

The effects of anisodamine and dobutamine on gut mucosal blood flow during gut ischemia/reperfusion

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Abstract

AIM: To determine if anisodamine is able to augment mucosal perfusion during gut I/R ischemia-reperfusion.

METHODS: A jejunal sac was formed in Sprague Dawley rat. A Laser Doppler probe and a tonometer were inserted into the sac which was filled with saline. The superior mesenteric artery was occluded (SMAO) for 60 minutes followed by 90 minutes of reperfusion. At the end of 60 minutes of SMAO, either 0.2mg/kg of anisodamine or dobutamine was injected into the jejunal sac. Laser Doppler mucosal blood flow and regional PrCO_2 (PrCO_2) measurements were made.

RESULTS: Mucosal blood flow was significantly increased at 30, 60 and 90 minutes of reperfusion (R_{30}, R_{60}, R_{90}) when intraluminal anisodamine or dobutamine was present compared to intraluminal saline only ($44 \pm 3.3\%$ or $48 \pm 4.1\%$ vs $37 \pm 2.6\%$ at R_{30} , $57 \pm 5.0\%$ or $56 \pm 4.7\%$ vs $45 \pm 2.7\%$ at R_{60} , $64 \pm 3.3\%$ or $56 \pm 4.2\%$ vs $48 \pm 3.4\%$ at R_{90} , respectively $P < 0.05$). Blood flow changes were also reflected by lowering of jejunal PrCO_2 measurements after intraluminal anisodamine or dobutamine compared with that of the saline controls ($41 \pm 3.1\text{mmHg}$ or $44 \pm 3.0\text{mmHg}$ vs $49 \pm 3.7\text{mmHg}$ at R_{30} , $38 \pm 3.7\text{mmHg}$ or $40 \pm 2.1\text{mmHg}$ vs $47 \pm 3.8\text{mmHg}$ at R_{60} , $34 \pm 2.1\text{mmHg}$ or $39 \pm 3.0\text{mmHg}$ vs $46 \pm 3.4\text{mmHg}$ at R_{90} , respectively, $P < 0.05$). Most interesting finding was that there were significantly higher mucosal blood flow and lower jejunal PrCO_2 in anisodamine group than those in dobutamine group at 90 minutes of reperfusion ($64 \pm 3.3\%$ vs $56 \pm 4.2\%$ for blood flow or $34 \pm 2.1\text{mmHg}$ vs $39 \pm 3.0\text{mmHg}$ for PrCO_2 , respectively, $P < 0.05$), suggesting that anisodamine had a more lasting effect on mucosal perfusion than dobutamine.

CONCLUSION: Intraluminal anisodamine and dobutamine can augment mucosal blood flow during gut I/R and alleviate mucosal acidosis. The results provided beneficial effects on the treatment of splanchnic hypoperfusion following traumatic or burn shock.

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INTRODUCTION

With remarkable advancement in our understanding of shock and greater ability to resuscitate patients from shock, few people today died of hypovolemic shock. However there still exists an inadequate

splanchnic perfusion, especially gut ischemia, despite apparent normalization of global hemodynamic parameters. Clinical and experimental studies have implicated gut hypoperfusion as an important inciting event which contributes to gut origin sepsis and multiple organ dysfunction^[1-10]. To improve gut perfusion, thus averting compensated shock, is still an important goal of resuscitation of shock^[11,12]. Anisodamine and dobutamine are commonly used as antishock drugs, as they can improve the microcirculation flow and splanchnic perfusion^[13-16]. Anisodamine, an anticholinergic drug extract from a Chinese herb *Anisodus tanguticus*, also possesses many other beneficial effects such as inhibition of thromboxane synthesis and protection of cell from reperfusion injury^[17-20]. Some researches indicated that anisodamine and dobutamine augmented gut perfusion during the shock^[21,22], but there was no report concerning their intraluminal effects on mucosal blood flow and metabolism in the gut. The purpose of this study is to investigate the effects of local administration of anisodamine and dobutamine on mucosal blood flow and PrCO_2 in a gut ischemia-reperfusion (I/R) rat model.

MATERIALS AND METHODS

Animal model

Male Sprague-Dawley rats weighing 350-450 grams were employed after acclimatization to the experimental environment. Rats were fasted overnight but allowed free access to water. Anesthesia was induced and maintained with 2% isoflurane and body temperature maintained at 37°C by the use of a warming blanket. Through an upper midline laparotomy, a segment of jejunum measuring 16cm in length was isolated 5cm distal to the ligament of Treitz with preservation of its mesentery. The isolated loop was closed at both ends with 3-0 silk ligatures^[23,24]. The superior mesenteric artery (SMA) was isolated at its origin and clamped for 60 minutes, followed by release of the clamp and restoration of blood to the intestine for 90 minutes to produce gut ischemia and reperfusion injury (I/R injury). At the time of release of the clamp, either 0.2mg/kg of anisodamine or dobutamine was injected into the jejunal sac. At the conclusion of the experiment, cardiac puncture and exsanguination were used to achieve euthanasia.

Animals were divided into three groups: Group one ($n=15$): I/R + anisodamine + saline; group two ($n=15$): I/R + dobutamine + saline and group three ($n=10$): I/R + saline as control.

Measurement of mucosal blood flow

A Teflon-coated laser optic flow probe (Peri flux PF409, flexible probe with 0.25mm fiber separation) was inserted through a small enterotomy at the proximal end of the jejunal sac and it was positioned along the antimesenteric border of the jejunum to the center of the sac. Mucosal blood flow was continuously recorded with a laser Doppler flow monitor (Peri Flux 4001 Master; Perimed, Jaarnfalla, Sweden). Blood flow measurements using Laser Doppler flow meters are not absolute but rather indicate flow in arbitrary perfusion units. Measurements were taken as the average flow over a five-minute period following an initial 30 minute period of stabilization. The quality of the signal was monitored by visualization on the computer screen so that motion artifact and noise were excluded from measurement^[25,26].

Measurement of PCO_2 ($PrCO_2$)

In the same animal, a 5F saline tonometer was inserted through a small enterotomy at the distal end of the sac and positioned to the center of the sac. The system was allowed to equilibrate for 30 minutes at which time a baseline $PrCO_2$ measurement was obtained by discarding the first 0.3ml saline from the tonometer balloon and using the remaining 0.7ml for analysis (model 1610 pH/blood gas analyzer, Milano, Italy), Regional PCO_2 ($PrCO_2$) was calculated using the following formula: $PrCO_2 = \text{measure } PCO_2 \times EF$. $EF = \text{equilibration factor}$, and based on the equilibration period for saline which gained from the handbook^[27-29].

Doppler measurements and $PrCO_2$ determinations were made every 30 minutes throughout the experimental period.

Statistics

Data were reported as mean \pm SEM and were analyzed by one-way analysis of variance (ANOVA) or student *t* test, significance was set at $P < 0.05$.

RESULTS

Laser doppler flow

Table 1 showed the effects of anisodamine and dobutamine on gut mucosal blood flow. Laser Doppler baseline flow (arbitrary units) was not different among groups. Flow dropped significantly during ischemia, with a reduction to 12% of baseline level, but notably not down to zero. With reperfusion, flow increased over time in all groups ($P < 0.05$) but did not reach baseline by 90 minutes. When compared to saline group, mucosal blood flow were higher in the anisodamine and dobutamine groups ($P < 0.05$) throughout the reperfusion period. Blood flow was also significantly higher at 90 minutes after reperfusion in anisodamine group compared with dobutamine animals ($P < 0.05$).

Table 1 Effects of anisodamine and dobutamine on gut mucosal blood flow (%)

Goups	B	I ₆₀	R ₃₀	R ₆₀	R ₉₀
Anisodamine	98 \pm 3.8	14 \pm 3.9	44 \pm 3.3 ^{ab}	57 \pm 5.0 ^{ab}	64 \pm 3.3 ^{abc}
Dobutamine	100 \pm 5.2	18 \pm 2.1	48 \pm 4.1 ^{ab}	56 \pm 4.7 ^{ab}	56 \pm 4.2 ^{ab}
Saline	103 \pm 6.9	16 \pm 3.4	37 \pm 2.6 ^a	45 \pm 2.7 ^a	48 \pm 3.4 ^a

Mean \pm SEM; ^a $P < 0.05$, vs I₆₀; ^b $P < 0.05$, vs saline; ^c $P < 0.05$, vs Dobutamine; B, baseline; I₆₀, ischemia 60 minutes; R₃₀, reperfusion 30 minutes; R₆₀, reperfusion 60 minutes; R₉₀, reperfusion 90 minutes.

Tonometry $PrCO_2$

Table 2 depicts jejunal $PrCO_2$ during I/R. Jejunal $PrCO_2$ did not differ in baseline among the groups, but it was significantly increased at 60 minutes of ischemia in all groups, amounting to 204% of baseline level. With reperfusion, $PrCO_2$ dropped over time in all groups ($P < 0.05$), but did not reach the baseline. When compared with saline group, $PrCO_2$ values were lower in the anisodamine and dobutamine groups ($P < 0.05$) throughout reperfusion period. There were no significant differences between anisodamine group and dobutamine animals at 30 and 60 minutes of reperfusion, but by 90 minutes of reperfusion anisodamine administration resulted in a significantly lower $PrCO_2$ than dobutamine ($P < 0.05$).

Table 2 Effects of anisodamine and dobutamine on $PrCO_2$ (mm Hg)

Goups	B	I ₆₀	R ₃₀	R ₆₀	R ₉₀
Anisodamine	28 \pm 2.2	55 \pm 3.9	41 \pm 3.1 ^{ab}	38 \pm 3.7 ^{ab}	34 \pm 2.1 ^{abc}
Dobutamine	31 \pm 3.8	61 \pm 7.8	44 \pm 3.0 ^a	40 \pm 2.1 ^{ab}	39 \pm 3.0 ^{ab}
Saline	26 \pm 2.5	57 \pm 5.4	49 \pm 3.7	47 \pm 3.8 ^a	46 \pm 3.4 ^a

Mean \pm SEM; ^a $P < 0.05$, vs I₆₀; ^b $P < 0.05$, vs saline; ^c $P < 0.05$, vs Dobutamine; B, baseline; I₆₀, ischemia 60 minutes; R₃₀, reperfusion 30 minutes; R₆₀, reperfusion 60 minutes; R₉₀, reperfusion 90 minutes

DISCUSSION

Both laboratory and clinical studies have demonstrated that splanchnic perfusion remains significantly impaired following resuscitation in traumatic, hemorrhagic and septic shock^[30-33]. Intravital video microscopic studies by Flynn *et al*^[34,35] have shown that although inflow and premucosal arterioles return to normal after resuscitation from hemorrhagic shock, there is a progressive arteriolar constriction resulting in a decrease in blood flow. There have been few reports about effects of intraluminal vasoactive agents on gut mucosal blood flow during both ischemia and reperfusion. It has been demonstrated that dobutamine could augment gut microcirculatory blood flow in septic shock^[36,37]. Observation in extensively burned patients by Sheng, *et al*^[22] have shown that intravenous administration of anisodamine 12 hours postburn resulted in a significant elevation in gastric pH and decrease in plasma level of TNF. In a porcine model of 30% TBSA full-thickness burn, anisodamine (0.4mg/kg) infused intravenously for one hour could increase portal blood flow, and it showed a positive correlation with intestinal pH^[38]. The use of Laser Doppler-measured tissue perfusion is a reliable technique that has been validated in the assessment of gastrointestinal mucosal blood flow, and the results were shown to correlate with other techniques of measurement of local blood flow^[33,38]. Recently, its use as a clinical tool for assessing jejunal mucosal perfusion had also been demonstrated^[39]. In this study with advanced Laser Doppler technique, we further demonstrated the beneficial effect of anisodamine on gut mucosal perfusion during I/R injury in comparison with dobutamine. The results showed that intraluminal anisodamine or dobutamine did increase mucosal blood flow throughout reperfusion as compared to intraluminal saline only. Blood flow augmentation was reflected by lower jejunal $PrCO_2$ measurements with intraluminal anisodamine or dobutamine. The most interesting finding was that there were significantly lower jejunal $PrCO_2$ and higher mucosal blood flow in anisodamine group than those in dobutamine at 90 minutes of reperfusion, suggesting that anisodamine had a more lasting effect on mucosal perfusion than dobutamine.

Tissue CO_2 gas tonometry provides an indirect measurement of perfusion and/or mucosal metabolic stress. Gastric tonometry, in particular, has been suggested as a tool to monitor splanchnic perfusion in experimental animals and critically ill patients^[40-43]. Though studies have demonstrated low gastric intramucosal pH (pHi) is a good predictor of poor outcome in critically ill patients, no improvement in outcome has been ascertained when patients are resuscitated based on the results of gastric tonometry^[44-46]. In this study we used 5F gut tonometry, in which the air pocket can be matched with rat small bowel sac, and the results are more accurate than gastric tonometry in detecting gut perfusion and metabolism^[47-50]. In our study significant increases in gut $PrCO_2$ following gut I/R were found, which could be markedly reduced by intraluminal anisodamine or dobutamine. These results might suggest that anisodamine or dobutamine is capable of augmenting mucosal blood flow, thus improving gut mucosal metabolism.

In conclusion, we have demonstrated in this laboratory model of gut ischemia and reperfusion that intraluminal anisodamine or dobutamine could augment mucosal blood flow, alleviate mucosal acidosis, improve metabolism in mucosal cell. These results provided reliable evidence to clinicians to adopt anisodamine or dobutamine in the treatment of splanchnic hypoperfusion, especially gut mucosal blood flow reduction following traumatic or burn shock.

REFERENCES

- 1 Heithan TH, Kone BC, Mercer DW, Moody FG, Weisbrodt NW, Moore FA. Postinjury multiple organ failure: The role of the gut. *Shock* 2001; 15:1-10
- 2 Moore FA. The role of the gastrointestinal tract in postinjury multiple organ

- failure. *Am J Surg* 1999;178:449-453
- 3 Zhang LY, Wang ZG, Zhu PF, Qin HJ. Gut barrier function disturbance posterior to hemorrhagic shock resuscitation in rats. *Shijie Huaren Xiaohua Zazhi* 2001;9:767-770
 - 4 Hu S, Sheng ZY, Zhou BT, Guo ZR, Lu JY, Xue LB, Jin H, Sun XQ, Sun SR, Li JY, Lu Y. Study on delay two-phase multiple organ dysfunction syndrome. *Chin Med J* 1998;111:101-108
 - 5 Hu S, Sheng ZY, Zhou BT, Xue LB, Jin H, Lu Y, Lin HY. Experimental study on hemodynamic changes in the development of multiple organ dysfunction syndrome. *Zhongguo Weizhongbing Jijiu Yixue* 1996;8:707-709
 - 6 Reed LL, Mangano R, Martin M, Hochman M, Kocka F, Barrett J. The effect of hypertonic saline resuscitation on bacterial translocation after hemorrhagic shock in rats. *Surgery* 1991;110:685-690
 - 7 Tamion F, Richard V, Lyoumi S, Daveau M, Bonmarchard G, Leroy T, Thuillez C, Lebreton JP. Gut ischemia and mesenteric synthesis of inflammatory cytokines after hemorrhagic or endotoxic shock. *Am J Physiol* 1997;273:G314-G321
 - 8 Moore EE, Moore FA, Franciose RJ, Kim FJ, Biffl, Banerjee A. Postischemia gut serves as a priming bed for circulating neutrophils that provoke multiple organ failure. *J Trauma* 1994;37:881-887
 - 9 Magnotti LL, Upperman JS, Xu DZ, Lu Q, Deitch EA. Gut derived mesenteric lymph but not portal blood increases endothelial cell permeability and promotes lung injury after hemorrhagic shock. *Ann Surg* 1998;228:518-527
 - 10 Schmidt H, Secchi A, Wellmann R, Bach A, Bohrer H, Gebhard MM, Martin E. Effect of endotoxemia on intestinal villus microcirculation in rats. *J Surg Res* 1996;61:521-526
 - 11 Fiddian-Green RG, Haglund U, Gutierrez, Shoemaker WC. Goals for the resuscitation on shock. *Crit Care Med* 1993;21:S25-S31
 - 12 Friedman G, Silva E, Vincent JL. Has the mortality of septic shock changed with time? *Crit Care Med* 1998;26:2078-2086
 - 13 Martin C, Viviani X, Arnaud S, Vialet R, Rougon T. Effects of norepinephrine plus dobutamine or norepinephrine alone on left ventricular performance of septic shock patients. *Crit Care Med* 1999;27:1708-1713
 - 14 Hoogenberg K, Smit AJ, Girbes AJ. Effects of low-dose dopamine on renal and systemic hemodynamics during incremental norepinephrine infusion in healthy volunteers. *Crit Care Med* 1998;26:260-265
 - 15 Backer DD, Zhang HB, Cherkhaoui S, Borgers M, Vincent JL. Effects of dobutamine on hepato-splanchnic hemodynamics in an experimental model of hyperdynamic endotoxic shock. *Shock* 2001;15:208-214
 - 16 Yang XH. Mechanism of the protective effect of anisodamine in gut ischemia injury. *Zhongguo Bingli Shengli Zazhi* 1988;4:134-136
 - 17 Huang YS, Li A, Yang ZC. Roles of thromboxane and its inhibitors anisodamine in burn shock. *Burns* 1990;16:249-253
 - 18 Shi LB, Peng SY, Meng XY, Liu YB, Peng CH. The role of platelet activating factor in hepatic ischemia-reperfusion injury and the protective effect of anisodamine. *Zhongguo Weizhongbing Jijiu Yixue* 2001;13:220-222
 - 19 Jang CG, Yang GT, Tang Y. Influence of Anisodamine on neuronal apoptosis after cerebral reperfusion in the rats. *Zhongguo Jijiu Zazhi* 2001;21:131-133
 - 20 Meng XK, Shi LB, Peng SY, Peng CH, Wu YL, Sheng HW. Experimental study of anisodamine against hepatic ischemia-reperfusion injury. *Zhongguo Jijiu Yixue* 2001;21:4-6
 - 21 Joly LM, Monchi M, Cariou A. Effects of dobutamine on gastric mucosal perfusion and hepatic metabolism in patients with septic shock. *Am J Respir Crit Care Med* 1999;160:1983-1986
 - 22 Sheng ZY, Gao WY, Guo ZR, He LX. Anisodamine restores bowel circulation in burn shock. *Burns* 1997;23:142-146
 - 23 Wilson TH. Methods. IN: Wilson TH, ed. *Intestinal Absorption*. Philadelphia: W. B. Saunders Co 1962:20-39
 - 24 Hu S, Kozar RA, Moore FA, Sheng ZY. Enteral feeding of glucose increases intestinal mucosal blood flow during intestinal ischemia/reperfusion injury. *Zhonghua Shaoshang Zazhi* 2001;17:139-141
 - 25 Wang P, Zhou M, Cioffi WG, Bland KI, Zheng F, Chaudry IH. Is prostacyclin responsible for producing the hyperdynamic response during early sepsis? *Crit Care Med* 2000;28:1534-1539
 - 26 Elizalde JJ, Hernandez C, Liach J, Monton C, Bordas JM, Pique JM, Torres A. Gastric intramucosal acidosis in mechanically ventilated patients: role of mucosal blood flow. *Crit Car Med* 1998;26:827-832
 - 27 Dawson AM, Trenchard D, Guz A. Small bowel tonometer: assessment of small gut mucosal oxygen tension in dog and man. *Nature* 1965;206:943-945
 - 28 Heinonen PO, Jousela IT, Blomqvist KA. Validation of air tonometric measurement of gastric regional concentration of CO₂ in critically ill septic patients. *Intensive Care Med* 1977;23:524-529
 - 29 Mckinley BA, Marvin RG, Moore FA. Gastric and intestinal mucosal regional PCO₂ following shock resuscitation: changes with small intestinal enteral feeding. *Shock* 1999;11:S72-73
 - 30 Fiddian-Green RG. Associations between intramucosal acidosis in the gut and organ failure. *Crit Care Med* 1993;21:S103-107
 - 31 Grossie B, Weisbrodt NW. Inhibition of small intestinal transit by ischemia/reperfusion in the rat. *Dig Dis Sci* 1998;43:1585-1587
 - 32 Wang P, Shou M, Rana MW. Differential alterations in microvascular perfusion in various organs during early and late sepsis. *Am J Physiol* 1992;263:G38-G43
 - 33 Wang P, Hauptman JG, Chaudry IH. Hemorrhage produces depression in microvascular blood flow that persists despite fluid resuscitation. *Circ Shock* 1990;32:307-318
 - 34 Flynn WJ, Cryer HG, Garrison RH. Pentoxifylline restores intestinal microvascular blood flow during resuscitated hemorrhagic shock. *Surgery* 1991;110:350-356
 - 35 Flynn WJ, Gosche JR, Garrison RN. Intestinal blood flow is restored with glutamine or glucose infusion after hemorrhage. *J Surg Res* 1992;52:499-504
 - 36 Gutierrez G, Clark C, Brown K SD, Price K, Ortiz L, Nelson C. Effect of dobutamine on oxygen consumption and gastric intramucosal pH in septic patients. *Am J Respir Crit Care Med* 1994;150:324-329
 - 37 Nevriere R, Chagnon JL, Vallet B, Lebleu N, Marechal X, Mathieu D, Wattel F, Dupuis B. Dobutamine improves gastrointestinal mucosal blood flow in a porcine model of endotoxic shock. *Crit Care Med* 1997;25:1371-1377
 - 38 Gao WY, Sheng ZY, Gou ZR, He LX, Xiong DX, Song HF, Zhang SX, Ma NS, Chang GY. Protective effect of anisodamine on intestine in the early period of burn shock. *Jiefangjun Yixue Zazhi* 1995;20:88-91
 - 39 Thoren A, Elam M, Ricksten S. Differential effects of dopamine, dopexamine, and dobutamine on jejunal mucosal perfusion early after cardiac surgery. *Crit Care Med* 2000;28:2338-2343
 - 40 Temmesfeld-Wollbruck B, Szalay A, Olschewski H. Advantage of buffered solutions or automated capnometry in air-filled balloons for use in gastric tonometry. *Intensive Care Med* 1997;23:423-427
 - 41 Knudson M, Bermudez KM, Doyle CA, Mackeres RC, Hopf HW, Morabito D. Use of tissue oxygen tension measurements during resuscitation from hemorrhagic shock. *J Trauma* 1997;42:608-611
 - 42 Hu S, Sheng ZY, Zhou BT, Xue LB, Jin H. Changes in gastrointestinal intramucosal pH in goats resuscitated from hypovolemic shock. *Zhongguo Weizhongbing Jijiu Yixue* 1997;9:708-710
 - 43 Marik P, Lorenzana A. Effect of tube feeding on the measurement of gastric intramucosal pH. *Crit Care Med* 1996;24:1498-1500
 - 44 Gomersall CD, Joynt GM, Freebairn RC, Hung V, Buckley CA, Oh TE. Resuscitation of critically ill patients based on the results of gastric tonometry: A prospective, randomized, controlled trial. *Crit Care Med* 2000;28:607-614
 - 45 Kirton OC, Windsor J, Wedderburn R, Hudson-Civetta J, Shatz DV, Mataragas NR, Civetta JM. Failure of splanchnic resuscitation in the acutely injured trauma patient correlates with multiple organ-system failure and length of stay in the ICU. *Chest* 1998;113:1064-1069
 - 46 Ivatury RR, Simon RJ, Islam S, Fueg A, Rohman M, Stahl WM. A prospective randomized study of resuscitation after major trauma: Global oxygen transport induces versus organ-specific gastric mucosal pH. *J Amer Col Surg* 1996;183:145-154
 - 47 Walley KR, Friesen BP, Humer MF, Phang PT. Small bowel tonometry is more accurate than gastric tonometry in detecting gut ischemia. *J Appl Physiol* 1998;85:1770-1777
 - 48 Barry B, Mallick A, Hartley G, Bodenham, Vucevic M. Comparison of air tonometry with gastric tonometry using saline and other equilibrating fluids: an in vivo and in vitro study. *Intensive Care Med* 1998;24:777-784
 - 49 Noone RB, Bolden JE, Mythen NG, Vaslef SN. Comparison of the response of saline tonometry and an automated gas tonometry device to a change in CO₂. *Crit Care Med* 2000;28:3728-3733
 - 50 Thorburn K, Hatherill M, Roberts PC, Durward A, Tibby SM, Murdoch IA. Evaluation of the 5-French saline paediatric gastric tonometer. *Intensive Care Med* 2000;26:973-980

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