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Enhanced Diastereoselectivity in β**-Mannopyranosylation through the Use of Sterically Minimal Propargyl Ether Protecting Groups**

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Abstract

2-*O*-Propargyl ethers are shown to be advantageous in the 4,6-*O*-benzylidene acetal directed βmannosylation reaction. The effect is most pronounced when the 3-*O*-protecting group is bulky silyl ether or a glycosidic bond, however, even with a 3-*O*-benzyl ether the use of a 2-*O*-propargyl ether results in a significant increase in diastereoselectivity. The beneficial effect of the propargyl ether is thought to be a combination of its minimal steric bulk, as determined by measurement of the steric A-value, and of its moderately disarming nature, as reflected in the p*K*a of propargyl alcohol. Conversely, the application of a 3-*O*-propargyl ether in the benzylidene acetal directed mannosylation has a detrimental effect on stereoselectivity, for which no explanation is at present available. Deprotection is achieved by base-catalyzed isomerization of the propargyl ether group to the corresponding allenyl ether, followed by oxidative cleavage with *N*-methylmorpholine *N*oxide and catalytic osmium tetroxide.

Graphical Abstract

Introduction

Protecting groups play a central role in carbohydrate chemistry,¹ with applications extending beyond the simple blocking of hydroxyl groups to the modulation of reactivity of both glycosyl donors and acceptors, and, critically, the control of anomeric stereochemistry. Indeed, the development of new protecting groups capable of rendering enhanced control of

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regioselectivity,² reactivity,³ and stereoselectivity,⁴ can be said to be one of the current frontiers of the discipline.

The influence of even remote protecting groups on the control of anomeric stereochemistry is illustrated by the 4,6-*O*-benzylidene protected β-mannosyl donors developed in this laboratory,⁵ in which the benzylidene acetal, or its surrogate, $4d,6$ is now understood to function by restricting the C5-C6 bond to the more disarming⁷ tg conformer,^{8,9} thereby limiting the lifetime of the transient contact ion pair¹⁰ that is in equilibrium with the covalent glycosyl triflate intermediate.^{5c,11}

However powerful this method may be in the synthesis of complex oligosaccharides containing the β-mannopyranoside and related linkages,^{12,13} it is not without limitations. Thus, the use of donors bearing bulky groups on *O*-3, either silyl ethers or glycosidic bonds, diminishes the selectivity of the mannosylation.¹⁴

Although the effect of the *O*-3 protecting group on anomeric stereoselectivity is not yet fully understood, we introduced the use of 2-*O*-propargyl ether as a means of overcoming the loss of selectivity as a result of the use of bulky groups at *O*-3.15 In this Article we examine in greater detail the potential of the readily cleavable, minimally sterically intrusive propargyl ether protecting group and show how, in conjunction with the correct choice of other protecting groups, it can lead to considerable enhancements in the stereoselectivity of mannopyranosylation reactions and even the very challenging rhamnopyranosylations.

Results and Discussion

The problem of diminished selectivity caused by bulky groups on *O*-3 was initially encountered in the synthesis of the common core pentasaccharide of the *N*-linked glycans,13a when coupling of the 2-*O*-benzyl-3-*O*-TBDMS mannosyl donor **2** with pentenyl glycoside acceptor **1** exhibited poor selectivity (77%, $\alpha:\beta = 1.8:1$). In contrast, with the 2-*O*-TBDMS-3-*O*-benzyl donor **3**, the selectivity was significantly better (72%, $\alpha:\beta = 1:3$), albeit still not at the high levels typically experienced with more standard 2,3-di-*O*-benzyl protected donors.¹⁶

A more critical manifestation of this problem presented itself during the synthesis of the alternating β-(1→3)-β-(1→4)-mannan common to *Rhodotorula glutinis*, *Rhodotorula mucilaginosa* and *Leptospira biflexa*. 14b Donors **4** and **5**, both displaying very bulky

glycosyl substituents on *O*-3, showed unusually poor β-selectivity in coupling reactions, thereby reducing the efficiency of the convergent synthesis of the target polysaccharide.

We hypothesized that the poor selectivity seen with donors **2**, **4**, and **5** was the result of steric buttressing between the *O*-2 and *O*-3 protecting groups, resulting in unusually high shielding of the β-face of the glycosyl donor.^{14a,15} Thus, as illustrated for the triflate derived from **2**, we reason that of the three possible staggered conformations around the *O-*3 substituent bond, (A) is disfavored by the steric interaction with the rigid benzylidene ring leading to the preferential population of conformers (**B**) and (**C**) in which the bulky silyl group is gauche to C-2 and its substituent (Fig. 1).

Viewed from the perspective of the *O*-2-substituent bond, the population of conformer (**D**) is likely extremely small due to high steric congestion. The bulky group on *O*-3 presumably destabilizes conformation (**E**) thus leaving (**F**) as the most populous state (Fig. 2). In conformer **F** the 2-*O*-benzyl ether is in close proximity to the β-face of the α-mannosyl triflate. This enhanced steric shielding retards attack on the β-face, either on the covalent triflate itself or on the transient contact ion pair arising from the covalent triflate, thereby resulting in the observed loss of β-selectivity.

In systems such as **2** this problem can be circumvented by the simple ruse of switching to a less bulky *O-*3 protecting group, however in target-directed convergent oligosaccharide synthesis there is no way to avoid the use of donors such as **4** and **5**. We reasoned that the unfavorable steric interaction in conformer **E** could be reduced by minimizing the size of the *O*-2 protecting group, which should have the effect of increasing the population of **E** at the expense of **F**. At the same time, the use of a protecting group with a low steric demand on O-2, should serve to minimize the detrimental effect of any residual population of conformer **F**. We were encouraged in this line of thinking, by van Boom's work on the successful βglycosylation of several acceptors by donor **6** with the relatively small 2-azido group.¹⁷ However, the size of the azido group cannot be viewed independently of its strongly disarming properties, thereby complicating the interpretation of this precedent. For similar reasons, we decided not to pursue the very small but also moderately disarming cyanate esters,^{4e} and focused instead on the allyl and propargyl ethers.

We began with the synthesis of the 3-*O*-silyl compounds **10** and **13** (Scheme 1) by standard means from the known thioglycoside 7.^{14b} In these syntheses the 3-*O*-silyl group was introduced after the allyl or propargyl ethers to preempt problems of silyl migration that were anticipated in the reverse protocol.

Donors **10** and **13** were then coupled to the acceptor **14** by our standard BSP/TTBP/Tf₂O protocol^{5c,16} leading to the yields and selectivities outlined in Table 1. Included in Table 1 for comparison is the previous coupling^{14a} of donor 2 to acceptor 1, by the directly analogous sulfoxide method.¹⁶

These results strongly support the hypothesis of the beneficial effect of reducing the bulk of the 2-*O* protecting group on the stereochemical outcome of the reaction, with the best anomeric ratio obtained with the smallest *O*-2 protecting group. To put the inverse relationship between the steric bulk of the *O*-2 protecting group and anomeric selectivity on a more secure footing we measured steric A-values for the propargyloxy, allyloxy, benzyloxy, and *tert*-butyldimethylsiloxy groups by the classical 1H VT-NMR method (Table 2).18 The observed trend in A-values fully supports the initial hypothesis, with the propargyl ether being significantly smaller than the allyl ether, which in turn is smaller than the benzyl ether. The A-value for the *tert*-butyldimethylsiloxy group determined here, and included for comparison purposes, is significantly greater than that previously measured by Eliel for the same group using an alternative 13 C-NMR method, 19 but is consistent with the general trend of coupling selectivities observed in this entire study.

In addition to the smaller size of the propargyl ether we also considered the possibility that it might exhibit an electron-withdrawing effect. Indeed, the sp-hybridization of the alkyne carbon renders the propargyloxy group moderately electron-withdrawing with respect to the other ethers studied, as seen from the p*K*a's of the corresponding alcohols (Table 2),²⁰ and it is likely that the beneficial effect of the 2-*O*-propargyl ethers arises from a combination of the minimal steric bulk and its moderately disarming property.

The coupling of donor **13** to two further substrates, again with excellent results (Table 3), confirmed the ability of the 2-*O*-propargyl ether protecting group to overcome the deleterious effects of a 3-*O*-silyl ether.

Attention was next focused on donors bearing a glycosidic bond at *O*-3, analogous to the problematic **4** and **5**. Furthermore, bearing in mind the potential for the eventual use in mannan synthesis, a glycosyl acceptor carrying a 2-*O*-propargyl ether was also prepared (Scheme 2).

Acceptor **12** was successfully coupled to the known donor **28**5c with α:β selectivity of 1:16 in 88% yield (Scheme 3). Per the protocol of van Boom, $14b,17,21$ triethyl phosphate was added after addition of the acceptor **12** to limit premature activation its thioglycoside functionality by any extraneous thiophiles. Coupling of disaccharide donor **29** to acceptor **27** then gave the mannotriose in 80% yield with an α:β ratio of 1:5, presenting a very significant improvement over the approximately 1:1 α:β ratio observed with donor **4** and a related acceptor.14b Additionally, the successful couplings employing compounds **12** and **27** illustrate that propargyl ethers are also suitable for protection of acceptors.

With a means to overcome the unfavorable effect of a bulky 3-O substituent in hand, we proceeded to undertake a broader investigation into the general effects of propargyl ethers on stereoselectivity in 4,6-*O*-benzylidene-directed β-mannosylation reactions. Specifically, we reasoned that while the steric buttressing effect discussed above and illustrated in Figs 1 and 3 will be maximized with a large group on *O*-3, it will necessarily be present with more common protecting groups on *O*-3, albeit to a lesser extent. Accordingly, the use of a 2-*O*propargyl even in conjunction with a 3-*O*-benzyl ether should lead to enhanced selectivity over the more typical 2,3-di-*O*-benzyl protected donors. To probe this idea, a series of four donors was prepared, using standard techniques from diol **31**14b via the known monobenzyl ethers **32** and **34**14b (Scheme 4).

Subsequent coupling of this series of donors to a standard acceptor **14** gave the results presented in Table 4. Comparison of entries 1 and 2 in Table 4 clearly demonstrates a 2-*O*propargyl ether lead to enhanced β-selectivity even with the 3-*O*-benzyl protected system, in accordance with the above stated hypothesis. The 3-*O*-propargyl donor **35** (Table 4, entry 3) gave surprisingly poor but reproducible results, for which we have no satisfactory explanation at the present time. It is clear, however, that the *O*-3 group plays a major role in these 4,6-*O*-benzylidene protected β-mannosylation reactions, and that the issue of steric bulk and buttressing discussed here is only one facet of the problem.²² Taking into account the selectivities obtained with donors **33** and **35**, it is clear that the modest 10:1 β:α selectivity obtained with the 2,3-di-*O*-propargyl protected donor **36** (Table 4, entry 4) is a compromise between the excellent selectivity obtained with a 2-*O*-propargyl group alone, and the obviously harmful effect of the 3-*O*-propargyl ether.

The very encouraging results obtained with donor **33**, featuring the combination of the 2-*O*propargyl and 3-*O*-benzyl ether protecting groups were then extended to encompass a broader range of typical acceptor alcohols (Table 5). In each excellent yields and β:α selectivies surpassing 20:1 were obtained.

While the use of allyl ethers as protecting groups is extremely widespread,^{1a, 23} that of propargyl ethers is novel, and required the investigation of suitable deprotection conditions. It has been reported that propargyl ethers may be cleaved with benzyltriethylammonium tetrathiomolybdate, 24 with low valent titanium in hot THF, 25 and by a nickel-catalyzed electrochemical protocol.26 However, based on experience in our laboratory with allyl ethers in oligosaccharide synthesis, $12c,d$, 27 we have preferred a method involving base-catalyzed isomerization to the corresponding allenyl ether followed by oxidative cleavage with catalytic osmium tetroxide in the presence of *N*-methyl morpholine *N*-oxide (NMNO) as reported by Mereyala and co-workers,²⁸ albeit under somewhat milder conditions. Thus, a representative series of propargyl ether protected saccharides was treated with potassium *tert*-butoxide in THF at room temperature, followed by exposure to catalytic $OsO₄$ in the presence of NMNO, also at room temperature, resulting in hydrolysis to the corresponding alcohols (Table 6).

Finally, we have briefly investigated the potential of the 2-*O*-propargyl ether protecting group in the synthesis of β-L-rhamnopyranosides, a cognate problem to that of the β-Dmannopyranosides but one which does not allow the use of the stereodirecting 4,6-*O*benzylidene acetal function in the donor. As part of our ongoing effort in this area, $4e,29$ we reported that the 3,4-*O*-carbonate protected rhamnosyl donor **60** gave moderate to good β:α selectivity (1.5:1 to β only) on coupling to various acceptors under the standard BSP/Tf₂O/ TTBP conditions, depending on the reactivity of the acceptor.4f

It was reasonable, therefore, to investigate the analogous 2-*O*-carbonate **64**, which was prepared as set out in Scheme 5 from the known^{4f} bisacetal **61**.

Activation of **64**, which proceeded smoothly under the standard conditions, was followed by the addition of methyl 2,3,6-tri-*O*-benzyl-α-D-glucopyranoside **14**, a member of the glucose 4-OH derivatives which are known to be relatively difficult to glycosylate, 30 giving the disaccharide **65** in 65% yield in the form of a 2:1 β:α mixture (Scheme 6). This represents only a modest improvement of selectivity over the 1.5:1 β:α ratio obtained on coupling of **60** with 14,^{4f} and discouraged us from further work with this donor. Presumably, the there is very little buttressing interaction between the tied back carbonate and the protecting group on *O*-2, as such the effect on stereoselectivity of changing the *O*-2 protecting from the benzyl ether to the propargyl ether is very small.

Overall, propargyl ethers are readily introduced and cleaved protecting groups for alcohols that bring about significant improvements in the diastereoselectivity of many mannosylation reactions, which we attribute to the combination of their minimal steric bulk and their modest disarming power. While we have focused on the application of this protecting group

to the solution of current problems in our laboratory, we anticipate that it will find broader application in organic synthesis, especially in situations in which the steric bulk of a protecting group is a factor.

Experimental Section

Phenyl 4,6-O-benzylidene-2-O-(prop-2-ynyl)-3-O-p-methoxybenzyl-1-thio-α**-Dmannopyranoside (11)**

To a stirred solution of phenyl 4,6-*O*-benzylidene-3-*O*-*p*-methoxybenzyl-1-thio-α-Dmannopyranoside (2.5 g, 5.5 mmol) in dry dimethylformamide (15 mL) at 0 ºC was added NaH 60% in oil (0.33 g, 8.3 mmol) and stirred for 15 min. Propargyl bromide (0.93 mL, 8.3 mmol) was added dropwise to the above reaction mixture and continued stirring for 3 h. The rection mixture was quenched by addition of methanol, diluted with CH_2Cl_2 (25 mL) and washed with sat. NaHCO₃. The organic layer was separated and dried over anhydrous Na2SO4 and concentrated under vaccum. The crude product was purified by flash column cromatography on silica gel (hexane:ethyl acetate; 8:1) to give 11 (2.46 g, 85%), $\lbrack \alpha \rbrack^{24.5}$ _D + 155.8 (*c*, 2.0, CHCl3); 1H NMR (500 MHz, CDCl3) δ: 2.4 (t, *J* = 2.4 Hz, 1H), 3.82 (s, 3H), 3.87 (t, *J* = 11.0 Hz, 1H), 3.98 (dd, *J* = 3.0, 10.0 Hz, 1H), 4.19–4.24 (m, 3H), 4.26–4.31 (m, 1H), 4.4 (dd, *J* = 0.5, 2.0 Hz, 2H), 4.70 (d, *J* = 12.0 Hz, 1H), 4.81 (d, *J* = 12.0 Hz, 1H), 5.61 (d, *J* = 1.5 Hz, 1H), 5.63 (s, 1H), 6.9 (d, *J* = 8.6 Hz, 2H), 7.3–7.45 (m, 10H), 7.5–7.56 (m, 2H); 13C NMR (125 MHz, CDCl3) δ: 55.3, 58.8, 65.4, 68.5, 72.9, 75.2, 75.7, 77.6, 79.0, 79.4, 87.4, 101.5, 113.8, 126.1, 127.6, 128.2, 128.8, 129.1, 129.2, 129.4, 130.2, 131.6, 133.7, 134.5, 137.5, 159.3 ESIHRMS Calcd for $C_{30}H_{30}O_6S$ [M+Na]⁺: 541.1661. Found 541.1658.

Phenyl 4,6-O-benzylidene-2-O-(prop-2-ynyl)-1-thio-α**-D-mannopyranoside (12)**

To stirred solution of 11 (0.47 g, 0.91 mmol) in CH₂Cl₂ (8 mL) and water (0.4 mL) was added DDQ (0.3 g, 1.3 mmol) at room temp. After 3 h, sat. NaHCO₃ was added, and the mixture was extracted with $CH₂Cl₂$. The extract was washed several times with sat NaHCO₃, and dried over Na₂SO₄. Evaporation of the solvent in vacuo gave an oil, which was chromatographed on a flash silica gel column (hexane:ethyl acetate; 4:1) to give **12** (0.34 g, 93%) as a white solid MP 128 °C [α]²⁷_D + 119 (*c*, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl3) δ 2.49 (t, *J* = 2.4 Hz, 1H), 2.5 (bs, 1H), 3.84 (t, *J* = 10.2 Hz, 1H), 3.9 (t, *J* = 9.6 Hz, 1H), 4.16 (dd, *J* = 3.6, 10.0 Hz, 1H), 4.21–4.24 (m, 2H), 4.27–4.32 (m, 1H), 4.34 (dd, *J* = 2.4, 16.1 Hz, 1H), 4.42 (dd, *J* = 2.4, 16.1 Hz, 1H), 5.59 (s, 1H), 5.68 (s, 1H), 7.32– 7.53 (m, 10H); 13C NMR (125 MHz, CDCl3) δ: 58.6, 64.7, 68.4, 68.9, 75.7, 78.9, 79.3, 79.4, 86.4, 102.2, 126.3, 127.7, 128.3, 129.2, 131.7, 133.8, 137.2. ESIHRMS Calcd for $C_{22}H_{22}O_5S$ [M+Na]⁺: 421.1086. Found 421.1095.

Phenyl 4,6-O-benzylidene-2-O-(prop-2-ynyl)-3-O-(2,3-di-O-benzyl-4,6-O-benzylidene-β**-Dmannopyranosyl)-1-thio-**α**-D-mannopyranoside 29**β **and the** α**-anomer 29**α

To a stirred solution of donor **28** (480 mg, 0.88 mmol), BSP (223 mg 1.06 mmol), TTBP (331 mg, 1.33 mmol), and 4 Å molecular sieves in CH₂Cl₂ (5 mL), at −60 °C under an Ar atmosphere, was added Tf₂O (195 μ L 1.15 mmol). After 30 min. the temperature was brought down to -78 °C, and then acceptor **12** (424 mg 1.06 mmol) in CH₂Cl₂ (3 mL), was

slowly added. The reaction mixture was stirred for 2h. at −78 ºC, and quenched by the addition of triethylphosphite (435 μ L, 2.7 mmol), and continued stirring for 1 h at −78 °C and allowed to reached room temperature. The reaction mixture was diluted with CH_2Cl_2 (10 mL) and molecular sieves were filtered off and washed with saturated NaHCO₃. The organic layer was separated, dried and concentrated. The crude was purified by radial chromatography (hexane:ethyl acetate; 8:1) to give **29**β and **29**α in 83% and 5% yield respectively. **29β:** [α]²⁴_D + 26.3 (*c*, 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ : 2.23 (t, *J* = 2.4 Hz, 1H), 3.29–3.34 (m, 1H), 3.6 (dd, *J* = 3.2, 9.7 Hz, 1H), 3.86 (t, *J* = 10.3 Hz, 1H), 3.93 (t, *J* = 10.3 Hz, 1H), 4.0 (d, *J* = 3.0 Hz, 1H), 4.13 (t, *J* = 9.7 Hz, 1H), 4.25–4.40 (m, 8H), 4.65 (d, *J* = 12.5, Hz, 1H), 4.76 (d, *J* = 12.5 Hz, 1H), 4.84 (s, 1H), 4.86 (d, *J* = 11.9 Hz, 1H), 4.98 (d, *J* = 11.8 Hz, 1H), 5.58 (s, 1H), 5.62 (s, 1H), 5.64 (s, 1H), 7.24–7.49 (m, 25H); 13C NMR (125 MHz, CDCl₃) δ: 57.5, 65.3, 67.8, 68.5, 68.6, 72.3, 73.4, 74.8, 75.6, 75.7, 76.5, 77.5, 77.6, 78.6, 79.0, 86.0, 98.9, 101.3, 101.9, 126.0, 126.2, 127.4, 127.6, 127.7, 127.8, 128.0, 128.1, 128.2, 128.3, 128.4, 128.8, 129.0, 129.2, 131.6, 133.6, 137.3, 137.6, 138.4, 138.7. ESIHRMS Calcd for C₄₉H₄₈O₁₀S [M+Na]⁺: 851.2866. Found 851.2875. **29α:** [α]²⁴_D + 76.4 (*c*, 1.0, CHCl3); 1H NMR (500 MHz, CDCl3) δ : 2.4 (t, *J* = 2.4 Hz, 1H), 3.8 (t, *J* = 10.5 Hz, 1H), 3.92–3.96 (m, 2H), 3.99–4.04 (m, 2H), 4.15 (t, *J* = 9.5 Hz, 1H), 4.2 (dd, *J* = 2.5, 16.1 Hz, 1H), 4.25–4.38 (m, 7H), 4.5 (d, *J* = 12.4 Hz, 1H), 4.62 (d, *J* = 12.4 Hz, 1H), 4.63 (d, *J* = 12.2 Hz, 1H), 4.7 (d, *J* = 12.2 Hz, 1H), 5.4 (d, *J* = 1.2 Hz, 1H), 5.57 (s, 1H), 5.59 (s, 1H), 5.67 (s, 1H), 7.15–7.52 (m, 25H); ¹³C NMR (125 MHz, CDCl₃) δ: 58.3, 64.7, 65.1, 68.5, 68.8, 72.1, 72.7, 72.8, 75.3, 75.7, 78.5, 79.0, 79.2, 86.9, 99.6, 101.4, 101.9, 125.9, 126.1, 127.5, 127.6, 127.7, 127.8, 128.2, 128.4, 128.8, 129.2, 129.3, 131.6, 133.5, 137.3, 137.7, 137.8, 138.5. ESIHRMS Calcd for C₄₉H₄₈O₁₀S [M+Na]⁺: 851.2866. Found 851.2874.

General procedure for deprotection of propargyl ethers

To a stirred solution of propargyl ether (1 mmol) in dry THF (5 mL) was added KO^tBu (1.1 mmol), stirring was continued at room temperature for 3–12 h untill TLC indicated completion. The reaction mixture was diluted with CH_2Cl_2 (10 mL). The organic phase was separated, washed with water, dried $(Na₂SO₄)$ and concentrated on a rotary evaporator to give the allenyl ethers in quantitative yields. A homogeneous solution of allenyl ethers (1 mmol) in acetone: water (4:1, 5 mL) was treated with $OsO₄$ (0.1 mmol) and *N*-methyl morpholine *N*-oxide (2 mmol) and the mixture was stirred for 3 h at room temperature. After completion of the reaction, acetone was removed under vaccum and the residue was dissolved in CH_2Cl_2 (10 mL) and washed with sat. NaHSO₃. The organic phase was separated, dried (Na_2SO_4) , and concentrated on a rotary evaporator. The residues were purified by flash or radial chromatography on silica gel to yield deprotected di and trisaccharides in 80–91%.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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Figure 1. Staggered Conformations about the C3-O3 Bond

Figure 2. Staggered Conformations about the C2-O2 Bond

Scheme 1. Synthesis of Donors **10** and **13**

Scheme 2. Synthesis of Acceptor **27**

Scheme 3. Synthesis of a Mannotriose using 2- *O*-Propargyl Ethers

Scheme 4. Synthesis of Donors **33**, **35**, and **36**

Scheme 5. Preparation of Rhamnosyl Donor **64** .

Scheme 6. Glycosylation in the Rhamnopyranose Series

Influence of the O2 Protecting Group on Selectivity

Steric A-Values and p*K*a's

RO $\frac{K}{100}$ RO

a) Measurement temp.

b) Equilbrium const.

 c_l) A = RTln*K*

Further Couplings to Donor **13**

Coupling of Mono and Di-*O*-proparyl Protected Donors to **14**

Coupling of **33** to Further Acceptors

Acceptor	Product
	(Yield, ratio)
HO, BnO BnO BnO OMe 41	Ph C BnO B_{BnO}^- BnO OMe 45 (90 %, β : α = 25:1)
OMe HO 23	Ph OMe BnO 46 (93%, $\beta:\alpha = 25:1$)
ÒН 42	Ph BnC 47 (91%, β : α = 20:1) C O
HO 'n 43	Ph [®] BnO 48 (90%, β : α = 23:1)
ΟН 44	Ph BnO 49 92 %, $\beta:\alpha = 22:1$

Cleavage of Propargyl Ethers.

