



## Genome Sequence of Rapid Beer-Spoiling Isolate *Lactobacillus brevis* BSO 464

Jordyn Bergsveinson, Vanessa Pittet,\* Emily Ewen, Nina Baecker, DBarry Ziola

Department of Pathology and Laboratory Medicine, College of Medicine, University of Saskatchewan, Saskatchewan, Canada

The genome of brewery-isolate *Lactobacillus brevis* BSO 464 was sequenced and assembly produced a chromosome and eight plasmids. This bacterium tolerates dissolved CO<sub>2</sub>/pressure and can rapidly spoil packaged beer. This genome is useful for analyzing the genetics associated with beer spoilage by lactic acid bacteria.

Received 9 October 2015 Accepted 16 October 2015 Published 3 December 2015

Citation Bergsveinson J, Pittet V, Ewen E, Baecker N, Ziola B. 2015. Genome sequence of rapid beer-spoiling isolate *Lactobacillus brevis* BSO 464. Genome Announc 3(6):e01411-15. doi:10.1128/genomeA.01411-15.

Copyright © 2015 Bergsveinson et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license. Address correspondence to Barry Ziola, b.ziola@usask.ca.

Prewery isolate *Lactobacillus brevis* BSO 464 (Lb464; obtained from the Brewing Research Foundation, Oxoid, UK) can resist the headspace pressure and dissolved CO<sub>2</sub> in packaged beer, making it a virulent beer-spoiler (1–3). Lb464 was grown overnight in MRS broth culture at 30°C and DNA was extracted with the UltraClean Microbial DNA Isolation Kit (Mo-Bio Laboratories, Carlsbad, CA). Heating at 10 min at 70°C was used prior to bead-beating to optimize cell lysis.

Sequencing was performed using the Roche 454 FLX platform at the National Research Council Plant Biotechnology Institute (NRC PBI; Saskatoon, SK). Two separate runs yielded 330,771 unpaired and 567,735 paired reads for ~30× coverage. Reads were assembled using Newbler GS *De Novo* assembler v2.5.3. Hawkeye (4) was used to visualize the 236 contigs across 17 scaffolds to identify contigs that could be potentially joined by sequencing of PCR amplicons (via the NRC PBI ABI 3700xl platform).

The Lb464 genome was assembled into a 2,503,991-bp circular chromosome (G+C content 45.7%) that had 8,461 bp cumulative of gaps due to transposase or repetitive regions not allowing PCRbased sequencing. Additionally, eight plasmids were assembled (G+C content ranged from 39.1% to 42.4%): pLb464-1 (15,324 bp), pLb464-2 (28,459 bp), pLb464-3 (22,411 bp), pLb464-4 (84,941 bp), pLb464-5 (10,867 bp), pLb464-6 (5018 bp), Lb464-7 (2353 bp), and pLb464-8 (49,835 bp, with a 1,000 bp gap due to repetitive regions) (2). Among sequenced lactobacilli, Lb464 has the second highest known plasmid number after L. brevis KB290 (KB290; GCA\_000359625.1) with nine plasmids (5). Mapping the Lb464 chromosome via DNA Plotter (6) revealed an atypical G+C-skew that has no symmetric bias in nucleotide composition of leading and lagging DNA strands relative to the replication origin (7). In Lb464, the AT-rich lagging strand encroaches into the normally G+C-rich leading strand by ~500 Kb. This could indicate a potential misassembly (inversion) of a ~500 Kb section of the Lb464 chromosome. However, the Lb464 G+C-skew is believed real due to similarity with the atypical chromosomal G+C-skew found in L. brevis ATCC 367<sup>T</sup> (Lb367; CP00416.1).

Lb464 genome annotation by the NCBI PGAP pathway (8)

produced 2,615 coding sequences, 6 rRNA operons, and 48 tRNAs (tRNAs for cysteine, histidine, and tryptophan are absent). One chromosomal clustered regularly interspaced short palindromic repeat (CRISPR) region is present, and 79 transposase genes of the ISL3 and IS30 family are found within the Lb464 genome, which is more than in Lb367 and KB290. Lb464 codes for several of the transcripts shown to be important in beer spoilage in a transcriptional study with Pediococcus claussenii (9), notably agmatine deiminase, putrescine carbamoyltransferase, and transporters malate, citrate, and other carbohydrate and nitrogen sources. Lb464 also contains the previously identified hop-tolerance genes hitA (10), horA (11), and horC (12) on pLb464-3, pLb464-1, and pLb464-2, respectively. However, only pLb464-2, pLb464-4, and pLb464-8, have been shown to be important for Lb464 growth in beer (2). Based on the PGAP annotation, these three plasmids code for 190 other proteins, including 106 hypothetical proteins, all of which need to be investigated for their contribution to the Lb464 beer-spoilage phenotype.

**Nucleotide sequence accession numbers.** The sequences for *Lactobacillus brevis* BSO 464 were deposited in GenBank under accession numbers CP005977, CP005978, CP005979, CP005980, CP005981, CP005982, CP005983, CP005984, and CP005985 for the chromosome and plasmids pLb464-1 to pLb464-8, respectively.

## **ACKNOWLEDGMENTS**

This study was funded by Discovery Grant 24067 awarded to B.Z. by the Natural Sciences and Engineering Research Council of Canada (NSERC). J.B. was supported by a Graduate Scholarship from the College of Graduate Studies at the University of Saskatchewan. She was also awarded the 2012 and 2013 Ecolab Scholarships, and the 2014 Roger C. Briess Scholarship by the American Society of Brewing Chemists (ASBC) Foundation. V.P. held an Alexander Graham Bell Canada Graduate Scholarship and a Michael Smith Foreign Study Supplement from NSERC, and was also the recipient of the MillerCoors, Anheuser-Busch InBev, and Brian Williams Graduate Scholarships from the ASBC Foundation. E.E. was the holder of an NSERC Undergraduate Student Research Award. N.B. was the recipient of a Barth-Haas Grant-in-Aid of Student Research.

<sup>\*</sup> Present address: Vanessa Pittet, Contango Strategies Ltd., Saskatoon, Saskatchewan, Canada.

## **REFERENCES**

- Bergsveinson J, Pittet V, Ziola B. 2012. RT-qPCR analysis of putative beer-spoilage gene expression during growth of *Lactobacillus brevis* BSO 464 and *Pediococcus claussenii* ATCC BAA-344<sup>T</sup> in beer. Appl Microbiol Biotechnol 96:461–470. http://dx.doi.org/10.1007/s00253-012-4334-3.
- Bergsveinson J, Baecker N, Pittet V, Ziola B. 2015. Role of plasmids in Lactobacillus brevis BSO 464 hop tolerance and beer spoilage. Appl Environ Microbiol 81:1234–1241. http://dx.doi.org/10.1128/AEM.02870-14.
- Bergsveinson J, Redekop A, Zoerb S, Ziola B. 2015. Dissolved carbon dioxide selects for lactic acid bacteria able to grow in and spoil packaged beer. J Am Soc Brew Chem 73:331–338. http://dx.doi.org/10.1094/ASBCJ -2015-0726-01.
- 4. Schatz MC, Phillippy AM, Shneiderman B, Salzberg SL. 2007. Hawkeye: an interactive visual analytics tool for genome assemblies. Genome Biol 8:R34. http://dx.doi.org/10.1186/gb-2007-8-3-r34.
- Fukao M, Oshima K, Morita H, Toh H, Suda W, Kim S, Suzuki S, Yakabe T, Hattori M, Yajima N. 2013. Genomic analysis by deep sequencing of the probiotic *Lactobacillus brevis* KB290 harboring nine plasmids reveals genomic stability. PLoS One 8:e60521. http://dx.doi.org/ 10.1371/journal.pone.0060521.
- Carver T, Thomson N, Bleasby A, Berriman M, Parkhill J. 2009. DNAPlotter: circular and linear interactive genome visualization. Bioinformatics 25:119–120. http://dx.doi.org/10.1093/bioinformatics/btn578.
- 7. Charneski CA, Honti F, Bryant JM, Hurst LD, Feil EJ, Petrov DA. 2011.

- Atypical AT skew in Firmicute genomes results from selection and not from mutation. PLoS Genet 7:e1002283. http://dx.doi.org/10.1371/journal.pgen.1002283.
- 8. Tautosova T, DiCuccio M, Badretdin A. 2013. Prokaryotic genome annotation pipeline. The NCBI handbook [Internet], 2nd ed. National Center for Biotechnology Information, Bethesda, MD: http://www.ncbi.nlm.nih.gov/books/NBK174280/.
- Pittet V, Phister TG, Ziola B. 2013. Transcriptome sequence and plasmid copy number analysis of the brewery isolate *Pediococcus claussenii* ATCC BAA-344<sup>T</sup> during growth in beer. PLoS One 8:e73627. http://dx.doi.org/ 10.1371/journal.pone.0073627.
- Sami M, Yamashita H, Hirono T, Kadokura H, Kitamoto K, Yoda K, Yamasaki M. 1997. Hop-resistant Lactobacillus brevis contains a novel plasmid harboring a multidrug resistance-like gene. J Ferment Bioeng 84:1–6. http://dx.doi.org/10.1016/S0922-338X(97)82778-X.
- Suzuki K, Iijima K, Ozaki K, Yamashita H. 2005. Isolation of a hopsensitive variant of *Lactobacillus lindneri* and identification of genetic markers for beer spoilage ability of lactic acid bacteria. Appl Environ Microbiol 71:5089–5097. http://dx.doi.org/10.1128/AEM.71.9.5089 -5097.2005
- Hayashi N, Ito M, Horiike S, Taguchi H. 2001. Molecular cloning of a putative divalent-cation transporter gene as a new genetic marker for the identification of *Lactobacillus brevis* strains capable of growing in beer. Appl Microbiol Biotechnol 55:596–603. http://dx.doi.org/10.1007/ s002530100600.