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## Role of intestinal microbiota in transplantation outcomes

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### Abstract

While allogeneic hematopoietic stem cell transplantations have a curative potential, infections and graft-versus-host disease remain significant problems. The intestinal microbiota can influence responses to cancer chemotherapy and the role of the microbiota in affecting allogeneic hematopoietic stem cell transplantation outcomes is increasingly appreciated. The following paper discusses the most recent developments in this area.

### Keywords

Allo-HSCT; bacteremia; diversity; intestinal microbiota; microbial dominance; metronidazole; mortality; survival

### Introduction

The intestinal microbiota comprises microbial populations that colonize the human gastrointestinal tract and has been shown to play a crucial role in human health by mediating resistance to infection [1–3]. Studies of the microbiome of humans have revealed that

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#### Conflict of interest

Eric G. Pamer: No relevant financial relationships with any commercial interest.

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healthy individuals harbor diverse microbial populations in the gut. The majority of bacterial taxa belong to the *Firmicutes* and *Bacteroidetes* phyla and bacteria belonging to the *Actinobacteria*, *Proteobacteria*, *Verrucomicrobia*, and *Fusobacteria* are also represented [1]. The composition of the intestinal microbiota differs from person to person, which can have implications in health and disease [4]. For example, the composition of the intestinal microbiota has been associated with malnutrition, obesity, and predisposition to rheumatoid arthritis [5–7]. Recently, two studies have indicated that the intestinal microbiota can influence the immune response to systemic cancer chemotherapy and disruption of the intestinal microbiome was associated with resistance to cancer therapy [8,9]. It has become clear that the intestinal microbiota not only mediates resistance to colonization by pathogens but also modulates immune function by promoting differentiation of different T-cell phenotypes. Given that infections and graft-versus-host disease are significant problems associated with allogeneic hematopoietic stem cell transplantation (allo-HSCT), the study of intestinal microbiota may provide clues to improve outcomes among allo-HSCT patients [10]. The following discussion provides a brief summary on the impact of intestinal microbiota in allo-HSCT outcomes.

### Intestinal microbiota and bacteremia in allo-HSCT

Although allo-HSCT represents a potentially curative treatment option for some patients with hematologic malignancies, infection and bacteremia are frequent complications [10]. A study of the fecal samples from five patients undergoing allo-HSCT showed that the composition of the intestinal microbiota can change dramatically following allo-HSCT (Figure 1) [11]. While all patients presented with a diverse intestinal microbiota before allo-HSCT, three of the patients showed dramatic fluctuations in the composition of the microbiota after allo-HSCT. In two of these patients, vancomycin-resistant *Enterococcus* (VRE) was shown to dominate the gastrointestinal tract before the onset of VRE bacteremia. However, the exact reason for this intestinal domination by VRE was not clear in this study.

To identify correlative factors that explain the loss of diversity of the intestinal microbiota and the intestinal dominance by certain microorganisms following allo-HSCT, a larger longitudinal study with 94 allo-HSCT patients was performed [12]. Fecal samples were collected from these 94 patients pre-allo-HSCT and for up to 35 days post-allo-HSCT. These samples were characterized for intestinal microbiota by 454 pyrosequencing of the V1-V3 region of bacterial 16S ribosomal RNA genes. Intestinal domination was defined as the endpoint in which a particular bacterial taxon attains 30% or greater relative abundance and is more abundant than any other population member within a single fecal specimen. As observed in Figure 2, the risk of intestinal domination with *Proteobacteria*, *Enterococcus*, and *Streptococcus* increased over time following allo-HSCT [12]. Notably, *Enterococcus* domination was associated with administration of metronidazole, which strongly inhibits obligate anaerobes. Similarly, *Streptococcus* domination was associated with beta-lactam antibiotics such as cephalosporins, beta-lactam/beta-lactamase combinations, and carbapenems. On the other hand, administration of fluoroquinolones such as ciprofloxacin and levofloxacin reduced the risk of domination with *Proteobacteria* (Table 1) [12]. Consequently, while domination with *Enterococcus* increased the risk of developing VRE

bacteremia, domination with *Proteobacteria* resulted in an increased likelihood of developing gram-negative bacteremia (Table 2) [12].

## Diversity of intestinal microbiota in allo-HSCT

Another important observation made in this study was the decrease in the diversity of the intestinal microbiota of the patients undergoing allo-HSCT. As shown in Figure 3, most of the patients demonstrate considerable diversity in the intestinal microbiota before undergoing allo-HSCT. However, as the patients underwent allo-HSCT and following administration of conditioning regimens, there was a decrease in the diversity of the intestinal microbiota. Certain patient samples continued to maintain diversity and hence the next step was to determine whether there was a correlation between the loss of diversity and some specific clinical parameters [12]. As suggested by Figure 4, diversity loss correlated with the administration of antibiotics during allo-HSCT [12]. High intestinal microbiota diversity was observed among patients receiving fewer antibiotics such as fluoroquinolone and vancomycin, which lack broad anaerobic coverage. Intermediate intestinal microbiota diversity loss was associated with the addition of some beta-lactam antibiotics and the greatest loss of intestinal microbiota diversity was correlated with antibiotics that specifically target anaerobic microorganisms (Figure 4) [12]. Classification of allo-HSCT patients into low, medium, and high microbiota diversity groups at the time of stem cell engraftment demonstrated that diversity correlates with survival (Figure 5) [13]. Univariate and multivariate analysis further corroborated that low diversity of the intestinal microbiota was an independent risk factor for mortality. Specific examination of transplant-related death confirmed that low diversity of the intestinal microbiota was associated with increased mortality (52%) as opposed to 8% probability of transplant-related death among patients with high intestinal microbiota diversity at the time of stem cell engraftment (Figure 6) [13].

## Summary

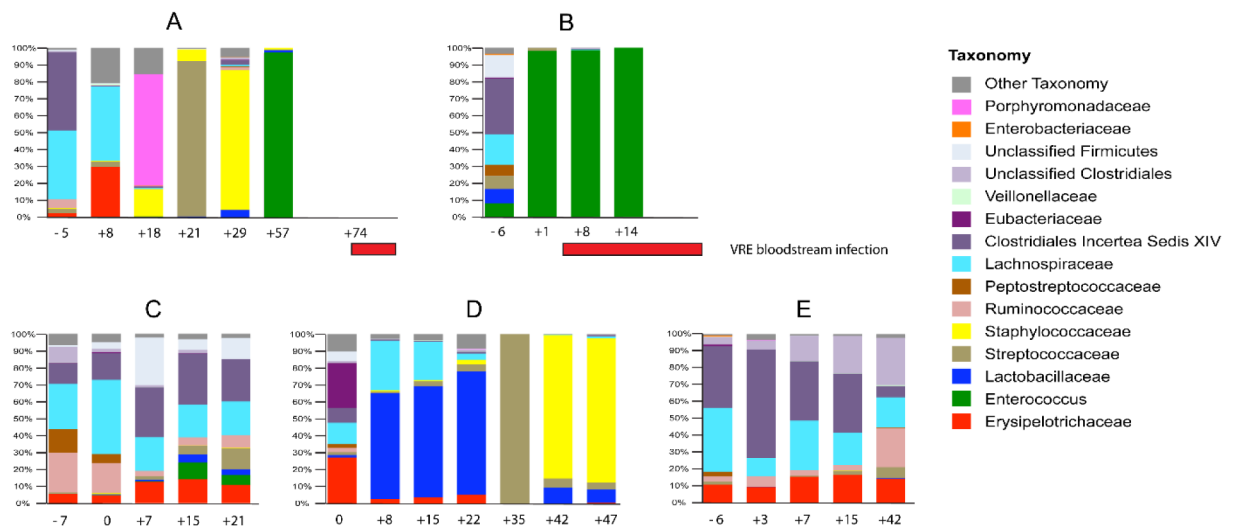
The intestinal microbiota stimulates immune development and defense against microbial pathogens. The diversity of the intestinal microbiota at the time of stem cell engraftment improves allo-HSCT outcomes. Decreased diversity and intestinal domination by *Enterococci* correlates with the spectrum of antibiotics administered to patients and leads to reduced survival. Ongoing studies are investigating whether reconstitution of intestinal microbiota using fecal microbial transplantation following allo-HSCT may provide an approach to optimize clinical outcomes and survival (clinicaltrials.gov identifier NCT02269150).

## Abbreviations

<b>allo-HSCT</b>	allogeneic HSCT
<b>HSCT</b>	hematopoietic stem cell transplantation
<b>VRE</b>	vancomycin-resistant <i>Enterococcus</i>

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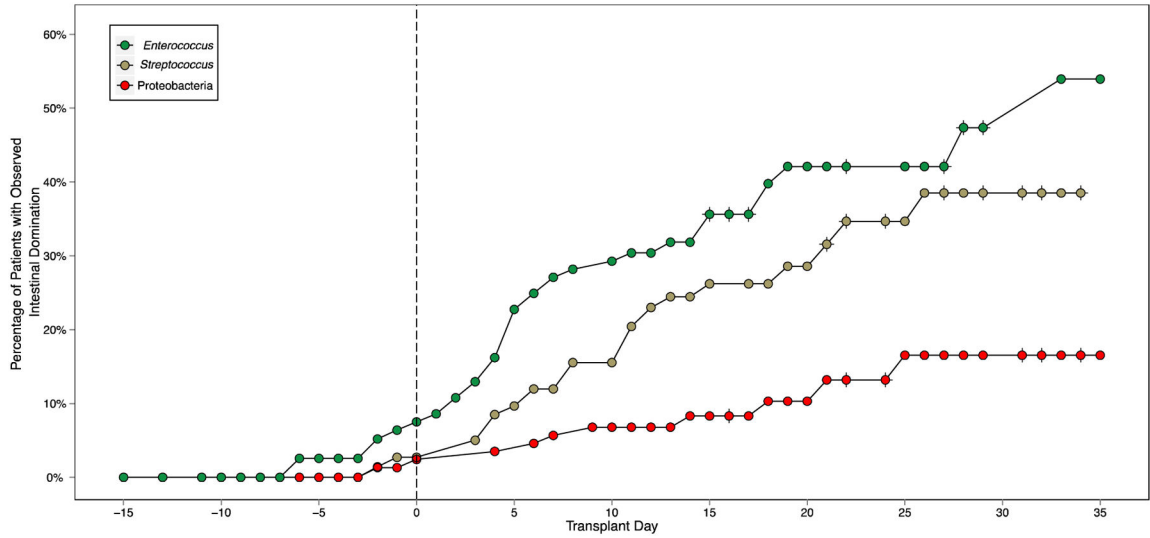
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**Figure 1.**

VRE dominates the intestinal microbiota in humans prior to invading the bloodstream [11]. Stool samples from 5 patients (A–E) were studied prior to and during the transplant period. Each bar on the graph depicts the microbiota of 1 stool sample. Days the samples were collected relative to the day of transplant are indicated along the x-axis. The red horizontal bars indicate the timing of vancomycin-resistant *Enterococcus* bloodstream infections in patients A and B.

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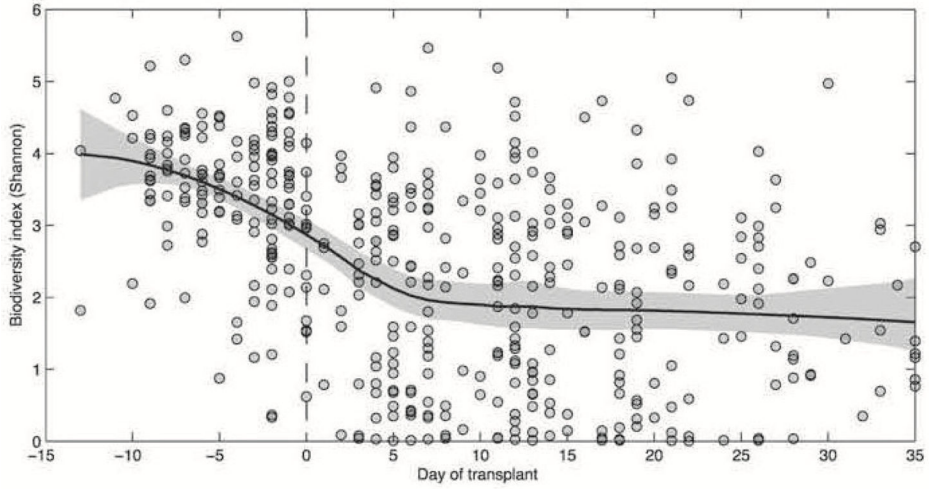


**Figure 2.**

Risk of intestinal domination of the microbiota increases over time following allo-HSCT [12].

This Kaplan-Meier plot shows intestinal domination by *Enterococcus* (top), *Streptococcus* (middle), and Proteobacteria (bottom) at various times throughout the transplant observation period.

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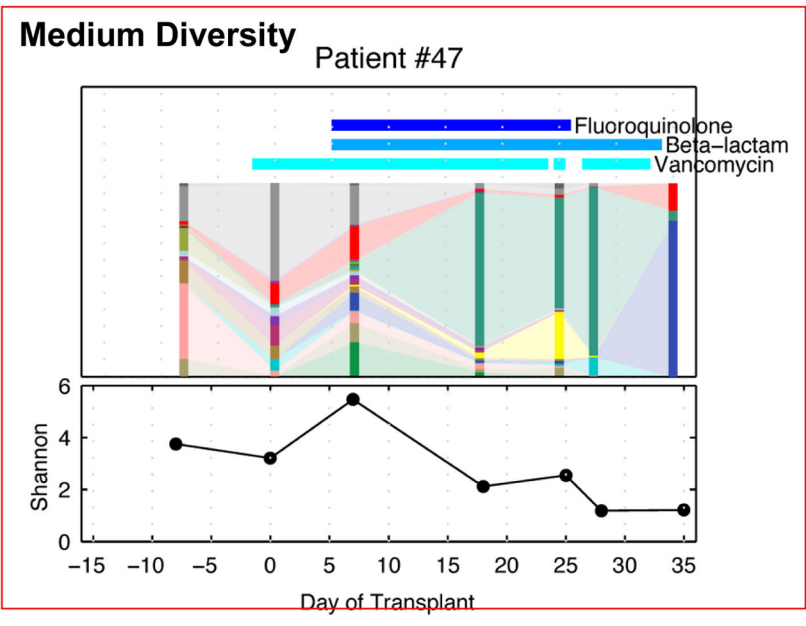
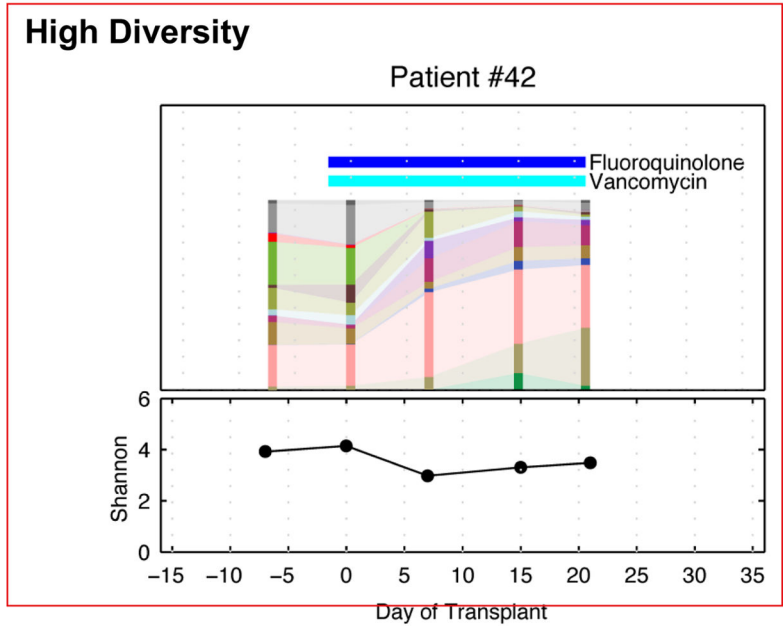


**Figure 3.**

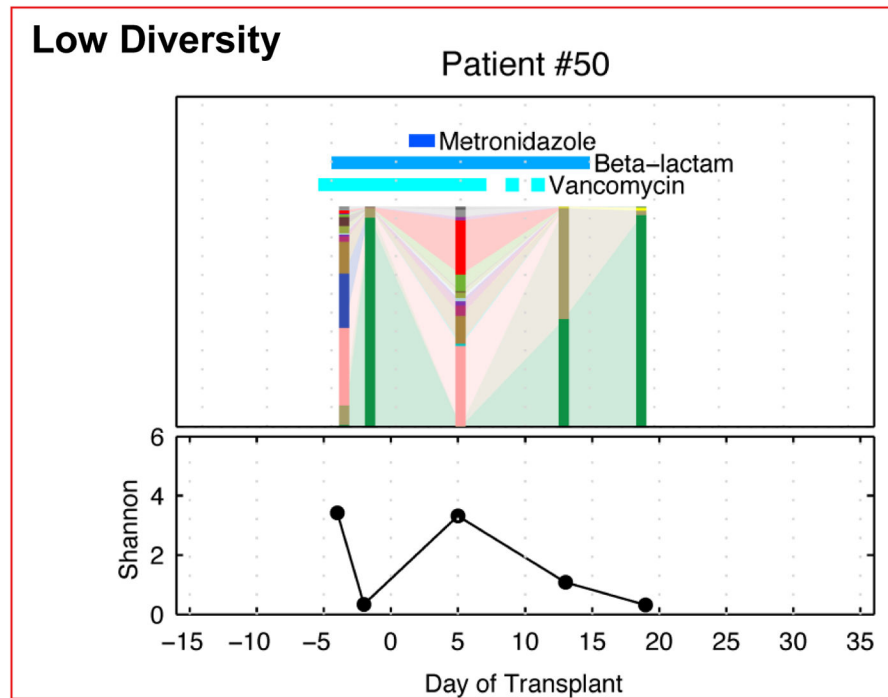
Intestinal microbiota diversity decreases following allo-HSCT [12].

A microbiota diversity trend, solid black line, was calculated for each fecal specimen of each patient during the transplant period using the Shannon diversity index. The gray area indicates the 95% confidence interval. Intestinal microbiota diversity decreased after allo-HSCT (Day 0).

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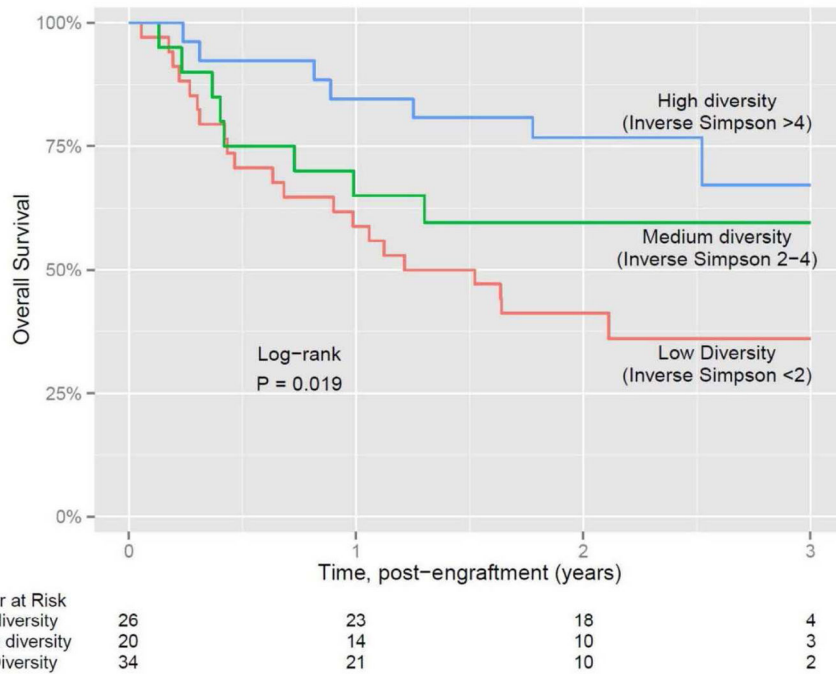


**Figure 4.**

Loss of microbiota diversity following allo-HSCT is variable [12].

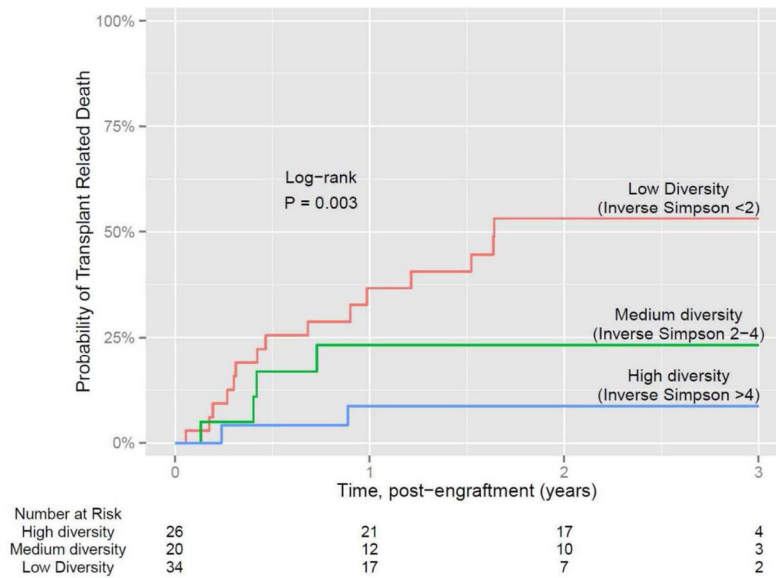
The microbiota diversity of 3 patients following allo-HSCT is depicted here. The vertical bars represent the microbial composition of a single fecal sample, and the horizontal bars indicate antibiotics administered concurrently. As can be seen, the diversity is variable from patient to patient.

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**Figure 5.**

After transplant engraftment: Intestinal microbial diversity correlates with survival [13]. This research was originally published in *Blood*. Taur Y, et al. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. *Blood* 2014;124:1174–1182. © the American Society of Hematology.



**Figure 6.**

Transplant-related mortality is markedly reduced in patients with a diverse microbiota following engraftment [13].

This research was originally published in *Blood*. Taur Y, et al. The effects of intestinal tract bacterial diversity on mortality following allogeneic hematopoietic stem cell transplantation. *Blood* 2014;124:1174–1182. © the American Society of Hematology.

Antibiotic risk factors for bacterial domination [12].  
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**Table 1**

Clinical Predictor	Enterococcus Domination		Streptococcus Domination		Proteobacteria Domination	
	Hazard ratio (95% CI)	P-value	Hazard ratio (95% CI)	P-value	Hazard ratio (95% CI)	P-value
Vancomycin	2.10 (0.67 – 10.14)	0.228	0.95 (0.33 – 3.75)	0.931	5.08 (0.52 – 693.3)	0.196
Metronidazole	<b>3.40 (1.66 – 6.75)</b>	<b>0.001</b>	1.94 (0.81 – 4.31)	0.130	1.73 (0.41 – 6.04)	0.425
Fluoroquinolones	1.09 (0.49 – 2.25)	0.824	1.19 (0.52 – 2.61)	0.673	<b>0.09 (0 – 0.75)</b>	<b>0.020</b>
Beta-lactam	1.19 (0.47 – 3.45)	0.724	3.56 (0.83 – 33.20)	0.094	0.64 (0.15 – 3.27)	0.574

**Table 2**

Domination as risk factors for subsequent bacteremia [12].

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Domination by	VRE bacteremia		Gram negative bacteremia	
	Haz Ratio (95% CI)	P-value	Haz Ratio (95% CI)	P-value
<i>Enterococcus</i>	9.47 (2.46 – 46.0)	0.001	1.53 (0.28 – 5.97)	0.583
<i>Streptococcus</i>	0.22 (0.00 – 1.77)	0.188	0.92 (0.10 – 4.17)	0.925
Proteobacteria	0.76 (0.01 – 6.20)	0.842	6.20 (1.15 – 23.37)	0.036