

Reply to “Immunosuppression in a Comparative Study of Feline Leukemia Virus Vaccines”

M. Patel,^a K. Carritt,^a J. Lane,^a H. Jayappa,^a M. Stahl,^b M. Bourgeois^b

Merck Animal Health, Elkhorn, Nebraska, USA^a; Merck Animal Health, Madison, New Jersey, USA^b

Our response to the [comments](#) on our recent publication in *Clinical and Vaccine Immunology* (1) is as follows. The authors state that the conclusions “are misleading and inconsistent with previous publications on the efficacy of canarypox-FeLV vaccine (2–5).” The first three references cited to support this claim did not use Purevax recombinant feline leukemia virus (FeLV) vaccine (which has a unique mutation within the envelope protein immunosuppressive domain) but used a vaccine not available in the United States (2–4). Later, the authors cite studies using the transdermal FeLV vaccine no longer available (5, 6). The only relevant paper cited using Purevax recombinant FeLV vaccine used a homologous challenge (Glasgow-1) 2 weeks postvaccination (7). In contrast, our study used a heterologous challenge (61E) 3 months postvaccination. Thus, while results from our study are inconsistent with publications on canarypox FeLV vaccines, only one paper cited is relevant. In that paper, a shorter challenge time frame and a homologous challenge are questionably relevant to vaccine field use.

The authors state that canarypox-vectored vaccines likely do not rely on antibodies and are dependent on a T-cell-mediated response; thus, immunosuppression may have a greater impact on Purevax recombinant FeLV vaccination. However, there are a number of studies showing that vaccination with killed, whole-virus adjuvanted vaccines does not induce virus-neutralizing antibodies before challenge, yet cats are still protected (8–10). In two of these studies, cats were immunosuppressed and were able to overcome infection (9, 10). Thus, T-cell-mediated responses should also be important for the immunosuppressed cats vaccinated with Nobivac Feline-2 FeLV. In addition, virus-neutralizing titers in this study were not evaluated; thus, the protective significance of the titers is unknown (1).

Cats that are at highest risk for FeLV infection (11) are often immunosuppressed. Elevated urine cortisol/creatinine ratios are seen in shelter and sick cats (12, 13), and concurrent illnesses are common (14, 15). Veterinary visits are stressful, and corticosteroids may be administered to vaccine reactors. Immunosuppression during FeLV challenge is an accepted methodology to ensure infection. Immune priming should not be impacted as long as sufficient time for the immune response (2 weeks) has elapsed before immunosuppression. In this study, immunosuppression occurred 3 months after booster vaccination. This has been supported in USDA licensing requirements for FeLV vaccines (16).

Thus, in this study, immunosuppression should have no effect on the formation of immunity. We do not know whether immunosuppression affects Purevax recombinant FeLV-vaccinated cats to a greater degree than Nobivac Feline-2 FeLV-vaccinated cats during challenge. However, if that is the case, this study shows that immunosuppressed cats are better protected upon FeLV exposure after vaccination with Nobivac Feline-2 FeLV than after vaccination with Purevax recombinant FeLV. The purpose of this study

was to compare the efficacies of two commercially available vaccines. As the same vaccination and study conditions were applied to both groups, the efficacy of the Nobivac Feline-2 FeLV vaccine was far superior to that of Purevax recombinant FeLV vaccine and the conclusions remain the same.

REFERENCES

1. Patel M, Carritt K, Lane J, Jayappa H, Stahl M, Bourgeois M. 2015. Comparative efficacy of feline leukemia virus (FeLV) inactivated whole-virus vaccine and canarypox virus-vectored vaccine during virulent FeLV challenge and immunosuppression. *Clin Vaccine Immunol* 22:798–805. <http://dx.doi.org/10.1128/CVI.00034-15>.
2. Tartaglia J, Jarrett O, Neil JC, Desmettre P, Paoletti E. 1993. Protection of cats against feline leukemia virus by vaccination with a canarypox virus recombinant, ALVAC-FL. *J Virol* 67:2370–2375.
3. Poulet H, Brunet S, Boularand C, Guiot AL, Leroy V, Tartaglia J, Minke J, Audonnet JC, Desmettre P. 2003. Efficacy of a canarypox virus-vectored vaccine against feline leukaemia. *Vet Rec* 153:141–145. <http://dx.doi.org/10.1136/vr.153.5.141>.
4. Hofmann-Lehmann R, Tandon R, Boretti FS, Meli ML, Willi B, Cattori V, Gomes-Keller MA, Ossent P, Golder MC, Flynn JN, Lutz H. 2006. Reassessment of feline leukaemia virus (FeLV) vaccines with novel sensitive molecular assays. *Vaccine* 24:1087–1094. <http://dx.doi.org/10.1016/j.vaccine.2005.09.010>.
5. Grosenbaugh DA, Leard T, Pardo MC, Motes-Kreimeyer L, Royston M. 2004. Comparison of the safety and efficacy of a recombinant feline leukemia virus (FeLV) vaccine delivered transdermally and an inactivated FeLV vaccine delivered subcutaneously. *Vet Ther* 5:258–262.
6. Grosenbaugh DA, Leard T, Pardo MC. 2006. Protection from challenge following administration of a canarypox virus-vectored recombinant feline leukemia virus vaccine in cats previously vaccinated with a killed virus vaccine. *J Am Vet Med Assoc* 228:726–727. <http://dx.doi.org/10.2460/javma.228.5.726>.
7. Schlecht-Louf G, Mangeney M, El-Garch H, Lacombe V, Poulet H, Heidmann T. 2014. A targeted mutation within the feline leukemia virus (FeLV) envelope protein immunosuppressive domain to improve a canarypox virus-vectored FeLV vaccine. *J Virol* 88:992–1001. <http://dx.doi.org/10.1128/JVI.02234-13>.
8. Torres AN, O’Halloran KP, Larson LJ, Schultz RD, Hoover EA. 2010. Feline leukemia virus immunity induced by whole inactivated virus vaccination. *Vet Immunol Immunopathol* 134:122–131. <http://dx.doi.org/10.1016/j.vetimm.2009.10.017>.
9. Pedersen NC. 1993. Immunogenicity and efficacy of a commercial feline leukemia virus vaccine. *J Vet Intern Med* 7:34–39. <http://dx.doi.org/10.1111/j.1939-1676.1993.tb03166.x>.
10. Jirjis FF, Davis T, Lane J, Carritt K, Sweeney D, Williams J, Wasmoen T. 2010. Protection against feline leukemia virus challenge for at least 2

Citation Patel M, Carritt K, Lane J, Jayappa H, Stahl M, Bourgeois M. 2015. Reply to “Immunosuppression in a comparative study of feline leukemia virus vaccines.” *Clin Vaccine Immunol* 22:1296–1297. doi:10.1128/CVI.00504-15.

Editor: M. F. Pasetti

Address correspondence to M. Bourgeois, Melissa.Bourgeois@merck.com.

This is a response to a letter by Poulet et al. (doi:10.1128/CVI.00497-15).

Copyright © 2015, American Society for Microbiology. All Rights Reserved.

- years after vaccination with an inactivated feline leukemia virus vaccine. *Vet Ther* 11:E1–E6.
11. Levy J, Crawford C, Hartmann K, Hofmann-Lehmann R, Little S, Sundahl E, Thayer V. 2008. American Association of Feline Practitioners' feline retrovirus management guidelines. *J Feline Med Surg* 10:300–316. <http://dx.doi.org/10.1016/j.jfms.2008.03.002>.
 12. McCobb EC, Patronek GJ, Marder A, Dinnage JD, Stone MS. 2005. Assessment of stress levels among cats in four animal shelters. *J Am Vet Med Assoc* 226:548–555. <http://dx.doi.org/10.2460/javma.2005.226.548>.
 13. Henry CJ, Clark TP, Young DW, Spano JS. 1996. Urine cortisol: creatinine ratio in healthy and sick cats. *J Vet Intern Med* 10:123–126.
 14. Digangi BA, Gray LK, Levy JK, Dubovi EJ, Tucker SJ. 2011. Detection of protective antibody titers against feline panleukopenia virus, feline herpesvirus-1, and feline calicivirus in shelter cats using a point-of-care ELISA. *J Feline Med Surg* 13:912–918. <http://dx.doi.org/10.1016/j.jfms.2011.07.009>.
 15. Zicola A, Saegerman C, Quatpers D, Viandier J, Thiry E. 2009. Feline herpesvirus 1 and feline calicivirus infections in a heterogeneous cat population of a rescue shelter. *J Feline Med Surg* 11:1023–1027. <http://dx.doi.org/10.1016/j.jfms.2009.05.023>.
 16. Shibley GP, Tanner JE, Hanna SA. 1991. United States Department of Agriculture licensing requirements for feline leukemia virus vaccines. *J Am Vet Med Assoc* 199:1402–1406.