LIVER CANCER •

Influence of hepatic arterial blockage on blood perfusion and VEGF, MMP-1 expression of implanted liver cancer in rats

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Abstract

AIM: To investigate the influence of hepatic arterial blockage on blood perfusion of transplanted cancer in rat liver and the expression of vascular endothelial growth factor (VEGF) and matrix metalloproteinase-1 (MMP-1), and to explore the mechanisms involved in transarterial embolization (TAE)induced metastasis of liver cancer preliminarily.

METHODS: Walker 256 carcinosarcoma was transplanted into rat liver to establish the liver cancer model. Hepatic arterial ligation (HAL) was used to block the hepatic arterial blood supply and simulate TAE. Blood perfusion of tumor in control, laparotomy control, and HAL group was analyzed by Hoechst 33342 labeling assay, the serum VEGF level was assayed by ELISA, the expression of VEGF and MMP-1 mRNA was detected by in situ hybridization.

RESULTS: Two days after HAL, the number of Hoechst 33342 labeled cells which represent the blood perfusion of tumor directly and hypoxia of tumor indirectly in HAL group decreased significantly compared with that in control group (329±29 ν s 384±19, P<0.01). The serum VEGF level in the HAL group increased significantly as against that of the control group (93ng·L⁻¹±44ng·L⁻¹ ν s 55ng·L⁻¹±19ng·L⁻¹, P<0.05). The expression of VEGF and MMP-1 mRNA in the tumor tissue of the HAL group increased significantly compared with that of the control and the laparotomy control groups (P<0.05). The blood perfusion data of the tumor, represented by the number of Hoechst 33342 labeled cells, showed a good linear inverse correlation with the serum VEGF level (r=-0.606, P<0.05) and the expression of VEGF mRNA in the tumor tissue (r=-0.338, P<0.01).

CONCLUSION: Blockage of hepatic arterial blood supply results in decreased blood perfusion and increased expression of metastasis-associated genes VEGF and MMP-1 of transplanted liver cancer in rats. Decreased blood perfusion and hypoxia may be the major cause of up-regulated expression of VEGF.

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INTRODUCTION

Primary liver cancer (PLC) is one of the most common cancers in Asia. Patients with PLC usually have very poor prognoses, and the overall 5year survival rate was not more than 5% worldwide^[1-3]. Surgery is considered as the only potential cure. However, the resection rate for PLC is only 10%-30%. Transarterial embolization (TAE) is one of the main non-surgical treatments for liver cancer in Asia via selectively blocking of the hepatic arterial blood supply of the tumor. However, a prospective controlled study showed that TAE enhanced the rate of lung metastases and accelerated metastases in patients with PLC^[4]. Other reports also showed that preoperative TAE for resectable PLC or TAE after curative resection of PLC could result in introhepatic recurrence or extrahepatic metastasis, and a shorter survival^[5,6]. Metastases induced by TAE will undoubtedly reduce the long-term efficacy of TAE for PLC, but the mechanisms responsible for that have not been previously reported and there is no good method to inhibit metastasis.

In the present study, we have observed the influence of hepatic arterial ligation (HAL) which simulate TAE on metastasis related genes expression in Walker 256 tumor transplanted in rat liver to explore the mechanisms involved in TAE-induced metastasis of liver cancer preliminarily.

MATERIALS AND METHODS

Tumor model and treatment schedule

An implanted liver cancer model was obtained from Shanghai Medical Industry Research Institute, China. Male pathogen-free Wistar rats, weighing 100-120g, were anesthetized with pentobarbital sodium (40mg/kg). Following midline laparotomy, Walker 256 carcinosarcoma (about 1mm³) was implanted into one hepatic lobe of the rat. Seven days later, the rats that developed tumors were divided randomly into 3 groups: a control group without any treatment (*n*=8), a laparotomy control group (*n*=4) which underwent laparotomy only, and a HAL group (*n*=8). In the HAL group, after anesthesia and laparotomy, the proper hepatic artery was ligated by a sewing line.

Staining of perfused vessels with Hoechst 33342

The perfusion marker Hoechst 33342 (Sigma, USA) was dissolved in sterile saline immediately before use. Two days after HAL, all rats from all three groups were anesthetized and then their abdominal cavity was opened. The inferior vena cava was punctured to collect 0.5mL blood and to inject 0.2mL Hoechst 33342 (at a concerntration of $6.25g\cdot L^{-1}$). The hepatic lobes bearing tumor were excised 1min after injection of Hoechst 33342 to prevent diffusion of Hoechst 33342 into adjacent non-perfused vascular structures. Hoechst 33342 is removed very rapidly from the circulation (half-time of 2min) and is very stable, once bound to DNA. Thus, Hoechst 33342 specifically labels the nuclei of endothelial cells and nuclei of the cells adjacent to the vessel walls, thereby delinealing the perfused vessels^[7-9]. The tumors were excised and frozen in liquid nitrogen and then stored at -80°C until tumors were sectioned. Frozen sections (5µm) were cut and Hoechst 33342 labeled cells were visualized and photographed

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under a fluorescence microscope equipped with a camera with 365nm excitation and 420nm emission filters showing blue fluorescence. Four high power fields under the fluorescence microscope were photographed and Hoechst 33342 labeled cells were counted. The mean number of labeled cells reflected the blood perfusion of tumor and has been shown to be inversely related to the level of hypoxia in tumor^[7].

In situ hybridization (ISH) detection

ISH kits (Boster, China) were used to detect vascular endothelial growth factor (VEGF) and matrix metalloprotenase-1 (MMP-1) mRNA expression. Experiments were performed following the manufacturer's instructions. Briefly, frozen sections (5μ m) were taken from the tumor and fixed with 40g-L⁻¹ polyformaldehyte in PBS (0.1mol-L^{-1}) for 30min, then digested in pepsin for 3min at 37 °C. The material was prehybridized for 2h and then hybridized with Digoxin labeled probe overnight at 40 °C. After hybridization, sections were washed with 2×SSC, 0.2×SSC, and then incubated with rabbit anti-Digoxin antibody, biotinylated secondary antibody IgG, SABC. Slides were stained with DAB and studied under a light microscope equipped with a camera. A positive stain should be present as intracytoplasmic. The level of mRNA expression was evaluated by two of the authors on blinded sections and was expressed as a positive staining rate.

Serum VEGF assay with ELISA

Blood samples collected from the inferior vena cava of rats were pooled, coagulated, and centrifuged at $1000 \times g$ for 10min. After the serum was aspirated, the VEGF level was assayed with an ELISA kit (MEGA, USA). Experiments were performed following the manufacturer's instructions.

Statistical analysis

The number of Hoechst 33342 labeled cells, the percentage of positive staining cells of ISH detection, and the serum VEGF level were expressed as mean \pm SD. The difference between mean values were compared by Student's *t* test. A *P* value less than 0.05 was considered to be statistically significant. Linear regression was used for the correlation assessment. Statistical analyses were performed using SPSS software.

RESULTS

Blood perfusion of tumor

The mean number of Hoechst 33342 labeled cells represents the blood perfusion of tumor, and indirectly represents the level of hypoxia in tumor. The lower number of labeled cells suggests an impaired blood perfusion and a higher level of hypoxia in tumor. The mean number of Hoechst 33342 labeled cells in the control, the laparotomy control, and the HAL groups were 384 ± 19 , 369 ± 27 , and 329 ± 29 , respectively. The number of labeled cells in the HAL group decreased significantly compared with that in the control and the laparotomy control groups (*P*<0.05, Table 1 and Figure 1). This suggests a decreased blood perfusion and a more serious hypoxia of tumor after HAL.

VEGF and MMP-1 mRNA expression

ISH showed both VEGF and MMP-1 mRNA expressed in the implanted tumor of control and laparotomy control groups. There was no significant difference between these two groups (P>0.05). The expression of VEGF and MMP-1 in the HAL group was elevated significantly compared with values in the control and the laparotomy control groups (P<0.05). Results of their expression in the different groups are shown in Table 1, and Figures 2 and 3.



Figure 1 Hoechst 33342 labeled cells. A: Control animals; B: HAL group, decreased number of labeled cells means decreased vascular supply. **Figure 2** VEGF mRNA in hybridization. A: Low expression in tumor of control animals; B: HAL group, a higher VEGF expression. **Figure 3** MMP-1 mRNA in hybridization. A: Low expression in tumor of control animals; B: HAL group, a higher MMP-1 expression. World J Gastroenterol

Serum VEGF level

The serum VEGF level was $55 \text{ng} \cdot \text{L}^{-1} \pm 19 \text{ng} \cdot \text{L}^{-1}$ in the control group, and $70 \text{ng} \cdot \text{L}^{-1} \pm 40 \text{ng} \cdot \text{L}^{-1}$ in the laparotomy control group, which was elevated slightly, but not significantly, as compared to the control group. While in the HAL group, the level of serum VEGF was $93 \text{ng} \cdot \text{L}^{-1} \pm 44 \text{ng} \cdot \text{L}^{-1}$, which was elevated significantly when compared with the control group (*P*<0.05, Table 1).

Table 1 Number of Hoechst labeled cells, serum VEGF, and VEGF, MMP-1 mRNA expression in tumor $(\bar{x}\pm s)$

Groups	n	Hoechst 33342 labeled cells	Serum VEGF (ng·L ⁻¹)	VEGF mRNA	MMP-1 mRNA
Control	8	384±19	55±19	0.34±0.14	0.24±0.17
Laparotomy control	4	369±27	70±40	0.19±0.17	0.30 ± 0.02
HAL	8	$329{\pm}29^{\rm bc}$	93±44 ^a	$0.51{\pm}0.15^{\rm ad}$	$0.47{\pm}0.11^{\rm bc}$

^aP<0.05, ^bP<0.01 vs control group; ^cP<0.05, ^dP<0.01 vs laparotomy control group.

Correlation between blood perfusion and VEGF, MMP-1 expression

There was a positive correlation between the serum VEGF level and the expression level of VEGF mRNA in tumor tissue (r=0.206, P<0.05). The blood perfusion data of tumor, represented by number of Hoechst 33342 labeled cells, showed a good linear inverse correlation with the serum VEGF level (r=-0.606, P<0.05) and VEGF mRNA expression in tumor tissue (r=-0.338, P<0.01). These data suggest that deteriorating blood perfusion and increased hypoxia of tumor correlated with a higher level of VEGF expression. There was no linear correlation between the number of Hoechst 33342 labeled cells and MMP-1 mRNA expression in tumor tissue.

DISCUSSION

TAE is an excellent debulking procedure and destroys malignant cells by selectively blocking the blood perfusion of the liver cancer. However, TAE can not block the blood supply of liver cancer and eliminate tumor cells completely, because the liver cancer receives the blood supply not only from the hepatic artery, but also from the port vein. Besides, it has been found that TAE could promote metastases of liver cancer^[4-6]. Thus, the long-term efficacy of TAE was disappointing: the 5-year survival rate was not above 20%^[10-14]. Even in several prospective randomized trials, TAE failed to improve significantly the survival of patients with PLC^[15,16]. Currently, the mechanisms involved in TAEinduced metastases of PLC are unknown. Understanding the mechanisms will facilitate the development of methods for blocking metastases and promoting long-term efficacy of TAE for treatment of liver cancer.

In recent years, many in vitro studies showed that hypoxia could enhance the metastatic ability of cancer cells by different mechanisms. It was reported that hypoxia stimulated carcinoma cell invasiveness by way of up-regulation of urokinase receptor expression^[17,18]. Hypoxia is the most important stimulus for the up-regulation of VEGF, one of the key cytokines for angiogenesis^[19-21]. Hypoxia could also induce the expression of other genes that promote angiogenesis and metastasis, such as basic fibroblast growth factor (bFGF) and angiogenin in tumor cells^[22-24]. Rofstad et al^[25] found that hypoxia could induce metastasis of human melanoma cells via VEGF-mediated angiogenesis. In vivo research also suggested that hypoxia may be one of the important stimuli of metastases. Hypoxia of tumor correlated with the expression of VEGF^[26-29], which was found to be correlated with metastases of the tumor and poor prognosis^[30-34]. Hypoxia in the squamous cell carcinoma of the uterine cervix with lymph node metastases was more serious than that in the carcinoma without lymph node metastases^[35]. Thus, we hypothesized that the inefficient vascular supply after TAE would result in hypoxia of some of the liver cancer cells, as it blocks the blood supply incompletely. We also suggest that hypoxia caused by

TAE may induce metastasis-associated factors, such as VEGF mediated metastases of liver cancer. It was found that the serum VEGF level in patients with liver cancer elevated 7 days after TAE therapy^[36]. In the present study, we blocked the hepatic arterial blood supply of implanted cancer in rat liver by HAL to simulate TAE therapy, and found that the blood perfusion decreased. This appears to suggest a more serious hypoxia. The expression of VEGF, both in serum and in tumor tissue, increased 2 days after the blockage of the arterial blood supply. Furthermore, we found that there was an inverse correlation between the blood perfusion of tumor and the serum VEGF level or VEGF mRNA expression in tumor tissues. These data indicate that hypoxia after HAL may be the major cause of the elevated expression of VEGF. The results of our study provide evidence for our hypothesis preliminarily. The hypoxia after TAE should be detected directly by hypoxyprobe-1[37,38], and if the hypoxia and expression of VEGF play a direct role in the mechanisms involved in TAE-induced metastases of liver cancer requires further studies. MMP is a secreted or transmembrane protein that is capable of digesting extracellular matrix and basement membrane and was associated with invasion and metastases of human tumor, such as hepatocellular carcinoma and colon cancer^[39-46]. It has been found that hypoxia could increase cellular invasiveness by inducing the expression of MMP^[47-50]; but there was no report about the change of MMP expression in liver cancer after TAE or HAL therapy. In the present study, we found that the expression of MMP-1 in tumor tissue increased 2 days after the blockage of the arterial blood supply, but there was no linear correlation between the blood perfusion of the tumor and MMP-1 mRNA expression in tumor tissue. Which is the cause of elevated MMP-1 expression and whether it played a direct role in the mechanisms involved in TAE-induced metastasis need further study.

In conclusion, the blood perfusion of implanted liver cancer in rats decreases and the expression of metastases-associated genes VEGF and MMP-1 increases after blockage therapy of the hepatic arterial blood supply. Decreased blood supply and hypoxia may be the main cause of VEGF expression. A more complete understanding of the mechanisms involved in TAE-induced metastasis may lead to an enhancement of the long-term effects of TAE on liver cancer.

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