

## Phase I Study of Navitoclax (ABT-263), a Novel Bcl-2 Family Inhibitor, in Patients With Small-Cell Lung Cancer and Other Solid Tumors

Leena Gandhi, D. Ross Camidge, Moacyr Ribeiro de Oliveira, Philip Bonomi, David Gandara, Divis Khaira, Christine L. Hamm, Evelyn M. McKeegan, Elizabeth Litvinovich, Philip M. Hemken, Caroline Dive, Sari H. Enschede, Cathy Nolan, Yi-Lin Chiu, Todd Busman, Hao Xiong, Andrew P. Krivoshek, Rod Humerickhouse, Geoffrey I. Shapiro, and Charles M. Rudin

From the Dana-Farber Cancer Institute, Boston, MA; University of Colorado Cancer Center, Aurora, CO; Northwest Medical Specialties, Tacoma, WA; Rush University Medical Center, Chicago; Abbott Laboratories, Abbott Park, IL; University of California Davis Cancer Center, Sacramento, CA; Sunterra Oncology Associates, Phoenix, AZ; Johns Hopkins University, Baltimore, MD; and Paterson Institute for Cancer Research, University of Manchester, United Kingdom.

Submitted August 27, 2010; accepted November 23, 2010; published online ahead of print at www.jco.org on January 31, 2011.

Supported by Abbott Laboratories and Genentech; and in part by the Alice and Steve Cutler Fund for Young Investigators at the Dana-Farber Cancer Institute (L.G.).

Authors' disclosures of potential conflicts of interest and author contributions are found at the end of this article.

Clinical Trials repository link available on JCO.org.

Corresponding author: Leena Gandhi, MD, PhD, Lowe Center for Thoracic Oncology and Early Drug Development Center, Dana-Farber Cancer Institute, 450 Brookline Ave, Boston, MA 02215; e-mail: Leena\_Gandhi@dfci.harvard.edu.

© 2011 by American Society of Clinical Oncology

0732-183X/11/2907-909/\$20.00

DOI: 10.1200/JCO.2010.31.6208

### ABSTRACT

#### Purpose

Resistance to chemotherapy-induced apoptosis represents a major obstacle to cancer control. Overexpression of Bcl-2 is seen in multiple tumor types and targeting Bcl-2 may provide therapeutic benefit. A phase I study of navitoclax, a novel inhibitor of Bcl-2 family proteins, was conducted to evaluate safety, pharmacokinetics, and preliminary efficacy in patients with solid tumors.

#### Patients and Methods

Patients enrolled to intermittent dosing cohorts received navitoclax on day -3, followed by dosing on days 1 to 14 of a 21-day cycle. Patients on continuous dosing received a 1-week lead-in dose of 150 mg followed by continuous daily administration. Blood samples were collected for pharmacokinetic analyses, biomarker analyses, and platelet monitoring.

#### Results

Forty-seven patients, including 29 with small-cell lung cancer (SCLC) or pulmonary carcinoid, were enrolled between 2007 and 2008, 35 on intermittent and 12 on continuous dosing cohorts. Primary toxicities included diarrhea (40%), nausea (34%), vomiting (36%), and fatigue (34%); most were grade 1 or 2. Dose- and schedule-dependent thrombocytopenia was seen in all patients. One patient with SCLC had a confirmed partial response lasting longer than 2 years, and eight patients with SCLC or carcinoid had stable disease (one remained on study for 13 months). Pro-gastrin releasing peptide (pro-GRP) was identified as a surrogate marker of *Bcl-2* amplification and changes correlated with changes in tumor volume.

#### Conclusion

Navitoclax is safe and well tolerated, with dose-dependent thrombocytopenia as the major adverse effect. Preliminary efficacy data are encouraging in SCLC. Efficacy in SCLC and the utility of pro-GRP as a marker of treatment response will be further evaluated in phase II studies.

*J Clin Oncol* 29:909-916. © 2011 by American Society of Clinical Oncology

### INTRODUCTION

The efficacy of many chemotherapeutic agents is dependent on activation of intrinsic apoptosis after DNA damage. Bcl-2 family proteins are central regulators of intrinsic apoptosis and overexpression of Bcl-2 in multiple solid tumor types has been hypothesized to play a role both in tumor cell survival as well as resistance to chemotherapy.<sup>1-3</sup> Bcl-2 overexpression is particularly frequent in small-cell lung cancer (SCLC),<sup>4-9</sup> where chemotherapy resistance represents a major obstacle to effective therapy. Bcl-2 upregulation has been seen in SCLC cell lines selected for chemotherapy resistance,<sup>10</sup> and has been

implicated in the mechanism of fibroblast growth factor 2- and Ets-mediated chemotherapy resistance in SCLC.<sup>11,12</sup>

Navitoclax is a potent and highly selective inhibitor of antiapoptotic members of the Bcl-2 family, with nanomolar affinity for Bcl-2, Bcl-xL, and Bcl-w. The potency and specificity of this agent are related to its unique mechanism of action, as a BH3 domain mimetic, to block the interaction of antiapoptotic family members with BH3 domain-containing pro-apoptotic proteins. Navitoclax is an orally available analog of ABT-737, first described to cause mechanism-based killing of multiple SCLC cell lines and regression of tumor xenografts.<sup>13,14</sup>

These effects were also seen with navitoclax<sup>15,16</sup> with significant tumor growth inhibition in nine of 11 tumor models and prolonged complete regression in some cases. Recent detailed mechanistic studies demonstrate that the cytotoxicity of ABT-737, unlike that of other small molecule inhibitors of Bcl-2 in clinical development including obatoclax, requires an intact intrinsic apoptotic pathway.<sup>17</sup> Preclinical data in animal models demonstrated marked and immediate thrombocytopenia with navitoclax that resolved on cessation of the drug. This is a mechanism-based toxicity induced by inhibition of Bcl-xL in circulating platelets,<sup>13,18</sup> which is required for platelet survival.<sup>19,20</sup> Here we describe a phase I dose-escalation study to evaluate the safety (with particular attention to platelet dynamics), pharmacokinetics, and preliminary efficacy of navitoclax administered on intermittent and continuous dosing schedules in patients with relapsed or refractory SCLC and other solid tumors.

In addition, potential biomarkers of response were evaluated, including amplification of *Bcl-2* in circulating tumor cells (CTCs), since amplification of a region of 18q that contains *Bcl-2* has recently been demonstrated to correlate with SCLC cell line sensitivity to ABT-737 in vitro.<sup>21</sup> Notably, this region contains not only *Bcl-2*, but also the gene for another potential marker of SCLC, pro-gastrin releasing peptide (pro-GRP). Pro-GRP is a tumor cell-secreted peptide that has previously been studied as a potential biomarker of disease progression and response to therapy in SCLC. Therefore, we also determined levels of circulating pro-GRP pre- and post-treatment, and correlated pretreatment levels with *Bcl-2* amplification, and change in levels during treatment with changes in tumor volume. Finally, caspase-mediated cleavage of the epithelial marker CK18, assessed with an antibody specific for the cleaved product, M30, was studied as another biomarker of navitoclax-induced apoptosis.

## PATIENTS AND METHODS

### Eligibility

Patients had histologically documented SCLC or other nonhematologic malignancies, age  $\geq 18$ , Eastern Cooperative Oncology Group performance status of  $\leq 2$ , measurable disease by Response Evaluation Criteria in Solid Tumors (RECIST version 1.0), and had received at least one prior chemotherapy regimen with documented progression. Patients with brain metastases were included if they had surgery and/or radiation therapy followed by 21 days of stable neurologic function and stable disease by imaging before the first dose of study drug. Additional inclusion criteria included adequate bone marrow, renal and hepatic function per local laboratory reference range, nonpregnant status, and a life expectancy of  $\geq 90$  days.

Patients were excluded if they had an underlying predisposing condition to or active bleeding; recent history of thrombocytopenia-associated bleeding; active immune thrombocytopenic purpura, autoimmune hemolytic anemia, or peptic ulcer disease; refractoriness to platelet transfusions within 1 year; need for full-dose anticoagulation, steroid, or aspirin therapy within 7 days.

The protocol was approved and monitored by all local institutional review boards; all patients provided written informed consent.

### Drug Administration and Definition of Dose-Limiting Toxicity

Patients enrolled in the intermittent dosing cohort received navitoclax on day -3 (lead-in dose), followed by dosing on days 1 to 14 of a 21-day cycle. In continuous dosing cohorts, patients received a 1-week lead-in dose of 150 mg, followed by 21 of 21 days of once daily dosing. Dose-limiting toxicity (DLT) was defined as any grade 3, 4, or 5 adverse event (AE) in cycle 1 considered possibly or probably related to navitoclax, with the following exceptions: grade 3 or 4 afebrile neutropenia, leukopenia, or lymphopenia;

alopecia; grade 3 nausea, vomiting, or diarrhea unless unresponsive to treatment; or grade 3 tumor lysis syndrome without clinical symptoms. Grade 2 or higher bleeding associated with thrombocytopenia of any grade and any unexpected grade 2 toxicity resulting in dose modification or delay of longer than 1 week were also considered DLTs. Any DLT required an interruption of dosing until the toxicity grade returned to  $\leq$  grade 1 or to baseline. After a DLT of thrombocytopenia, ABT-263 could be reintroduced only if the platelet count rose to  $\geq 50,000/\mu\text{L}$  ( $\leq$  grade 2). In those situations, the dose level was determined on an individual basis by the investigator and the Abbott Medical Monitor.

### Safety and Efficacy Assessments

Patients were evaluated with history, physical exam, vital signs, CBC, chemistries, urinalysis, and Eastern Cooperative Oncology Group performance status weekly through cycle 2 and on day 1 of each subsequent cycle. Incidence and severity of AEs were collected at each study visit and graded according to National Cancer Institute Common Toxicity Criteria for Adverse Events version 3. Radiographic tumor assessments were performed at baseline, after the second and fourth cycles, and after every 3 cycles thereafter. Response was measured according to standard RECIST. While all patients were considered evaluable for safety, patients with lower than 80% dosing compliance during cycle 1 were considered unevaluable for response. Additional clinical parameters were measured as described in Appendix (online only).

### Pharmacokinetics

For patients on 14 of 21-day dosing cohorts, blood samples were collected on day -3 before cycle 1 for pharmacokinetic monitoring at time 0, 2, 3, 4, 6, 8, 24, 48, and 72 hours. For patients on 21 of 21-day dosing cohorts, blood samples were collected on cycle 1, day 1 at time 0, 2, 3, 4, 6, 8, and 24 hours. For all patients, collections were also performed on cycle 1 day 14 (blood samples at 0, 2, 3, 4, 6, 8 hours). In addition, predose samples were collected beyond cycle 1 for all patients. Plasma concentration measurements are detailed in the Appendix (online only).

### Biomarker Assessment

Blood samples for CTCs and the plasma marker Pro-GRP were collected as described in Appendix (online only). CTCs were analyzed via the CellSearch

**Table 1.** Baseline Patient Demographics and Clinical Characteristics (N = 47)

Characteristic	No.	%
Median age, years	62	
Sex		
Male	27	57
Female	20	43
ECOG performance status		
0	16	35
1	26	56
2	27	56
No. of prior therapies		
1-2	24	51
$\geq 3$	23	49
Median time since last therapy, days	49	
Range	12-445	
Tumor type		
SCLC	26	55
Atypical pulmonary carcinoid	3	6
Other	18	38
Dosing schedule		
14/21	35	
SCLC	17	
Atypical pulmonary carcinoid	3	
21/21	12	
SCLC	4	

Abbreviations: ECOG, Eastern Cooperative Oncology Group; SCLC, small-cell lung cancer.

system (Veridex, Raritan, NJ). Blood (10 mL) collected in CellSearch tubes was processed within 72 hours after collection. Fluorescent in situ hybridization (FISH) was performed as previously described<sup>21</sup> using a Vysis LSI Bcl-2 (orange) probe and chromosome 18 probe (green) developed by Abbott Molecular. Similar probes were used to assess *Bcl-2* amplification in tumor biopsies. Pro-GRP was measured using ARCHITECT ELISA kits (Abbott Diagnostics, Abbott Park, IL). Serum samples were analyzed for M30 and M65 (Peviva, Bromma, Sweden) using previously described assays validated to good clinical laboratory practice.<sup>22,23</sup>

**Statistical Analyses**

This study was designed using an adaptation of the continuous reassessment method (CRM) for dose escalation<sup>24-26</sup> (further described in online-only Appendix). Two stages of dose escalation were planned. In stage I, dosing began at 10 mg with a cohort of three patients, with a plan thereafter for single patient cohorts and doubling of the dose in subsequent cohorts until a DLT was observed. Once a DLT was observed, stage II began with cohort sizes of three patients and CRM was employed to select the next dose, but with the constraint that escalation will not exceed the greater of 100 mg or 40%. Descriptive statistics summarized the demographics, safety data, and pharmacokinetics. Correlations between median *Bcl-2* copy number, pro-GRP, M30, dose, and best tumor response were made using the JMP 8.0 statistical software (JMP, Cary, NC).

**RESULTS**

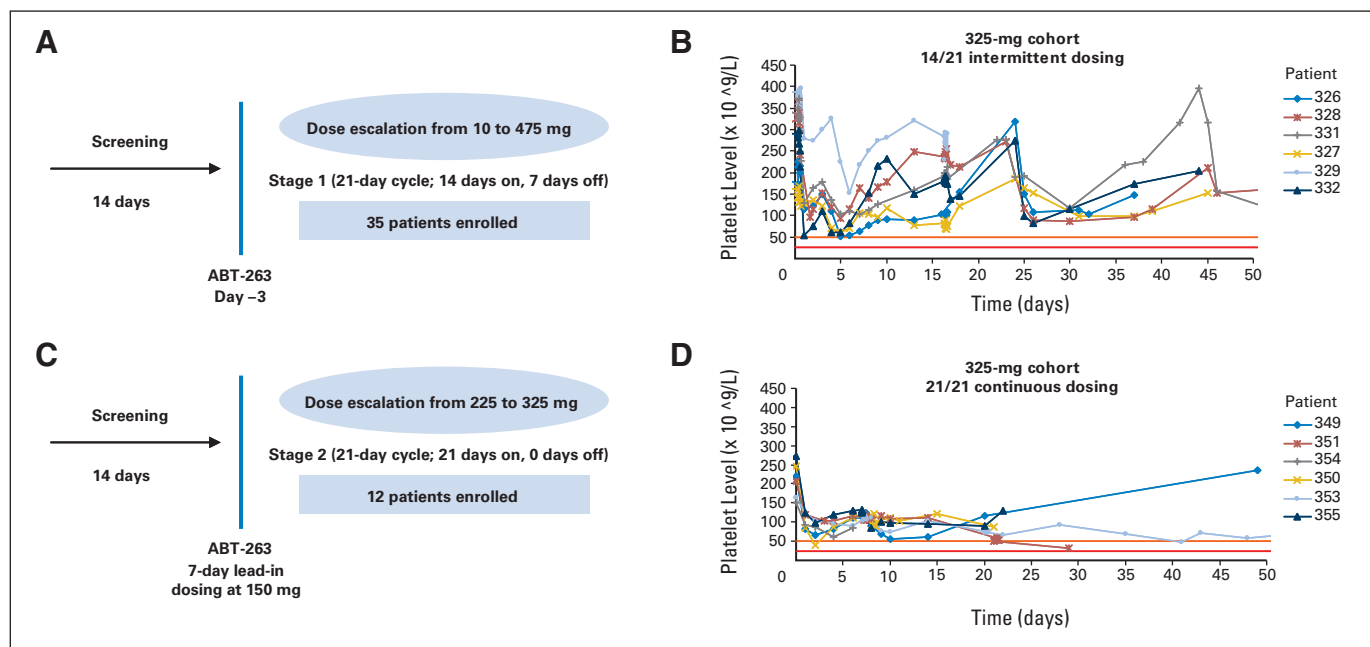
**Dose Escalation**

Forty-seven patients were enrolled in this study from April 2007 to May 2008. Table 1 presents the baseline characteristics of this patient population, more than 60% of whom had SCLC or atypical pulmonary carcinoid. The median duration on study was 38 days

(range, 17 to 882 days). Nine patients were considered unevaluable for response due to lower than 80% dosing compliance in cycle 1, which was primarily related to missed doses secondary to nausea, vomiting, or diarrhea. The most common reasons for study discontinuation were radiographic progressive disease, declining performance status, and withdrawal of consent. Thirty-five patients were treated on intermittent dosing cohorts ranging from 10 to 475 mg, with a day -3 lead-in dose, followed by dosing on days 1 through 14 of a 21-day cycle (Fig 1A). The kinetics of thrombocytopenia paralleled that seen in animal models. Platelet counts for all patients dropped to a nadir within 24 to 72 hours of dosing on day -3, consistent with peripheral destruction on Bcl-xL inhibition. However, with marrow compensation, platelet counts subsequently showed partial recovery during ongoing dosing and then recovered close to baseline during the week off drug (Fig 1B). A recurrent variable decline in platelet count occurred when drug was restarted.

Given the kinetics of thrombocytopenia and platelet recovery, with recovery even during continued dosing, the protocol was amended to evaluate whether a low lead-in dose for 7 days before therapeutic dosing could prime the marrow to upregulate platelet production, to allow for higher continuous dosing and to minimize subsequent platelet variability. A lead-in dose of 150 mg was chosen, as this was the dose level at which the mean maximal platelet drop of approximately 60% from baseline was expected based on observations in other concurrent studies of navitoclax as a single agent.

Based on a maximum-tolerated dose estimation of 350 mg using intermittent dosing, the daily equivalent dosing of 225 mg was the



**Fig 1.** Platelet dynamics with different dosing schedules. (A) Dosing schema for intermittent dosing. (B) Platelet levels over time in six representative patients on intermittent dosing cohorts. Day 0 represented in this figure is study day -3. Patient 326 was a 60-year-old man with non-small-cell lung cancer (NSCLC); patient 327 was an 80-year-old man with pancreatic cancer; patient 328 was a 68-year-old man with NSCLC; patient 329 was a 64-year-old man with small-cell lung cancer (SCLC); patient 331 was a 60-year-old woman with neuroendocrine carcinoid; patient 332 was a 62-year-old man with SCLC. (C) Dosing schema for continuous dosing. (D) Platelet levels over time in six representative patients on continuous dosing cohorts. Day 0 represented in this figure is the lead-in period day 1. Red and orange lines represent the boundaries of a dose-limiting toxicity (grade 3, 25,000 to 50,000); 325 mg was chosen as a representative dose for comparing the two different dosing schedules. Patient 349 was a 65-year-old man with SCLC; patient 350 was a 67-year-old man with SCLC; patient 351 was a 57-year-old woman with SCLC; patient 353 was a 63-year-old man with SCLC; patient 354 was a 65-year-old woman with SCLC; patient 355 was a 60-year-old man with SCLC.

**Table 2.** Pharmacokinetic Parameters of Navitoclax After Oral Administration (14/21-day dosing schedule)

Study Day by Dose (mg)	No.	$T_{max}$ (hours)		$C_{max}$ ( $\mu\text{g/mL}$ )		$AUC_{0-24}$ ( $\mu\text{g} \times \text{hr/mL}$ )		$AUC_{0-inf}$ ( $\mu\text{g} \times \text{hr/mL}$ )		CL/F (L/h)		$t_{1/2}^*$ (hours)	
		Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD	Mean	SD
10													
-3	4	6.0	1.6	0.13	0.04	1.4	0.4	2.0	0.6†	5.3	1.4†	15.3	2.2†
14	4	6.0	0.0	0.12	0.07	—	—	—	—	—	—	—	—
20													
-3	4	6.3	2.4	0.38	0.05	4.9	0.9	7.1	1.9	3.0	0.8	13.5	6.6
14	4	7.0	1.2	0.46	0.14	—	—	—	—	—	—	—	—
30													
-3	6	6.0	1.3	0.67	0.29	7.3	3.4	10.1	4.6	5.0	5.7	14.1	10.4
14	5	4.4	2.1	0.81	0.21	—	—	—	—	—	—	—	—
130													
-3	4	11.0	8.7	1.84	1.22	25.4	12.8	42.0	12.4	3.3	0.8	16.9	1.2
14	4	6.0	1.6	2.81	1.75	—	—	—	—	—	—	—	—
225													
-3	3	5.7	2.5	2.97	1.24	39.5	18.8	53.0	24.6	5.3	3.3	12.8	0.8
14	4	7.5	1.0	5.10	0.85	—	—	—	—	—	—	—	—
325													
-3	6	10.0	6.9	2.55	1.46	40.9	25.8	80.5	44.3‡	5.9	4.7†	15.8	3.1†
14	8	5.8	2.3	4.31	2.14	—	—	—	—	—	—	—	—
425													
-3	5	5.0	2.0	3.43	1.75	53.7	30.8	99.2	69.5	7.5	6.4	15.0	3.4
14	3	3.7	4.0	2.87	0.78	—	—	—	—	—	—	—	—
475													
-3	3	12.0	10.4	3.76	1.71	66.8	35.6	81.0	29.7§	6.3	2.3§	15.7	0.2§
14	1	6.0		5.59		—	—	—	—	—	—	—	—

NOTE. All doses were taken under nonfasting condition.

Abbreviations:  $T_{max}$ , time to maximum observed plasma concentration;  $C_{max}$ , maximum plasma concentration; AUC, area under the curve; inf, infinity; CL/F, apparent oral clearance;  $t_{1/2}$ , terminal half-life; SD, standard deviation.

\*Harmonic mean  $\pm$  pseudo SD.

†N = 3.

‡N = 5.

§N = 2.

starting point for the dose escalation in continuous dosing cohorts (Fig 1C), where less variability in platelet dynamics over time were observed (Fig 1D). Twelve additional patients were treated on continuous dosing cohorts before an estimated maximum-tolerated dose of 325 mg was reached.

### Pharmacokinetics

Table 2 presents the pharmacokinetic profile of navitoclax exposure after a single lead-in dose (day -3) or after the day 14 dose on the 14 of 21-day dosing schedule, demonstrating that exposure was dose-proportional. Peak concentrations ( $C_{max}$ ) were observed approximately 7 hours postdose with a half-life of approximately 15 hours. Peak-to-trough plasma concentration ratio was close to two-fold at steady-state and overall the interpatient variability in exposure was 40%. Dosage at or above 225 mg met the minimum plasma exposure predicted to be in the therapeutic range based on animal models.

### Safety

The most frequent AEs excluding thrombocytopenia were diarrhea (40%), vomiting (36%), nausea (34%), and fatigue (34%), as detailed in Table 3. The majority of these were grade 1 or 2 (Appendix Table A1, online only). While all patients experienced some degree of thrombocytopenia, only 15% met criteria for AEs. In addition, three

patients were removed from study for severe AEs that were deemed to be possibly or probably related to study drug. These included fatal respiratory failure in one patient that occurred at the lowest dosing level during cycle 1; left ventricular systolic dysfunction with a drop from 55% to 70% at baseline to 40% during cycle 1 and resolution to 55% 1 week after stopping drug; and asymptomatic lipase elevation from a baseline of grade 2 to grade 3 with normalization to baseline within 1 week off drug.

### Antitumor Activity

Of the 38 patients who were evaluable for response (23 with SCLC or pulmonary carcinoid), eight had stable disease (five SCLC and three atypical pulmonary carcinoid) and one patient with SCLC remained on study for 13 months. One patient with SCLC had a partial response that has been sustained for longer than 35 months and remains on study; this patient had a localized recurrence after first-line treatment with progressive disease on second-line therapy before study entry. Overall, among patients with disease control, the median number of prior therapies was three (range, one to five therapies). The majority of patients with disease control were those treated at the highest dose levels (Fig 2) and the median duration of disease control was 5 months (range, 2 to 35 months). Dosing schedule details for patients with disease control are presented in Appendix Table A2 (online only).

**Table 3.** Most Common AEs

AE	Patients for All Grades (N = 47)		14/21-Day Dosing Cohorts by Grade (mg)												21/21-Day Dosing by Grade (mg)			
			10-30 (n = 16)		130 (n = 4)		225 (n = 3)		325 (n = 6)		425 (n = 5)		475 (n = 3)		225 (n = 6)		325 (n = 6)	
			1/2	3/4	1/2	3/4	1/2	3/4	1/2	3/4	1/2	3/4	1/2	3/4	1/2	3/4	1/2	3/4
Lymphopenia	2	4.3											1 (3)				1	
Neutropenia	1	2.1											1 (3)				1	
Thrombocytopenia	7	14.9								1 (4)			1 (4)		1 (4)	1	1 (3)	
Diarrhea	19	40.4			2		1		4			4		2		2	4	
Enteritis	1	2.1															1	
Nausea	16	34	2		1		1		1			3	1	2		1	4	
Vomiting	17	36.2	3			1		3		2	1	1		2		2	4	
Fatigue	16	34	2			2 (3)		3		2	2 (3)	1		3		3	1	
Decreased appetite	9	19	2					1		2				3		3	1	
Weight decrease	2	4.3						1					1 (3)					
Hypophosphataemia	1	2.1											1 (3)					
Hemoptysis	1	2.1											1 (3)					
Respiratory failure	1	2.1		1 (5)														

NOTE. All AEs are reported if > 10% or if grade 3, 4, or 5. If grade 3, 4, or 5, the specific grade is reported in parentheses. Abbreviation: AE, adverse event.

### Biomarker Analyses

To determine whether *Bcl-2* amplification was associated with response, *Bcl-2* copy number was assessed in blood samples from 46 patients when feasible. Twenty-one patients had eight or more CTCs detected and in these, FISH for *Bcl-2* was performed. Figure 3A shows an example of a patient with amplification of *Bcl-2* in CTCs, but not in peripheral blood mononuclear cells (PBMC). In patients where *Bcl-2* amplification was performed, *Bcl-2* copy number was compared to circulating pro-GRP levels. There was a direct correlation between median *Bcl-2* copy number and circulating pro-GRP ( $R^2 = 0.98$ ), consistent with previous studies demonstrating coamplification of these two genes<sup>21</sup> (Fig 3B). When only patients with SCLC or neuroendocrine tumors were analyzed, the same correlation was seen ( $R^2 = 0.95$ , data not shown). Overall, pro-GRP levels declined or stabilized with increasing navitoclax dose (Fig 3C), but more importantly, the change in pro-GRP levels correlated with best percentage tumor change (Fig 3D;  $R^2 = 0.76$ ).

A dose-dependent transient increase in circulating M30, a serum marker for tumor cell apoptosis, was also observed in dose cohorts  $\geq 130$  mg ( $R^2 = 0.48$ ;  $P = .0012$ ; Fig 3E). In most instances, the rise in M30 was rapid, occurring within 6 hours after the first dose. Evidence of apoptosis was sustained through 14 days of oral dosing, mirroring the biomarker behavior in a SCLC human xenograft preclinical model (data not shown).<sup>27</sup>

## DISCUSSION

Deregulation of apoptosis is a hallmark of cancer and several studies have suggested that antiapoptotic gene activity is critical for continued survival and proliferation.<sup>12,28</sup> *Bcl-2* is attractive therapeutic target in SCLC and other tumors, but, to date, anti-*Bcl-2* therapies have not demonstrated significant clinical efficacy. Trials of an antisense oligonucleotide to *Bcl-2* mRNA were largely unsuccessful, but downregulation of *Bcl-2* in clinical specimens was limited and did not correlate with response.<sup>29,30</sup> In addition, the *Bcl-2* family contains both antiapoptotic members (ie, *Bcl-2*,

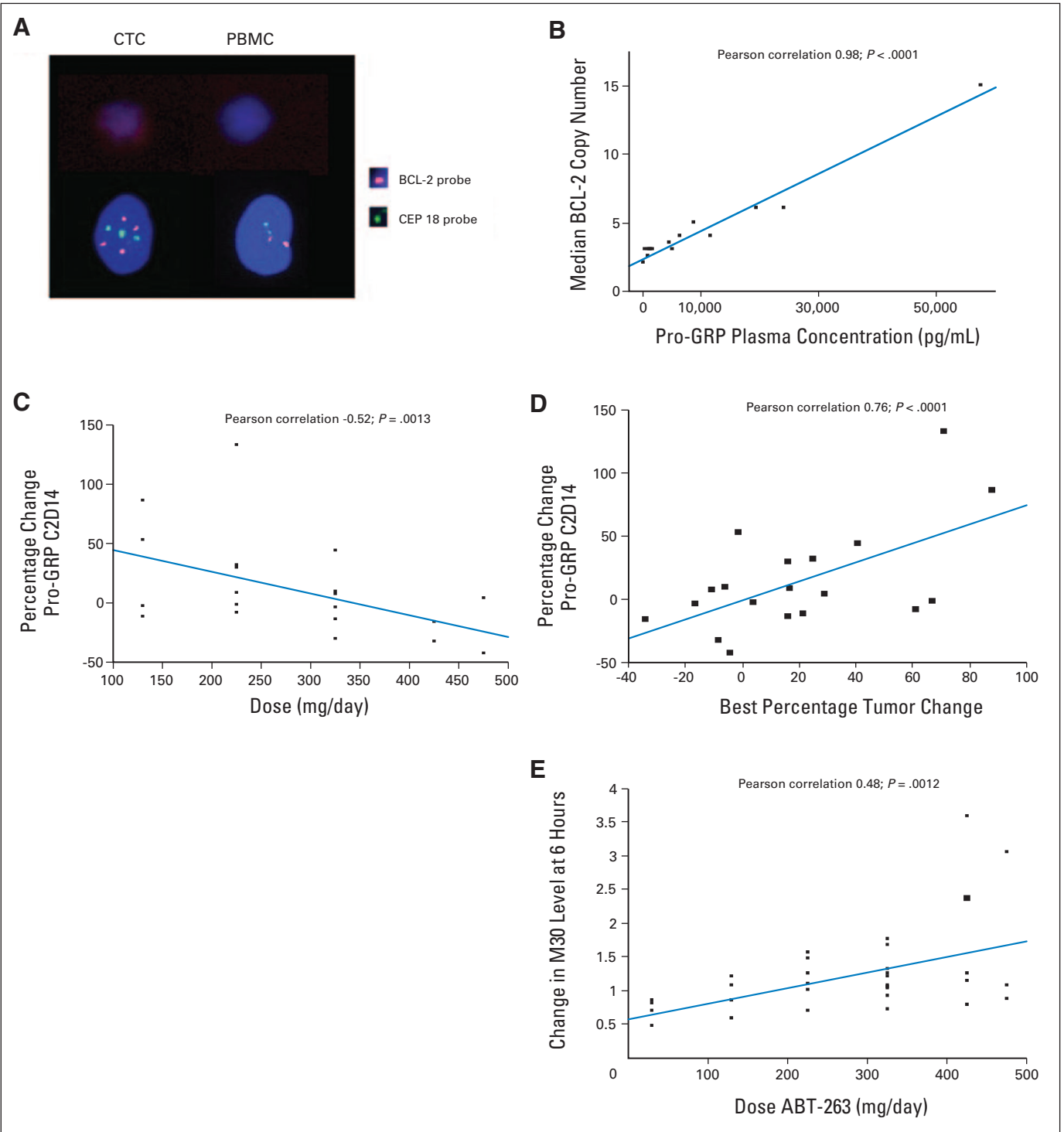
*Bcl-xL*, *Bcl-w*, *Mcl-1*, and *Bcl-A2*), pro-apoptotic multidomain members (ie, *Bak*, *Bax*), and BH3-only domain members (ie, *Bim*, *Bid*, *Puma*, *Bad*, *Bik*, *Noxa*, and *Bmf*), and many agents that target *Bcl-2* do so with a lack of specificity among this group.<sup>31</sup>

Navitoclax has a unique mechanism of action that disrupts the regulatory step that prevents pro-apoptotic *Bcl-2* family members from initiating apoptotic pathways. Although this study was open to all solid tumors, given the level of preclinical activity seen in SCLC models, enrollment was enriched for patients with small cell or other neuroendocrine tumors.

Navitoclax was overall well-tolerated and the majority of treatment-related AEs (ie, diarrhea, nausea, vomiting, and fatigue) other than thrombocytopenia were grade 1 or 2 and manageable. Thrombocytopenia was experienced by all patients on this study, as expected for a drug that inhibits *Bcl-xL* function. Consistent with a peripheral apoptotic process that did not affect platelet production or function, a relative increase in large platelet forms was noted among the study population. Although intermittent dosing was associated with significant intracycle variability of platelets, changing to a continuous dosing schedule after a priming lead-in dose effectively mitigated the variability in platelet counts. Thrombocytopenia developed and resolved in a predictable fashion without clinical sequelae and, based on dose- and schedule dependence, behaves as a pharmacodynamic marker of on-target drug activity.

In order to evaluate the basis for response, ongoing evaluations of *Bcl-2* in tumor tissue and blood are underway. Importantly, we were able to demonstrate that CTCs were readily detectable, that *Bcl-2* amplification was present in CTCs and that *Bcl-2* gene copy number correlated directly with plasma pro-GRP levels. The data indicate that the easily identifiable plasma protein pro-GRP can serve as an accurate surrogate for *Bcl-2* amplification. Furthermore, the direct correlation with best tumor response seen in patients treated at sufficient dose levels indicates that pro-GRP can serve as a pharmacodynamically relevant biomarker of response to anti-*Bcl-2* therapy. Measures of





**Fig 3.** Biomarkers of navitoclax activity and tumor response. (A) Fluorescent in situ hybridization analysis of Bcl-2 in a patient with small-cell lung cancer demonstrating amplified signal (four copies) of Bcl-2 (orange) and three copies of CEP 18 (green) in circulating tumor cells (CTCs), but no amplification (two copies of both) in peripheral blood mononuclear cells (PBMC). (B) Pro-gastrin releasing peptide (pro-GRP) plasma concentration (pg/mL) plotted against mean Bcl-2 copy number. (C) Relative change in pro-GRP plasma concentration with different dose levels. (D) The relationship between best tumor percentage change is plotted against percentage change of pro-GRP from baseline to cycle 2 day 14. (E) Changes in circulating M30 levels with increasing dose as measured in cycle 1, 6 hours after first exposure.

**Employment or Leadership Position:** Evelyn M. McKeegan, Abbott Laboratories (C); Philip M. Hemken, Abbott Laboratories (C); Andrew P. Krivoshik, Abbott Laboratories (C); Sari H Enschede, Abbott Laboratories (C); Cathy Nolan, Abbott Laboratories (C); Yi-Lin Chiu,

Abbott Laboratories (C); Todd Busman, Abbott Laboratories (C); Hao Xiong, Abbott Laboratories (C); Rod Humerickhouse, Abbott Laboratories (C) **Consultant or Advisory Role:** Philip Bonomi, Abbott Laboratories (C); David Gandara, Amgen (C), Biodesix (C), Boehringer

Ingelheim (C), Bristol-Myers Squibb/ImClone (C), GlaxoSmithKline (C), Genentech (C), Merck (C), Novartis (C), sanofi-aventis (C), Response Genetics (C), AstraZeneca (C), Eli Lilly (U); Charles M. Rudin, Syndax Pharmaceuticals (C), OSI Pharmaceuticals (C), Genentech (C)  
**Stock Ownership:** Evelyn M. McKeegan, Abbott Laboratories; Philip M. Hemken, Abbott Laboratories; Andrew P. Krivoshik, Abbott Laboratories; Sari H Enschede, Abbott Laboratories; Yi-Lin Chiu, Abbott Laboratories; Todd Busman, Abbott Laboratories; Hao Xiong, Abbott Laboratories; Rod Humerickhouse, Abbott Laboratories  
**Honoraria:** Philip Bonomi, Abbott Laboratories  
**Research Funding:** Philip Bonomi, Abbott Laboratories; David Gandara, Abbott Laboratories, Bristol-Myers Squibb/ImClone, Genentech, Eli Lilly, Merck, Novartis, Pfizer; Divis Khaira, Abbott Laboratories, Schering-Plough; Caroline Dive, Abbott Laboratories  
**Expert Testimony:** None  
**Other Remuneration:** None

## AUTHOR CONTRIBUTIONS

**Conception and design:** Leena Gandhi, Christine L. Hann, Philip M. Hemken, Caroline Dive, Andrew P. Krivoshik, Sari H. Enschede, Cathy

Nolan, Yi-Lin Chiu, Hao Xiong, Rod Humerickhouse, Geoffrey I. Shapiro, Charles M. Rudin

**Financial support:** Andrew P. Krivoshik, Rod Humerickhouse

**Administrative support:** Cathy Nolan

**Provision of study materials or patients:** Leena Gandhi, D. Ross Camidge, Moacyr Ribeiro de Oliveira, Philip Bonomi, Divis Khaira, Christine L. Hann, Andrew P. Krivoshik, Rod Humerickhouse, Geoffrey I. Shapiro, Charles M. Rudin

**Collection and assembly of data:** Leena Gandhi, Moacyr Ribeiro de Oliveira, Divis Khaira, Evelyn M. McKeegan, Elizabeth Litvinovich, Caroline Dive, Andrew P. Krivoshik, Cathy Nolan, Todd Busman, Rod Humerickhouse, Charles M. Rudin

**Data analysis and interpretation:** Leena Gandhi, D. Ross Camidge, Moacyr Ribeiro de Oliveira, David Gandara, Divis Khaira, Evelyn M. McKeegan, Elizabeth Litvinovich, Andrew P. Krivoshik, Sari H. Enschede, Cathy Nolan, Yi-Lin Chiu, Todd Busman, Hao Xiong, Rod Humerickhouse, Geoffrey I. Shapiro, Charles M. Rudin

**Manuscript writing:** All authors

**Final approval of manuscript:** All authors

## REFERENCES

1. Reed JC: Bcl-2: Prevention of apoptosis as a mechanism of drug resistance. *Hematol Oncol Clin North Am* 9:451-473, 1995
2. Ohmori T, Podack ER, Nishio K, et al: Apoptosis of lung cancer cells caused by some anti-cancer agents (MMC, CPT-11, ADM) is inhibited by bcl-2. *Biochem Biophys Res Commun* 192:30-36, 1993
3. Kirkin V, Joos S, Zornig M: The role of Bcl-2 family members in tumorigenesis. *Biochim Biophys Acta* 1644:229-249, 2004
4. Ben-Ezra JM, Kornstein MJ, Grimes MM, et al: Small cell carcinomas of the lung express the Bcl-2 protein. *Am J Pathol* 145:1036-1040, 1994
5. Eerola AK, Tormanen U, Rainio P, et al: Apoptosis in operated small cell lung carcinoma is inversely related to tumour necrosis and p53 immunoreactivity. *J Pathol* 181:172-177, 1997
6. Brambilla E, Negoescu A, Gazzeri S, et al: Apoptosis-related factors p53, Bcl2, and Bax in neuroendocrine lung tumors. *Am J Pathol* 149:1941-1952, 1996
7. Kaiser U, Schilli M, Haag U, et al: Expression of bcl-2-protein in small cell lung cancer. *Lung Cancer* 15:31-40, 1996
8. Breton C, Story MD, Meyn RE: Bcl-2 expression correlates with apoptosis induction but not loss of clonogenic survival in small cell lung cancer cell lines treated with etoposide. *Anticancer Drugs* 9:751-757, 1998
9. Jiang SX, Sato Y, Kuwano S, et al: Expression of bcl-2 oncogene protein is prevalent in small cell lung carcinomas. *J Pathol* 177:135-138, 1995
10. Sartorius UA, Krammer PH: Upregulation of Bcl-2 is involved in the mediation of chemotherapy resistance in human small cell lung cancer cell lines. *Int J Cancer* 97:584-592, 2002
11. Pardo OE, Arcaro A, Salerno G, et al: Fibroblast growth factor-2 induces translational regulation of Bcl-XL and Bcl-2 via a MEK-dependent pathway: Correlation with resistance to etoposide-induced apoptosis. *J Biol Chem* 277:12040-12046, 2002
12. Certo M, Del Gaizo Moore V, Nishino M, et al: Mitochondria primed by death signals determine cellular addiction to antiapoptotic BCL-2 family members. *Cancer Cell* 9:351-365, 2006
13. Oltsdorf T, Elmore SW, Shoemaker AR, et al: An inhibitor of Bcl-2 family proteins induces regression of solid tumours. *Nature* 435:677-681, 2005
14. Hann CL, Daniel VC, Sugar EA, et al: Therapeutic efficacy of ABT-737, a selective inhibitor of BCL-2, in small cell lung cancer. *Cancer Res* 68:2321-2328, 2008
15. Shoemaker AR, Mitten MJ, Adickes J, et al: Activity of the Bcl-2 family inhibitor ABT-263 in a panel of small cell lung cancer xenograft models. *Clin Cancer Res* 14:3268-3277, 2008
16. Tse C, Shoemaker AR, Adickes J, et al: ABT-263: A potent and orally bioavailable Bcl-2 family inhibitor. *Cancer Res* 68:3421-3428, 2008
17. Vogler M, Weber K, Dinsdale D, et al: Different forms of cell death induced by putative BCL2 inhibitors. *Cell Death Differ* 16:1030-1039, 2009
18. Roberts AW, Wilson WH, Gandhi L, et al: Ongoing phase 1 studies of ABT-263: Mitigating Bcl-XL induced thrombocytopenia with lead-in and continuous dosing. *J Clin Oncol* 27:147s, 2009 (suppl; abstr 3505)
19. Mason KD, Carpinelli MR, Fletcher JI, et al: Programmed anuclear cell death delimits platelet life span. *Cell* 128:1173-1186, 2007
20. Zhang H, Nimmer PM, Tahir SK, et al: Bcl-2 family proteins are essential for platelet survival. *Cell Death Differ* 14:943-951, 2007
21. Olejniczak ET, Van Sant C, Anderson MG, et al: Integrative genomic analysis of small-cell lung carcinoma reveals correlates of sensitivity to bcl-2 antagonists and uncovers novel chromosomal gains. *Mol Cancer Res* 5:331-339, 2007
22. Cummings J, Ward TH, LaCasse E, et al: Validation of pharmacodynamic assays to evaluate the clinical efficacy of an antisense compound (AEG 35156) targeted to the X-linked inhibitor of apoptosis protein XIAP. *Br J Cancer* 92:532-538, 2005
23. Cummings J, Ranson M, Lacasse E, et al: Method validation and preliminary qualification of pharmacodynamic biomarkers employed to evaluate the clinical efficacy of an antisense compound (AEG35156) targeted to the X-linked inhibitor of apoptosis protein XIAP. *Br J Cancer* 95:42-48, 2006
24. Piantadosi S, Fisher JD, Grossman S: Practical implementation of a modified continual reassessment method for dose-finding trials. *Cancer Chemother Pharmacol* 41:429-436, 1998
25. O'Quigley J, Shen LZ: Continual reassessment method: A likelihood approach. *Biometrics* 52:673-684, 1996
26. Goodman SN, Zahurak ML, Piantadosi S: Some practical improvements in the continual reassessment method for phase I studies. *Stat Med* 14:1149-1161, 1995
27. Micha D, Cummings J, Shoemaker A, et al: Circulating biomarkers of cell death after treatment with the BH-3 mimetic ABT-737 in a preclinical model of small-cell lung cancer. *Clin Cancer Res* 14:7304-7310, 2008
28. Del Gaizo Moore V, Letai A: Rational design of therapeutics targeting the BCL-2 family: Are some cancer cells primed for death but waiting for a final push? *Adv Exp Med Biol* 615:159-175, 2008
29. Tolcher AW, Chi K, Kuhn J, et al: A phase II, pharmacokinetic, and biological correlative study of oblimersen sodium and docetaxel in patients with hormone-refractory prostate cancer. *Clin Cancer Res* 11:3854-3861, 2005
30. Rudin CM, Salgia R, Wang X, et al: Randomized phase II Study of carboplatin and etoposide with or without the bcl-2 antisense oligonucleotide oblimersen for extensive-stage small-cell lung cancer: CALGB 30103. *J Clin Oncol* 26:870-876, 2008
31. Vogler M, Dinsdale D, Dyer MJ, et al: Bcl-2 inhibitors: Small molecules with a big impact on cancer therapy. *Cell Death Differ* 16:360-367, 2009
32. Tahir SK, Yang X, Anderson MG, et al: Influence of Bcl-2 family members on the cellular response of small-cell lung cancer cell lines to ABT-737. *Cancer Res* 67:1176-1183, 2007
33. Lin X, Morgan-Lappe S, Huang X, et al: 'Seed' analysis of off-target siRNAs reveals an essential role of Mcl-1 in resistance to the small-molecule Bcl-2/Bcl-XL inhibitor ABT-737. *Oncogene* 26:3972-3979, 2007
34. Hauck P, Chao BH, Litz J, et al: Alterations in the Noxa/Mcl-1 axis determine sensitivity of small cell lung cancer to the BH3 mimetic ABT-737. *Mol Cancer Ther* 8:883-892, 2009
35. van Delft MF, Wei AH, Mason KD, et al: The BH3 mimetic ABT-737 targets selective Bcl-2 proteins and efficiently induces apoptosis via Bak/Bax if Mcl-1 is neutralized. *Cancer Cell* 10:389-399, 2006