

REVIEW

The peroxisome proliferator-activated receptors in cardiovascular diseases: experimental benefits and clinical challenges

Wai San Cheang¹, Xiao Yu Tian², Wing Tak Wong² and Yu Huang¹

¹Shenzhen Research Institute, Institute of Vascular Medicine and Li Ka Shing Institute of Health Sciences, Chinese University of Hong Kong, Hong Kong, China, and ²Department of Cardiovascular Sciences, Houston Methodist Research Institute, Houston, TX, USA

Correspondence

Yu Huang, School of Biomedical Sciences, Chinese University of Hong Kong, Hong Kong, China.
E-mail: yu-huang@cuhk.edu.hk

Received

20 August 2014

Revised

24 October 2014

Accepted

20 November 2014

The peroxisome proliferator-activated receptors, PPAR α , PPAR β/δ and PPAR γ , are ligand-activated transcriptional factors belonging to the nuclear receptors superfamily and they are known to play important roles in glucose and lipid metabolism. Experimental studies in animal models of metabolic diseases have also revealed that activation of PPARs protects against the vascular complications of diabetes, hypertension, atherosclerosis, myocardial infarction and stroke, through exerting their anti-inflammatory, anti-atherogenic and antioxidant effects. In clinical trials and post-market surveillance, agonists of PPARs have been shown to effectively prevent cardiovascular events. However, adverse effects, particularly for PPAR γ agonists, are also observed with the use of investigational PPAR agonists and even some approved drugs. Further exploration of underlying mechanisms is needed to develop novel ways of PPAR activation without causing serious side effects. This article reviews the cardiovascular effects of PPARs, with emphasis on the therapeutic potential of PPAR agonists in combating metabolic vascular diseases.

LINKED ARTICLES

This article is part of a themed section on Chinese Innovation in Cardiovascular Drug Discovery. To view the other articles in this section visit <http://dx.doi.org/10.1111/bph.2015.172.issue-23>

Abbreviations

Ang II, angiotensin II; EC, endothelial cell; RXR, retinoid X receptor; TCM, traditional Chinese medicine; VSMC, vascular smooth muscle cell

Tables of Links

TARGETS
Nuclear hormone receptors^a
PPAR α , NR1C1
PPAR β/δ , NR1C2
PPAR γ , NR1C3
RXR, retinoid X receptor
Enzymes^b
Akt
Casein kinase 2
Cdk5, cyclin-dependent kinase 5
Endothelial NOS
Glycogen synthase kinase-3
GPCR^c
5-HT _{2B} receptor

LIGANDS	
15-deoxy- $\Delta^{12,14}$ -PGJ ₂	GW2331
5-hydroxy-L-tryptophan	Imiglitazar
9-HODE, 9-hydroxyoctadecadienoic acid	LTB ₄
Alelitazar	LY465608
Angiotensin II	Metformin
Bezafibrate	MK-767
Ciprofibrate	PGD ₂
Clofibrate	Pioglitazone
Endothelin-1	Prostacyclin
Fenofibrate	Ragaglitazar
Gemfibrozil	Rosiglitazone
GW0742	Troglitazone
GW1516	VEGF

These Tables list key protein targets and ligands in this article which are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson *et al.*, 2014) and are permanently archived in the Concise Guide to PHARMACOLOGY 2013/14 (^{a,b,c}Alexander *et al.*, 2013a,b,c).

Introduction

The physiological and pathophysiological roles of the nuclear receptors, PPAR α , PPAR β/δ and PPAR γ , have been much less studied in terms of their effects on the vascular system, compared with their functions in glucose and lipid metabolism. PPARs are expressed in endothelial cells (ECs), vascular smooth muscle cells (VSMCs) and macrophages. Activation of PPARs in the vasculature is mostly anti-inflammatory and antioxidant. It is therefore protective against endothelial dysfunction, atherosclerosis and vascular remodelling *in vivo*. The ongoing clinical use or trials of synthetic PPAR agonists support the predicted vascular benefits of PPAR activation although some adverse effects have been reported. This review will focus on how PPAR α , PPAR β/δ and PPAR γ regulate vascular function through different targets in ECs, VSMCs and macrophages. In particular, we wish to provide an overview of the clinically relevant aspects and controversies of the use of PPAR agonists for the treatment of cardiovascular complications associated with metabolic disorders. This information increases the potential therapeutic value of PPAR agonists against vascular dysfunction and justifies further development of novel PPAR agonists.

Transcriptional regulation and physiological functions of PPARs

PPARs heterodimerize with retinoid X receptors (RXR) and bind to specific DNA regions of target genes (AGGTCAXAG-GTCA, with X being a random nucleotide) termed as peroxisome proliferator hormone response elements (Gearing

et al., 1993). PPAR-RXR heterodimers remain bound with co-repressor protein complex in the absence of ligand but ligand activation triggers conformational changes of PPAR-RXR which releases the co-repressor complex and, subsequently, the heterodimer assembles with co-activators to activate the transcription of target genes (DiRenzo *et al.*, 1997). Furthermore, activated PPARs can interact with other transcription factors, such as AP-1 and NF- κ B, resulting in transcriptional repression (Delerive *et al.*, 1999). These transcriptional regulation and key functions of PPARs are summarized in Figure 1.

The endogenous ligands for PPARs are fatty acids and eicosanoids. Both PPAR α and PPAR δ are activated by a wide variety of saturated and unsaturated fatty acids while PPAR γ interacts only with polyunsaturated fatty acids (Xu *et al.*, 1999). Differences also exist between eicosanoids as activators of PPARs. For instance, LTB₄ and 8(S)-hydroxyeicosanoid, 5,8,11,13-tetraenoic acid (8(S)-HETE) are PPAR α ligands; 9- and 13-hydroxyoctadecadienoic acids (9- and 13-HODEs), and 15-deoxy- $\Delta^{12,14}$ -PGJ₂ are agonists for PPAR γ whereas prostacyclin, PGA₁, and PGD₂ are agonists for PPAR δ (Forman *et al.*, 1997; Kliewer *et al.*, 1997).

PPARs have different levels of expression in different tissues. PPAR α is predominantly expressed in cells with high rates of fatty acid catabolism such as liver, heart, kidney and skeletal muscle (Braissant *et al.*, 1996) to regulate fatty acid oxidation systems. PPAR γ is mainly associated with adipose tissue to regulate adipocyte differentiation and thus, lipid and energy storage (Rosen and Spiegelman, 2001). PPAR δ is ubiquitously expressed at much higher levels than PPAR γ and PPAR α , markedly in brain, adipose tissue and skin (Kliewer *et al.*, 1992). It is implicated in fatty acid oxidation and energy dissipation, improving lipid profiles and reducing adiposity (Wang *et al.*, 2003).

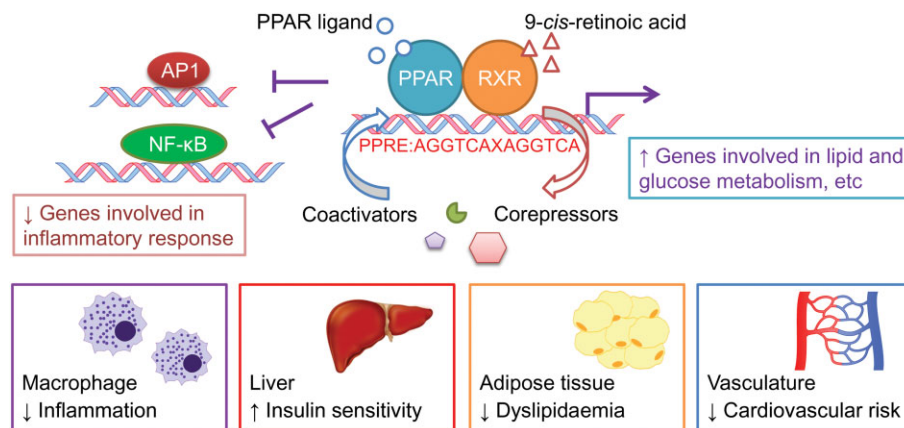


Figure 1

Transcriptional regulation and key functions of PPAR family. Ligand binding to PPAR recruits coactivators in replacement of corepressors to induce transcription and inhibits other transcriptional factors such as AP-1 and NF- κ B, regulating different functions.

Cardiovascular effects of PPAR α activation and deletion

PPAR α activation limits inflammatory responses in ECs and VSMCs (Plutzky, 2011) as well as macrophages (Neve *et al.*, 2001) by inhibiting NF- κ B and AP-1 and improves lipid profile, thus contributing to the anti-atherogenic action of PPAR α agonists. PPAR α inhibits the activities of NF- κ B and AP-1 through direct interactions with p65 and c-Jun subunits, respectively (Delerive *et al.*, 1999), or through induction of I κ B α which sequesters NF- κ B in the cytoplasm (Delerive *et al.*, 2000). PPAR α agonists were developed to treat dyslipidaemia. For example, fibric acid derivatives (fibrates), including clofibrate, ciprofibrate, fenofibrate and gemfibrozil, reduce triglyceride and raise high-density lipoprotein (HDL) with modest effect to lower low-density lipoprotein (Remick *et al.*, 2008). Fenofibrate retards the development of atherosclerosis in ApoE $^{-/-}$ and LDLR $^{-/-}$ mice (Duez *et al.*, 2002; Srivastava *et al.*, 2006). However, atherosclerosis develops more slowly in PPAR α $^{-/-}$ mice (Tordjman *et al.*, 2007), while another study shows that PPAR α deficiency in macrophages increases atherosclerosis (Babaev *et al.*, 2007). Such conflicting data may raise concerns over the relevance of the choice of mouse models used to reveal the roles of molecular targets in the development of atherosclerosis. Using tissue-specific gene deletion, mice are probably better than global knockout mice as more selective activity can be investigated with less effects on the whole body. Deletion of PPAR α further increased the plasma level of triglyceride in ApoE $^{-/-}$ mice which might cause increased atherosclerosis (Babaev *et al.*, 2007). Interestingly, fenofibrate exerts some PPAR α -independent actions in human microvascular ECs, reducing endothelin-1 expression via inhibition of glycogen synthase kinase-3 (Glineur *et al.*, 2013).

Clinical perspective of PPAR α agonists

Several clinical studies evaluated the vascular effects of fibrates. In these trials, gemfibrozil alone reduces cardiovas-

cular events such as coronary heart disease, myocardial infarction and stroke in type 2 diabetic patients without any statin use (Asztalos *et al.*, 2008). By contrast, the more potent PPAR α agonist, fenofibrate, shows no overall benefit in reducing macrovascular events in diabetic patients (Group *et al.*, 2010). On the other hand, fenofibrate prevents microvascular complications such as diabetic retinopathy (Chen *et al.*, 2013). This agonist also ameliorates coronary flow velocity reserve and arterial stiffness in patients with hypertriglyceridaemia (Wang *et al.*, 2013). It appears that fenofibrate may preferentially regulate those cellular targets which control for microvascular dysfunction. In particular, many clinical trials have found that fibrates are more effective in patients with high triglyceride and low-HDL cholesterol (Fruchart *et al.*, 1998; Tenkanen *et al.*, 2006), both of which are risk factors for cardiovascular diseases, implying a higher effectiveness to protect vascular function in such patients. Patients with high triglyceride and low-HDL cholesterol have not been the basis of an entire clinical trial, so the vasoprotective effect of fibrates may have been underestimated in the trials so far (Tenkanen *et al.*, 2006; Scott *et al.*, 2009).

Four fibrates are available for clinical application to treat hyperlipidaemia. The US Food and Drug Administration (FDA) approved fenofibrate and gemfibrozil but not ciprofibrate and bezafibrate while the latter two are available elsewhere except in the United States. Treating mice and rats with fibrates increases incidence of hepatocellular carcinogenesis but no significant toxic or carcinogenic effects have been reported in patients (Gonzalez and Shah, 2008). The underlying mechanism attributed to this differential effect among species is currently unclear but the different levels of PPAR α expression might play a role. Notably, mouse liver expresses about 10-fold more PPAR α than human liver, implying that PPAR α signalling might be much stronger in mice and thus recruit more molecular targets to cause more severe toxicity. Furthermore, the hepatocarcinogenicity of fibrates in rodents may be independent of their binding to PPAR. Fibrates are known to affect NADH cytochrome c reductase activity, disrupting mitochondrial electron respiratory chain (Scatena *et al.*, 2003). Because peroxisomal

β -oxidation is more active in rodents than in humans (Perrone *et al.*, 1998), this process may be a factor in the higher level of hepatocarcinogenicity in rodents. Identifying the mechanisms behind adverse drug effects is inherently difficult but new synthetic agonists should be modified to be more targeted, with minimal side effects. Correct dosing is also a critical factor but it is important to achieve the beneficial effects without severe side effects. More potent and specific PPAR α agonists including AVE8134, DRF10945, GW590735 and LY518674 have been developed and currently under investigation in phase II trials.

PPAR γ linking to cardiovascular complications in diabetes and obesity

Extensive studies support that PPAR γ activation by thiazolidinediones (TZDs), such as troglitazone, rosiglitazone and pioglitazone, confers vascular benefits in diabetic animals. Pioglitazone (Huang *et al.*, 2008) and rosiglitazone (Wong *et al.*, 2011) improve endothelial function in diabetic and obese mice by stimulating NO production and lowering oxidative stress. Existing *in vivo* data indicate that PPAR γ activation by both pharmacological ligands (Chen *et al.*, 2001) and genetic overexpression (Hu *et al.*, 2010) attenuates the occurrence of atherosclerosis in ApoE^{-/-} mice. PPAR γ activation in monocytes/macrophages is anti-inflammatory by reducing inflammatory molecules and by inducing the generation of IL-1 receptor antagonist (Jiang *et al.*, 1998; Meier *et al.*, 2002). VSMC-specific PPAR γ knockout aggravates angiotensin II (Ang II)-induced vascular remodelling and impairment of endothelium-dependent relaxations (Marchesi *et al.*, 2012), atherosclerosis (Chang *et al.*, 2012) and vascular calcification (Woldt *et al.*, 2012) in mice, thus revealing a vasoprotective property of PPAR γ agonists.

TZDs suppress the vasoconstrictor effects of endothelin-1 (Tian *et al.*, 2010), Ang II (Yuen *et al.*, 2011) and 5-HT_{2B} receptor agonists (Liu *et al.*, 2012), suggesting a promising strategy to delay the development of hypertension and vascular dysfunction. However, contradictory findings on BP regulation

are reported in animals with VSMC-selective PPAR γ deletion. Hypertension [systolic BP: ~130 vs. 120 mmHg (day) and ~145 vs. 135 mmHg (night) for transgenic vs. control mice] (Halabi *et al.*, 2008) was accompanied by increased RhoA and Rho kinase activity (Pelham *et al.*, 2012). However, in another study, hypotension (systolic: 96 \pm 27 vs. 123 \pm 14 mmHg and diastolic: 84 \pm 15 vs. 102 \pm 14 mmHg for knockout vs. control mice in daytime) was described, accompanied with reduced vasoconstriction and increased vasodilatation in aortas (Chang *et al.*, 2009). These contradictory results between nuclear receptor deficiency and agonists might be due to compensation mechanisms and/or the absence of active repression of gene transcription by unliganded nuclear receptors. Apart from these considerations, the discrepancies between findings of genetic deletion and pharmacological activation may be due to the PPAR-independent activities of PPAR ligands, summarized in Table 1. Pioglitazone restores blood flow recovery in ischaemic muscle of diabetic mice via the Akt-VEGF pathway (Biscetti *et al.*, 2009). Troglitazone causes dephosphorylation of endothelial NOS at Ser¹¹⁶ to increase endothelial NO (Cho *et al.*, 2004), inhibits ECs proliferation by suppressing casein kinase 2 (Lee *et al.*, 2006b) and triggers apoptosis of VSMCs by activating the Gadd45 and p53 pathway (Okura *et al.*, 2000).

Rosiglitazone increases angiogenesis and functional recovery after focal cerebral ischaemia (Chu *et al.*, 2006) and pioglitazone enhances angiogenesis and blood flow recovery in hind limb ischaemia in diabetic mice through activating endothelial NO synthesis (Huang *et al.*, 2008). Besides, PPAR γ activation favours angiogenesis through increasing endothelial progenitor cell (EPC) formation (Tousoulis *et al.*, 2008) and inhibiting EPC apoptosis (Zhang *et al.*, 2013). Of interest, EC-specific PPAR γ ^{-/-} mice show dyslipidaemia and rosiglitazone therapy is unable to lower serum levels of free fatty acid and triglyceride (Kanda *et al.*, 2009), implying that the PPAR γ in ECs plays a significant role in lipid metabolism which may in turn affect vascular functions; and thus endothelial PPAR γ could be a therapeutic target.

The promising data on vasoprotective effects of TZDs in animal studies led to several clinical trials in patients. A meta-analysis confirmed the prevention of diabetes-

Table 1

PPAR-independent effects of clinically used PPAR ligands in the vascular system

Agonist	Model	Pathway	Effect	Reference
PPAR α				
Fenofibrate	Human microvascular ECs (HMEC-1)	Inhibit glycogen synthase kinase-3	Reduce endothelin-1 expression	Glineur <i>et al.</i> , 2013
PPAR γ				
Pioglitazone	Streptozotocin-diabetic mice with hind limb ischaemia	Stimulate Akt-VEGF pathway	Improve blood flow recovery in ischaemic muscle	Biscetti <i>et al.</i> , 2009
Troglitazone	Bovine aortic endothelial cells	Induce endothelial NOS dephosphorylation at Ser ¹¹⁶	Increase NO production	Cho <i>et al.</i> , 2004
	HUVECs	Suppress casein kinase 2 activity	Inhibit proliferation of ECs	Lee <i>et al.</i> , 2006b
	Rat aortic VSMCs	Activate Gadd45 and p53 pathway	Induce apoptosis of VSMCs	Okura <i>et al.</i> , 2000

associated vascular complications by pioglitazone (Lincoff *et al.*, 2007) and another clinical study also shows that pioglitazone promoted endothelial function in non-diabetic patients at high cardiovascular risk (Campia *et al.*, 2006). In the PROactive clinical trial, pioglitazone reduced non-fatal myocardial infarction and stroke, but it did not affect the risk of coronary revascularization while increasing leg revascularization in type 2 diabetic patients (Wilcox *et al.*, 2008). Pioglitazone also improved aortic elasticity and decreased inflammation and disease progression, with minimal safety concerns, in patients with rheumatoid arthritis (Marder *et al.*, 2013). A number of clinical studies now show that TZDs reduce BP independent of their insulin-sensitizing properties, suggesting a direct vascular benefit (Giles and Sander, 2007). In addition, PPAR γ agonists inhibit atherosclerosis in diabetic patients (Nissen *et al.*, 2008).

Deciphering the adverse effects of PPAR γ agonists

TZDs were launched in the late 1990s, as an effective means of restoring insulin sensitivity in type 2 diabetes. However, troglitazone was removed from the market in 2000 because of liver failure in diabetic patients (Kohlroser *et al.*, 2000). Rosiglitazone and pioglitazone currently remain marketed but their application is seriously questioned as adverse effects have been reported. Meta-analyses from randomized trials and Medicare databases show that rosiglitazone was associated with increased risk of stroke, myocardial infarction, heart failure and death (Nissen and Wolski, 2007; Graham *et al.*, 2010). As a result, rosiglitazone has also been withdrawn from Europe by the European Medicines Agency. However, the FDA removed restrictions in November 2013, based on all available data including the re-adjudicated RECORD trial showing no increase of cardiovascular incidence with rosiglitazone therapy (Mahaffey *et al.*, 2013). Likewise, pioglitazone has been associated with bone fracture, heart failure and bladder cancer (Vallarino *et al.*, 2013). Meanwhile, new selective PPAR γ modulators are being synthesized and tested for potential therapeutic efficacy and safety. For example, a PPAR γ ligand SR1664 improved insulin sensitivity without causing significant adverse effects on fluid retention or bone formation (Choi *et al.*, 2011). SR1664, unlike classical PPAR γ agonists, has a unique mode of binding to PPAR γ with high affinity, blocking the Cdk5-mediated phosphorylation of PPAR γ but not acting as a transcriptional agonist. This difference in action might explain the fewer side effects of this drug. It may therefore be possible to develop a new class of antidiabetic drug with fewer side effects by specifically targeting the Cdk-mediated phosphorylation of PPAR γ . Additionally, the development of tissue-specific agonists may enhance the therapeutic benefits and reduce deleterious effects.

PPAR δ in vascular dysfunction

Several PPAR δ agonists consistently correct dyslipidaemia and improve insulin and glucose tolerance in animal models of

obesity and diabetes (Wang *et al.*, 2003; Lee *et al.*, 2006a). Dyslipidaemia and inflammation in macrophage are common contributors to the development of atherosclerosis. PPAR δ agonists favourably modulate lipid metabolism and attenuate inflammation, thus reducing the susceptibility to atherosclerosis in both ApoE $^{-/-}$ and LDLR $^{-/-}$ mice (Barish *et al.*, 2008; Bojic *et al.*, 2014). Conversely, PPAR $\delta^{-/-}$ bone marrow transplants reduce atherosclerotic lesion area in LDLR $^{-/-}$ mice because of the elevated availability of inflammatory suppressor Bcl-6 (Lee *et al.*, 2003). This discrepancy is probably related to the different actions of PPAR δ activation in different cell types.

PPAR δ agonists directly induce endothelium-dependent relaxation when used at higher concentrations (Jimenez *et al.*, 2010) and can restore endothelial function in animal models of both type 1 (Quintela *et al.*, 2012) and type 2 (Tian *et al.*, 2012) diabetes through increasing NO bioavailability and suppressing generation of NADPH oxidase-derived superoxide anions. In hypertensive rats, GW0742 prevented the development of endothelial dysfunction and hypertension by enhancing endothelial NOS activity, and expression of antioxidant genes and regulators of G protein-coupled signalling proteins (RGS) 5, limiting NADPH oxidase activity and reducing expressions of proinflammatory and pro-atherogenic genes (Zarzuelo *et al.*, 2011; 2013). A recent study also suggested that PPAR δ plays a crucial role for vasoprotective effect of the commonly used antidiabetic drug metformin in obese mice (Cheang *et al.*, 2014).

Clinical translation of PPAR δ agonists

Clinical trials on selected PPAR δ agonists have assessed both metabolic and vascular outcomes. No severe side effects have been so far reported, except for GW1516 which induced cancer in several organs in rodents (Gupta *et al.*, 2004; Pollock *et al.*, 2010) and its development has been stopped. MBX-8025, another PPAR δ agonist, normalized lipid profiles and C-reactive protein levels in patients with the metabolic syndrome and has completed a phase II clinical trial (Choi *et al.*, 2012). More long-term clinical studies are needed to further evaluate the safety profile before any of these PPAR δ ligands can be marketed for clinical treatment of metabolic and vascular complications. To conclude, protective effects of PPAR δ agonists against metabolic and vascular diseases are strongly indicated by result from animal models but, unlike the other isoforms, none of the PPAR δ agonists has been approved for clinical use. Furthermore, PPAR δ is ubiquitously expressed and any differential mechanism of PPAR δ action in different tissues should be explored for developing new PPAR δ agonists with improved efficacy and safety. Developing such agonists with high tissue specificity may reduce adverse effects.

Dual PPAR agonism

Because both fibrates and TZDs exerted favourable effects on lipid and glucose metabolism as well as inhibitory effects on

vascular inflammation, interest has grown to develop and explore the clinical outcomes of dual PPAR α/γ agonists. Indeed, dual PPAR α/γ agonists such as ragaglitazar (Chakrabarti *et al.*, 2003) and muraglitazar (Mittra *et al.*, 2007) provide better glycaemic control than conventional TZDs. Unfortunately, the development of diverse dual PPAR α/γ agonists has not met with the anticipated success. Their development was halted in late-phase trials because of reported side effects such as increased cardiovascular risk (muraglitazar), carcinogenicity (ragaglitazar and MK-767), liver toxicity (imiglitazar) and renal injury (tesaglitazar) (Fievet *et al.*, 2006). The effect of dual PPAR α/γ agonists on atherogenesis is now also being debated. For instance, GW2331 (Claudel *et al.*, 2001), LY465608 (Zuckerman *et al.*, 2002) and tesaglitazar (Zadelaar *et al.*, 2006) attenuated atherosclerosis in ApoE^{-/-} mice however, treatment with compound 3q ((S)-3-(4-(2-carbazol(phenoxazin)-9-yl-ethoxy)phenyl)-2-ethoxy-propionic acid) increases atherosclerotic plaque formation associated with an increased oxidative stress in these mice (Calkin *et al.*, 2007). The reasons for these disparate findings are unclear, possibly related to the different potency of these drugs to activate PPAR α and PPAR γ together with differential transcriptional outcomes. For instance, muraglitazar and tesaglitazar preferentially act on PPAR γ , possibly accounting for some of the adverse effects. Therefore, a balanced PPAR α/γ agonist, aleglitazar, was developed, but disappointingly again, its development is terminated at phase III AleCardio trial because of safety concerns with increased risk for bone fractures, heart failure and gastrointestinal bleeding (Lincoff *et al.*, 2013). This outcome has strongly discouraged the further development of safer dual PPAR α/γ ligands. Dual PPAR α/γ agonists primarily acting on PPAR α may provide another option in the reduction of side effects. On the other hand, dual PPAR α/δ agonists are less investigated with GFT505 being shown to favourably modulate lipid and glucose metabolism in patients with metabolic syndrome (Cariou *et al.*, 2011). However, it has only completed a phase II clinical trial, so its safety profile remains to be evaluated.

Pan-PPAR activator

Bezafibrate is a pan-PPAR activator activating all three PPAR subtypes. It has been approved by the FDA for the treatment of hyperlipidaemia, despite the reports, in some countries, of renal and hepatobiliary disorders. With beneficial effects on glucose and insulin tolerance, bezafibrate decreased the incidence of and delayed the onset of type 2 diabetes in patients with impaired fasting glucose concentrations (Tenenbaum *et al.*, 2004) and in obese patients (Tenenbaum *et al.*, 2005a). Moreover, bezafibrate decreased the risk of myocardial infarction and cardiac mortality in patients with metabolic syndrome and with a history of recent myocardial infarction and/or stable angina (Tenenbaum *et al.*, 2005b). The benefits of bezafibrate appear to justify the development of newer pharmacological agents targeting more than one PPAR isoform. Consequently, several pan-PPAR agonists were synthesized. However, the majority of them (DRL 11605, GW-625019, indeglitazar, netoglitazone, sipoglitazar and

sodelglitazar) have been terminated because of serious safety concerns.

Traditional Chinese compounds acting as PPAR ligands

Traditional Chinese medicine (TCM) compounds have long been used against metabolic disease and the related cardiovascular complications. TCM is an attractive resource in the design of new PPAR agonists to reduce cardiovascular risks. Screening the TCM database containing more than 30 000 candidates, two TCM compounds, (S)-tryptophan-betaxanthin and berberrubine, have been identified as potential lead compounds targeting more than one PPAR (Chen *et al.*, 2012). By *in silico* identification, two other TCM candidates, 5-hydroxy-L-tryptophan and abrine, were found to bind to PPAR γ (Chen and Chen, 2014). In addition, honokiol from TCM Magnolia bark was also *in silico* predicted to bind to the ligand-binding domain of PPAR γ and prevented hyperglycaemia in diabetic mice (Atanasov *et al.*, 2013). Formononetin, the predominant component of San-ao decoction, which is one of the most popular TCM formulae for asthma, was demonstrated to activate PPAR γ (Zhou *et al.*, 2009). Through their action on PPARs, such TCM compounds may prevent inflammatory diseases, hyperglycaemia and cardiovascular diseases, apart from their originally known therapeutic functions. They warrant further investigation as novel therapeutic agents. More importantly, TCM is deployed as many different combinations of compounds to treat human diseases and the investigation of the active components together with mechanisms of action is crucial for drug development. The efficacy and safety of identified compounds can be ensured and improved using modern analytical and chemical techniques.

Conclusion

The results of many animal studies taken together have demonstrated that activation of PPAR α , PPAR γ and PPAR δ with their corresponding agonists produced clear vasoprotective effects, including inhibition of vascular inflammation, oxidative stress and atherogenic development. These findings stimulated intense interest in using PPAR agonists as another therapeutic approach to vascular dysfunction. Disappointingly, a number of drugs targeting PPARs have failed clinical trials because of safety concerns. Only drugs that activate PPAR α (ciprofibrate, fenofibrate and gemfibrozil) and PPAR γ (pioglitazone and rosiglitazone) and one pan-PPAR agonist (bezafibrate) have reached the market, so far. PPAR α agonists and bezafibrate are available for the treatment of hyperlipidaemia associated with metabolic syndrome while PPAR γ agonists are used to correct hyperglycaemia in type 2 diabetic patients. Clinical studies reveal that several PPAR agonists have not achieved the expected levels of cardiovascular benefits or that the benefits are significantly offset by unwanted outcomes such as malignancy, liver and renal toxicity and increased cardiovascular risk. The cardiovascular benefits and side effects are summarized in Figure 2. Despite these

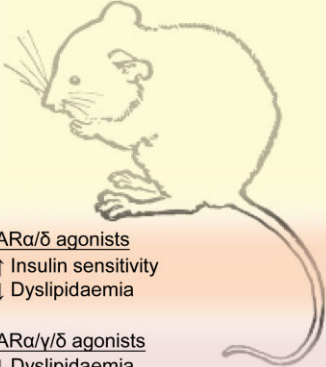

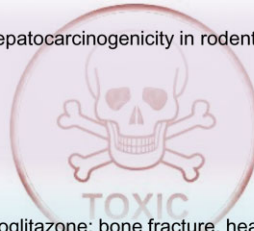
Experimental benefits	Clinical benefits	Side effects
<p>PPARα agonists</p> <ul style="list-style-type: none"> • ↓ Dyslipidaemia • ↓ Oxidative stress • ↓ Inflammation • ↓ Cardiac remodelling in hypertension • ↓ Blood pressure • ↓ Atherosclerosis <p>PPARγ agonists</p> <ul style="list-style-type: none"> • ↑ Insulin sensitivity • ↑ NO, vasorelaxation • ↓ Ang II and ET-1, vasoconstriction • ↓ Oxidative stress • ↓ Inflammation • ↓ Aneurysm formation • ↓ Pulmonary arterial hypertension • ↓ Atherosclerosis <p>PPARδ agonists</p> <ul style="list-style-type: none"> • ↓ Dyslipidaemia • ↓ Oxidative stress • ↓ Inflammation • ↑ NO, vasorelaxation • ↓ Hypertension • ↓ Atherosclerosis <p>PPARα/γ agonists</p> <ul style="list-style-type: none"> • ↑ Insulin sensitivity • ↓ Atherosclerosis <p>PPARα/δ agonists</p> <ul style="list-style-type: none"> • ↑ Insulin sensitivity • ↓ Dyslipidaemia <p>PPAR$\alpha/\gamma/\delta$ agonists</p> <ul style="list-style-type: none"> • ↓ Dyslipidaemia • ↑ Insulin sensitivity 	<p>• ↓ Dyslipidaemia</p> <p>• Fenofibrate: ↓ coronary flow velocity reverse, arterial stiffness, and diabetic retinopathy</p> <p>• Gemfibrozil: ↓ coronary heart disease, myocardial infarction and stroke</p> <p>• ↑ Insulin sensitivity</p> <p>• Pioglitazone: ↓ myocardial infarction, stroke, atherosclerosis and inflammation; ↑ aortic elasticity</p> <p>• ↓ Dyslipidaemia</p> <p>• ↑ Insulin sensitivity</p> <p>• ↑ Insulin sensitivity</p> <p>• ↓ Dyslipidaemia</p> <p>• ↑ Insulin sensitivity</p> <p>• ↓ Dyslipidaemia</p> <p>• ↑ Insulin sensitivity</p> <p>• Bezaifibrate: ↓ myocardial infarction and cardiac mortality</p> 	<p>• Hepatocarcinogenicity in rodents</p> <p>• Pioglitazone: bone fracture, heart failure and bladder cancer</p> <p>• Troglitazone: liver failure – removed from market, 2000</p> <p>• GW1516: cancers in organs in rodents – terminated at phase IV, 2006</p> <p>• Aleglitazar : bone fractures, heart failure and gastrointestinal bleeding – terminated at phase III, 2013</p> <p>• Imiglitazar: liver toxicity – terminated at phase III, 2005</p> <p>• MK-767: vascular tumours in mice – terminated at phase III, 2003</p> <p>• Muraglitazar: cardiovascular risk – terminated at phase III, 2006</p> <p>• Ragaglitazar: bladder tumours in mice – terminated at phase III, 2004</p> <p>• Tesaglitazar: kidney toxicity – terminated at phase III, 2006</p> <p>• Bezaifibrate: renal and hepatobiliary disorders</p> 

Figure 2

Cardiovascular effects of PPAR agonists in animal and clinical studies. The list shows the cardiovascular benefits shown in rodents (experimental benefits) and in patients with metabolic syndrome (clinical benefits) and the adverse effects of agonists for PPARs.

disadvantages, PPARs are important players in the control of lipid and glucose homeostasis. Therefore, new PPAR ligands are being developed, in the hope that these new drugs will produce fewer side effects. With deeper understanding of the regulatory mechanisms of PPAR activation, for example, co-repressor/co-activators of PPARs, epigenetic modifiers and intermediate metabolites that may activate certain enzymes or induce post-translational modification, new insights could be useful in developing novel or tissue-targeted PPAR agonists with more tolerable side effects for better prevention and

management of cardiovascular events associated with metabolic diseases.

Acknowledgements

This study is supported by Hong Kong Research Grants Council (CUHK2/CRF/12G and T12/402/13N) and Natural Science Foundation of China (91339117 and 2012CB517805).

Conflict of interest

None declared.

References

- Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013a). The Concise Guide to PHARMACOLOGY 2013/14: Nuclear Hormone Receptors. *Br J Pharmacol* 170: 1652–1675.
- Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013b). The Concise Guide to PHARMACOLOGY 2013/14: Enzymes. *Br J Pharmacol* 170: 1797–1867.
- Alexander SPH, Benson HE, Faccenda E, Pawson AJ, Sharman JL, Spedding M *et al.* (2013c). The Concise Guide to PHARMACOLOGY 2013/14: G Protein-Coupled Receptors. *Br J Pharmacol* 170: 1459–1581.
- Asztalos BF, Collins D, Horvath KV, Bloomfield HE, Robins SJ, Schaefer EJ (2008). Relation of gemfibrozil treatment and high-density lipoprotein subpopulation profile with cardiovascular events in the Veterans Affairs High-Density Lipoprotein Intervention Trial. *Metabolism* 57: 77–83.
- Atanasov AG, Wang JN, Gu SP, Bu J, Kramer MP, Baumgartner L *et al.* (2013). Honokiol: a non-adipogenic PPAR γ agonist from nature. *Biochim Biophys Acta* 1830: 4813–4819.
- Babaev VR, Ishiguro H, Ding L, Yancey PG, Dove DE, Kovacs WJ *et al.* (2007). Macrophage expression of peroxisome proliferator-activated receptor- α reduces atherosclerosis in low-density lipoprotein receptor-deficient mice. *Circulation* 116: 1404–1412.
- Barish GD, Atkins AR, Downes M, Olson P, Chong LW, Nelson M *et al.* (2008). PPAR δ regulates multiple proinflammatory pathways to suppress atherosclerosis. *Proc Natl Acad Sci U S A* 105: 4271–4276.
- Biscetti F, Straface G, Arena V, Stigliano E, Pecorini G, Rizzo P *et al.* (2009). Pioglitazone enhances collateral blood flow in ischemic hindlimb of diabetic mice through an Akt-dependent VEGF-mediated mechanism, regardless of PPAR γ stimulation. *Cardiovasc Diabetol* 8: 49.
- Bojic LA, Burke AC, Chhoker SS, Telford DE, Sutherland BG, Edwards JY *et al.* (2014). Peroxisome proliferator-activated receptor delta agonist GW1516 attenuates diet-induced aortic inflammation, insulin resistance, and atherosclerosis in low-density lipoprotein receptor knockout mice. *Arterioscler Thromb Vasc Biol* 34: 52–60.
- Braissant O, Fougere F, Scotto C, Dauca M, Wahli W (1996). Differential expression of peroxisome proliferator-activated receptors (PPARs): tissue distribution of PPAR- α , - β , and - γ in the adult rat. *Endocrinology* 137: 354–366.
- Calkin AC, Allen TJ, Lassila M, Tikellis C, Jandeleit-Dahm KA, Thomas MC (2007). Increased atherosclerosis following treatment with a dual PPAR agonist in the ApoE knockout mouse. *Atherosclerosis* 195: 17–22.
- Campia U, Matuskey LA, Panza JA (2006). Peroxisome proliferator-activated receptor- γ activation with pioglitazone improves endothelium-dependent dilation in nondiabetic patients with major cardiovascular risk factors. *Circulation* 113: 867–875.
- Cariou B, Zaïr Y, Staels B, Bruckert E (2011). Effects of the new dual PPAR α/δ agonist GFT505 on lipid and glucose homeostasis in abdominally obese patients with combined dyslipidemia or impaired glucose metabolism. *Diabetes Care* 34: 2008–2014.
- Chakrabarti R, Vikramadithyan RK, Misra P, Hiriyan J, Raichur S, Damarla RK *et al.* (2003). Ragaglitazar: a novel PPAR α PPAR γ agonist with potent lipid-lowering and insulin-sensitizing efficacy in animal models. *Br J Pharmacol* 140: 527–537.
- Chang L, Villacorta L, Zhang J, Garcia-Barrio MT, Yang K, Hamblin M *et al.* (2009). Vascular smooth muscle cell-selective peroxisome proliferator-activated receptor-deletion leads to hypotension. *Circulation* 119: 2161–2169.
- Chang L, Villacorta L, Li R, Hamblin M, Xu W, Dou C *et al.* (2012). Loss of perivascular adipose tissue on peroxisome proliferator-activated receptor- γ deletion in smooth muscle cells impairs intravascular thermoregulation and enhances atherosclerosis. *Circulation* 126: 1067–1078.
- Cheang WS, Tian XY, Wong WT, Lau CW, Lee SS, Chen ZY *et al.* (2014). Metformin protects endothelial function in diet-induced obese mice by inhibition of endoplasmic reticulum stress through 5' adenosine monophosphate-activated protein kinase-peroxisome proliferator-activated receptor δ pathway. *Arterioscler Thromb Vasc Biol* 34: 830–836.
- Chen KC, Chen CY (2014). *In silico* identification of potent PPAR- γ agonists from traditional Chinese medicine: a bioactivity prediction, virtual screening, and molecular dynamics study. *Evid Based Complement Alternat Med* 2014: 192452.
- Chen KC, Chang SS, Huang HJ, Lin TL, Wu YJ, Chen CY (2012). Three-in-one agonists for PPAR- α , PPAR- γ , and PPAR- δ from traditional Chinese medicine. *J Biomol Struct Dyn* 30: 662–683.
- Chen Y, Hu Y, Lin M, Jenkins AJ, Keech AC, Mott R *et al.* (2013). Therapeutic effects of PPAR α agonists on diabetic retinopathy in type 1 diabetes models. *Diabetes* 62: 261–272.
- Chen Z, Ishibashi S, Perrey S, Osuga J, Gotoda T, Kitamine T *et al.* (2001). Troglitazone inhibits atherosclerosis in apolipoprotein E-knockout mice: pleiotropic effects on CD36 expression and HDL. *Arterioscler Thromb Vasc Biol* 21: 372–377.
- Cho DH, Choi YJ, Jo SA, Jo I (2004). Nitric oxide production and regulation of endothelial nitric-oxide synthase phosphorylation by prolonged treatment with troglitazone: evidence for involvement of peroxisome proliferator-activated receptor (PPAR) γ -dependent and PPAR γ -independent signaling pathways. *J Biol Chem* 279: 2499–2506.
- Choi JH, Banks AS, Kamenecka TM, Busby SA, Chalmers MJ, Kumar N *et al.* (2011). Antidiabetic actions of a non-agonist PPAR γ ligand blocking Cdk5-mediated phosphorylation. *Nature* 477: 477–481.
- Choi YJ, Roberts BK, Wang X, Geaney JC, Naim S, Wojnoonski K *et al.* (2012). Effects of the PPAR- δ agonist MBX-8025 on atherogenic dyslipidemia. *Atherosclerosis* 220: 470–476.
- Chu K, Lee ST, Koo JS, Jung KH, Kim EH, Sinn DI *et al.* (2006). Peroxisome proliferator-activated receptor- γ -agonist, rosiglitazone, promotes angiogenesis after focal cerebral ischemia. *Brain Res* 1093: 208–218.
- Claudiel T, Leibowitz MD, Fievet C, Tailleux A, Wagner B, Repa JJ *et al.* (2001). Reduction of atherosclerosis in apolipoprotein E knockout mice by activation of the retinoid X receptor. *Proc Natl Acad Sci U S A* 98: 2610–2615.
- Delerive P, De Bosscher K, Besnard S, Vanden Berghe W, Peters JM, Gonzalez FJ *et al.* (1999). Peroxisome proliferator-activated receptor α negatively regulates the vascular inflammatory gene response by negative cross-talk with transcription factors NF- κ B and AP-1. *J Biol Chem* 274: 32048–32054.

- Delerive P, Gervois P, Fruchart JC, Staels B (2000). Induction of I κ B α expression as a mechanism contributing to the anti-inflammatory activities of peroxisome proliferator-activated receptor- α activators. *J Biol Chem* 275: 36703–36707.
- DiRenzo J, Soderstrom M, Kurokawa R, Ogliastro MH, Ricote M, Ingrey S *et al.* (1997). Peroxisome proliferator-activated receptors and retinoic acid receptors differentially control the interactions of retinoid X receptor heterodimers with ligands, coactivators, and corepressors. *Mol Cell Biol* 17: 2166–2176.
- Duez H, Chao YS, Hernandez M, Torpier G, Poulain P, Mundt S *et al.* (2002). Reduction of atherosclerosis by the peroxisome proliferator-activated receptor α agonist fenofibrate in mice. *J Biol Chem* 277: 48051–48057.
- Fievet C, Fruchart JC, Staels B (2006). PPAR α and PPAR γ dual agonists for the treatment of type 2 diabetes and the metabolic syndrome. *Curr Opin Pharmacol* 6: 606–614.
- Forman BM, Chen J, Evans RM (1997). Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors α and δ . *Proc Natl Acad Sci U S A* 94: 4312–4317.
- Fruchart JC, Brewer B, Leitersdorf E, Brochier M, Brown G, Calvo C *et al.* (1998). Consensus for the use of fibrates in the treatment of dyslipoproteinemia and coronary heart disease. *Am J Cardiol* 81: 912–917.
- Gearing KL, Gottlicher M, Teboul M, Widmark E, Gustafsson JA (1993). Interaction of the peroxisome-proliferator-activated receptor and retinoid X receptor. *Proc Natl Acad Sci U S A* 90: 1440–1444.
- Giles TD, Sander GE (2007). Effects of thiazolidinediones on blood pressure. *Curr Hypertens Rep* 9: 332–337.
- Glineur C, Gross B, Neve B, Rommens C, Chew GT, Martin-Nizard F *et al.* (2013). Fenofibrate inhibits endothelin-1 expression by peroxisome proliferator-activated receptor α -dependent and independent mechanisms in human endothelial cells. *Arterioscler Thromb Vasc Biol* 33: 621–628.
- Gonzalez FJ, Shah YM (2008). PPAR α : mechanism of species differences and hepatocarcinogenesis of peroxisome proliferators. *Toxicology* 246: 2–8.
- Graham DJ, Ouellet-Hellstrom R, MaCurdy TE, Ali F, Sholley C, Worrall C *et al.* (2010). Risk of acute myocardial infarction, stroke, heart failure, and death in elderly Medicare patients treated with rosiglitazone or pioglitazone. *JAMA* 304: 411–418.
- Group AS, Ginsberg HN, Elam MB, Lovato LC, Crouse JR 3rd, Leiter LA *et al.* (2010). Effects of combination lipid therapy in type 2 diabetes mellitus. *N Engl J Med* 362: 1563–1574.
- Gupta RA, Wang D, Katkuri S, Wang H, Dey SK, DuBois RN (2004). Activation of nuclear hormone receptor peroxisome proliferator-activated receptor- δ accelerates intestinal adenoma growth. *Nat Med* 10: 245–247.
- Halabi CM, Beyer AM, De Lange WJ, Keen HL, Baumbach GL, Faraci FM *et al.* (2008). Interference with PPAR γ function in smooth muscle causes vascular dysfunction and hypertension. *Cell Metab* 7: 215–226.
- Hu Q, Zhang XJ, Liu CX, Wang XP, Zhang Y (2010). PPAR γ 1-induced caveolin-1 enhances cholesterol efflux and attenuates atherosclerosis in apolipoprotein E-deficient mice. *J Vasc Res* 47: 69–79.
- Huang PH, Sata M, Nishimatsu H, Sumi M, Hirata Y, Nagai R (2008). Pioglitazone ameliorates endothelial dysfunction and restores ischemia-induced angiogenesis in diabetic mice. *Biomed Pharmacother* 62: 46–52.
- Jiang C, Ting AT, Seed B (1998). PPAR- γ agonists inhibit production of monocyte inflammatory cytokines. *Nature* 391: 82–86.
- Jimenez R, Sanchez M, Zarzuelo MJ, Romero M, Quintela AM, Lopez-Sepulveda R *et al.* (2010). Endothelium-dependent vasodilator effects of peroxisome proliferator-activated receptor β agonists via the phosphatidylinositol-3 kinase-Akt pathway. *J Pharmacol Exp Ther* 332: 554–561.
- Kanda T, Brown JD, Orasanu G, Vogel S, Gonzalez FJ, Sartoretto J *et al.* (2009). PPAR γ in the endothelium regulates metabolic responses to high-fat diet in mice. *J Clin Invest* 119: 110–124.
- Kliwer SA, Umeson K, Mangelsdorf DJ, Evans RM (1992). Retinoid X receptor interacts with nuclear receptors in retinoic acid, thyroid hormone and vitamin D3 signalling. *Nature* 355: 446–449.
- Kliwer SA, Sundseth SS, Jones SA, Brown PJ, Wisely GB, Koble CS *et al.* (1997). Fatty acids and eicosanoids regulate gene expression through direct interactions with peroxisome proliferator-activated receptors α and γ . *Proc Natl Acad Sci U S A* 94: 4318–4323.
- Kohlroser J, Mathai J, Reichheld J, Banner BF, Bonkovsky HL (2000). Hepatotoxicity due to troglitazone: report of two cases and review of adverse events reported to the United States Food and Drug Administration. *Am J Gastroenterol* 95: 272–276.
- Lee CH, Chawla A, Urbiztondo N, Liao D, Boisvert WA, Evans RM (2003). Transcriptional repression of atherogenic inflammation: modulation by PPAR δ . *Science* 302: 453–457.
- Lee CH, Olson P, Hevener A, Mehl I, Chong LW, Olefsky JM *et al.* (2006a). PPAR δ regulates glucose metabolism and insulin sensitivity. *Proc Natl Acad Sci U S A* 103: 3444–3449.
- Lee KS, Park JH, Lee S, Lim HJ, Jang Y, Park HY (2006b). Troglitazone inhibits endothelial cell proliferation through suppression of casein kinase 2 activity. *Biochem Biophys Res Commun* 346: 83–88.
- Lincoff AM, Wolski K, Nicholls SJ, Nissen SE (2007). Pioglitazone and risk of cardiovascular events in patients with type 2 diabetes mellitus: a meta-analysis of randomized trials. *JAMA* 298: 1180–1188.
- Lincoff AM, Tardif JC, Neal B, Nicholls SJ, Ryden L, Schwartz GG *et al.* (2013). Evaluation of the dual peroxisome proliferator-activated receptor α/γ agonist aleglitazar to reduce cardiovascular events in patients with acute coronary syndrome and type 2 diabetes mellitus: rationale and design of the AleCardio trial. *Am Heart J* 166: 429–434.
- Liu Y, Tian XY, Mao G, Fang X, Fung ML, Shyy JY *et al.* (2012). Peroxisome proliferator-activated receptor- γ ameliorates pulmonary arterial hypertension by inhibiting 5-hydroxytryptamine 2B receptor. *Hypertension* 60: 1471–1478.
- Mahaffey KW, Hafley G, Dickerson S, Burns S, Tourt-Uhlig S, White J *et al.* (2013). Results of a reevaluation of cardiovascular outcomes in the RECORD trial. *Am Heart J* 166: 240–249.e1.
- Marchesi C, Rehman A, Rautureau Y, Kasal DA, Briet M, Leibowitz A *et al.* (2012). Protective role of vascular smooth muscle cell PPAR in angiotensin II-induced vascular disease. *Cardiovasc Res* 97: 562–570.
- Marder W, Khalatbari S, Myles JD, Hench R, Lustig S, Yalavarthi S *et al.* (2013). The peroxisome proliferator activated receptor- γ pioglitazone improves vascular function and decreases disease activity in patients with rheumatoid arthritis. *J Am Heart Assoc* 2: e000441.

- Meier CA, Chicheportiche R, Juge-Aubry CE, Dreyer MG, Dayer JM (2002). Regulation of the interleukin-1 receptor antagonist in THP-1 cells by ligands of the peroxisome proliferator-activated receptor gamma. *Cytokine* 18: 320–328.
- Mitra S, Sangle G, Tandon R, Sharma S, Roy S, Khanna V *et al.* (2007). Increase in weight induced by muraglitazar, a dual PPARalpha/gamma agonist, in db/db mice: adipogenesis/or oedema? *Br J Pharmacol* 150: 480–487.
- Neve BP, Corseaux D, Chinetti G, Zawadzki C, Fruchart JC, Duriez P *et al.* (2001). PPARalpha agonists inhibit tissue factor expression in human monocytes and macrophages. *Circulation* 103: 207–212.
- Nissen SE, Wolski K (2007). Effect of rosiglitazone on the risk of myocardial infarction and death from cardiovascular causes. *N Engl J Med* 356: 2457–2471.
- Nissen SE, Nicholls SJ, Wolski K, Nesto R, Kupfer S, Perez A *et al.* (2008). Comparison of pioglitazone vs glimepiride on progression of coronary atherosclerosis in patients with type 2 diabetes: the PERISCOPE randomized controlled trial. *JAMA* 299: 1561–1573.
- Okura T, Nakamura M, Takata Y, Watanabe S, Kitami Y, Hiwada K (2000). Troglitazone induces apoptosis via the p53 and Gadd45 pathway in vascular smooth muscle cells. *Eur J Pharmacol* 407: 227–235.
- Pawson AJ, Sharman JL, Benson HE, Faccenda E, Alexander SP, Buneman OP *et al.*; NC-IUPHAR (2014). The IUPHAR/BPS Guide to PHARMACOLOGY: an expert-driven knowledge base of drug targets and their ligands. *Nucl. Acids Res.* 42 (Database Issue): D1098–1106.
- Pelham CJ, Ketsawatsomkron P, Groh S, Grobe JL, de Lange WJ, Ibeawuchi SR *et al.* (2012). Cullin-3 regulates vascular smooth muscle function and arterial blood pressure via PPAR γ and RhoA/Rho-kinase. *Cell Metab* 16: 462–472.
- Perrone CE, Shao L, Williams GM (1998). Effect of rodent hepatocarcinogenic peroxisome proliferators on fatty acyl-CoA oxidase, DNA synthesis, and apoptosis in cultured human and rat hepatocytes. *Toxicol Appl Pharmacol* 150: 277–286.
- Plutzky J (2011). The PPAR-RXR transcriptional complex in the vasculature: energy in the balance. *Circ Res* 108: 1002–1016.
- Pollock CB, Rodriguez O, Martin PL, Albanese C, Li X, Kopelovich L *et al.* (2010). Induction of metastatic gastric cancer by peroxisome proliferator-activated receptor δ activation. *PPAR Res* 2010: 571783.
- Quintela AM, Jimenez R, Gomez-Guzman M, Zarzuelo MJ, Galindo P, Sanchez M *et al.* (2012). Activation of peroxisome proliferator-activated receptor- β/δ (PPAR β/δ) prevents endothelial dysfunction in type 1 diabetic rats. *Free Radic Biol Med* 53: 730–741.
- Remick J, Weintraub H, Setton R, Offenbacher J, Fisher E, Schwartzbard A (2008). Fibrate therapy: an update. *Cardiol Rev* 16: 129–141.
- Rosen ED, Spiegelman BM (2001). PPARgamma: a nuclear regulator of metabolism, differentiation, and cell growth. *J Biol Chem* 276: 37731–37734.
- Scatena R, Bottoni P, Vincenzoni F, Messana I, Martorana GE, Nocca G *et al.* (2003). Bezafibrate induces a mitochondrial derangement in human cell lines: a PPAR-independent mechanism for a peroxisome proliferator. *Chem Res Toxicol* 16: 1440–1447.
- Scott R, O'Brien R, Fulcher G, Pardy C, d'Emden M, Tse D *et al.* (2009). Effects of fenofibrate treatment on cardiovascular disease risk in 9,795 individuals with type 2 diabetes and various components of the metabolic syndrome the Fenofibrate Intervention and Event Lowering in Diabetes (FIELD) study. *Diabetes Care* 32: 493–498.
- Srivastava RA, Jahagirdar R, Azhar S, Sharma S, Bisgaier CL (2006). Peroxisome proliferator-activated receptor-alpha selective ligand reduces adiposity, improves insulin sensitivity and inhibits atherosclerosis in LDL receptor-deficient mice. *Mol Cell Biochem* 285: 35–50.
- Tenenbaum A, Motro M, Fisman EZ, Schwammenthal E, Adler Y, Goldenberg I *et al.* (2004). Peroxisome proliferator-activated receptor ligand bezafibrate for prevention of type 2 diabetes mellitus in patients with coronary artery disease. *Circulation* 109: 2197–2202.
- Tenenbaum A, Motro M, Fisman EZ, Adler Y, Shemesh J, Tanne D *et al.* (2005a). Effect of bezafibrate on incidence of type 2 diabetes mellitus in obese patients. *Eur Heart J* 26: 2032–2038.
- Tenenbaum A, Motro M, Fisman EZ, Tanne D, Boyko V, Behar S (2005b). Bezafibrate for the secondary prevention of myocardial infarction in patients with metabolic syndrome. *Arch Intern Med* 165: 1154–1160.
- Tenkanen L, Manttari M, Kovanen PT, Virkkunen H, Manninen V (2006). Gemfibrozil in the treatment of dyslipidemia: an 18-year mortality follow-up of the Helsinki Heart Study. *Arch Intern Med* 166: 743–748.
- Tian J, Wong WT, Tian XY, Zhang P, Huang Y, Wang N (2010). Rosiglitazone attenuates endothelin-1-induced vasoconstriction by upregulating endothelial expression of endothelin B receptor. *Hypertension* 56: 129–135.
- Tian XY, Wong WT, Wang NP, Lu Y, Cheang WS, Liu J *et al.* (2012). PPAR δ activation protects endothelial function in diabetic mice. *Diabetes* 61: 3285–3293.
- Tordjman KM, Semenkovich CF, Coleman T, Yudovich R, Bak S, Osher E *et al.* (2007). Absence of peroxisome proliferator-activated receptor-alpha abolishes hypertension and attenuates atherosclerosis in the Tsukuba hypertensive mouse. *Hypertension* 50: 945–951.
- Tousoulis D, Andreou I, Antoniadis C, Tentolouris C, Stefanadis C (2008). Role of inflammation and oxidative stress in endothelial progenitor cell function and mobilization: therapeutic implications for cardiovascular diseases. *Atherosclerosis* 201: 236–247.
- Vallarino C, Perez A, Fusco G, Liang H, Bron M, Manne S *et al.* (2013). Comparing pioglitazone to insulin with respect to cancer, cardiovascular and bone fracture endpoints, using propensity score weights. *Clin Drug Investig* 33: 621–631.
- Wang G, He L, Liu J, Yu J, Feng X, Li F *et al.* (2013). Coronary flow velocity reserve is improved by PPAR- α agonist fenofibrate in patients with hypertriglyceridemia. *Cardiovasc Ther* 31: 161–167.
- Wang YX, Lee CH, Tiep S, Yu RT, Ham J, Kang H *et al.* (2003). Peroxisome-proliferator-activated receptor delta activates fat metabolism to prevent obesity. *Cell* 113: 159–170.
- Wilcox R, Kupfer S, Erdmann E (2008). Effects of pioglitazone on major adverse cardiovascular events in high-risk patients with type 2 diabetes: results from PROspective pioglitAzone Clinical Trial In macro Vascular Events (PROactive 10). *Am Heart J* 155: 712–717.
- Woldt E, Terrand J, Mlih M, Matz RL, Bruban V, Coudane F *et al.* (2012). The nuclear hormone receptor PPAR γ counteracts vascular calcification by inhibiting Wnt5a signalling in vascular smooth muscle cells. *Nat Commun* 3: 1077.
- Wong WT, Tian XY, Xu AM, Yu J, Lau CW, Hoo RLC *et al.* (2011). Adiponectin is required for PPAR gamma-mediated improvement of endothelial function in diabetic mice. *Cell Metab* 14: 104–115.
- Xu HE, Lambert MH, Montana VG, Parks DJ, Blanchard SG, Brown PJ *et al.* (1999). Molecular recognition of fatty acids by peroxisome proliferator-activated receptors. *Mol Cell* 3: 397–403.

- Yuen CY, Wong WT, Tian XY, Wong SL, Lau CW, Yu J *et al.* (2011). Telmisartan inhibits vasoconstriction via PPAR γ -dependent expression and activation of endothelial nitric oxide synthase. *Cardiovasc Res* 90: 122–129.
- Zadelaar AS, Boesten LS, Jukema JW, van Vlijmen BJ, Kooistra T, Emeis JJ *et al.* (2006). Dual PPAR α /gamma agonist tesaglitazar reduces atherosclerosis in insulin-resistant and hypercholesterolemic ApoE*3Leiden mice. *Arterioscler Thromb Vasc Biol* 26: 2560–2566.
- Zaruelo MJ, Jimenez R, Galindo P, Sanchez M, Nieto A, Romero M *et al.* (2011). Antihypertensive effects of peroxisome proliferator-activated receptor- β activation in spontaneously hypertensive rats. *Hypertension* 58: 733–743.
- Zaruelo MJ, Gomez-Guzman M, Jimenez R, Quintela AM, Romero M, Sanchez M *et al.* (2013). Effects of peroxisome proliferator-activated receptor- β activation in endothelin-dependent hypertension. *Cardiovasc Res* 99: 622–631.
- Zhang HF, Wang L, Yuan HJ, Ma YH, Wang YF, Hu ZY *et al.* (2013). PPAR- γ agonist pioglitazone prevents apoptosis of endothelial progenitor cells from rat bone marrow. *Cell Biol Int* 37: 430–435.
- Zhou L, Tang YP, Gao L, Fan XS, Liu CM, Wu DK (2009). Separation, characterization and dose-effect relationship of the PPAR γ -activating bio-active constituents in the Chinese herb formulation 'San-Ao decoction. *Molecules* 14: 3942–3951.
- Zuckerman SH, Kauffman RF, Evans GF (2002). Peroxisome proliferator-activated receptor alpha, gamma coagonist LY465608 inhibits macrophage activation and atherosclerosis in apolipoprotein E knockout mice. *Lipids* 37: 487–494.