methods in different human tumor subtypes, especially for the prognosis of patients that have taken alpha 1-adrenoceptor antagonists and radiotherapy [44, 45]. Further, considering the inflammation associated function of such gene, such mutations and expression alteration may contribute to the process of tumorigenesis through two individual regulation mechanisms: proliferation associated pathways that involve NF-kappa-B and specific immune response associated pathways in tumor microenvironment [46]. In addition to the NF-kappa-B pathway, the JNK pathway is also a specific pathway in tumor initiation and progression, including PC [47, 48]. A specific regulator of Rho protein exchange reactions which is crucial for JNK pathway, ARHGDIA (see row 2 of Table 1, with betweenness 94 and permutation FDR 0.012), was also identified using our method and functions in several types of tumors [49, 50]. In addition, this apoptosis inhibitory protein has been confirmed to control Rho protein homeostasis and participate in the initiation and progression of PC through the apoptosis associated GDP/GTP exchange reaction via RhoA-Rho pathway [51]. Apoptosis inhibition widely exists in tumor tissues and is an effective way to induce malignant cells, implying the potential relationship between ARHGDIA and pancreatic cancer [52, 53]. As we all know, pathways that contribute to the proliferation and invasion of cancer cells (JNK-STAT, RhoA-Rho signaling pathway, etc.) have all been confirmed to involve phosphorylation and dephosphorylation process which may be regulated by the survival

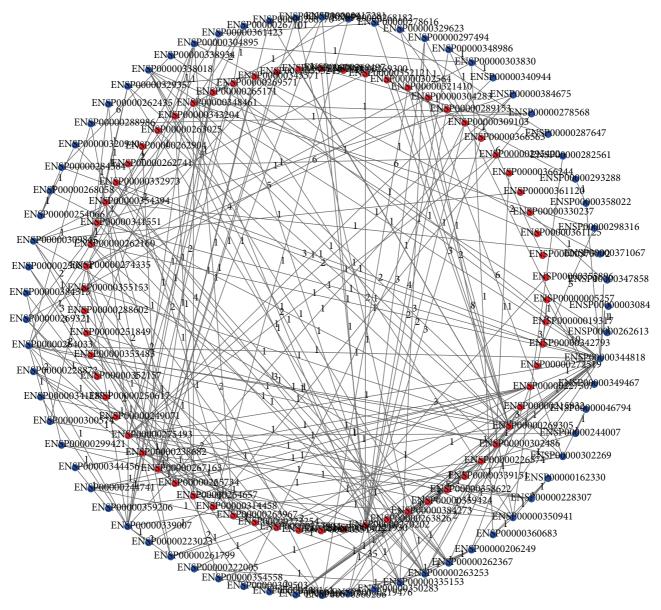


FIGURE 1: A graph consisting of 2,080 shortest paths. The nodes on the inner circle (red nodes) represent 65 PC-related genes, while the nodes on the outer circle (blue nodes) represent 69 shortest path genes. The numbers on the edges represent the weights of the edges.

associated GDP/GTP exchange reaction, further validating the potential relationship between ARHGDIA and pancreatic cancer [54–58]. What is more, regarded as a therapeutic target of pancreatic cancer, ARHGDIA (also known as RhoGDI) has been actually proved to be associated with the proliferation and invasion process of the pancreatic tumorigenesis which may further affect the prognosis of various patients [51]. The expression quantity of such gene, ARHGDIA, as the specific negative regulator of Rho protein exchange reactions, has been widely confirmed to be downregulated in tumors, which further induce the activation of survival associated Rho GTPases and promote the progression of various tumors [55–57]. As for specific mutations that may affect the biological function of such gene, coincidentally, specific triple Y156F/S101A/S174A-RhoGDI mutation has been confirmed

to be related to the progression of pancreatic cancer [58]. Furthermore, we also obtained the gene XIAP as one of the candidate genes in our list (see row 3 of Table 1, with betweenness 64 and permutation FDR 0.006) that encodes a functional apoptosis suppressor protein. As the most potent apoptosis suppressor, XIAP interacts with several caspases directly and prevents the apoptosis process [59–61]. In PC, XIAP has been proved to be critical for the progression and prognosis of this disease via several classical pathways (Erk, PTEN/PI3K/AKT, etc.) [62–64]. What is more, the overexpression of XIAP has been confirmed to be associated with poor prognosis of pancreatic cancer. Therefore, such protein has already been applied as a useful laboratory test parameter and a functional therapeutic target of pancreatic cancer [64, 65]. As an immune associated functional gene,

TABLE 1: Detailed information on the thirteen candidate genes.

Row number	Ensembl ID	Gene symbol	Full name	Betweenness	Permutation FDR
1	ENSP00000216797	NFKBIA	Nuclear Factor of Kappa Light Polypeptide Gene Enhancer in B-Cells Inhibitor, Alpha	73	0.002
2	ENSP00000269321	ARHGDIA	Rho GDP Dissociation Inhibitor (GDI) Alpha	94	0.012
3	ENSP00000347858	XIAP	X-Linked Inhibitor of Apoptosis, E3 Ubiquitin Protein Ligase	64	0.006
4	ENSP00000250894	MAPK8IP3	Mitogen-Activated Protein Kinase 8 Interacting Protein 3	64	<0.001
5	ENSP00000262613	SLC9A3R1	Solute Carrier Family 9, Subfamily A (NHE3, Cation Proton Antiporter 3), Member 3 Regulator 1	128	0.031
6	ENSP00000384515	PARVB	Parvin, Beta	64	< 0.001
7	ENSP00000254066	RARA	Retinoic Acid Receptor, Alpha	24	0.005
8	ENSP00000299421	ILK1	Integrin-linked kinase 1	64	0.03
9	ENSP00000338934	EZR	Ezrin	128	0.026
10	ENSP00000309845	HRAS	Harvey Rat Sarcoma Viral Oncogene Homolog	113	0.033
11	ENSP00000335153	HSP90AA1	Heat Shock Protein 90 kDa Alpha (Cytosolic), Class A Member 1	230	0.007
12	ENSP00000348986	INS-IGF2	INS-IGF2 Readthrough	64	0.031
13	ENSP00000284384	PRKCA	Protein Kinase C, Alpha	175	< 0.001

the polymorphism of XIAP may contribute to the complex and personalized immune response to pancreatic cancers. Some of the functional variants have already been proved to be directly associated with the prognosis of various subtypes of pancreatic cancer, indicating the potential significant role of XIAP in pancreatic cancer [62, 66].

In addition to inhibitory genes, a group of scaffold genes was also among the thirteen candidate genes. MAPK8IP3 (see row 4 of Table 1, with betweenness 64 and permutation FDR <0.001), which is also known as JIP3, is a scaffold protein involved in the JNK pathway [67]. Associated with the MAP kinase cascade, it may be related to PC via potential downstream mechanisms [67-69]. With high tissue specificity, such gene has been reported to actually induce the excessive proliferation of pancreatic cells, which may further initiate the biological process of pancreatic cancer [70]. MAPK8IP3 is also one of the crucial components of JNK pathway which has been overactivated in tumors including pancreatic tumor [71, 72]. During the initiation and invasion stages of pancreatic cancer, MAPK8IP3 has been reported to be overexpressed which may further contribute to the excessive proliferation of pancreatic cells in clinical cases [70]. Mutations have also been identified in MAPK8IP3 which has been reported to partially interfere with its own functions under pathological conditions [73, 74]. Some of the specific mutations of MAPK8IP3 have been reported to influence the adhesion and invasion process of various cancer cells including the pancreatic cancer cells, validating the crucial role of MAPK8IP3 in pancreatic cancer [75, 76]. As another functional scaffold protein that participates in the interaction between the cell membrane and cytoskeleton,

our candidate gene SLC9A3R1 (see row 5 of Table 1, with betweenness 128 and permutation FDR 0.031) regulates the subcellular location and function of SLC9A3 and plays a specific role in the Wnt pathway [77–79]. Known as NHERF1, SLC9A3R1 acts as a scaffold protein in many types of tumors depending on its specific role in Wnt pathway, especially in PC [78, 80, 81]. During the course of pancreatic cancer raising, SLC9A3R1 has been confirmed to be overexpressed and involve the poor prognosis of such disease [81, 82]. Just like other candidates encoding scaffold proteins, SLC9A3R1 has also been proved to contain specific nonsynonymous mutations in various cancer subtypes, especially in pancreatic cancer [83, 84]. Coding an adaptor protein, the candidate gene PARVB (see row 6 of Table 1, with betweenness 64 and permutation FDR <0.001) is critical to certain specific biological processes in cancer, such as ERK signaling pathway and focal adhesion which both have been identified in pancreatic cancer [85, 86]. Therefore, this gene is critical for tumorigenesis and may have a specific role in PC [87]. As a crucial adaptor protein, the overexpression of PARVB has been proved to be associated with high cell proliferation rate which indicates worsening clinical prognosis of tumors, including pancreatic cancer [86, 88]. The polymerase of such gene, PARVB, has also been identified and confirmed to be associated with a group of severe diseases including cancers [87]. Such abnormal expression level and high polymerase in tumor cells suggest that PARVB may play a crucial role in the specific pathogenic process of pancreatic cancer.

Six remaining genes may also be associated with PC in their respective way. RARA (see row 7 of Table 1, with betweenness 24 and permutation FDR 0.005) encodes

a specific receptor for retinoic acid, which may further regulate germ cell development during spermatogenesis and the expression of specific genes through recruiting the chromatin complex (KMT2E, MLL5, etc.) [89-91]. As we all know, the stem-cell-like features of tumor cells may be associated with the potential invasion and proliferation ability of such tumor cells [92-94]. In pancreatic cancer, it has been confirmed that the more "stem-like" the cancer cell is, the more malignant the cancer is [95]. Similar to its specific function in germ cells, RARA is related to the regulation of stem- cell-like features in PC, which implies that it may further affect the prognosis of such a severe malignancy [96]. The overexpression of RARA has been reported to be related to specific biological behaviors of tumors such as EMT (epithelial-to-mesenchymal) and invasion, which indicates poor prognosis of pancreatic cancers in clinical cases [97]. Although few mutations have been identified in RARA, crucial fusion gene variants have been widely identified in cancers which may be analyzed later. As we have mentioned above, SLC9A3R1 has been reported to regulate cellular morphology as a scaffold protein [78]. Similarly, the candidate gene ILK1 (see row 8 of Table 1, with betweenness 64 and permutation FDR 0.005) also mediates cell architecture, signaling transduction and cell adhesion via integrin-mediated signaling transduction, especially in tumor tissues [98, 99]. Although studies have not explained clearly how ILK1 actually contributes to the process of pancreatic cancer, it has been shown to participate in several types of tumors and may be associated with several unique molecules (GSK, AKT, PTEN, etc.) that all have been confirmed to be critical to PC [98-103]. Recently, such gene has been reported to be associated with the expression of TGF- α , which may partially explain the underlying carcinogenic mechanism of such gene [104]. What is more, such gene has been confirmed to be overexpressed during the whole biological process of tumorigenesis including the proliferation, migration, and invasion of pancreatic cancers [104-106]. Further, specific mutations have also been identified in such gene. Integrinlinked kinase 1 (ILK1) and HSP90 (HSP90AA1, which is also in our list and will be analyzed later) have both been confirmed to contain quite a lot of tumor-associated variants. Such mutations of ILK1 have been reported to be functional pancreatic cancer biomarkers [107-109]. All in all, ILK1 has been proved to regulate the tumorigenesis process of PC through a specific TGF- α -associated mechanism. As a functional SLC9A3R1-associated protein, a specific protein ezrin is encoded by one of our candidate genes EZR (see row 9 of Table 1, with betweenness 128 and permutation FDR 0.026) [110]. Crucial for cytoplasmic peripheral membrane, EZR is associated with the actin cytoskeleton and regulates the surface expression of actin [111, 112]. It has been widely reported that ezrin is critical for the proliferation, metastasis, and invasion process of several tumor subtypes, especially of pancreatic cancer [113, 114]. The overexpression of EZR has been widely reported to be associated with the initiation, proliferation, and metastasis processes of tumor, which strongly affect the patients' prognosis especially in pancreatic cancer [115-117]. EZR, as the crucial scaffold protein which may interact with SLC9A3R1 that we have mentioned above, has also been reported to contain specific mutations that may

contribute to the tumorigenesis process of various cancer subtypes including pancreatic cancer [118]. All in all, EZR may be a potential pancreatic-cancer-associated gene and may contribute to similar pathways with SL9A3R1 as we have mentioned above [110].

Further, we also predicted one of the most famous oncogenes HRAS (see row 10 of Table 1, with betweenness 113 and permutation FDR 0.033), which binds GDP/GTP and exhibits intrinsic GTPase activity [119]. Associated with several tumors and similar to its homologues (r-RAS, k-Ras, etc.), HRAS is a powerful oncogene that can initiate tumors through inducing excessive growth factor activation in cells and promoting malignant proliferation of tumor cells, especially in tumors in situ [120-122]. In PC, such gene has also been regarded as a main oncogene and is widely reported to be overexpressed during the entire clinical course of pancreatic cancer, validating our prediction of the potential pancreatic cancer-associated genes [123]. Several genetic alterations in HRAS (mutations, CNVs, etc.) have been identified and reported to be associated with such a severe type of cancer [124–126]. Recently, some of the crucial mutations have already been confirmed to directly contribute to the tumorigenesis of pancreatic cancer, indicating the underlying relationship between mutations of candidate genes and the process of tumorigenesis [127]. Heat shock proteins compose a unique group of proteins produced by cells to resist harmful and stressful conditions, including the tumor microenvironment, which contains less oxygen and a lower pH [128-130]. This cluster of proteins has been confirmed to be functional in tumor microenvironment and promote the progression of various tumor subtypes [131, 132]. One of the candidate genes, HSP90AA1 (see row 11 of Table 1, with betweenness 230 and permutation FDR 0.007), also encodes a specific heat shock protein, heat shock protein 90 kDa alpha, which has been reported to be expressed in the cytoplasm. Several types of heat shock proteins (such as HSP20, HSP70, and HSP90), including our predicted protein, are not only related to tumor genesis but specifically contribute to PC [133-135]. Heat shock proteins have also been confirmed to be overexpressed in pancreatic cancer, including HSP90 which is encoded by our predicted gene [136-140]. The overexpression of heat shock proteins has been confirmed to actually contribute to the tumorigenesis process of pancreatic cancer which validates our predication of HSP90AA1 as a functional candidate gene [133]. As a hormone-associated gene, our candidate gene INS-IGF2 (see row 12 of Table 1, with betweenness 64 and permutation FDR 0.031) is a readthrough gene of INS and IGF2. This gene may act as a posttranscriptional regulatory factor for the two genes INS and IGF2, which has been shown to be associated with several pancreatic-associated diseases, such as diabetes [141-143]. As a specific fusion gene, INS-IGF2 participates in several cancer types, especially in prostate cancer [144-146]. The fusion of a tumor-associated gene and a pancreas-related gene, INS-IGF2, probably participates in PC progression and invasion specifically. Recently, such fusion gene INS-IGF2 is reported to be specifically expressed in pancreatic islets especially in patients with autoimmune diseases under pathological conditions [147, 148]. Since

immune response is quite significant for tumor surveillance, INS-IGF2 is definitely associated with the unique tumorassociated immune response and may further contribute to the prognosis of pancreatic cancer [148]. Although genes like RARA (mostly identified as FIP1L1/PML-RARA fusion gene) (we have mentioned above) and INS-IGF2 have not been reported to contain specific mutations that may be associated with pancreatic cancer, such fusion genes have both been confirmed to have abnormal gene functions which may directly contribute to the tumorigenesis of various cancer subtypes, including pancreatic cancer [149, 150]. The last candidate gene PRKCA (see row 13 of Table 1, with betweenness 175 and permutation FDR <0.001) encodes a specific kinase that has been reported in several types of tumors [151, 152]. As a widely reported oncogene, PRKCA contributes to the phosphorylation process of crucial tumor-associated proteins [153]. The overexpression of PRKCA has been proved to enhance the transformed proliferation and invasion process of pancreatic cancer which further definitely influences the prognosis of such disease [154, 155]. While at the same time, genes like PRKCA which encodes functional kinase may be strongly affected by genetic alterations especially by some specific nonsynonymous mutations [75, 156]. Therefore, the polymerase of gene PRKCA may definitely affect the function of protein kinase C which has also been identified in pancreatic cancer [156, 157]. However, recent studies show that, at least in vitro, PRKCA has a specific function in tumor suppression [156]. The complex role of PKC in cancer may reflect the complicated interactive relationship between PRKCA and PC [156, 158]. All in all, our predicted gene PRKCA has been confirmed to be definitely associated with pancreatic cancer, while, at the same time, the various functions of PRKCA imply the complex role of such gene in pancreatic cancer.

Consequently, all candidate genes obtained using our method play a critical role in tumor initiation and progression. Because the detailed regulation mechanisms of only few genes remain unclear, those candidate genes may be genes that are critical to PC. PC features genetic heterogeneity, and many familial hereditary related genes remain ambiguous [16]. Therefore, in addition to the confirmed genes, such as KRAS and p16/INK41 [8, 9], more genes must be screened and validated to clearly demonstrate the mechanisms of pancreatic adenocarcinoma. In conclusion, based on reported PC-related genes, the proposed method was effective at predicting candidate tumor-associated genes in PC.

4. Conclusions

In this study, we proposed a computational method to mine new candidate genes related to pancreatic cancer, which utilized protein-protein interaction information. The analyses of the obtained candidate genes indicate that they may be novel PC-related genes. Hopefully, this contribution will promote studies on pancreatic cancer and provide new hope for designing effective treatments.

Conflict of Interests

The authors declare that there is no conflict of interests regarding the publication of this paper.

Acknowledgments

This study was supported by the National Basic Research Program of China (2011CB510101 and 2011CB510102), the National Natural Science Foundation of China (31371335), and the Innovation Program of Shanghai Municipal Education Commission (12ZZ087).

References

- [1] M. Hidalgo, S. Cascinu, J. Kleeff et al., "Addressing the challenges of pancreatic cancer: future directions for improving outcomes," *Pancreatology*, vol. 15, no. 1, pp. 8–18, 2015.
- [2] C. Verbeke, M. Löhr, J. Severin Karlsson, and M. Del Chiaro, "Pathology reporting of pancreatic cancer following neoadjuvant therapy: challenges and uncertainties," *Cancer Treatment Reviews*, vol. 41, no. 1, pp. 17–26, 2015.
- [3] J. Freilich, T. Strom, G. Springett et al., "Age and resected pancreatic cancer outcomes," *International Journal of Radiation Oncology Biology Physics*, vol. 90, no. 1, supplement, p. S356, 2014.
- [4] S. Mizuno, Y. Nakai, H. Isayama et al., "Smoking, family history of cancer, and diabetes mellitus are associated with the age of onset of pancreatic cancer in japanese patients," *Pancreas*, vol. 43, no. 7, pp. 1014–1017, 2014.
- [5] G. Preziosi, J. A. Oben, and G. Fusai, "Obesity and pancreatic cancer," *Surgical Oncology*, vol. 23, no. 2, pp. 61–71, 2014.
- [6] S. Deng, S. Zhu, B. Wang et al., "Chronic pancreatitis and pancreatic cancer demonstrate active epithelial-mesenchymal transition profile, regulated by miR-217-SIRT1 pathway," *Cancer Letters*, vol. 355, no. 2, pp. 184–191, 2014.
- [7] K. S. Snima, R. S. Nair, S. V. Nair, C. R. Kamath, and V.-K. Lakshmanan, "Combination of anti-diabetic drug metformin and boswellic acid nanoparticles: a novel strategy for pancreatic cancer therapy," *Journal of Biomedical Nanotechnology*, vol. 11, no. 1, pp. 93–104, 2015.
- [8] J. Lüttges, A. Reinecke-Lüthge, B. Möllmann et al., "Duct changes and K-ras mutations in the disease-free pancreas: analysis of type, age relation and spatial distribution," *Virchows Archiv*, vol. 435, no. 5, pp. 461–468, 1999.
- [9] D. Georgiadou, T. N. Sergentanis, S. Sakellariou et al., "Cyclin D1, p16^{INK4A} and p27^{Kipl} in pancreatic adenocarcinoma: assessing prognostic implications through quantitative image analysis," *Acta Pathologica, Microbiologica, Et Immunologica Scandinavica*, vol. 122, no. 12, pp. 1230–1239, 2014.
- [10] Z. Wang, S. Ali, S. Banerjee et al., "Activated K-Ras and INK4a/Arf deficiency promote aggressiveness of pancreatic cancer by induction of EMT consistent with cancer stem cell phenotype," *Journal of Cellular Physiology*, vol. 228, no. 3, pp. 556–562, 2013.
- [11] L. Zhang, W. Liu, Y. Qin, and R. Wu, "Expression of TGF- β 1 in Wilms' tumor was associated with invasiveness and disease progression," *Journal of Pediatric Urology*, vol. 10, no. 5, pp. 962–968, 2014.

- [12] Y. El-Gohary, S. Tulachan, P. Guo et al., "Smad signaling pathways regulate pancreatic endocrine development," *Develop*mental Biology, vol. 378, no. 2, pp. 83–93, 2013.
- [13] J. A. Veenstra, "Immunocytochemical demonstration of a homology in peptidergic neurosecretory cells in the suboesophageal ganglion of a beetle and a locust with antisera to bovine pancreatic polypeptide, FMRFamide, vasopressin and α -MSH," *Neuroscience Letters*, vol. 48, no. 2, pp. 185–190, 1984.
- [14] R. Das Gupta and R. Kolodner, "Identification of postmeiotic segregation (PMS) mutants as novel alleles of mismatch repair (MMR) genes," *The FASEB Journal*, vol. 11, no. 9, p. 1971, 1997.
- [15] T. Golan, Z. S. Kanji, R. Epelbaum et al., "Overall survival and clinical characteristics of pancreatic cancer in BRCA mutation carriers," *British Journal of Cancer*, vol. 111, no. 6, pp. 1132–1138, 2014.
- [16] A. P. Klein, K. A. Brune, G. M. Petersen et al., "Prospective risk of pancreatic cancer in familial pancreatic cancer kindreds," *Cancer Research*, vol. 64, no. 7, pp. 2634–2638, 2004.
- [17] D. Szklarczyk, A. Franceschini, S. Wyder et al., "STRING v10: protein-protein interaction networks, integrated over the tree of life," *Nucleic Acids Research*, vol. 43, no. 1, pp. D447–D452, 2015
- [18] M. Kanehisa and S. Goto, "KEGG: kyoto encyclopedia of genes and genomes," *Nucleic Acids Research*, vol. 28, no. 1, pp. 27–30, 2000
- [19] M. Deng, K. Zhang, S. Mehta, T. Chen, and F. Sun, "Prediction of protein function using protein-protein interaction data," *Journal of Computational Biology*, vol. 10, no. 6, pp. 947–960, 2003.
- [20] L. Hu, T. Huang, X. Shi, W.-C. Lu, Y.-D. Cai, and K.-C. Chou, "Predicting functions of proteins in mouse based on weighted protein-protein interaction network and protein hybrid properties," *PLoS ONE*, vol. 6, no. 1, Article ID e14556, 2011.
- [21] P. Gao, Q.-P. Wang, L. Chen, and T. Huang, "Prediction of human genes regulatory functions based on proteinprotein interaction network," *Protein and Peptide Letters*, vol. 19, no. 9, pp. 910–916, 2012.
- [22] K.-L. Ng, J.-S. Ciou, and C.-H. Huang, "Prediction of protein functions based on function-function correlation relations," *Computers in Biology and Medicine*, vol. 40, no. 3, pp. 300–305, 2010
- [23] R. Sharan, I. Ulitsky, and R. Shamir, "Network-based prediction of protein function," *Molecular Systems Biology*, vol. 3, article 88, 2007.
- [24] P. Bogdanov and A. K. Singh, "Molecular function prediction using neighborhood features," *IEEE/ACM Transactions on Com*putational Biology and Bioinformatics, vol. 7, no. 2, pp. 208–217, 2010.
- [25] M. D. M. AbdulHameed, G. J. Tawa, K. Kumar et al., "Systems level analysis and identification of pathways and networks associated with liver fibrosis," *PLoS ONE*, vol. 9, no. 11, Article ID e112193, 2014.
- [26] L. Chen, C. Chu, J. Lu, X. Kong, T. Huang, and Y. D. Cai, "A computational method for the identification of new candidate carcinogenic and non-carcinogenic chemicals," *Molecular BioSystems*, vol. 11, no. 9, pp. 2541–2550, 2015.
- [27] T. Gui, X. Dong, R. Li, Y. Li, and Z. Wang, "Identification of hepatocellular carcinoma-related genes with a machine learning and network analysis," *Journal of Computational Biology*, vol. 22, no. 1, pp. 63–71, 2015.

- [28] L. Chen, C. Chu, X. Kong, G. Huang, T. Huang, and Y. Cai, "A hybrid computational method for the discovery of novel reproduction-related genes," *PLoS ONE*, vol. 10, no. 3, Article ID e0117090, 2015.
- [29] B.-Q. Li, J. Zhang, T. Huang, L. Zhang, and Y.-D. Cai, "Identification of retinoblastoma related genes with shortest path in a protein-protein interaction network," *Biochimie*, vol. 94, no. 9, pp. 1910–1917, 2012.
- [30] J. B. M. Craven, Markov Networks for Detecting Overlapping Elements in Sequence Data, MIT Press, 2005.
- [31] F. Huang, J. Tang, X. Zhuang et al., "MiR-196a promotes pancreatic cancer progression by targeting nuclear factor kappa-Binhibitor alpha," *PLoS ONE*, vol. 9, no. 2, Article ID e87897, 2014.
- [32] P.-Y. Chang, K. Draheim, M. A. Kelliher, and S. Miyamoto, "NFKB1 is a direct target of the TAL1 oncoprotein in human T leukemia cells," *Cancer Research*, vol. 66, no. 12, pp. 6008–6013, 2006.
- [33] N. Fullard, C. L. Wilson, and F. Oakley, "Roles of c-Rel signalling in inflammation and disease," *International Journal of Biochemistry and Cell Biology*, vol. 44, no. 6, pp. 851–860, 2012.
- [34] Y. Ito, E. Kikuchi, N. Tanaka et al., "Down-regulation of NF kappa B activation is an effective therapeutic modality in acquired platinum-resistant bladder cancer," *BMC Cancer*, vol. 15, article 324, 2015.
- [35] L. Lessard, A.-M. Mes-Masson, L. Lamarre, L. Wall, J.-B. Lattouf, and F. Saad, "NF- κ B nuclear localization and its prognostic significance in prostate cancer," *BJU International*, vol. 91, no. 4, pp. 417–420, 2003.
- [36] T. Gilmore, M.-E. Gapuzan, D. Kalaitzidis, and D. Starczynowski, "Rel/NF- κ B/I κ B signal transduction in the generation and treatment of human cancer," *Cancer Letters*, vol. 181, no. 1, pp. 1–9, 2002.
- [37] B. Haefner, "NF-kappa B: arresting a major culprit in cancer," Drug Discovery Today, vol. 7, no. 12, pp. 653–663, 2002.
- [38] H. Kinugasa, K. Nouso, K. Miyahara et al., "Detection of Kras gene mutation by liquid biopsy in patients with pancreatic cancer," *Cancer*, vol. 121, no. 13, pp. 2271–2280, 2015.
- [39] A. Purohit, M. Varney, S. Rachagani, M. Ouellette, S. K. Batra, and R. K. Singh, "CXCR2 signaling axis- a helping hand to KRAS(G12D) mutation in pancreatic cancer initiation," *Cancer Research*, vol. 74, article 4874, 2014.
- [40] H. Hayashi, H. Ueno, Y. Sakamoto et al., "Gene mutation profile of pancreatic cancer in Japanese patients and its association with prognosis," *Annals of Oncology*, vol. 25, 2014.
- [41] K. Saito, Y. Nakai, H. Isayama et al., "Associations between K-ras mutation, smoking, and prognosis of pancreatic cancer," *Journal of Clinical Oncology*, vol. 32, supplement 3, abstract 298, 2014.
- [42] M. Bredel, H. Kim, T. K. Nanda et al., "Deletion of the tumor suppressor NFKBIA in triple-negative breast cancer," *International Journal of Radiation Oncology, Biology, Physics*, vol. 87, no. 2, p. S98, 2013.
- [43] G. R. Harsh, D. M. Scholtens, A. K. Yadav et al., "NFKBIA in glioblastomas: tumor suppressor and potent predictor of outcome," *Journal of Neurosurgery*, vol. 115, p. A404, 2011.
- [44] T. Mukogawa, F. Koyama, M. Tachibana et al., "Adenovirusmediated gene transduction of truncated IκBα enhances radiosensitivity in human colon cancer cells," *Cancer Science*, vol. 94, no. 8, pp. 745–750, 2003.
- [45] J. V. Partin, I. E. Anglin, and N. Kyprianou, "Quinazoline-based α 1-adrenoceptor antagonists induce prostate cancer cell apoptosis via TGF- β signalling and I κ B α induction," *British Journal of Cancer*, vol. 88, no. 10, pp. 1615–1621, 2003.

- [46] K. M. Reid-Lombardo, B. L. Fridley, W. R. Bamlet, J. M. Cunningham, M. G. Sarr, and G. M. Petersen, "Survival is associated with genetic variation in inflammatory pathway genes among patients with resected and unresected pancreatic cancer," *Annals of Surgery*, vol. 257, no. 6, pp. 1096–1102, 2013.
- [47] S. Suzuki, M. Okada, K. Shibuya et al., "JNK suppression of chemotherapeutic agents-induced ROS confers chemoresistance on pancreatic cancer stem cells," *Oncotarget*, vol. 6, no. 1, pp. 458–470, 2015.
- [48] X. P. Yuan, M. Dong, X. Li, and J. P. Zhou, "GRP78 promotes the invasion of pancreatic cancer cells by FAK and JNK," *Molecular* and Cellular Biochemistry, vol. 398, no. 1-2, pp. 55–62, 2014.
- [49] L. Liang, Q. Li, L. Y. Huang et al., "Loss of ARHGDIA expression is associated with poor prognosis in HCC and promotes invasion and metastasis of HCC cells," *International Journal of Oncology*, vol. 45, no. 2, pp. 659–666, 2014.
- [50] M. Dal Bo, F. Pozzo, R. Bomben et al., "ARHGDIA, a mutant TP53-associated Rho GDP dissociation inhibitor, is over-expressed in gene expression profiles of TP53 disrupted chronic lymphocytic leukaemia cells," *British Journal of Haematology*, vol. 161, no. 4, pp. 596–599, 2013.
- [51] M. A. Harding and D. Theodorescu, "RhoGDI signaling provides targets for cancer therapy," *European Journal of Cancer*, vol. 46, no. 7, pp. 1252–1259, 2010.
- [52] D. Dixit, V. Sharma, S. Ghosh, V. S. Mehta, and E. Sen, "Inhibition of Casein kinase-2 induces p53-dependent cell cycle arrest and sensitizes glioblastoma cells to tumor necrosis factor (TNFα)-induced apoptosis through SIRT1 inhibition," Cell Death and Disease, vol. 3, no. 2, article e271, 2012.
- [53] Q. Liu, X.-Y. Zhao, R.-Z. Bai et al., "Induction of tumor inhibition and apoptosis by a candidate tumor suppressor gene DRRI on 3p21.1," *Oncology Reports*, vol. 22, no. 5, pp. 1069–1075, 2009.
- [54] M. J. Marinissen, M. Chiariello, T. Tanos, O. Bernard, S. Narumiya, and J. S. Gutkind, "The small GTP-binding protein RhoA regulates c-jun by a ROCK-JNK signaling axis," *Molecular Cell*, vol. 14, no. 1, pp. 29–41, 2004.
- [55] W. P. Bozza, Y. Zhang, K. Hallett, L. A. Rivera Rosado, and B. Zhang, "RhoGDI deficiency induces constitutive activation of Rho GTPases and COX-2 pathways in association with breast cancer progression," *Oncotarget*, 2015.
- [56] S. Hooshmand, A. Ghaderi, K. Yusoff, K. Thilakavathy, R. Rosli, and Z. Mojtahedi, "Differentially expressed proteins in ER⁺ MCF7 and ER⁻ MDA-MB-231 human breast cancer cells by RhoGDI-α silencing and overexpression," Asian Pacific Journal of Cancer Prevention, vol. 15, no. 7, pp. 3311–3317, 2014.
- [57] B. Zhang, "Rho GDP dissociation inhibitors as potential targets for anticancer treatment," *Drug Resistance Updates*, vol. 9, no. 3, pp. 134–141, 2006.
- [58] Z. X. Wang and D. C. Thurmond, "Differential phosphorylation of RhoGDI mediates the distinct cycling of Cdc42 and Rac1 to regulate second-phase insulin secretion," *Journal of Biological Chemistry*, vol. 285, no. 9, pp. 6186–6197, 2010.
- [59] G. Jiao, W. Guo, T. Ren et al., "BMPR2 inhibition induced apoptosis and autophagy via destabilization of XIAP in human chondrosarcoma cells," *Cell Death and Disease*, vol. 5, no. 12, Article ID e1571, 2014.
- [60] R. Hu, Y. Yang, Z. Liu et al., "The XIAP inhibitor Embelin enhances TRAIL-induced apoptosis in human leukemia cells by DR4 and DR5 upregulation," *Tumor Biology*, vol. 36, no. 2, pp. 769–777, 2015.

- [61] Z. Cao, X. Li, J. Li, W. Luo, C. Huang, and J. Chen, "X-linked inhibitor of apoptosis protein (XIAP) lacking RING domain localizes to the nuclear and promotes cancer cell anchorage-independent growth by targeting the E2F1/Cyclin E axis," *Oncotarget*, vol. 5, no. 16, pp. 7126–7137, 2014.
- [62] X. P. Yi, T. Han, Y. X. Li, X. Y. Long, and W. Z. Li, "Simultaneous silencing of XIAP and survivin causes partial mesenchymalepithelial transition of human pancreatic cancer cells via the PTEN/PI3K/Akt pathway," *Molecular Medicine Reports*, vol. 12, pp. 601–608, 2015.
- [63] L.-P. Cao, J.-L. Song, X.-P. Yi, and Y.-X. Li, "Double inhibition of NF-κB and XIAP via RNAi enhances the sensitivity of pancreatic cancer cells to gemcitabine," *Oncology Reports*, vol. 29, no. 4, pp. 1659–1665, 2013.
- [64] Z.-H. Wang, H. Chen, H.-C. Guo et al., "Enhanced antitumor efficacy by the combination of emodin and gemcitabine against human pancreatic cancer cells via downregulation of the expression of XIAP in vitro and in vivo," *International Journal of Oncology*, vol. 39, no. 5, pp. 1123–1131, 2011.
- [65] H. Zai, X. Yi, Y. Li, C. Jiang, and X. Lü, "Simultaneous inhibition of XIAP and survivin expression on EMT and invasion of human pancreatic cancer cells," *Journal of Central South University*, vol. 37, no. 9, pp. 883–888, 2012.
- [66] R. Yu, S. M. Albarenque, R. H. Cool, W. J. Quax, A. Mohr, and R. M. Zwacka, "DR4 specific TRAIL variants are more efficacious than wild-type TRAIL in pancreatic cancer," *Cancer Biology and Therapy*, vol. 15, no. 12, pp. 1658–1666, 2014.
- [67] T. Sun, N. Yu, L.-K. Zhai et al., "C-Jun NH2-terminal kinase (JNK)-interacting protein-3 (JIP3) regulates neuronal axon elongation in a kinesin- and JNK-dependent manner," *Journal* of *Biological Chemistry*, vol. 288, no. 20, pp. 14531–14543, 2013.
- [68] H. Matsuura, H. Nishitoh, K. Takeda et al., "Phosphorylation-dependent scaffolding role of JSAP1/JIP3 in the ASK1-JNK signaling pathway. A new mode of regulation of the MAP kinase cascade," *The Journal of Biological Chemistry*, vol. 277, no. 43, pp. 40703–40709, 2002.
- [69] N. Skrypek, R. Vasseur, A. Vincent, B. Duchêne, I. Van Seuningen, and N. Jonckheere, "The oncogenic receptor ErbB2 modulates gemcitabine and irinotecan/SN-38 chemoresistance of human pancreatic cancer cells via hCNT1 transporter and multidrug-resistance associated protein MRP-2," *Oncotarget*, vol. 6, no. 13, pp. 10853–10867, 2015.
- [70] J. Størling, N. Allaman-Pillet, A. E. Karlsen, N. Billestrup, C. Bonny, and T. Mandrup-Poulsen, "Antitumorigenic effect of proteasome inhibitors on insulinoma cells," *Endocrinology*, vol. 146, no. 4, pp. 1718–1726, 2005.
- [71] C. L. Standen, N. J. Kennedy, R. A. Flavell, and R. J. Davis, "Signal transduction cross talk mediated by Jun N-terminal kinase-interacting protein and insulin receptor substrate scaffold protein complexes," *Molecular and Cellular Biology*, vol. 29, no. 17, pp. 4831–4840, 2009.
- [72] P. P. Ongusaha, H. H. Qi, L. Raj et al., "Identification of ROCK1 as an upstream activator of the JIP-3 to JNK signaling axis in response to UVB damage," *Science Signaling*, vol. 1, no. 47, p. ral4, 2008.
- [73] S. N. Rao and V. N. Balaji, "Molecular docking studies on JNK inhibitors at the allosteric JNK-JIP interaction site," *Current Science*, vol. 102, no. 7, pp. 1009–1016, 2012.
- [74] W. Engström and M. Granerus, "Expression of JNK-interacting protein JIP-1 and insulin-like growth factor II in Wilms tumour cell lines and primary Wilms tumours," *Anticancer Research*, vol. 29, no. 7, pp. 2467–2472, 2009.

- [75] T. Takino, M. Nakada, H. Miyamori et al., "JSAP1/JIP3 cooperates with focal adhesion kinase to regulate c-Jun N-terminal kinase and cell migration," *The Journal of Biological Chemistry*, vol. 280, no. 45, pp. 37772–37781, 2005.
- [76] T. Matsuguchi, A. Masuda, K. Sugimoto, Y. Nagai, and Y. Yoshikai, "JNK-interacting protein 3 associates with Toll-like receptor 4 and is involved in LPS-mediated JNK activation," *The EMBO Journal*, vol. 22, no. 17, pp. 4455–4464, 2003.
- [77] N. Broere, M. Chen, A. Cinar et al., "Defective jejunal and colonic salt absorption and altered Na⁺/H⁺ exchanger 3 (NHE3) activity in NHE regulatory factor 1 (NHERF1) adaptor protein-deficient mice," *Pflügers Archiv—European Journal of Physiology*, vol. 457, no. 5, pp. 1079–1091, 2009.
- [78] A. Bellizzi, M. R. Greco, R. Rubino et al., "The scaffolding protein NHERF1 sensitizes EGFR-dependent tumor growth, motility and invadopodia function to gefitinib treatment in breast cancer cells," *International Journal of Oncology*, vol. 46, no. 3, pp. 1214–1224, 2015.
- [79] B. Hopwood, A. Tsykin, D. M. Findlay, and N. L. Fazzalari, "Microarray gene expression profiling of osteoarthritic bone suggests altered bone remodelling, WNT and transforming growth factor-β/bone morphogenic protein signalling," *Arthri*tis Research and Therapy, vol. 9, no. 5, article R100, 2007.
- [80] Y. Jiang, S. Wang, J. Holcomb et al., "Crystallographic analysis of NHERF1-PLCbeta3 interaction provides structural basis for CXCR2 signaling in pancreatic cancer," *Biochemical and Biophysical Research Communications*, vol. 446, no. 2, pp. 638–643, 2014.
- [81] S. Wang, Y. Wu, Y. Hou et al., "CXCR2 macromolecular complex in pancreatic cancer: a potential therapeutic target in tumor growth," *Translational Oncology*, vol. 6, no. 2, pp. 216–225, 2013.
- [82] A. Mangia, C. Saponaro, A. Malfettone et al., "Involvement of nuclear NHERF1 in colorectal cancer progression," *Oncology Reports*, vol. 28, no. 3, pp. 889–894, 2012.
- [83] Y. Hayashi, J. R. Molina, S. R. Hamilton, and M.-M. Georgescu, "NHERF1/EBP50 is a new marker in colorectal cancer," *Neoplasia*, vol. 12, no. 12, pp. 1013–1022, 2010.
- [84] Y. Y. Jiang, S. O. Wang, J. Holcomb et al., "Crystallographic analysis of NHERF1-PLC β 3 interaction provides structural basis for CXCR2 signaling in pancreatic cancer," *Biochemical and Biophysical Research Communications*, vol. 446, no. 2, pp. 638–643, 2014.
- [85] T. Fukuda, L. Guo, X. Shi, and C. Wu, "CH-ILKBP regulates cell survival by facilitating the membrane translocation of protein kinase B/Akt," *The Journal of Cell Biology*, vol. 160, no. 7, pp. 1001–1008, 2003.
- [86] A. Eslami, K. Miyaguchi, K. Mogushi et al., "PARVB overexpression increases cell migration capability and defines high risk for endophytic growth and metastasis in tongue squamous cell carcinoma," *British Journal of Cancer*, vol. 112, no. 2, pp. 338– 344, 2015.
- [87] J. L. Sepulveda and C. Wu, "The parvins," *Cellular and Molecular Life Sciences*, vol. 63, no. 1, pp. 25–35, 2006.
- [88] G. Rosenberger, I. Jantke, A. Gal, and K. Kutsche, "Interaction of α PIX (ARHGEF6) with β -parvin (PARVB) suggests an involvement of α PIX in integrin-mediated signaling," *Human Molecular Genetics*, vol. 12, no. 2, pp. 155–167, 2003.
- [89] N. A. Nejad, F. Amidi, M. A. Hoseini et al., "Male germ-like cell differentiation potential of human umbilical cord Wharton's jelly-derived mesenchymal stem cells in co-culture with human placenta cells in presence of BMP4 and retinoic acid," *Iranian Journal of Basic Medical Sciences*, vol. 18, no. 4, pp. 325–333, 2015.

- [90] S. S. W. Chung, X. Wang, S. S. Roberts, S. M. Griffey, P. R. Reczek, and D. J. Wolgemuth, "Oral administration of a retinoic Acid receptor antagonist reversibly inhibits spermatogenesis in mice," *Endocrinology*, vol. 152, no. 6, pp. 2492–2502, 2011.
- [91] S. Flajollet, B. Lefebvre, C. Cudejko, B. Staels, and P. Lefebvre, "The core component of the mammalian SWI/SNF complex SMARCD3/BAF60c is a coactivator for the nuclear retinoic acid receptor," *Molecular and Cellular Endocrinology*, vol. 270, no. 1-2, pp. 23–32, 2007.
- [92] Q. H. Wu, R. J. Guo, M. Lin, B. Zhou, and Y. F. Wang, "MicroRNA-200a inhibits CD133/1+ ovarian cancer stem cells migration and invasion by targeting E-cadherin repressor ZEB2," *Gynecologic Oncology*, vol. 122, no. 1, pp. 149–154, 2011.
- [93] F. Yu, J. Li, H. Chen et al., "Kruppel-like factor 4 (KLF4) is required for maintenance of breast cancer stem cells and for cell migration and invasion," *Oncogene*, vol. 30, no. 18, pp. 2161–2172, 2011.
- [94] K. M. Bae, N. N. Parker, Y. Dai, J. Vieweg, and D. W. Siemann, "E-cadherin plasticity in prostate cancer stem cell invasion," *American Journal of Cancer Research*, vol. 1, pp. 71–84, 2011.
- [95] A. Yasuda, H. Sawai, H. Takahashi et al., "The stem cell factor/c-kit receptor pathway enhances proliferation and invasion of pancreatic cancer cells," *Molecular Cancer*, vol. 5, article 46, 2006.
- [96] M. Herreros-Villanueva, T. K. Er, and L. Bujanda, "Retinoic acid reduces stem cell-like features in pancreatic cancer cells," *Pancreas*, vol. 44, no. 6, pp. 918–924, 2015.
- [97] A. Doi, K. Ishikawa, N. Shibata et al., "Enhanced expression of retinoic acid receptor alpha (RARA) induces epithelial-tomesenchymal transition and disruption of mammary acinar structures," *Molecular Oncology*, vol. 9, no. 2, pp. 355–364, 2015.
- [98] H.-J. Yang, Y.-B. Zheng, T. Ji et al., "Overexpression of ILK1 in breast cancer associates with poor prognosis," *Tumor Biology*, vol. 34, no. 6, pp. 3933–3938, 2013.
- [99] C. Leung-Hagesteijn, A. Mahendra, I. Naruszewicz, and G. E. Hannigan, "Modulation of integrin signal transduction by ILKAP, a protein phosphatase 2C associating with the integrin-linked kinase, ILK1," *The EMBO Journal*, vol. 20, no. 9, pp. 2160–2170, 2001.
- [100] J. Sanghera, P. Costello, L. Kojic et al., "Integrin-linked kinase 1 (ILK1): a 'hot' therapeutic target," *Clinical Cancer Research*, vol. 7, pp. 3766s–3767s, 2001.
- [101] D. Bang, W. Wilson, M. Ryan, J. J. Yeh, and A. S. Baldwin, "GSK- 3α promotes oncogenic KRAS function in pancreatic cancer via TAK1-TAB stabilization and regulation of noncanonical NF- κ B," *Cancer Discovery*, vol. 3, no. 6, pp. 690–703, 2013.
- [102] P. Shi, T. Yin, F. Zhou, P. Cui, S. Gou, and C. Wang, "Valproic acid sensitizes pancreatic cancer cells to natural killer cellmediated lysis by upregulating MICA and MICB via the PI3K/Akt signaling pathway," BMC Cancer, vol. 14, article 370, 2014.
- [103] J. Y. C. Chow, M. Ban, H. L. Wu et al., "TGF- β downregulates PTEN via activation of NF- κ B in pancreatic cancer cells," *American Journal of Physiology: Gastrointestinal and Liver Physiology*, vol. 298, no. 2, pp. G275–G282, 2010.
- [104] L.-H. Chang, S.-L. Pan, C.-Y. Lai, A.-C. Tsai, and C.-M. Teng, "Activated PAR-2 regulates pancreatic cancer progression through ILK/HIF-α-induced TGF-α expression and MEK/VEGF-A-mediated angiogenesis," *The American Journal of Pathology*, vol. 183, no. 2, pp. 566–575, 2013.
- [105] A. H. Huang, S. H. Pan, W. H. Chang et al., "PARVA promotes metastasis by modulating ILK signalling pathway in lung

- adenocarcinoma," *PLoS ONE*, vol. 10, no. 3, Article ID e0118530, 2015.
- [106] Z. P. Yan, H. Z. Yin, R. Wang et al., "Overexpression of integrin-linked kinase (ILK) promotes migration and invasion of colorectal cancer cells by inducing epithelial-mesenchymal transition *via* NF-κB signaling," *Acta Histochemica*, vol. 116, no. 3, pp. 527–533, 2014.
- [107] A. Traister, M. Walsh, S. Aafaqi et al., "Mutation in integrinlinked kinase (ILK^{R211A}) and heat-shock protein 70 comprise a broadly cardioprotective complex," *PLoS ONE*, vol. 8, no. 11, Article ID e77331, 2013.
- [108] J. A. Gilbert, L. J. Adhikari, R. V. Lloyd, T. R. Halfdanarson, M. H. Muders, and M. M. Ames, "Molecular markers for novel therapeutic strategies in pancreatic endocrine tumors," *Pancreas*, vol. 42, no. 3, pp. 411–421, 2013.
- [109] F. Bergmann, S. Aulmann, B. Sipos et al., "Acinar cell carcinomas of the pancreas: a molecular analysis in a series of 57 cases," *Virchows Archiv*, vol. 465, no. 6, pp. 661–672, 2014.
- [110] L. Mery, B. Strauss, J. F. Dufour, K. H. Krause, and M. Hoth, "The PDZ-interacting domain of TRPC4 controls its localization and surface expression in HEK293 cells," *Journal of Cell Science*, vol. 115, no. 17, pp. 3497–3508, 2002.
- [111] I. Del Giudice, M. Messina, S. Chiaretti et al., "Behind the scenes of non-nodal MCL: downmodulation of genes involved in actin cytoskeleton organization, cell projection, cell adhesion, tumour invasion, TP53 pathway and mutated status of immunoglobulin heavy chain genes," *British Journal of Haema*tology, vol. 156, no. 5, pp. 601–611, 2012.
- [112] X. L. Li, X. Lu, S. Parvathaneni et al., "Identification of RECQ1-regulated transcriptome uncovers a role of RECQ1 in regulation of cancer cell migration and invasion," *Cell Cycle*, vol. 13, no. 15, pp. 2431–2445, 2014.
- [113] M. D. Pastor, A. Nogal, S. Molina-Pinelo et al., "Identification of proteomic signatures associated with lung cancer and COPD," *Journal of Proteomics*, vol. 89, pp. 227–237, 2013.
- [114] J. D. Figueroa, S. S. Han, M. Garcia-Closas et al., "Genomewide interaction study of smoking and bladder cancer risk," *Carcinogenesis*, vol. 35, pp. 1737–1744, 2014.
- [115] J. H. Zhou, Y. J. Feng, K. T. Tao et al., "The expression and phosphorylation of ezrin and merlin in human pancreatic cancer," *International Journal of Oncology*, vol. 45, no. 6, pp. 2059–2067, 2014.
- [116] Y. X. Meng, Z. H. Lu, S. N. Yu, Q. A. Zhang, Y. H. Ma, and J. Chen, "Ezrin promotes invasion and metastasis of pancreatic cancer cells," *Journal of Translational Medicine*, vol. 8, article 61, 2010.
- [117] H. M. Kocher, J. Sandle, T. A. Mirza, N. F. Li, and I. R. Hart, "Ezrin interacts with cortactin to form podosomal rosettes in pancreatic cancer cells," *Gut*, vol. 58, no. 2, pp. 271–284, 2009.
- [118] Y. Oda, S. Aishima, K. Morimatsu et al., "Differential ezrin and phosphorylated ezrin expression profiles between pancreatic intraepithelial neoplasia, intraductal papillary mucinous neoplasm, and invasive ductal carcinoma of the pancreas," *Human Pathology*, vol. 44, no. 8, pp. 1487–1498, 2013.
- [119] J. V. Thevathasan, E. Tan, H. Zheng et al., "The small GTPase HRas shapes local PI3K signals through positive feedback and regulates persistent membrane extension in migrating fibroblasts," *Molecular Biology of the Cell*, vol. 24, no. 14, pp. 2228–2237, 2013.
- [120] H. Fensterer, K. Giehl, M. Buchholz et al., "Expression profiling of the influence of RAS mutants on the TGFB1-induced

- phenotype of the pancreatic cancer cell line PANC-1," *Genes Chromosomes & Cancer*, vol. 39, no. 3, pp. 224–235, 2004.
- [121] F. Schönleben, W. Qiu, J. D. Allendorf, J. A. Chabot, H. E. Remotti, and G. H. Su, "Molecular analysis of *PIK3CA*, *BRAF*, and *RAS* oncogenes in periampullary and ampullary adenomas and carcinomas," *Journal of Gastrointestinal Surgery*, vol. 13, no. 8, pp. 1510–1516, 2009.
- [122] J. Sayyah, A. Bartakova, N. Nogal, L. A. Quilliam, D. G. Stupack, and J. H. Brown, "The Ras-related protein, Rap1A, mediates thrombin-stimulated, integrin-dependent glioblastoma cell proliferation and tumor growth," *The Journal of Biological Chemistry*, vol. 289, no. 25, pp. 17689–17698, 2014.
- [123] J.-S. Park, K.-M. Lim, S. G. Park et al., "Pancreatic cancer induced by in vivo electroporation-enhanced sleeping beauty transposon gene delivery system in mouse," *Pancreas*, vol. 43, no. 4, pp. 614–618, 2014.
- [124] T. Shichinohe, N. Senmaru, K. Furuuchi et al., "Suppression of pancreatic cancer by the dominant negative ras mutant, N116Y," *The Journal of Surgical Research*, vol. 66, no. 2, pp. 125–130, 1996.
- [125] V. Sirivatanauksorn, Y. Sirivatanauksorn, and N. R. Lemoine, "Molecular pattern of ductal pancreatic cancer," *Langenbeck's Archives of Surgery*, vol. 383, no. 2, pp. 105–115, 1998.
- [126] A. Nagy, L. Kozma, I. Kiss et al., "Copy number of cancer genes predict tumor grade and survival of pancreatic cancer patients," *Anticancer Research*, vol. 21, no. 2, pp. 1321–1326, 2001.
- [127] T. Seufferlein, J. Van Lint, S. Liptay, G. Adler, and R. M. Schmid, "Transforming growth factor α activates Ha-Ras in human pancreatic cancer cells with Ki-ras mutations," *Gastroenterology*, vol. 116, no. 6, pp. 1441–1452, 1999.
- [128] T. J. Haggerty, I. S. Dunn, L. B. Rose, E. E. Newton, F. Pandolfi, and J. T. Kurnick, "Heat shock protein-90 inhibitors enhance antigen expression on melanomas and increase T cell recognition of tumor cells," *PLoS ONE*, vol. 9, no. 12, Article ID e114506, 2014.
- [129] Y. J. Zhou and R. J. Binder, "The heat shock protein-CD91 pathway mediates tumor immunosurveillance," *OncoImmunology*, vol. 3, no. 4, Article ID e28222, 2014.
- [130] K. Dodd, S. Nance, M. Quezada et al., "Tumor-derived inducible heat-shock protein 70 (HSP70) is an essential component of anti-tumor immunity," *Oncogene*, vol. 34, pp. 1312–1322, 2015
- [131] N. Qiao, Y. Zhu, H. Li, Z. Qu, and Z. Xiao, "Expression of heat shock protein 20 inversely correlated with tumor progression in patients with ovarian cancer," *European Journal of Gynaecological Oncology*, vol. 35, no. 5, pp. 576–579, 2014.
- [132] U. Ramp, C. Mahotka, S. Heikaus et al., "Expression of heat shock protein 70 in renal cell carcinoma and its relation to tumor progression and prognosis," *Histology and Histopathol*ogy, vol. 22, no. 10–12, pp. 1099–1107, 2007.
- [133] J. J. Hyun, H. S. Lee, B. Keum et al., "Expression of heat shock protein 70 modulates the chemoresponsiveness of pancreatic cancer," *Gut and Liver*, vol. 7, no. 6, pp. 739–746, 2013.
- [134] Y. Kuramitsu, Y. Wang, K. Taba et al., "Heat-shock protein 27 plays the key role in gemcitabine-resistance of pancreatic cancer cells," *Anticancer Research*, vol. 32, no. 6, pp. 2295–2299, 2012.
- [135] Z. Zhang, K. Kawamura, Y. Jiang et al., "Heat-shock protein 90 inhibitors synergistically enhance melanoma differentiation-associated gene-7-mediated cell killing of human pancreatic carcinoma," *Cancer Gene Therapy*, vol. 20, no. 12, pp. 663–670, 2013.

- [136] T. N. MacKenzie, N. Mujumdar, S. Banerjee et al., "Triptolide induces the expression of miR-142-3p: a negative regulator of heat shock protein 70 and pancreatic cancer cell proliferation," *Molecular Cancer Therapeutics*, vol. 12, no. 7, pp. 1266–1275, 2013.
- [137] Y. K. Yu, A. Hamza, T. Zhang et al., "Withaferin A targets heat shock protein 90 in pancreatic cancer cells," *Biochemical Pharmacology*, vol. 79, no. 4, pp. 542–551, 2010.
- [138] S. Mori-Iwamoto, K. Taba, Y. Kuramitsu et al., "Interferon-y down-regulates heat shock protein 27 of pancreatic cancer cells and helps in the cytotoxic effect of gemcitabine," *Pancreas*, vol. 38, no. 2, pp. 224–226, 2009.
- [139] S. A. Lang, C. Moser, A. Gaumann et al., "Targeting heat shock protein 90 in pancreatic cancer impairs insulin-like growth factor-I receptor signaling, disrupts an interleukin-6/signal-transducer and activator of transcription 3/hypoxia-inducible factor-Iα autocrine loop, and reduces orthotopic tumor growth," *Clinical Cancer Research*, vol. 13, no. 21, pp. 6459–6468, 2007.
- [140] V. Dudeja, P. Phillips, R. Sharif, Y. Zhan, R. Dawra, and A. Saluja, "Heat shock protein 70 (HSP70) confers resistance to apoptosts in pancreatic cancer cells by lysosomal stabilization," *Pancreas*, vol. 33, no. 4, p. 457, 2006.
- [141] W. Fendler, I. Klich, A. Cieślik-Heinrich, K. Wyka, A. Szadkowska, and W. Młynarski, "Increased risk of type 1 diabetes in Polish children—association with INS-IGF2 5'VNTR and lack of association with HLA haplotype," *Endokrynologia Polska*, vol. 62, no. 5, pp. 436–442, 2011.
- [142] G. Sutherland, G. Mellick, J. Newman et al., "Haplotype analysis of the IGF2-INS-TH gene cluster in Parkinson's disease," American Journal of Medical Genetics. Part B: Neuropsychiatric Genetics, vol. 147, no. 4, pp. 495–499, 2008.
- [143] C. Polychronakos, A. Kukuvitis, N. Giannoukakis, and E. Colle, "Parental imprinting effect at the IPIS-IGF2 diabetes susceptibility locus," *Diabetologia*, vol. 38, no. 6, pp. 715–719, 1995.
- [144] G. Y. F. Ho, A. Melman, S.-M. Liu et al., "Polymorphism of the insulin gene is associated with increased prostate cancer risk," *British Journal of Cancer*, vol. 88, no. 2, pp. 263–269, 2003.
- [145] I. A. Eaves, S. T. Bennett, P. Forster et al., "Transmission ratio distortion at the INS-IGF2 VNTR," *Nature Genetics*, vol. 22, no. 4, pp. 324–325, 1999.
- [146] C. A. Haiman, Y. Han, Y. Feng et al., "Genome-wide testing of putative functional exonic variants in rlationship with breast and prostate cancer risk in a multiethnic population," *PLoS Genetics*, vol. 9, no. 3, Article ID e1003419, 2013.
- [147] N. Kanatsuna, A. Delli, C. Andersson et al., "Doubly reactive INS-IGF2 autoantibodies in children with newly diagnosed autoimmune (type 1) diabetes," *Scandinavian Journal of Immunology*, vol. 82, pp. 361–369, 2015.
- [148] N. Kanatsuna, J. Taneera, F. Vaziri-Sani et al., "Autoimmunity against INS-IGF2 protein expressed in human pancreatic islets," *Journal of Biological Chemistry*, vol. 288, no. 40, pp. 29013– 29023, 2013.
- [149] G. Paroni, M. Fratelli, G. Gardini et al., "Synergistic antitumor activity of lapatinib and retinoids on a novel subtype of breast cancer with coamplification of ERBB2 and RARA," *Oncogene*, vol. 31, no. 29, pp. 3431–3443, 2012.
- [150] A. Saumet, G. Vetter, M. Bouttier et al., "Transcriptional repression of microRNA genes by PML-RARA increases expression of key cancer proteins in acute promyelocytic leukemia," *Blood*, vol. 113, no. 2, pp. 412–421, 2009.

- [151] T. A. Bailey, H. Luan, E. Tom et al., "A kinase inhibitor screen reveals protein kinase C-dependent endocytic recycling of ErbB2 in breast cancer cells," *The Journal of Biological Chemistry*, vol. 289, no. 44, pp. 30443–30458, 2014.
- [152] H. Qi, B. Sun, X. Zhao et al., "Wnt5a promotes vasculogenic mimicry and epithelial-mesenchymal transition via protein kinase Cα in epithelial ovarian cancer," *Oncology Reports*, vol. 32, no. 2, pp. 771–779, 2014.
- [153] A. Doller, C. Winkler, I. Azrilian et al., "High-constitutive HuR phosphorylation at Ser 318 by PKC delta propagates tumor relevant functions in colon carcinoma cells," *Carcinogenesis*, vol. 32, no. 5, pp. 676–685, 2011.
- [154] A. M. Butler, M. L. S. Scotti Buzhardt, S. Li, K. E. Smith, A. P. Fields, and N. R. Murray, "Protein kinase C Zeta regulates human pancreatic cancer cell transformed growth and invasion through a STAT3-dependent mechanism," *PLoS ONE*, vol. 8, no. 8, Article ID e72061, 2013.
- [155] Y. Chen, G. Z. Yu, D. H. Yu, and M. H. Zhu, "PKCα-induced drug resistance in pancreatic cancer cells is associated with transforming growth factor-β1," *Journal of Experimental & Clinical Cancer Research*, vol. 29, no. 1, article 104, 2010.
- [156] C. E. Antal, A. M. Hudson, E. Kang et al., "Cancer-associated protein kinase C mutations reveal kinase's role as tumor suppressor," *Cell*, vol. 160, no. 3, pp. 489–502, 2015.
- [157] M. Linch, M. Sanz-Garcia, E. Soriano et al., "A cancer-associated mutation in atypical protein kinase C_i occurs in a substratespecific recruitment motif," Science Signaling, vol. 6, no. 293, article ra82, 2013.
- [158] P. Storz, "Targeting protein kinase C subtypes in pancreatic cancer," *Expert Review of Anticancer Therapy*, vol. 15, no. 4, pp. 433–438, 2015.