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# **The Perplexing Paradox of Paraquat: the Case for Host-Based Susceptibility and Postulated Neurodegenerative Effects**

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### **Abstract**

Paraquat is an herbicide used extensively in agriculture, and has also been proposed to be a risk factor for Parkinson's disease. To date, experimental, clinical, and epidemiological data on paraquat neurotoxicity have been equivocal. In this short review, we discuss some technical and biological mechanisms that contribute to inconsistencies regarding paraquat neurotoxicity. We hypothesize that individual genetic variations in susceptibility generate major differences in neurotoxic risk and functional outcome. Identifying these heritable sources of variation in host susceptibility, and their role in complex gene-environment interactions, is crucial to identify risk biomarkers, and to devise better prevention and treatment for those exposed to paraquat and other potential neurotoxicants.

#### **Keywords**

host susceptibility; complex traits; toxicogenetics; paraquat; Parkinson's disease; neurodegeneration; iron

# **Introduction**

It is becoming increasingly evident that many neurodegenerative diseases can be classified as familial vs. sporadic. Familial forms are usually manifest at earlier stages of life, and can have high heritability. Consequently, these forms have been more tractable for genetic analyses, thus well-defined gene variants have been linked to risk of Alzheimer's (1) and Parkinson's diseases (2). Conversely, the etiology of sporadic neurodegenerative diseases is

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far more complex, and almost certainly involves gene-environment interactions (3). Moreover, heritable factors likely involve multiple genes making the scientific investigation into the etiology of these diseases challenging since not all "susceptible" genotypes are exposed to environments that contain risk factors. Sporadic Parkinson's disease (sPD) provides a useful example. Suggested environmental risk factors for sPD include rural living and the drinking of well water (4). Rural living also is physically associated with agriculture and the consequent exposure to chemicals used to control weeds, fungi, and insects. Some of these chemicals have been shown to be associated with increased risk of neurodegenerative diseases. Identifying those individuals carrying toxicant-sensitive genotypes is particularly challenging, but can provide the foundation for making inroads into the investigation into host susceptibility.

One such chemical, paraquat, is an herbicide that is used nearly worldwide in agriculture, and in other scenarios where weed control is desired. When handled improperly or ingested deliberately, it can cause acute pulmonary damage and death. Possibly of even broader impact is the fact that chronic exposure to paraquat has also been associated with a higher risk of sPD, although inconsistencies in epidemiological and animal studies have led to controversy. Our goal is to review the literature that may guide future research directions that can test the hypothesis that there is a relationship between paraquat exposure and risk of sPD.

#### **Is paraquat a red herring?**

In an article entitled "Paraquat: the red herring of Parkinson's disease research," Miller (5) argued that the risk for developing PD following paraquat exposure was over-stated. His view was that although paraquat is structurally similar to the well-known dopamine toxicant MPP+, there were few data linking paraquat to disruption of mitochondrial complex I. In addition, he stated that in animal models paraquat causes no more than 25% loss of dopamine neurons in the substantia nigra pars compacta (SNc), whereas early symptoms of sPD require upwards of 70% loss of these neurons. Others have rebutted this assertion noting that not only has paraquat been demonstrated to destroy dopamine neurons in rodents, but paraquat toxicity is likely a complex interaction of both biological and environmental factors  $(6,7)$ . Lopachin and Gavin (7) further noted that MPP<sup>+</sup> toxicity may not be exclusively related to mitochondrial toxicity, and there is little evidence that paraquat and MPP+ are toxicologically similar despite their structural similarity. Indeed, Choi et al. (8) have shown that, at least in cell culture, complex I inhibition by paraquat is not required for dopaminergic neuron destruction by paraquat. Nevertheless, Miller's perspective (5) has been influential, possibly due to inconsistencies in epidemiological data about the influence of paraquat on risk of sPD. We offer an alternate viewpoint that is based on a widely-held hypothesis that sPD is a complex disease influenced by the interaction of genetic and environmental factors. We hypothesize that the genetic make-up of PQ-exposed animals (human or laboratory) plays a major role in host-based susceptibility (i.e., the variability in the risk of sPD in paraquat-exposed individuals).

# **Sporadic Parkinson's disease is a complex trait influenced by multiple genes and environmental factors**

After Alzheimer's disease, PD is the second most common neurodegenerative disorder, afflicting more than 1% of individuals over 65 years of age (9). The primary symptoms are resting tremor, progressive rigidity, hyperkinesias, and finally, difficulty in initiating voluntary movement and postural instability. The pathobiological hallmarks of PD include dopaminergic (DA) neuron loss in the substantia nigra pars compacta (SNc), and  $>60\%$ decrease in striatal dopamine concentrations at first clinical diagnosis. The suggested causes are many, and many investigators (e.g., 10;11) have hypothesized that sPD is a complex trait involving multiple cellular processes, each of which is modulated by multiple gene variants acting in concert with environmental factors, together causing a net increase or decrease in risk.

#### **sPD is of particular concern, although not limited to, rural communities**

In 1993, the National Institutes of Health launched a large study of agricultural health that continues to the present. Participating agencies include the National Cancer Institute, the National Institute of Environmental Health Sciences, the Environmental Protection Agency and the national Institute for Occupational Safety and Health. Although most of the research published from this effort concerns pesticide exposure and various cancers, the NIEHS epidemiology group led by Kamel has addressed the association between exposure to pesticides and neurodegenerative disease, including Parkinson's disease (12–15). This group has advanced our knowledge about the associated risk and most recently showed that identification of subpopulations within epidemiological studies (16) can elucidate the basis for individual differences in susceptibility to paraquat neurotoxicity.

A meta-analysis of more than 200 studies examining risk factors for sPD revealed significantly increased risk for sPD from pesticides, rural living, and consumption of well water (4;17). Exposure to insecticides and herbicides used in agriculture has been implicated in increased sPD risk. One of the most commonly used herbicides (although now banned in much of Northern Europe) is paraquat (PQ). PQ has a long environmental half-life and has been implicated as a risk factor for sPD. In the environment, PQ leaches into ground water and can remain at 80% retention in river water for nearly two years (18;19).

#### **Is exposure to PQ a risk factor for sPD?**

The evidence for PQ as risk for sPD is mixed, both in clinical-epidemiological and preclinical studies. Among nine epidemiological studies, six showed that PQ increased risk (20–25), whereas three did not (17;26;27). Berry et al. (28) were highly critical of both epidemiological and preclinical studies that had associated PQ exposure and sPD, arguing that the epidemiological studies failed to show a convincing link between PQ and sPD because of poor definition of exposure. They also argued that preclinical studies forced a comparison with the toxicology of MPTP, and ignored toxicokinetic issues necessary for PQ to enter neurons (28).

## **Why the disparity?**

Epidemiological studies tend to sample populations broadly, and may therefore yield either Type I or Type II errors when subgroups of the sampled population respond differently to pathogens, toxicants, etc. This may be relevant in evaluating the PQ-induced risk for sPD. Indeed, in a recent study, Goldman et al. (16) showed that individuals homozygous for a null mutation in the glutathione S-transferase theta 1 gene and who were exposed to PQ had an odds ratio (OR) of 11.1 for developing sPD as compared to an OR of 1.5 for those not carrying the mutation and exposed to PQ. Although the number of subjects was small, the data implies that there subpopulations of humans (and mice?) that are differentially susceptible to the neurotoxic effects of PQ (and probably other toxicants as well). The task then becomes one of identifying those susceptible and those factors that confer individual differences in risk.

#### **What about the preclinical research?**

The principal pathophysiological sign of PD is loss of dopamine neuron perikarya in the SNc, with consequent loss of DA in the striatum. MPTP, a pro-neurotoxicant that targets the SNc DA neurons, is commonly used to model the disease *in vivo* and *in vitro*. MPTP intoxication provides a superb model of the symptoms and signs of PD in humans and nonhuman primates (29–31), and also responds essentially identically to approved symptomatic antiparkinson drugs (32). Thus, it has become the research standard often used to evaluate both the effects of other toxicants, and also therapeutic interventions. Although heuristically useful, the MPTP model has, at least to date, failed to translate into successful diseasemodifying therapies for sPD, the reasons for which are not relevant here. What is important is how this may educate us on the role of environmental chemicals as a contributor to sPD, specifically in that it is highly likely that there are individual differences in toxic responses to MPTP. Thus, we have shown recently that MPTP neurotoxicity varied dramatically among inbred mouse strains (11;33). These data suggest that responses to PQ also are very dependent on factors such as age, strain, species, etc. Can this explain why a recent study by Breckenridge et al. (34) did not find PQ-related toxicity in C57BL/6J mice, whereas other studies have reported PQ-associated loss of DA neurons (35–38), as well as loss of striatal DA function and associated behavioral problems (36)?

#### **Why the disparity?**

All of these studies used the same C57BL/6 strain. Breckenridge et al. (34), however, used mice that were only two months old (equivalent to late adolescence in humans), and found no effects of PQ. Conversely, studies in which older animals were used have consistently reported PQ-induced effects on dopamine neurons, including animals that were 3–4 mo (37), 4–6 mo (38), and 10 mo of age (36). McCormack et al. (35) tested mice at 6 weeks, 5, or 18 months of age for PQ toxicity (three weekly injections of 10 mg/kg ip), and found that the number of dopamine neurons in the SNc decreased significantly with age, with the greatest loss observed in the 18 mo animals. Since age is the most critical variable in sPD, it is not surprising that it might also be a factor in response to toxicants that might be related to sPD etiology (11;33).

#### **Host characteristics – the key to understanding the complexity of sPD**

The epidemiological work of Goldman et al. (16) is illustrative of how identification of host characteristics can elucidate the relative risk for sPD in those exposed to PQ. Similarly, in animal models the age of the animal appears to be a critical factor in susceptibility to PQ neurotoxicity. What other host characteristics may influence PQ toxicity? Recently, we demonstrated that strain differences in reduction in tyrosine hydroxylase-positive (TH+) neurons in the SNc by PQ is related to PQ-induced iron increases in the ventral midbrain, including the SNc (37). Iron has been shown to act synergistically with PQ in nigrostriatal neuron loss (39). Among the many proposed mechanisms of PQ neurotoxicity, one important factor may be genetic differences in how PQ disrupts iron regulation in the SNc. We hypothesize that those individuals whose exposure to PQ increases iron in the SNc will be most susceptible to PQ-induced parkinsonian toxicity.

#### **Iron regulation as a major factor in the etiology of PQ-induced sPD**

Peng et al. (40) demonstrated that PQ applied to a primary culture of mesencephalic neurons did not affect survival of TH+ neurons, yet when the culture consisted of neurons and glia, PQ caused a 40%% reduction in TH+ neurons. Adding iron to the culture medium increased this effect by 10%. This suggests that glia may be critical for PQ neurotoxicity and that iron exacerbates this toxicity. Moreover, Peng et al. (40) also showed that microglial activation *per se* results from iron-related oxidative stress. Inhibiting this oxidative stress by inactivating superoxide reduced the loss of PQ- and PQ+Fe treated neurons from the neuronglia primary culture (40). More recently, Rappold et al. (41) showed that PQ exists in two oxidative states,  $PQ^{2+}$  and  $PQ^{+}$ , and that the reduced form enters cells via the dopamine transporter (DAT) more readily than does  $PQ^{2+}$ . Moreover, Rappold et al. (41) also found that microglia reduce  $PQ^{2+}$  using NADPH oxidase in humans and microglial nitric oxide synthase in mice.

It appears, then, that a limiting factor for PQ neurotoxicity may be the availability of iron. Iron activates microglia that in turn reduce  $PQ^{2+}$  to  $PQ^{+}$  which can enter neurons via DAT and by organic cation transporter-3 (41). Rhodes and Ritz (42) have proposed that the genetics of iron regulation may play a major role in the degree of sPD risk resulting from PQ exposure. Relevant to this, Wu et al. (43) reported that there was increased iron concentrations in the SN (pars reticulata), globus pallidus, and red nucleus in humans who had suffered acute PQ poisoning at 6, 12, and 24 mo following intoxication, although the increase was significant in the globus pallidus and SNpr only at the last time point. These findings suggest the importance of determining whether acute PQ poisoning may disrupt iron homeostasis over the long term.

#### **Iron accumulation in the SNc per se is associated with sPD**

Iron regulation in the brain (as well as other tissues) is highly complex and requires the participation of many proteins that sequester, change oxidative state, transport and otherwise control availability of free or loosely bound iron. The genetics and genomics of these proteins provide ample opportunity for individual differences in iron regulation, especially in the SNc. Moreover, iron deposition in the SNc increases progressively with age in

humans (44), rats (45) and mice (46). There is agreement about such changes even using different methodologies such as MRI imaging (47), sonography (48), and postmortem examination. Notably, Sofic et al. (49) showed in postmortem samples that a progressive iron increase in the SNc is a feature of advanced PD. What is not known is whether increases in iron concentration in regions like the SNc are a cause or a consequence (or both) of sPD.

## **The biology of PQ neurotoxicity**

SPD is a highly complex disease and many factors likely contribute to its etiology. Although we recognize that there are several likely pathways, one major pathway is by way of cooperation between iron and PQ. In our model, in susceptible individuals, PQ disrupts iron regulation in the SNc, causing an increase in iron. This iron influx in turn activates microglia, which reduce  $PQ^{2+}$  to  $PQ^{+}$ , thus facilitating uptake into DA neurons to produce ROS and cell damage/death (41). We have demonstrated genetic differences in the capacity of PQ to disrupt iron regulation in mice (37), and we propose that the same genetic differences in PQ disruption of SNc iron regulation occurs in humans and thus is another key to understanding individual differences in susceptibility to PQ neurotoxicity. Of course, iron regulation is only one of many suspected host characteristics. Others include polymorphisms in the dopamine transporter gene (50) and in the divalent metal transporter 1 gene (51).

#### **How can we use this information in future PD related research?**

The first lesson to be learned here is how to identify those individuals who are at relatively greater risk for PQ-related neurological damage – and not just paraquat but other toxicants as well. Genotyping has become relatively inexpensive and markers identified from genome-wide association studies and from complementary animal studies will prove to be instrumental in prevention (52). The use of genetically defined mice can prove useful in identifying candidate genes as the mouse and human genomes are more than 90% syntenic.

In parallel to studies in animals, it is timely to determine how genetic makeup influences individual susceptibility to environmental toxicants that play a role in the development of sPD, and its progression in human. This shall pave a critical step for personalized prevention and possible treatment. Of course, such studies may also offer insight into general mechanisms of sPD. In addition to there are multifactorial etiological factors (both environmental and genetic) that are involved, there are, however, several other challenges to studying the interaction of genetic and environmental factors in neurodegenerative diseases like PD in humans. These include: 1) a long preclinical period prior to the onset of clinical symptoms and 2) a lack of reliable human biochemical, molecular, or imaging biomarkers.

Imaging using both radioligands that label dopamine terminals and magnetic resonance imaging (MRI) has been applied extensively to study PD-related pathological changes in human subjects (53–55). In particular, diffusion tensor imaging (DTI), by measuring microstructural disorganization, has shown promise as a tool of PD-related changes (53;56;57). In the MPTP mouse model, DTI changes have been associated with dopamine neuron loss in the SN (58). Furthermore, several human studies have demonstrated reduced

FA values in the SN of early PD patients, consistent with the notion that DTI changes may be able to detect nigral changes *in vivo* (47;55). Recently, we have reported that subtoxic, chronic pesticide exposure of asymptomatic agriculture workers leads to DTI changes in the SN similar to those seen in PD patients (59), suggesting that such changes may represent the consequence of pesticide-induced toxicity to nigral circuitry. These findings are consistent with the previously proposed role of pesticide exposure in initiating or accelerating pathological processes that are similar to those occurring in PD. More importantly, the study demonstrated that DTI could detect microstructural changes in the SN possibly caused by pesticides. This approach may be useful in determining if other environmental chemicals alter brain microstructure, and/or can assist in focusing on specific chemicals that affect PD susceptibility.

#### **Overall significance**

sPD is more prevalent in rural areas than in urban areas and is most likely the result of geneenvironment interactions. Not only is potential exposure to pesticides and some other environmental chemicals more common in rural areas, many individuals living on farms or in the country also drink well water that ranges widely in iron concentration. Iron homeostasis involves many proteins whose expression varies widely in different individuals (60–65). We hypothesize that iron may be an important factor in the toxicity of PQ, and that a systems genetics approach can help to elucidate some of the critical mechanisms. Although the interaction of genetic and environment factors is widely thought to be important for sPD, it has created a complex experimental template. Identification of the genetic mechanisms underlying individual differences in susceptibility to PQ neurotoxicity has great promise in developing new strategies for the prevention and treatment of environmentally-related sPD, and experimental tools may now exist to, at least partially, untangle these factors.

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#### **References**

- 1. Bertram L, Lill CM, Tanzi RE. The genetics of Alzheimer disease: back to the future. Neuron. 2010; 68:270–281. [PubMed: 20955934]
- 2. Trinh J, Farrer M. Advances in the genetics of Parkinson disease. Nat. Rev. Neurol. 2013; 9:445– 454. [PubMed: 23857047]
- 3. Kieburtz K, Wunderle KB. Parkinson's disease: evidence for environmental risk factors. Mov Disord. 2013; 28:8–13. [PubMed: 23097348]
- 4. Noyce AJ, Bestwick JP, Silveira-Moriyama L, Hawkes CH, Giovannoni G, Lees AJ, Schrag A. Meta-analysis of early nonmotor features and risk factors for Parkinson disease. Ann. Neurol. 2012; 72:893–901. [PubMed: 23071076]
- 5. Miller GW. Paraquat: the red herring of Parkinson's disease research. Toxicol. Sci. 2007; 100:1–2. [PubMed: 17934192]
- 6. Cory-Slechta DA, Thiruchelvam M, Di Monte DA. Letter regarding:"Paraquat: the red herring of Parkinson's disease research". Toxicol. Sci. 2008; 103:215–216. [PubMed: 18162474]

- 8. Choi WS, Kruse SE, Palmiter RD, Xia Z. Mitochondrial complex I inhibition is not required for dopaminergic neuron death induced by rotenone, MPP+, or paraquat. Proc. Natl. Acad. Sci. U. S. A. 2008; 105:15136–15141. [PubMed: 18812510]
- 9. Moore DJ, West AB, Dawson VL, Dawson TM. Molecular pathophysiology of Parkinson's disease. Annu. Rev. Neurosci. 2005; 28:57–87. [PubMed: 16022590]
- 10. Gwinn-Hardy K. Genetics of parkinsonism. Mov Disord. 2002; 17:645–656. [PubMed: 12210852]
- 11. Jones BC, Lu L, Williams RW, Unger EL, Yin L. Response to Breckenridge et al. Neurotoxicology. 2014 in press.
- 12. Kamel F, Boyes WK, Gladen BC, Rowland AS, Alavanja MC, Blair A, Sandler DP. Retinal degeneration in licensed pesticide applicators. Am. J. Ind. Med. 2000; 37:618–628. [PubMed: 10797505]
- 13. Kamel F, Tanner C, Umbach D, Hoppin J, Alavanja M, Blair A, Comyns K, Goldman S, Korell M, Langston J, Ross G, Sandler D. Pesticide exposure and self-reported Parkinson's disease in the agricultural health study. Am. J. Epidemiol. 2007; 165:364–374. [PubMed: 17116648]
- 14. Kamel F, Umbach DM, Bedlack RS, Richards M, Watson M, Alavanja MC, Blair A, Hoppin JA, Schmidt S, Sandler DP. Pesticide exposure and amyotrophic lateral sclerosis. Neurotoxicology. 2012; 33:457–462. [PubMed: 22521219]
- 15. Kamel F, Goldman SM, Umbach DM, Chen H, Richardson G, Barber MR, Meng C, Marras C, Korell M, Kasten M, Hoppin JA, Comyns K, Chade A, Blair A, Bhudhikanok GS, Webster RG, William LJ, Sandler DP, Tanner CM. Dietary fat intake, pesticide use, Parkinson's disease. Parkinsonism. Relat Disord. 2013
- 16. Goldman SM, Kamel F, Ross GW, Bhudhikanok GS, Hoppin JA, Korell M, Marras C, Meng C, Umbach DM, Kasten M, Chade AR, Comyns K, Richards MB, Sandler DP, Blair A, Langston JW, Tanner CM. Genetic modification of the association of paraquat and Parkinson's disease. Mov Disord. 2012; 27:1652–1658. [PubMed: 23045187]
- 17. Gatto NM, Cockburn M, Bronstein J, Manthripragada AD, Ritz B. Well-water consumption and Parkinson's disease in rural California. Environ. Health Perspect. 2009; 117:1912–1918. [PubMed: 20049211]
- 18. Wang YS, Yen JH, Hsieh YN, Chen YL. Dissipation of 2,4-D glyphosate and paraquat in river water. Water, Air, & Soil Pollution. 1994; 72:1–4.
- 19. Fernandez M, Ibanez M, Pico Y, Manes J. Spatial and temporal trends of paraquat, diquat, and difenzoquat contamination in water from marsh areas of the valencian community (Spain). Arch. Environ. Contam Toxicol. 1998; 35:377–384. [PubMed: 9732466]
- 20. Hertzman C, Wiens M, Bowering D, Snow B, Calne D. Parkinson's disease: a case-control study of occupational and environmental risk factors. Am. J. Ind. Med. 1990; 17:349–355. [PubMed: 2305814]
- 21. Liou HH, Tsai MC, Chen CJ, Jeng JS, Chang YC, Chen SY, Chen RC. Environmental risk factors and Parkinson's disease: a case-control study in Taiwan. Neurology. 1997; 48:1583–1588. [PubMed: 9191770]
- 22. Landrigan PJ, Sonawane B, Butler RN, Trasande L, Callan R, Droller D. Early environmental origins of neurodegenerative disease in later life. Environ. Health Perspect. 2005; 113:1230–1233. [PubMed: 16140633]
- 23. Tanner CM, Kamel F, Ross GW, Hoppin JA, Goldman SM, Korell M, Marras C, Bhudhikanok GS, Kasten M, Chade AR, Comyns K, Richards MB, Meng C, Priestley B, Fernandez HH, Cambi F, Umbach DM, Blair A, Sandler DP, Langston JW. Rotenone, paraquat, and Parkinson's disease. Environ. Health Perspect. 2011; 119:866–872. [PubMed: 21269927]
- 24. Wang A, Costello S, Cockburn M, Zhang X, Bronstein J, Ritz B. Parkinson's disease risk from ambient exposure to pesticides. Eur. J. Epidemiol. 2011; 26:547–555. [PubMed: 21505849]
- 25. McCormack AL, Atienza JG, Johnston LC, Andersen JK, Vu S, Di Monte DA. Role of oxidative stress in paraquat-induced dopaminergic cell degeneration. J Neurochem. 2005; 93:1030–1037. [PubMed: 15857406]

- 26. Elbaz A, Clavel J, Rathouz PJ, Moisan F, Galanaud JP, Delemotte B, Alperovitch A, Tzourio C. Professional exposure to pesticides and Parkinson disease. Ann. Neurol. 2009; 66:494–504. [PubMed: 19847896]
- 27. Firestone JA, Lundin JI, Powers KM, Smith-Weller T, Franklin GM, Swanson PD, Longstreth WT Jr, Checkoway H. Occupational factors and risk of Parkinson's disease: A population-based casecontrol study. Am. J. Ind. Med. 2010; 53:217–223. [PubMed: 20025075]
- 28. Berry C, La VC, Nicotera P. Paraquat and Parkinson's disease. Cell Death. Differ. 2010; 17:1115– 1125. [PubMed: 20094060]
- 29. Burns RS, Chiueh CC, Markey SP, Ebert MH, Jacobowitz DM, Kopin IJ. A primate model of parkinsonism: selective destruction of dopaminergic neurons in the pars compacta of the substantia nigra by N-methyl-4- phenyl-1,2,3,6-tetrahydropyridine. Proc. Natl. Acad. Sci. U. S. A. 1983; 80:4546–4550. [PubMed: 6192438]
- 30. Davis GC, Williams AC, Markey SP, Ebert MH, Caine ED, Reichert CM, Kopin IJ. Chronic Parkinsonism secondary to intravenous injection of meperidine analogues. Psychiatry Res. 1979; 1:249–254. [PubMed: 298352]
- 31. Langston JW, Ballard P, Tetrud JW, Irwin I. Chronic Parkinsonism in humans due to a product of meperidine-analog synthesis. Science. 1983; 219:979–980. [PubMed: 6823561]
- 32. Mailman, RB.; Huang, X. Dopamine receptor pharmacology. In: Koller, WC.; Melamed, E., editors. Parkinson's disease and related disorders, Part 1. Elsevier; 2007. p. 77-105.
- 33. Jones BC, Miller DB, O'Callaghan JP, Lu L, Unger EL, Alam G, Williams RW. Systems analysis of genetic variation in MPTP neurotoxicity in mice. Neurotoxicology. 2013; 37:26–34. [PubMed: 23558233]
- 34. Breckenridge CB, Sturgess NC, Butt M, Wolf JC, Zadory D, Beck M, Mathews JM, Tisdel MO, Minnema D, Travis KZ, Cook AR, Botham PA, Smith LL. Pharmacokinetic, neurochemical, stereological and neuropathological studies on the potential effects of paraquat in the substantia nigra pars compacta and striatum of male C57BL/6J mice. Neurotoxicology. 2013; 37:1–14. [PubMed: 23523781]
- 35. McCormack AL, Thiruchelvam M, Manning-Bog AB, Thiffault C, Langston JW, Cory-Slechta DA, Di Monte DA. Environmental risk factors and Parkinson's disease: selective degeneration of nigral dopaminergic neurons caused by the herbicide paraquat. Neurobiol. Dis. 2002; 10:119–127. [PubMed: 12127150]
- 36. Li X, Yin J, Cheng CM, Sun JL, Li Z, Wu YL. Paraquat induces selective dopaminergic nigrostriatal degeneration in aging C57BL/6 mice. Chin Med. J. (Engl.). 2005; 118:1357–1361. [PubMed: 16157030]
- 37. Yin L, Lu L, Prasad K, Richfield EK, Unger EL, Xu J, Jones BC. Genetic-based, differential susceptibility to paraquat neurotoxicity in mice. Neurotoxicol. Teratol. 2011; 33:415–421. [PubMed: 21371552]
- 38. Jiao Y, Lu L, Williams RW, Smeyne RJ. Genetic dissection of strain dependent paraquat-induced neurodegeneration in the substantia nigra pars compacta. PLoS.One. 2012; 7:e29447. [PubMed: 22291891]
- 39. Peng J, Peng L, Stevenson FF, Doctrow SR, Andersen JK. Iron and paraquat as synergistic environmental risk factors in sporadic Parkinson's disease accelerate age-related neurodegeneration. J. Neurosci. 2007; 27:6914–6922. [PubMed: 17596439]
- 40. Peng J, Stevenson FF, Oo ML, Andersen JK. Iron-enhanced paraquat-mediated dopaminergic cell death due to increased oxidative stress as a consequence of microglial activation. Free Radic. Biol. Med. 2009; 46:312–320. [PubMed: 19027846]
- 41. Rappold PM, Cui M, Chesser AS, Tibbett J, Grima JC, Duan L, Sen N, Javitch JA, Tieu K. Paraquat neurotoxicity is mediated by the dopamine transporter and organic cation transporter-3. Proc. Natl. Acad. Sci. U. S. A. 2011; 108:20766–20771. [PubMed: 22143804]
- 42. Rhodes SL, Ritz B. Genetics of iron regulation and the possible role of iron in Parkinson's disease. Neurobiol. Dis. 2008; 32:183–195. [PubMed: 18675357]
- 43. Wu B, Song B, Tian S, Huo S, Cui C, Guo Y, Liu H. Central nervous system damage due to acute paraquat poisoning: a neuroimaging study with 3.0 T MRI. Neurotoxicology. 2012; 33:1330– 1337. [PubMed: 22947519]

- 44. Daugherty A, Raz N. Age-related differences in iron content of subcortical nuclei observed in vivo: a meta-analysis. Neuroimage. 2013; 70:113–121. [PubMed: 23277110]
- 45. Hunter RL, Liu M, Choi DY, Cass WA, Bing G. Inflammation and age-related iron accumulation in F344 rats. Curr. Aging Sci. 2008; 1:112–121. [PubMed: 20021380]
- 46. Kaur D, Peng J, Chinta SJ, Rajagopalan S, Di Monte DA, Cherny RA, Andersen JK. Increased murine neonatal iron intake results in Parkinson-like neurodegeneration with age. Neurobiol. Aging. 2007; 28:907–913. [PubMed: 16765489]
- 47. Du G, Lewis MM, Sen S, Wang J, Shaffer ML, Styner M, Yang QX, Huang X. Imaging nigral pathology and clinical progression in Parkinson's disease. Mov Disord. 2012; 27:1636–1643. [PubMed: 23008179]
- 48. Double KL, Todd G, Duma SR. Pathophysiology of transcranial sonography signal changes in the human substantia nigra. Int. Rev. Neurobiol. 2010; 90:107–120. [PubMed: 20692497]
- 49. Sofic E, Paulus W, Jellinger K, Riederer P, Youdim MB. Selective increase of iron in substantia nigra zona compacta of parkinsonian brains. J Neurochem. 1991; 56:978–982. [PubMed: 1704426]
- 50. Ritz BR, Manthripragada AD, Costello S, Lincoln SJ, Farrer MJ, Cockburn M, Bronstein J. Dopamine transporter genetic variants and pesticides in Parkinson's disease. Environ. Health Perspect. 2009; 117:964–969. [PubMed: 19590691]
- 51. Garcia-Closas M, Hall P, Nevanlinna H, Pooley K, Morrison J, Richesson DA, Bojesen SE, Nordestgaard BG, Axelsson CK, Arias JI, Milne RL, Ribas G, Gonzalez-Neira A, Benitez J, Zamora P, Brauch H, Justenhoven C, Hamann U, Ko YD, Bruening T, Haas S, Dork T, Schurmann P, Hillemanns P, Bogdanova N, Bremer M, Karstens JH, Fagerholm R, Aaltonen K, Aittomaki K, von SK, Blomqvist C, Mannermaa A, Uusitupa M, Eskelinen M, Tengstrom M, Kosma VM, Kataja V, Chenevix-Trench G, Spurdle AB, Beesley J, Chen X, Devilee P, van Asperen CJ, Jacobi CE, Tollenaar RA, Huijts PE, Klijn JG, Chang-Claude J, Kropp S, Slanger T, Flesch-Janys D, Mutschelknauss E, Salazar R, Wang-Gohrke S, Couch F, Goode EL, Olson JE, Vachon C, Fredericksen ZS, Giles GG, Baglietto L, Severi G, Hopper JL, English DR, Southey MC, Haiman CA, Henderson BE, Kolonel LN, Le ML, Stram DO, Hunter DJ, Hankinson SE, Cox DG, Tamimi R, Kraft P, Sherman ME, Chanock SJ, Lissowska J, Brinton LA, Peplonska B, Klijn JG, Hooning MJ, Meijers-Heijboer H, Collee JM, van den Ouweland A, Uitterlinden AG, Liu J, Lin LY, Yuqing L, Humphreys K, Czene K, Cox A, Balasubramanian SP, Cross SS, Reed MW, Blows F, Driver K, Dunning A, Tyrer J, Ponder BA, Sangrajrang S, Brennan P, McKay J, Odefrey F, Gabrieau V, Sigurdson A, Doody M, Struewing JP, Alexander B, Easton DF, Pharoah PD. Heterogeneity of breast cancer associations with five susceptibility loci by clinical and pathological characteristics. PLoS. Genet. 2008; 4:e1000054. [PubMed: 18437204]
- 52. Ermann J, Glimcher LH. After GWAS: mice to the rescue? Curr. Opin. Immunol. 2012; 24:564– 570. [PubMed: 23031443]
- 53. Chan LL, Rumpel H, Yap K, Lee E, Loo HV, Ho GL, Fook-Chong S, Yuen Y, Tan EK. Case control study of diffusion tensor imaging in Parkinson's disease. J. Neurol. Neurosurg. Psychiatry. 2007; 78:1383–1386. [PubMed: 17615165]
- 54. Martin WR, Wieler M, Gee M. Midbrain iron content in early Parkinson disease: a potential biomarker of disease status. Neurology. 2008; 70:1411–1417. [PubMed: 18172063]
- 55. Vaillancourt DE, Spraker MB, Prodoehl J, Abraham I, Corcos DM, Zhou XJ, Comella CL, Little DM. High-resolution diffusion tensor imaging in the substantia nigra of de novo Parkinson disease. Neurology. 2009; 72:1378–1384. [PubMed: 19129507]
- 56. Peran P, Cherubini A, Assogna F, Piras F, Quattrocchi C, Peppe A, Celsis P, Rascol O, Demonet JF, Stefani A, Pierantozzi M, Pontieri FE, Caltagirone C, Spalletta G, Sabatini U. Magnetic resonance imaging markers of Parkinson's disease nigrostriatal signature. Brain. 2010; 133:3423– 3433. [PubMed: 20736190]
- 57. Du G, Lewis MM, Styner M, Shaffer ML, Sen S, Yang QX, Huang X. Combined R2\* and Diffusion Tensor Imaging Changes in the Substantia Nigra in Parkinson's Disease. Mov Disord. 2011
- 58. Boska MD, Hasan KM, Kibuule D, Banerjee R, McIntyre E, Nelson JA, Hahn T, Gendelman HE, Mosley RL. Quantitative diffusion tensor imaging detects dopaminergic neuronal degeneration in a murine model of Parkinson's disease. Neurobiol. Dis. 2007; 26:590–596. [PubMed: 17428671]

- 59. Du G, Lewis MM, Sterling NW, Kong L, Chen H, Mailman RB, Huang X. Microstructural changes in the substantia nigra of asymptomatic agriculture workers. Neurobehav. Tox. Teratol. 2014 in press.
- 60. Connor JR. Iron transport proteins in the diseased brain. J Neurol. Sci. 2003; 207:112–113. [PubMed: 12614944]
- 61. Connor JR. Iron acquisition and expression of iron regulatory proteins in the developing brain: manipulation by ethanol exposure, iron deprivation and cellular dysfunction. Dev. Neurosci. 1994; 16:233–247. [PubMed: 7768202]
- 62. Connor JR, Benkovic SA. Iron regulation in the brain: histochemical, biochemical, and molecular considerations. Ann. Neurol. 1992; 32(Suppl):S51–S61. [PubMed: 1510381]
- 63. Liu Y, Connor JR. Iron and ER stress in neurodegenerative disease. Biometals. 2012; 25:837–845. [PubMed: 22526559]
- 64. Snyder AM, Connor JR. Iron, the substantia nigra and related neurological disorders. Biochim. Biophys. Acta. 2009; 1790:606–614. [PubMed: 18778755]
- 65. Tandara L, Salamunic I. Iron metabolism: current facts and future directions. Biochem. Med. (Zagreb.). 2012; 22:311–328. [PubMed: 23092063]