Original article:

ANTIHYPERCHOLESTEROLEMIC EFFECT OF *CLEOME ARABICA L* ON HIGH CHOLESTEROL DIET INDUCED DAMAGE IN RATS

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ABSTRACT

Dietary cholesterol is known to be one of the main risk factors that accelerate oxidation process leading to hypercholesterolemia and attendant cardiovascular diseases. The purpose of this study, carried out on adult male Wistar rats, was to evaluate the inhibitory effects of supplementation with aqueous of *Cleome arabica* leaf extract on hypercholesterolemia. After 3 months of treatment, animals were sacrificed by decapitation. Blood serum was obtained by centrifugation. Under our experimental conditions, administration of *Cleome arabica* leaf extract decreased the total cholesterol (TC), LDL-cholesterol (LDL-chol) and triglycerides (TG) levels by 27 %, 52 %, 37 %, respectively, and reduced SGOT, SGPT, LDH and PAL levels in blood serum compared to untreated hypercholesterolemic rats. TBARS concentrations decreased by 21 % in liver, 22 % in heart and 30 % in kidney in a group of rats treated with cholesterol and *Cleome arabica* (Chol C.ar) compared to a Chol-treated group. The same treatment with *Cleome arabica* leaf extract increased superoxide dismutase and enhanced glutathione peroxidase activity. Catalase activity was found to increase in liver, heart and kidney by 17 %, 16 % and 23 %, respectively, in the C.ar Chol-treated group. The protective effect of *Cleome arabica* on hypercholesterolemia inducing oxidative stress in several organs was mainly attributed to antioxidant properties. The latter were due to the presence of phenolic acids and flavonoids shown by the obtained HPLC profiles.

Keywords: hypercholesterolemic rat, *Cleome arabica*, lipid peroxidation (TBARS), antioxidant enzymes

INTRODUCTION

Hypercholesterolemia is one of the major risk factors for cardiovascular diseases. It also causes oxidative stress resulting in increased lipid peroxidation in multiple organs (Cai et al., 2014). Previous studies also indicate that hypercholesterolemia is causally linked to a significant increase in reactive oxygen species with concomitant decrease in thiol levels in heart tissues (Sudhahar et al., 2007; Ouslimani et al., 2005). Hypercholesterolemic diet leads to various spontaneous arterial vasoconstrictions including renal artery vasoconstriction associated with decreased glomerular filtration rate in normal rats (Campos et al., 1999). Moreover, severe liver damage, perturbations of cholesterol metabolism and hyperglycemia have been associated with hypercholesterolemia.

Over the past several years, a growing interest has been focused on dietary regimen and, more particularly, natural products because of their capacity to reduce toxicity as well as their protective effects on oxidative stress and related health problems including age-associated diseases and cancer.

In this respect, Cleome arabica has attracted a great deal of attention in recent studies conducting antioxidant-related research. This plant belongs to the family Capparaceae which has several species widely grown in tropical and subtropical regions of the world, more specifically in North Africa (Ladhari et al., 2013). It has been investigated and shown to serve several important activities for medical purposes such as treatment of inflammation (Tsichritzis et al., 1993), rheumatism (Ahmad et al., 1990) and cytotoxicity (Nagaya et al., 1997). Cleome arabica has been shown to scavenge free radical species and modulate lipid peroxidation levels by stimulating the defense system (Selloum et al., 1997). Cleome arabica contains a large variety of antioxidant compounds such as syringic acid, vanillic acid, quercetin and kaempferol (Ladhari et al., 2013).

The main objective of the present experimental study is to investigate whether a *Cleome arabica*-enriched dietary regimen exerts a preventive effect on hypercholesterolemia inducing hepatic, renal and cardiac toxicity.

Serum TC levels, LDL, HDL-cholesterol (HDL-chol), TG,SGOT, SGPT, LDH, PAL, urea, uric acid, creatinine, lipid peroxidation level and activities of antioxidant enzymes (catalase, superoxide-dismutase and glutathione-peroxidase) in liver, heart and kidney were measured.

A phytochemical study of the *Cleome arabica* leaf extract was carried out in order to identify some antioxidative substances.

MATERIAL AND METHODS

Preparation of the aqueous Cleome arabica leaf extract

The leaves and stems of *Cleome arabica* were collected from the Gafsa region located in south-western Tunisia. They were dried in the shade at a research laboratory for one week then ground. Extraction of *Cleome arabica* was carried out by immersing 5 g of the plant in 100 ml distilled water for 5 to 10 min followed by filtration and gavage administration.

Experimental rats and diets

Healthy adult male rats of the Wistar strain (n=28) weighing 100 ± 5 g were used for the present study. The animals were purchased from Central Pharmacy of Tunisia, Tunis, Tunisia. The animals were randomly divided into four groups of seven rats each. Each group was housed in a spacious indoor cage with a relative humidity of 60 % at room temperature of 24 °C. Before treatment, rats were allowed to adapt to their new environment for 7 days. The animals were kept under photoperiods (a 12 h light/dark cycle) for 3 months.¹ The rats were fed for 3 months as follows:

- Group 1: control group (T) fed with normal diet.
- Group 2: Chol group fed with 1 % cholesterol-enriched diet in combination with 0.5 % cholic acid(Chol).
- Group 3:C.ar group fed with normal diet + *Cleome arabica* (C.ar) lyophilized aqueous extract (0.5 %).
- Group 4: Chol C.ar group fed with 1 % cholesterol-enriched diet in combination with 0.5 % cholic acid + *Cleome arabica* (C.ar) lyophilized aqueous extract (0.5 %) (Chol+C.ar).

¹ All the experimental procedures were carried out in compliance with Guidelines on the Use of Living Animals in Scientific Investigations.

Blood samples

After 3 months, rats from each group were rapidly sacrificed by decapitation in order to minimize the handling stress. Blood samples were collected from jugular vein in dried tubes and centrifuged at 1500 g for 15 min at 4 °C. Serum samples were collected for biochemical analyses to determine the TC rate, HDL-chol, LDL-chol and TG. Liver, heart and kidney were also quickly excised, cleaned of fat and stored at -80 °C until use.

Extraction and analysis of phenolic ' compounds

 $\hat{2}$ g of the air-dried plant material were mixed with 10 ml of 80 % aqueous methanol (v/v) and gently agitated at room temperature for 10 min, followed by a centrifugation for 5 min at 12000 rpm. Another successive extraction with 0,5 ml of supernatant and 0,5 ml of acetone was conducted at room temperature for 30 min.

HPLC analysis conditions

A Varian Prostar HPLC system equipped with a ProStar 230 ternary pump, a manual injector and a ProStar 330 diode array detector were used. The chromatographic analyses were performed on a 5 µm particle C-18 reversed-phase column (Varian, 250×4.6 mm). Final chromatographic conditions were a gradient elution made up of solvent A: 0.05 % acetic acid aqueous solution and solvent B: methanol. At t=0, the mobile phase consisted of 65 % A and 35 % B, and it changed with a linear gradient to 50 % A and 50 % B in 30 min. A change in the methanol under observation occurred during each 5min interval until it reached 20% A and 80 % B within a total duration of HPLCbased analysis of 40 min. The flow rate amounted to 1 ml/min with an injection volume of 20 µl at 25 °C. UV detection was performed at 365 nm for four flavonoids (luteolin, quercetin, kaempferol and isorhamnetin) and at 290 nm of phenolic acids. Identification of such compounds was carried out by comparing the retention time and mass

spectra of the peaks in the injected sample extracts to those of HPLC standard compounds.

Biochemical assays

The total cholesterol level, HDL, LDLchol, TG as well as activities of glutamicoxaloacetic transaminase (SGOT), glutamicpyruvic transaminase (SGPT), alkaline phosphatase (PAL), lactate dehydrogenase (LDH), urea, uric acid and creatinine (Crea) were determined by kit methods (Spinreact).

In accordance with Yagi (1976), the lipid peroxidation level was measured as thiobarbituric acid reactive substances (TBARS). In this study, 125 µl of supernatant (S1 liver, heart or kidney) were mixed with 175 µl of 20 % trichloroacetic acid containing 1 % butyl-hydroxytoluene and centrifuged ($1000 \times g$, 10 min, 4 °C). Then, 200 µl of supernatant (S2) was mixed with 40 μ l of HCl (0.6 M) and 160 μ l of thiobarbituric acid (0.72 mM), and the mixture was heated at 80 °C for 10 min. Absorbance was measured at 530 nm. The amount of TBARS was calculated using an extinction coefficient of $156 \text{ mM}^{-1} \text{ cm}^{-1}$ and expressed in nmol/mg protein.

Superoxide dismutase (SOD) activity was determined by measuring its ability to inhibit the photoreduction of nitroblue tetrazolium (NBT) using Hara et al.,'s method (1972). One unit of SOD represents the amount capable of inhibiting the photoreduction of NBT by 50 %. The activity was expressed as units/mg protein, at 25 °C.

Glutathione peroxidase (GPx) was determined by the method of Flohe and Gunzler (1984). One unit of GPx was defined as oxidation by H_2O_2 of 1 µmol of reduced glutathione peroxide per min at a pH of 7 and a temperature of 25 °C.

Catalase (CAT) activity was measured according to Aebi(1984). The reaction mixture (1 ml) contained 100 mM phosphate buffer (pH=7), 100 mM H₂O₂ and 20 μ l (about 1-1.5 mg of protein) of liver, heart and kidney. H₂O₂ decomposition at 25 °C was followed by measuring the decrease in absorbance at 240 nm for 1 min. Enzyme activity was calculated using an extinction coefficient of 0.043 mM^{-1} cm⁻¹ and expressed in international units (I.U.), i.e. in µmoles H₂O₂ destroyed/min/mg protein. Tissue extract protein content was determined according to Lowry's method (Lowry et al., 1951) using bovine serum albumin as standard.

Statistical analysis

Results are expressed as mean \pm standard deviation (SD) and the statistical significance was assessed by Student's t-test. P < 0.05 was considered statistically significant. The limit of statistical significance was set at P < 0.05 between both the *Cleome arabica*-treated and untreated groups.

RESULTS

Body weight, food and cholesterol intake

In this study, as shown in Table 1, the dietary cholesterol induced an increase in body weight compared to the control group (T) (P < 0.05) after a 3-month treatment.

Total cholesterol, LDL, HDL-chol and triglycerides in blood serum

The obtained results showed a significant increase in the total cholesterol, LDL-chol and triglycerides levels, and a significant decrease in HDL-chol in the hypercholesterolemic rats compared to controls. After three months of treatment with *Cleome arabica* leaf extract, these biomarkers were found to revert to almost control values (Table 1).

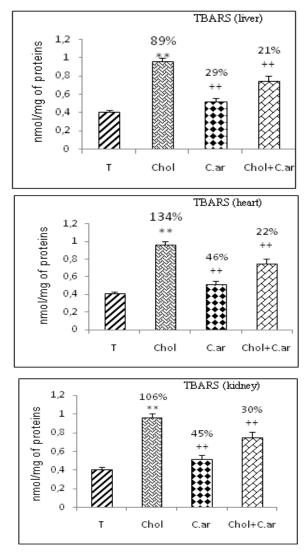


Figure 1: Lipid peroxidation level (expressed as TBARS, nmol/mg protein) in liver, heart and kidney after 3 months of treatment in controls (T), (Chol) cholesterol-treated rats, (Chol C.ar) cholesterol-treated rats supplemented with *Cleome arabica* leaf extract, (C.ar) rats supplemented with *Cleome arabica* leaf extract

 Table 1: Body weight, TC, HDL, LDL-chol and TG in Control (T) and experimental rats after 3 months of treatment

Rats	т	Chol	C.ar	Chol+C.ar
Body weight(g)	158,14 ± 5,01	275,57 ± 4,32**	158,54 ± 3,28 ⁺⁺	$160,85 \pm 4,16^{++}$
TC(mmol/L)	$1,59 \pm 0,06$	2,35 ± 0,08**	$1,52 \pm 0,02^{++}$	$1,69 \pm 0,02^{++}$
HDL-chol(mmol/L)	0,67 ± 0,01	0,34 ± 0,02**	$0,68 \pm 0,03^{++}$	$0,48 \pm 0,02^{++}$
LDL-chol(mmol/L)	0,71 ± 0,03	1,72 ± 0,10**	$0,77 \pm 0,01^{++}$	$1,08 \pm 0,04^+$
TG(mmol/L)	$0,66 \pm 0,06$	1,34 ± 0,05**	$0,67 \pm 0,05^{++}$	$0,63 \pm 0,11^{++}$

Values are mean ± SEM of 7 rats per group. Both groups were fed the cholesterol-enriched diet and either treated (Chol C.ar) or untreated (Chol) with *Cleome arabica*.

** P < 0.01 compared with control (T); $^+$ P < 0.05 compared with Chol; $^{++}$ P < 0.01 compared with Chol.

Serum markers of liver and kidney damage

Glutamic-oxaloacetic transaminase(SGOT), glutamic-pyruvic transaminase (SGPT), alkalinephosphatase (ALP) and lactate dehydrogenase (LDH) were released into the blood when certain organs or tissues were injured. As shown in Figure 1, these activities were significantly greater in hypercholesterolemic rats than in controls (T). Hypercholesterolemia also induced a clearly noticeable kidney damage due to a significant increase in urea, uric acid and creatinine levels in serum (Figure 2). All these biomarkers reverted to almost normal values in the case of Cleome arabica-treated hypercholesterolemic rats.

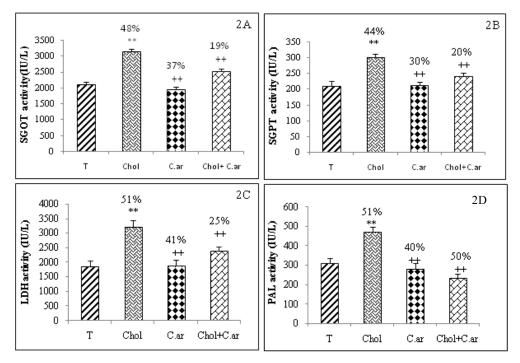


Figure 2: Effect of hypercholesterolemia and Cleome arabica on SGOT (2A), SGPT (2B), LDH (2C) and PAL (2D) activities.

Values are mean ± SEM; n = 7

Chol compared to control group (T): P < 0.05; ** $P \le 0.01$ compared to control group (T) compared to Chol group: P < 0.05; ⁺⁺ $P \le 0.01$ compared to hypercholesterolemic group (Chol)

Oxidative damage

In the case of rats receiving dietary cholesterol, TBARS levels in hepatic, cardiac and renal tissues increased by 89%, 134% and 106 %, respectively, when compared to controls (Figure 3). The administration of Cleome arabica leaf extract resulted in a significant reduction in these TBARS levels.

Antioxidant activities

The enzyme activities that had a protective effect against ROS, i.e. CAT and GPx, were found to decrease by 26 %, 22 % in liver, 16 %, 21 % in heart and by 31 %, 41 % in kidney. Table 2 shows that SOD activity decreased by 47 % in liver, 19 % in heart and by 26% in kidney of hypercholesterolemic rats compared to controls. These changes, which revealed a failing defense against oxidative stress, were largely corrected in animals treated with Cleome arabica.

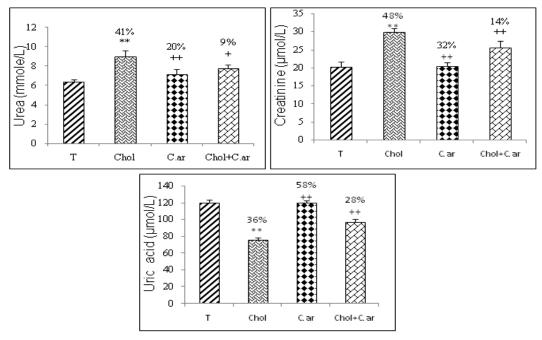


Figure 3: Effects of hypercholesterolemic diet, *Cleome arabica* on urea (mmol/L), creatinine (μ mol/L) and uric acid (μ mol/L) levels in blood.

Values are the mean of 7 measurements ± SD.

*P < 0.05 compared with control (T); **P < 0.01 compared with control

⁺ P < 0.05 compared with hypercholesterolemic rats (Chol); ⁺⁺P < 0.01 compared with (Chol).

Enzymes	Т	Chol	C.ar	Chol+C.ar
Liver (IU/mg protein) GPx SOD CAT	15,91 ± 1,31 1, 51 ± 0,01 165,14 ± 2,05	12,26 ± 0,34 ^{**} 0,79 ± 0,03 ^{**} 120,97 ± 1,07 ^{**}	15,28 ± 1,30 ⁺⁺ 1,52 ± 0,05 ⁺⁺ 167 ± 1,51 ⁺⁺	$15,35 \pm 1,14^{++}$ 1, 35 ± 0,03 ⁺ 142 ± 1,92 ⁺
Heart (IU/mg protein) GPx SOD CAT	13,32 ± 1,15 9,17 ± 0,38 823,06 ± 12,01	$10,40 \pm 0,07^{**}$ 7,39 ± 0,40 ^{**} 684,87 ± 17,07 ^{**}	13,50 ± 1,20 ⁺⁺ 8,17 ± 0,5 ⁺⁺ 817,18 ± 12,51 ⁺⁺	$13,09 \pm 1,67^{+}$ 7,65 ± 0,19 795,36 ± 15,92 ⁺⁺
Kidney(IU/mg protein) GPx SOD CAT	11,13 ± 0,09 8,42 ± 0,07 330,78 ± 3,15	$6,48 \pm 0,40^{14}$ $6,23 \pm 0,05^{14}$ $227,41 \pm 4,75^{14}$	$\begin{array}{c} 10,8 \pm 0,98^{++} \\ 7,67 \pm 0,14^{++} \\ 300 \pm 3,51^{++} \end{array}$	$8,43 \pm 0,29^{++}$ $6,13 \pm 0,09^{+}$ $280,60 \pm 2,01^{++}$

Table 2: Antioxidant enzyme activity in hepatic, cardiac and renal tissues

Values are means \pm SEM of 7 rats per group. Both groups were fed the cholesterol-enriched diet and either treated (C.ar +HC) or untreated (Chol) with *Cleome arabica*.

*P < 0.05 compared with control (T); **P < 0.01 compared with control (T)

⁺P < 0.05 compared with hypercholesterolemic rats (Chol); ⁺⁺P < 0.01 compared with (Chol).

HPLC analysis of Cleome arabica phenolic acids and flavonoids

As shown in Figure 4, the HPLC analysis of *Cleome arabica* phenolic acids in 290 nm revealed the presence of phenolic acids. Three phenolic acids were identified in FPA: protocatechic, gallic, vanillic and syringic acids. The HPLC elution profile of flavonoids (Figure 5) also illustrated seven peaks of unknown compounds and four flavonoids in 365 nm: quercetin, luteolin, kaempferol, isorhamnetin. Phenolic acids and flavonoids contributed to the beneficial effect of *Cleome arabica* leaf extract *in vivo*.

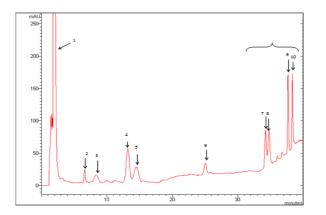


Figure 4: HPLC elution profile of phenolic acids (λ = 290 nm) from lyophilized aqueous extract of *Cleome arabica*

Peaks (1) acetone peak, (2) protocatechic acid, (3) gallic acid, (4) 4-hydroxybenzoic acid, (5) vanillic acid, (6) syringic acid, (7, 8, 9 and 10) unknown compounds. The HPLC analyses were performed using a Varian ProStar HPLC equipped with a reverse phase C-18 column (Varian, 150 mm×4.6 mm, particle size 5 μ m) in conjunction with a gradient elution (A, 0.05 % CH₃COOH aqueous solution; B, methanol): t₀ = 65 %A and 35 %B; 30 min = 50 %A and 50 % B; 40 min = 20 %A and 80 %B. The flow rate was 1 ml min⁻¹ and the injection volume was 20 μ l at 25 C.

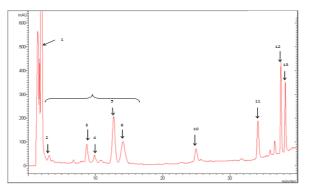


Figure 5: HPLC elution profile of flavonoids (λ = 365 nm) from lyophilized aqueous *Cleome arabica* leaf extract

Peaks (1) acetone peak, (2–6) unknown compounds, (10) quercetin, (11) luteolin, (12) kaempferol, (13) isorhamnetin. The HPLC analysis were performed using a Varian ProStar HPLC equipped with a reverse phase C-18 column (Varian, 150 mm×4.6 mm, particle size 5 μ m) in conjunction with a gradient elution (A, 0.05 % CH₃COOH aqueous solution; B, methanol): t₀ = 65 %A and 35 %B; 30 min = 50 %A and 50 %B; 40 min = 20 %A and 80 % B.

DISCUSSION

The aim of this experimental study was to determine the protective effect of Cleome arabica on hypercholesterolemia in liver, heart and kidney. The biochemical parameters and activities of antioxidant enzymes were determined. The obtained results indicated that hypercholesterolemia led to statistically significant changes in biochemical parameters. It was found that an increase in total plasma cholesterol, LDL-chol, TG, SGOT, SGPT, LDH, PAL, urea and creatinine occurred with a decrease in HDL-chol and uric acid value compared to control group. Hypercholesterolemia caused elevated cholesterol level in serum as well as in multiple cells such as erythrocytes, platelets and endothelial cells. This increased cholesterol level was found to activate these cells and lead to enhanced production of reactive oxygen species (ROS) (Kamesh and Sumathi. 2012). This mechanism induced alteration of vascular function, cell proliferation and apoptotic/neurotic process. Louala et al., (2013) and Milagro et al. (2006) reported that hypercholesterolemia increases lipid peroxidation in liver cells then stimulates lipogenesis in adipose tissue. Several studies have revealed that high cholesterol level can cause cardiac damage (Johnkennedy et al., 2011). Markedly elevated level of serum cardiac marker enzymes such as SGOT and LDH in HDF-induced rats is due to peroxide formation generated by hypercholesterolemia in the form of ROS (Sudhahar et al., 2007). Hypercholesterolemia also contributes to renal, glomerular, interstitial and vascular damage. In the present study, it was found that hypercholesterolemia did not only alter the liver, cardiac and renal biomarkers but also inhibited antioxidative defense mechanisms. In fact, the lipid peroxidation levels increased with decrease in SOD, CAT and GPx levels in heart, liver and kidney due to hypercholesterolemia. This led to aggravation of oxidative stress by reducing cellular glutathione levels, as observed in previous studies (Cai et al., 2014).

Findings reported in several studies suggest that hypercholesterolemia-induced organ damage is probably associated with the accumulation of reactive oxygen species (Li et al., 2014). Other findings relate hypercholesterolemia-induced damage in tissues to overproduction of free radicals and ROS resulting in enhanced lipid peroxidation, damage to DNA, protein degradation, and exhaustion of the antioxidative defense systems (Kamesh and Sumathi, 2012).

Under our experimental conditions, Cleome arabica leaf extract administered by gavage corrected all the investigated parameters that had shifted to pathological values due to hypercholesterolemia. The decrease in TC, LDL-chol and TG levels in serum by 27 %,52 %, 37 %, respectively, compared to untreated hypercholesterolemic rats could be explained by the regulation of lipid metabolism. As has already been reported in previous research, Cleome arabica has an antioxidant activity owing to the presence of its phenolics and flavonoid compounds. Moreover, Cleome arabica protected liver, heart and kidney functions. It enabled reversion of these major organs' dysfunction indices such as SGOT, SGPT, LDH, PAL, urea, uric acid and creatinine activities almost back to normal values.

A similar result was obtained by supplementation with certain dietary antioxidants rich in polyphenols and flavonoids extracted from *Cleome arabica* (Djeridane et al., 2010). Indeed, flavonoids can increase HDLchol and decrease LDL cholesterol oxidation. The defense mechanism against hypercholesterolemia consists of both antioxidants synthesized in the tissues and exogenous antioxidants supplied with diet. Interestingly, our results have demonstrated that *Cleome arabica* exerts a beneficial effect on body

weight of hypercholesterolemic rats. This inhibitory activity by Cleome arabica leaf extract against oxidative stress could be explained by the presence of polyphenols and flavonoids in Cleome arabica leaf extract. This result is in agreement with previous data reporting that *Cleome arabica* leaf extract contains various flavonoids such as luteolin, kaempferol and isorhamnetin (Ladhari et al., 2013). Actually, flavonoids existing in Cleome arabica leaf extract reduced lipid peroxidation and enhanced the expression of intracellular endogenous antioxidants such as SOD, GPx and catalase by maintaining their activities at a higher level than cholesteroltreated rats. This is due to the ability of such enzymes to scavenge free radicals, as confirmed by Cai et al. (2014). Furthermore, the polyphenols and flavonoids present in Cleome arabica leaf extract can be considered as a highly effective antioxidant able to protect biological systems against the oxidative stress known as an important pathophysiological factor for a variety of diseases including hypercholesterolemia, cancer, diabetes, cardiovascular disorders and rheumatoid arthritis (Unnikrishnan et al., 2014; Duchnowicz et al., 2012).

Under our experimental conditions, *Cleome arabica* was found to be a valuable source of natural health-promoting antioxidants and ingredients such as phenolic acids (protocatechic acid, gallic acid, 4-hydroxybenzoic acid, vanillic acid, syringic acid and flavonoids (quercetin, luteolin, kaempferol, isorhamnetin). A number of studies have highlighted the relation between the chemical structure of phenolic acids and their ability to reduce free radicals by the release of a proton from the hydroxyl group against oxidative stress inducing damage in multiple tissues (Hfaiedh et al., 2014).

In conclusion, the strong therapeutic effect and potential of *Cleome arabica* are arguably associated with its potent hypocholesterolemic properties. *Cleome arabica* was found to be able to significantly achieve almost the same degree of improvement in cholesterol-related pathological conditions in rat liver, heart and kidney.

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Competing interests

The authors declare that they have no competing interests.

REFERENCES

Aebi H. Catalase in vitro. Methods Enzymol. 1984; 105:121-6.

Ahmad I, Malik MI, Iqbal K, Ahmad K, Naz S. Efficacy of formalinized liver organ vaccine against Angara disease in broilers. Veterinary Arhiv. 1990;60: 131-8.

Cai J, Yang L, He HJ, Xu T, Liu HB, Wu Q et al. Antioxidant capacity responsible for a hypocholesterolemia is independent of dietary cholesterol in adult rats fed rice protein. Gene. 2014;533:57-66.

Campos SB, Ori M, Dórea EL, Seguro AC. Protective effect of L-arginine on hypercholesterolemia-enhanced renal ischemic injury. Atherosclerosis. 1999;143: 327-34.

Djeridane A, Yousfi M, Brunel JM, Stocker P. Isolation and characterization of a new steroid derivative as a powerful antioxidant from *Cleome arabica* in screening the in vitro antioxidant capacity of 18 Algerian medicinal plants. Food Chem Toxicol. 2010;48: 2599-606.

Duchnowicz P, Bors M, Podsędek A, Koter-Michalak M, Broncel M.Effect of polyphenols extracted from Brassica vegetables on erythrocyte membranes (in vitro study). Environ Toxicol Pharmacol.2012;34: 783-90.

Flohe L, Gunzler WA. Assays of glutathione peroxidase. Methods Enzymol. 1984;105:114-21.

Hara P,Misra, Fridovich I. The role of superoxide anion in the autooxidation of epinephrine and a simple assay for superoxide dismutase. Biol Chem. 1972; 247:3170-5.

Hfaiedh N, Dhibi S, Mbarki S, Murat JC, Elfeki AF. Beneficial effects of Fenugreek seeds (*Trigonella foenum graecum L*) added in the diet of diabetic rats. J Adv Chem 2014; 8:1556-65.

Johnkennedy N, Adamma E, Nnedimma NC. Hypolipidemic effects of aqueous extract of *Acalypha capitata* leaves in rats fed on high cholesterol diet. Asian Pac J Trop Biomed. 2011;1(Suppl 2):S183-5.

Kamesh V, Sumathi T.Antihypercholesterolemic effect of *Bacopa monniera* linn. on high cholesterol diet induced hypercholesterolemia in rats. Asian Pac J Trop Med. 2012;949:949-55.

Ladhari A,Omezzine F, DellaGreca M, Zarrelli A, Zuppolini S. Phytotoxic activity of Cleome arabica L. and its principal discovered active compounds. South Afr J Bot2013;88:341-51.

Li H, Horke S, Förstermann U.Vascular oxidative stress, nitric oxide, and atherosclerosis. Atherosclerosis.2014;237:208-19.

Louala S, Benyahia-Mostefaoui A, Lamri-Senhadji MY. Energy restriction reduces oxidative stress in the aorta and heart and corrects the atherogenic risk in obese rat. Ann Cardiol Angeiol (Paris). 2013;62:155-60.

Lowry OH, Rosenbrough NJ, Farr AL, Randall R. Protein measurement with the folin phenol reagent. Biol Chem. 1951;193:265-75.

Milagro I, Campion J, Martinez JA.Weight gain induced by high-fat feeding involves increased liver oxidative stress. Obesity. 2006;14:1118-23.

Nagaya H, Tobita Y, Nagae T, Itokawa H, Takaya K, Halim AF et al. Cytotoxic triterpenes from *Cleome africana*. Phytochemistry 1997;44:1115-9.

Ouslimani N, Peynet J, Bonnefont-Rousselot D, Therond P, Legrand A, Beaudeux JL. Metformin decreases intracellular production of reactive oxygen species in aortic endothelial cells. Metabolism 2005;54:829-34.

Selloum L, Sebihi L ,Mekhalfia A, Mahdadi R, Senator A.Antioxidant activity of *Cleome arabica* leaves extract. Biochem Soc Trans.1997;25:S608.

Sudhahar V, Ashok-Kumar S, Mythili Y, Varalakshmi P. Remedial effect of lupeol and its ester derivative on hypercholesterolemia-induced oxidative and inflammatory. Nutr Res. 2007;27:778-87. Tsichritzis F, Abdel-Mogip M, Jakupovic J. Dammarane triterpènes from *Cleome africana*. Phytochemistry. 1993;33:423-5.

Unnikrishnan MK, Veerapur V, Nayak Y, Mudgal PP, Mathew G. Antidiabetic, antihyperlipidemic and antioxidant effects of the flavonoids. In: Watson RR, Preedy VR, Zibadi S (eds.): Polyphenols in human health and disease, Vol. 1: Polyphenols in chronic diseases and their mechanisms of action (pp 143-61). San Diego, CA: Academic Press, 2014.

Yagi K. A simple fluorometric assay for lipoperoxide in blood plasma. Biochem Med.1976;15:212-6.