

Original Article

Alterations in vitamin D signaling pathway in gastric cancer progression: a study of vitamin D receptor expression in human normal, premalignant, and malignant gastric tissue

Yanghui Wen^{1*}, Mingxu Da^{2*}, Yongbin Zhang², Lingzhi Peng¹, Jibin Yao², Yaoxing Duan²

¹Clinical Medical College of Ningxia Medical University, Yinchuan 750004, P.R. China; ²Department of Surgical Oncology, Gansu Province People's Hospital, Lanzhou 730000, P.R. China. *Equal contributors.

Received August 22, 2015; Accepted September 25, 2015; Epub October 1, 2015; Published October 15, 2015

Abstract: Amount of studies in cells and animal models have proved vitamin D has multifarious antitumor effects. However, epidemiological studies showed inconsistent result on gastric cancer. The antitumor role is mainly mediated by the vitamin D receptor (VDR). Our hypothesis is that VDR may be abnormally (poorly) expressed in gastric cancer tissue. Present study is aimed at discovering and analyzing VDR expression in a series of human gastric tissues, including normal, premalignant, and malignant gastric tissue, and correlated VDR to the clinicopathological parameters of gastric cancer patients. VDR expression was detected by immunohistochemistry. The χ^2 test was used to analyze the VDR expression as well as the relationship between VDR and the clinicopathological factors of gastric cancer patients. Compared with normal (82.61%) and premalignant tissues (73.64%), VDR was lower expressed in cancer tissues (57.61%), with a statistically significant difference ($P = 0.001$). Among cancer tissues, VDR was higher expressed in well and moderate differentiated tissues contrasted with tissues with poor differentiation, and higher expressed in small tumors (< 5 cm) compared with large tumors (≥ 5 cm), with a statistically significant difference respectively ($P = 0.016$, $P = 0.009$). A decline linear trend appeared when analyzing the statistical difference of VDR expression among normal, premalignant, and malignant gastric tissues. VDR expression has been on the decline from the premalignant stage, finally low expressed in gastric cancer tissues, especial in poorly differentiated tissues. VDR could be a potential prognostic factor for patients with gastric cancer.

Keywords: Vitamin D, antitumor, gastric cancer, vitamin D receptor, immunohistochemistry

Introduction

Although incidence of gastric cancer is declining in some regions of the world, because of most cases being diagnosed in advanced stages with poor prognosis and limited treatment options, it is still the fourth (after lung, breast and colorectal) most common malignancy and the second (after lung cancer) leading cause of death among all cancers worldwide [1]. Early diagnosis and treatment is absolutely important.

In recent years, with the development of epidemiology and molecular biology, 1,25-dihydroxyvitamin D₃, the active metabolite of vitamin D, has been found broadly antitumor effects, including anti-proliferative effects, induction of

apoptosis, stimulation of differentiation, inhibition of invasion and metastasis, inhibition of angiogenesis, and also anti-inflammatory effects. Some specific signaling pathways that regulate tumor growth by 1,25-dihydroxyvitamin D₃ are particularly showed in breast, colon and prostate cancer cells. Although it still needs some large-scale and long-term human randomized controlled trials to confirm, recent result from amount of preclinical studies in cells and animal models, some observational studies and smaller interventional studies immensely support the anticancer effects of 1,25-dihydroxyvitamin D₃. It suggests that avoiding deficiency or adding vitamin D supplements might be an economical and safe way to reduce cancer incidence, or participant into the treatment of tumor as a new chemotherapy drug [2-5].

Vitamin D receptor expression in a series of gastric tissues

In gastric cancer, several preclinical studies in cells proved the antitumor effects of 1,25-dihydroxyvitamin D₃ [6-11], but epidemiological studies have little support for its protective effects on gastric cancer [12-14]. We have known the biological function of 1,25-dihydroxyvitamin D₃, especially anticancer effects, is largely mediated by the vitamin D receptor [2, 3, 15], which was detected and originally identified as a chromatin-associated protein by Haussler [16] in 1969. VDR is a member of the steroid and thyroid hormone receptor superfamily [17], and also a member of transcription family [18], when free 1,25-dihydroxyvitamin D₃ binds to the VDR, causing phosphorylation of the receptor, the ligand-activated VDR interacts with the retinoid X receptor (RXR) to form a heterodimer, and get its translocation to the nucleus, then the 1,25-dihydroxyvitamin D₃-VDR-RXR complex binds to the vitamin D response elements (VDREs) in multiple regulatory regions located in the promoters of target genes or at distal sites, and this causes the recruitment of co-activators or co-repressors, which leads to positive or negative transcriptional regulation of gene expression [15, 19, 20]. Kallay, E. et al. [21] found VDR gene knockout mice show hyperplasia and increased mitotic activity in the descending colon, suggesting a role for 1,25-dihydroxyvitamin D₃ mediated signaling way in tumor suppression. Several studies by Zinser GM et al. [22-24] proved the importance of VDR in cancer and that the VDR signalling may be required to suppress tumorigenesis. The anticancer mechanism of 1,25-dihydroxyvitamin D₃ mediated by VDR via the gene pathways has been concluded in a recent review that discussing the anticancer role of 1,25-dihydroxyvitamin D₃ by Feldman et al. [3].

In general, VDR is an important determinant of tumor cells response to 1,25-dihydroxyvitamin D₃. Even though there is adequate or higher serum 25-hydroxyvitamin D in cancer patients or high risk individuals, if the expression of VDR is in a lower level, the anticancer effect of 1,25-dihydroxyvitamin D₃ could weaken. Based on the inconformity between epidemiological data and preclinical study results of the anticancer effects mediated by 1,25-dihydroxyvitamin D₃ on gastric cancer, we hold a bold speculation here, the VDR may be abnormally (poorly) expressed in gastric cancer tissue. We try to illustrate it from the VDR expression status, so

that can better understand the prevention and treatment value of 1,25-dihydroxyvitamin D₃ in gastric cancer.

Present study is the first detection and analysis of VDR expression in a series of human gastric tissue types. The local VDR expression was examined in normal, premalignant (chronic atrophic gastritis, intestinal metaplasia, atypical hyperplasia) and malignant gastric tissue simultaneously by immunohistochemical technique. In addition, the relationship between VDR and clinicopathological factors of gastric cancer patients was analyzed.

Materials and methods

Tissue samples selection

All the samples were collected from a cohort of patients enrolled in Gansu Province People's Hospital from September 2013 to February 2015. It contains gastric cancer tissue and matched para-carcinoma tissue from 92 patients who underwent radical surgery for gastric carcinoma at the Surgical Oncology Department, and 148 cases premalignant gastric tissue, including chronic atrophic gastritis (n = 50), intestinal metaplasia (n = 52), and atypical hyperplasia tissues (n = 46), which were obtained from gastroscopic with biopsies at the Endoscopic Center of Gansu Province People's Hospital. Among of them, the patients who received surgical tumor resection have been recorded clinical and pathological data clearly, without preoperative radiochemotherapy or other therapies. The staging of gastric cancer was according to the American Joint Committee on Cancer (ACJJ, 7th edition) [25]. The matched normal gastric tissues were isolated ≥ 5 cm away from the cancer lesions. This study was approved by the Ethics Committee of Gansu Province People's Hospital (syll2015009), and all patients gave their informed consent prior to inclusion in the study.

VDR immunohistochemistry

The detection of VDR expression by Immunohistochemistry was carried out with the rabbit anti-human VDR monoclonal antibodies purchased from Sigma-Aldrich, Inc. All the specimens have been formalin-fixed and paraffin-embedded (FFPE) in advance, prepared for future use. Four-micrometer-thick sections

Vitamin D receptor expression in a series of gastric tissues

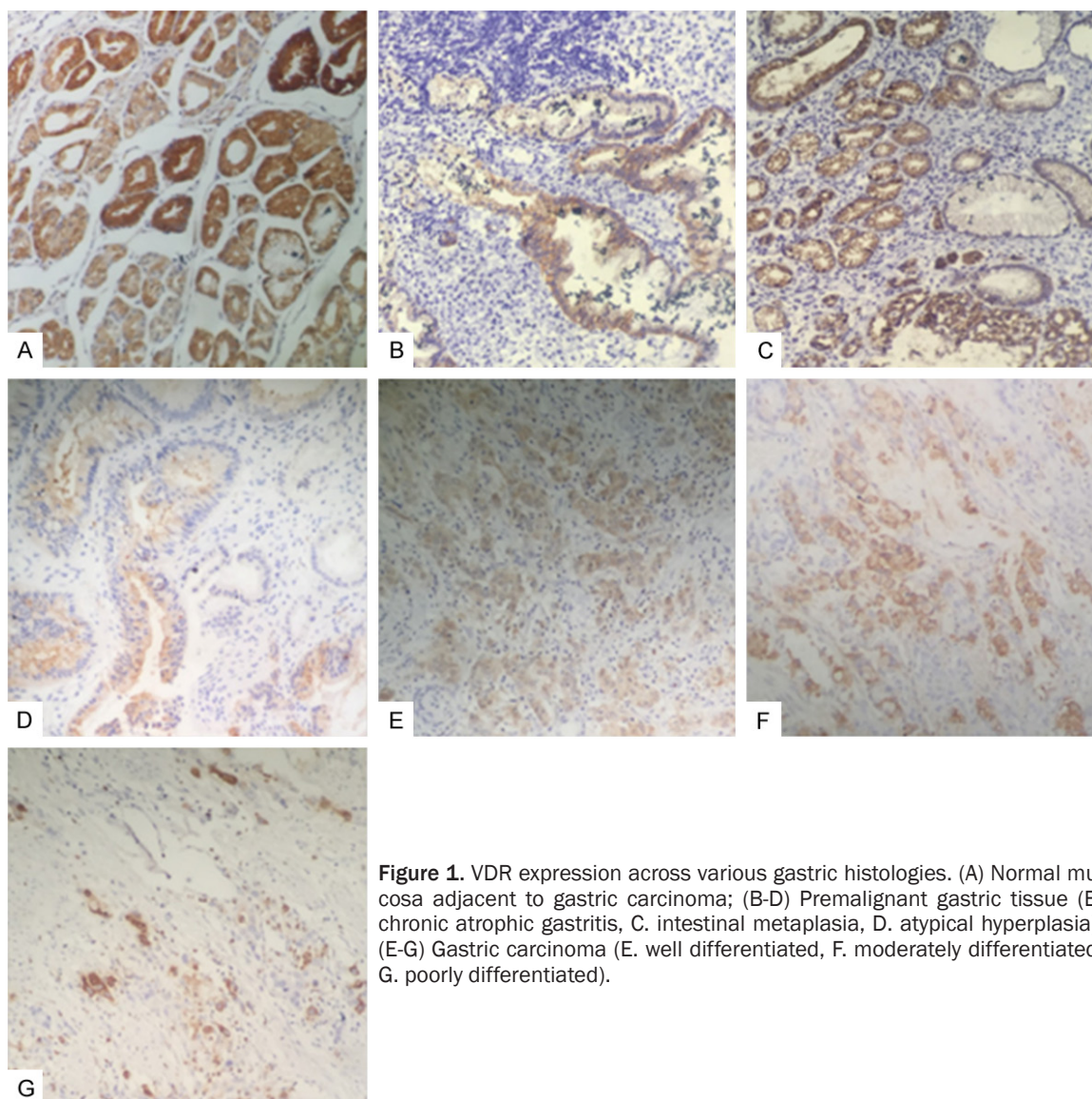


Figure 1. VDR expression across various gastric histologies. (A) Normal mucosa adjacent to gastric carcinoma; (B-D) Premalignant gastric tissue (B. chronic atrophic gastritis, C. intestinal metaplasia, D. atypical hyperplasia); (E-G) Gastric carcinoma (E. well differentiated, F. moderately differentiated, G. poorly differentiated).

were cut and mounted on slides, after placed at room temperature for 60 min, tissue sections were deparaffinized by xylene 10 min per time for twice, followed by 5 times 10 min per time in gradient ethanol (100%, 95%, 85%, 75%, 50%) for rehydration, then rinsed with water and PBS 5 min per time successively, add 0.3% H₂O₂ into the section followed by incubation in wet box for 10 min at room temperature, then rinsed with PBS 3 times 5 min each, slides were placed into citrate buffer heated to boiling by microwave and stay 8 min for antigen retrieval, when the slides were naturally cooled, washing with PBS 2 times 5 min reduplicative, then sections were blocked with 10% normal mouse serum for 45 min. Rabbit anti-human VDR anti-

body (1:50 dilution) were added, and slides were incubated overnight at 4°C. On the next day, washing the slides in PBS firstly, then biotinylated goat anti-rabbit secondary antibody was applied, incubate the sections at room temperature for 30 min subsequently, followed by washing with PBS again. Adding DBP into the slides for incubation, observing and grasping the degree of dyeing under a microscope, then sections were washed and counterstained with hematoxylin, using hydrochloric acid alcohol for differentiation. According the reverse order of rehydration above-mentioned for dehydration, then hyalinizing the slides with xylene, finally, add a coverslip to the slide for microscopic examination.

Vitamin D receptor expression in a series of gastric tissues

Table 1. VDR expression in normal, premalignant, and malignant gastric tissues

Histological type	NO. of cases	VDR			p
		(+)	(-)	PR (%)	
Normal gastric tissue	92	76	16	82.61	0.001*
Premalignant gastric tissue	148	109	39	73.64	
Malignant gastric tissue	92	53	39	57.61	

*Indicates statistical significance

Evaluation of immunostaining

Under the microscope, slides were examined and evaluated for both staining intensity and percentage of positive cells referring to a scoring method described in a similar study recently published [26]. The staining intensity score was classified as 1 (weak), 2 (moderate), and 3 (strong). The percentage of positive cells were scored as 0 ($\leq 5\%$), 1 (6-25%), 2 (26-50%), 3 (51-75%), and 4 (76-100%). The staining intensity and percentage of staining cells were then multiplied to generate the immunoreactivity score for each case, ranging from 0 to 12. Tissue sections with an immunoreactivity score ≥ 4 were considered to be positive (high) expression, while a score < 4 was considered as negative (low) expression. Pictures were taken of representative areas containing the diagnosis of interest for each sample and blindly scored by three observers.

Statistical analysis

Our study was designed to detect and analyze the VDR expression in various human gastric tissues, to test and verify the hypothesis of VDR abnormally (poorly) expressed in gastric cancer tissue. SPSS 16.0 statistical software was used for analysis. The χ^2 test were used to analyze the statistical difference among normal, premalignant, and malignant gastric tissues. The relationship between VDR expression and clinicopathological factors of gastric cancer patients was statistically analyzed by χ^2 test either. In all analyses, a *p* value < 0.05 was considered to be statistically significant.

Results

The distribution of VDR

VDR was expressed in the various gastric tissues, at different amounts. And appeared to be localized in the cytoplasm and perinuclear

regions, with nuclear staining absent (**Figure 1**).

Tumor expression of VDR

Compared with normal and premalignant tissues, VDR was lower expressed in gastric cancer tissue. The PR (positive rate) of VDR expression in tumor tissues was 57.61% (53/92), obviously lower than the adjacent normal tissues (82.61%, 76/92) and premalignant tissues (70.41%, 109/148), with a significantly statistical difference ($P = 0.001$) (**Table 1**).

Correlation between VDR and clinicopathological factors

No statistical differences have been found between VDR expression and gender or age of gastric cancer patients, tumor location, histological type, TNM stage, and lymph node metastasis except differentiated degree and tumor size. The PR of VDR expression was 70.00% (28/40) in well differentiated cancer tissues, and 65.00% (13/20) in moderate differentiated cancer tissues, but only 37.50% (12/32) in poor differentiated tissues, with a significantly statistical difference ($P = 0.016$). The PR of VDR was 66.13% (41/61) in small tumors (< 5 cm), only 38.71% (12/31) in large tumors (≥ 5 cm), with a significantly statistical difference either ($P = 0.009$) (**Table 2**).

VDR expression across all samples

Along with the pathological changes of tissues in the light of the gastric cancer progression pattern, which was put forward by Corea et al. [27] and widely accepted by most scholars, the expression of VDR showed a decline linear trend (**Figure 2**). The PR of VDR expression in normal tissues was 82.61% (76/92), then 73.64% (109/148) and 57.61% (53/92) in premalignant and gastric cancer tissues respectively, with a statistically significant difference ($P = 0.001$) (**Table 1**). In the premalignant tissues, no statistically difference ($P = 0.888$) was found among chronic atrophic gastritis (76.00%, 38/50), intestinal metaplasia (73.07%, 38/52) and atypical hyperplasia tissues (71.73%, 33/46) (**Table 3**).

Discussion

Since Colston et al. [28] found 1,25-dihydroxyvitamin D_3 can inhibit the proliferation of

Vitamin D receptor expression in a series of gastric tissues

Table 2. Correlation of VDR expression with the clinicopathological parameters of gastric cancer patients

Clinicopathological factors	Discription	NO. of cases	VDR			P
			(+)	(-)	PR (%)	
Age (years)	< 55	40	24	16	60.00	0.684
	≥ 55	52	29	23	55.77	
Gender	Male	63	35	28	55.56	0.557
	Female	29	18	11	62.07	
Tumor location	Cardia	13	8	5	61.54	0.757
	Fundic, body, antral	79	45	34	56.96	
Tumor size (cm)	< 5	61	41	20	67.21	0.009*
	≥ 5	31	12	19	38.71	
Differentiation degree	High	40	28	12	70.00	0.016*
	Middle	20	13	7	65.00	
	Low	32	12	20	37.50	
Pathological type	Pathological type	37	23	14	62.16	0.606
	Polypoid adenocarcinoma	23	14	9	60.87	
	Mucinous adenocarcinoma	24	13	11	54.17	
	Signet-ring cell carcinoma	8	3	5	37.50	
Lymphnode metastasis	Yes	58	30	28	51.72	0.136
	No	34	23	11	67.65	
TNM stage	I+II	38	23	15	60.53	0.635
	III+IV	54	30	24	55.56	

*Indicates statistical significance.

human melanoma cells in 1981, and Abe et al. [29] reported mouse myeloid leukemia cells can be induced to differentiate into macrophages in vitro by 1,25-dihydroxyvitamin D₃ in the same year, the anticancer effects of 1,25-dihydroxyvitamin D₃ have been shown in vitro and in vivo among various malignancies, recently reviewed by Feldman et al. [3]. In general, accumulating results from preclinical and some clinical studies have strongly suggested the anticancer action of 1,25-dihydroxyvitamin D₃.

Specific to gastric cancer, several studies in vitro proved 1,25-dihydroxyvitamin D₃ have the anticarcinoma effects of anti-proliferation, promoting apoptosis [7, 8, 11], regulating cell via-

bility [10], inhibiting gastric cancer cell growth and inducing cell cycle arrest [7, 9]. However, epidemiological studies have little support for its protective effects on gastric cancer [13, 14]. Chen, W. et al. [12] found no relationship between serum vitamin D status and the risk of gastric cancer in a cohort of Chinese patients. Genomic and proteomic screening approaches have identified the antitumor effects of 1,25-dihydroxyvitamin D₃ is particularly mediated by VDR [3, 11]. In other words, VDR is an important determinant of tumor cell response to 1,25-dihydroxyvitamin D₃. Therefore, we hold a bold hypothesis that VDR may be abnormally (poorly) expressed in gastric cancer tissue here, trying to explain the inconsistent conclusion between epidemiological data and preclin-

Vitamin D receptor expression in a series of gastric tissues

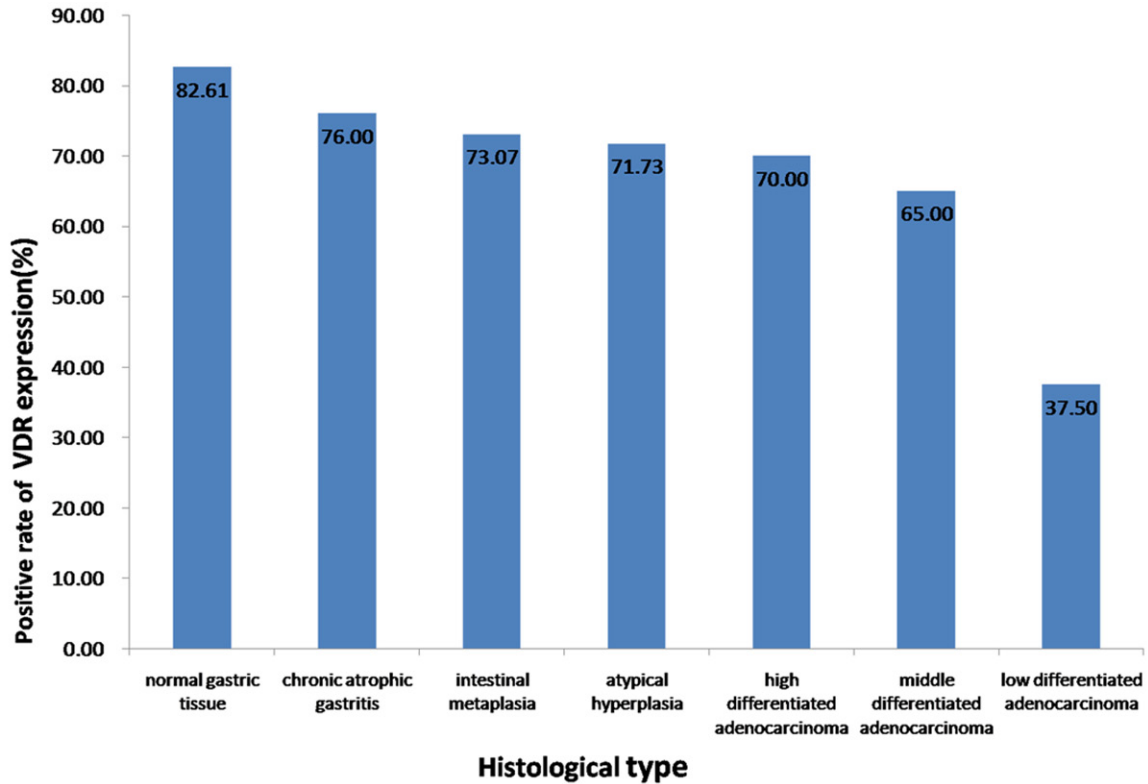


Figure 2. VDR was expressed in the various gastric tissues at different amounts, which was showed a declined linear trend with the pathological change from normal to premalignant (chronic atrophic gastritis, intestinal metaplasia, atypical hyperplasia), then malignant as well as the differentiation degree of gastric carcinoma (well differentiated, moderately differentiated, poorly differentiated).

Table 3. VDR expression in the various types of premalignant gastric tissues

Histological type	NO. of cases	VDR			p
		(+)	(-)	PR (%)	
Chronic atrophic gastritis	50	38	12	76.00	0.888
Intestinal metaplasia	52	38	14	73.07	
Atypical hyperplasia	46	33	13	71.74	

ical study results of the anticancer effects of 1,25-dihydroxyvitamin D₃ on gastric cancer from the VDR expression status.

To verify our conjecture, we retrieved literatures on VDR distribution in human tissues in advance, different types of tissue or cell have diverse VDR expression [30]. The VDR is over-expressed or repressed in several histological types of cancer, demonstrating tissue-type variations in 1,25-dihydroxyvitamin D₃ signaling [2]. F.C. et al. reviewed the Yin and Yang of VDR signaling in neoplastic progression [31]. Some studies have described the VDR was

expressed in human normal gastric tissue [32, 33]. Here we detected the expression of VDR in a series of human gastric tissues, including normal, premalignant, and malignant gastric tissues. This study first illustrated that VDR expression differs in various types of gastric tissue by immunohistochemical detection. We found VDR expression does declined in tumor cells and revealed a decline trend along with the advance of gastric cancer. Furthermore, VDR was highly expressed in more tumors with high or middle differentiation degree, but poorly expressed in low differentiated cancer tissues, and higher expressed in small tumors contrasted with large tumors. It suggests VDR could be a potential prognostic factor for patients with gastric cancer. On account of the samples were collected from the patients who accepted radical surgery for gastric carcinoma in our department in recent one and a half years, this cohort of patients are still in a timely follow-up status, the relationship between VDR

Vitamin D receptor expression in a series of gastric tissues

expression and the five-year survival rate of the patients will be statistically analyzed in the near future, which may further declare its role as a potential prognostic factor.

Even though there is no study covering the dose-response relationship between the VDR and 1,25-dihydroxyvitamin D₃ so far. On the basis of the VDR pathway inducing the anticancer action of 1,25-dihydroxyvitamin D₃, and the result from our study that VDR abnormally expressed in gastric cancer tissue, we suggest 1,25-dihydroxyvitamin D₃ may better provide a therapeutic choice on the patients with well or moderate differentiated degree, which have a higher VDR level, but not poorly differentiated degree group, which has a lower VDR level, supposedly not enough to induce 1,25-dihydroxyvitamin D₃ play its anticancer role well. In the case of how much 1,25-dihydroxyvitamin D₃ or its analogue should be used for tumor treatment, it is a huge research project, which needs some large-scale and long-term human randomized controlled trials to complete.

As to the cancer prevention value of 1,25-dihydroxyvitamin D₃, similarly, no exactly conclusion has been published and no right dose have been recommended, especially the inconsistent results reported by IOM and Endocrine Society [34, 35]. We just suggest the population in the regions with high incidence of gastric cancer, especially those who cannot receive adequate sunlight exposure to natural sunlight appropriately as a way for reducing the incidence of gastric cancer at present. It is worth stressing that our study shows VDR has been declined in precancerous conditions of gastric cancer, we particularly suggest 1,25-dihydroxyvitamin D₃, as a new anticancer agent, if possible, should better be used in the patients who stay in the early stage of gastric cancer progression, such as intestinal metaplasia and atypical hyperplasia.

Clearly, our study is limited to detecting the VDR expression by immunohistochemistry, and theoretically assess the anticancer effects of 1,25-dihydroxyvitamin D₃ only from the VDR expression status. The activity of CYP24A1, which catabolizes 1,25-dihydroxyvitamin D₃, also influences the anticancer effect of 1,25-dihydroxyvitamin D₃ [3, 36], has not been involved in present study. Further studies of VDR expression and the anticancer effects of

1,25-dihydroxyvitamin D₃ on gastric cancer still be needed.

In summary, this is the first study to detect and analyze the expression of the VDR in a series of human gastric tissues. We found VDR was expressed in the various types of gastric tissues, but in different quantity. VDR expression has been declined from the premalignant stage, finally low expressed in gastric cancer tissues, especial in poorly differentiated tissues. That suggests 1,25-dihydroxyvitamin D₃, as a new anticancer agent, may better be selectively used in the gastric cancer patients with well or moderate differentiated degree for treatment; or be added to the patients who stay in a premalignant status for gastric cancer prevention, which have a higher VDR level, to induce 1,25-dihydroxyvitamin D₃ to play its anticancer role well. In addition, except histology differentiated degree, we correlated the expression of VDR to the tumor size. VDR could be a potential prognostic factor for patients with gastric cancer.

Disclosure of conflict of interest

None.

Address correspondence to: Mingxu Da, Department of Surgical Oncology, Gansu Province People's Hospital, 204 Donggang West Road, Lanzhou 730000, P.R. China. E-mail: hxdamingxu@hotmail.com

References

- [1] Piazzuelo MB and Correa P. Gastric cancer: Overview. *Colomb Med (Cali)* 2013; 44: 192-201.
- [2] Deeb KK, Trump DL and Johnson CS. Vitamin D signalling pathways in cancer: potential for anticancer therapeutics. *Cancer* 2007; 7: 684-700.
- [3] Feldman D, Krishnan AV, Swami S, Giovannucci E and Feldman BJ. The role of vitamin D in reducing cancer risk and progression. *Nat Rev Cancer* 2014; 14: 342-357.
- [4] Wolozynska-Read A, Johnson CS and Trump DL. Vitamin D and cancer: clinical aspects. *Best Pract Res Clin Endocrinol Metab* 2011; 25: 605-615.
- [5] Leyssens C, Verlinden L and Verstuyf A. Anti-neoplastic effects of 1,25(OH)2D3 and its analogs in breast, prostate and colorectal cancer. *Endocr Relat Cancer* 2013; 20: R31-47.
- [6] Chang S, Gao L, Yang Y, Tong D, Guo B, Liu L, Li Z, Song T, Huang C. miR-145 mediates the an-

Vitamin D receptor expression in a series of gastric tissues

- triproliferative and gene regulatory effects of vitamin D₃ by directly targeting E2F3 in gastric cancer cells. *Oncotarget* 2015; 6: 7675-85.
- [7] Bao A, Li Y, Tong Y, Zheng H, Wu W and Wei C. 1,25-Dihydroxyvitamin D₃ and cisplatin synergistically induce apoptosis and cell cycle arrest in gastric cancer cells. *Int JMol Med* 2014; 33: 1177-1184.
- [8] Wang W, Zhao CH, Zhang N and Wang J. Vitamin D analog EB1089 induces apoptosis in a subpopulation of SGC-7901 gastric cancer cells through a mitochondrial-dependent apoptotic pathway. *Nutr Cancer* 2013; 65: 1067-1075.
- [9] Park MR, Lee JH, Park MS, Hwang JE, Shim HJ, Cho SH, Chung IJ, Bae WK. Suppressive effect of 19-nor-1 α -25-dihydroxyvitamin D₂ on gastric cancer cells and peritoneal metastasis model. *J Korean Med Sci* 2012; 27: 1037-1043.
- [10] Baek S, Lee YS, Shim HE, Yoon S, Baek SY, Kim BS, Oh SO. Vitamin D₃ regulates cell viability in gastric cancer and cholangiocarcinoma. *Anat Cell Biol* 2011; 44: 204-209.
- [11] Pan L, Matloob AF, Du J, Pan H, Dong Z, Zhao J, Feng Y, Zhong Y, Huang B, Lu J. Vitamin D stimulates apoptosis in gastric cancer cells in synergy with trichostatin A/sodium butyrate-induced and 5-aza-2'-deoxycytidine-induced PTEN upregulation. *FEBS J* 2010; 277: 989-999.
- [12] Chen W, Dawsey SM, Qiao YL, Mark SD, Dong ZW, Taylor PR, Zhao P, Abnet CC. Prospective study of serum 25(OH)-vitamin D concentration and risk of oesophageal and gastric cancers. *Br J Cancer* 2007; 97: 123-128.
- [13] Helzlsouer KJ and Gallicchio L. Shedding light on serum vitamin D concentrations and the risk of rarer cancers. *Anticancer Agents Med Chem* 2013; 13: 65-69.
- [14] Trowbridge R, Mittal SK and Agrawal DK. Vitamin D and the epidemiology of upper gastrointestinal cancers: a critical analysis of the current evidence. *Cancer Epidemiol Biomarkers Prev* 2013; 22: 1007-1014.
- [15] Haussler MR, Jurutka PW, Mizwicki M and Norman AW. Vitamin D receptor (VDR)-mediated actions of 1 α ,25(OH)₂vitamin D₃: genomic and non-genomic mechanisms. *Best Pract Res Clin Endocrinol Metab* 2011; 25: 543-559.
- [16] Haussler MR and Norman AW. Chromosomal receptor for a vitamin D metabolite. *Proc Natl Acad Sci U S A* 1969; 62: 155-162.
- [17] Evans RM. The steroid and thyroid hormone receptor superfamily. *Science* 1988; 240: 889-895.
- [18] Carlberg C and Campbell MJ. Vitamin D receptor signaling mechanisms: integrated actions of a well-defined transcription factor. *Steroids* 2013; 78: 127-136.
- [19] Haussler MR, Whitfield GK, Kaneko I, Haussler CA, Hsieh D, Hsieh JC, Jurutka PW. Molecular mechanisms of vitamin D action. *Calcif Tissue Int* 2013; 92: 77-98.
- [20] Pike JW and Meyer MB. Fundamentals of vitamin D hormone-regulated gene expression. *J Steroid Biochem Molecul Biol* 2014; 144 Pt A: 5-11.
- [21] Kallay E, Pietschmann P, Toyokuni S, Bajna E, Hahn P, Mazzucco K, Bieglmayer C, Kato S, Cross HS. Characterization of a vitamin D receptor knockout mouse as a model of colorectal hyperproliferation and DNA damage. *Carcinogenesis* 2001; 22: 1429-1435.
- [22] Zinser GM, Sundberg JP and Welsh J. Vitamin D(3) receptor ablation sensitizes skin to chemically induced tumorigenesis. *Carcinogenesis* 2002; 23: 2103-2109.
- [23] Zinser GM and Welsh J. Vitamin D receptor status alters mammary gland morphology and tumorigenesis in MMTV-neu mice. *Carcinogenesis* 2004; 25: 2361-2372.
- [24] Zinser GM, Suckow M and Welsh J. Vitamin D receptor (VDR) ablation alters carcinogen-induced tumorigenesis in mammary gland, epidermis and lymphoid tissues. *J Steroid Biochem Mol Biol* 2005; 97: 153-164.
- [25] Washington K. 7th edition of the AJCC cancer staging manual: stomach. *Ann Surg Oncol* 2010; 17: 3077-3079.
- [26] Wang K, Dong M, Sheng W, Liu Q, Yu D, Dong Q, Li Q, Wang J. Expression of Vitamin D Receptor as a Potential Prognostic Factor and Therapeutic Target in Pancreatic Cancer. *Histopathology* 2015; 67: 386-97.
- [27] Correa P. A human model of gastric carcinogenesis. *Cancer Res* 1988; 48: 3554-3560.
- [28] Colston K, Colston MJ and Feldman D. 1,25-dihydroxyvitamin D₃ and malignant melanoma: the presence of receptors and inhibition of cell growth in culture. *Endocrinology* 1981; 108: 1083-1086.
- [29] Abe E, Miyaura C, Sakagami H, Takeda M, Konno K, Yamazaki T, Yoshiki S, Suda T. Differentiation of mouse myeloid leukemia cells induced by 1 α ,25-dihydroxyvitamin D₃. *Proc Natl Acad Sci U S A* 1981; 78: 4990-4994.
- [30] Wang Y, Zhu J and DeLuca HF. Where is the vitamin D receptor? *Arch Biochem Biophys* 2012; 523: 123-133.
- [31] Campbell FC, Xu H, El-Tanani M, Crowe P and Bingham V. The yin and yang of vitamin D receptor (VDR) signaling in neoplastic progression: operational networks and tissue-specific growth control. *Biochem Pharmacol* 2010; 79: 1-9.
- [32] Trowbridge R, Mittal SK, Sharma P, Hunter WJ and Agrawal DK. Vitamin D receptor expression in the mucosal tissue at the gastroesoph-

Vitamin D receptor expression in a series of gastric tissues

- ageal junction. *Exp Mol Pathol* 2012; 93: 246-249.
- [33] Berger U, Wilson P, McClelland RA, Colston K, Hausler MR, Pike JW, Coombes RC. Immunocytochemical detection of 1,25-dihydroxyvitamin D receptors in normal human tissues. *J Clin Endocrinol Metab* 1988; 67: 607-613.
- [34] Holick MF, Binkley NC, Bischoff-Ferrari HA, Gordon CM, Hanley DA, Heaney RP, Murad MH, Weaver CM; Endocrine Society. Evaluation, treatment, and prevention of vitamin D deficiency: an Endocrine Society clinical practice guideline. *J Clin Endocrinol Metab* 2011; 96: 1911-1930.
- [35] Rosen CJ, Abrams SA, Aloia JF, Brannon PM, Clinton SK, Durazo-Arvizu RA, Gallagher JC, Gallo RL, Jones G, Kovacs CS, Manson JE, Mayne ST, Ross AC, Shapses SA, Taylor CL. IOM committee members respond to Endocrine Society vitamin D guideline. *J Clin Endocrinol Metab* 2012; 97: 1146-1152.
- [36] Bikle DD. Vitamin D metabolism, mechanism of action, and clinical applications. *Chem Biol* 2014; 21: 319-329.