

RESEARCH ARTICLE

Impact of *IL28B*, *APOH* and *ITPA* Polymorphisms on Efficacy and Safety of TVR- or BOC-Based Triple Therapy in Treatment-Experienced HCV-1 Patients with Compensated Cirrhosis from the ANRS CO20-CUPIC Study



click for updates

Frédégonde About^{1,2}, Tiphaine Oudot-Mellakh^{3,4}, Jonathan Niay^{3,4}, Pascaline Rabiéga⁵, Vincent Pedergnana^{1,2,6}, Darragh Duffy^{7,8}, Philippe Sultanik^{9,10}, Carole Cagnot¹¹, Fabrice Carrat^{5,12}, Patrick Marcellin¹³, Fabien Zoulim^{14,15,16}, Dominique Larrey¹⁷, Christophe Hézode^{18,19}, Hélène Fontaine^{9,10}, Jean-Pierre Bronowicki²⁰, Stanislas Pol^{9,10}, Matthew L. Albert^{7,8,10}, Ioannis Theodorou^{3,4}, Aurélie Cobat^{1,2,*}, Laurent Abel^{1,2,21}*, ANRS CO20-CUPIC study group[†]

OPEN ACCESS

Citation: About F, Oudot-Mellakh T, Niay J, Rabiéga P, Pedergnana V, Duffy D, et al. (2015) Impact of *IL28B*, *APOH* and *ITPA* Polymorphisms on Efficacy and Safety of TVR- or BOC-Based Triple Therapy in Treatment-Experienced HCV-1 Patients with Compensated Cirrhosis from the ANRS CO20-CUPIC Study. PLoS ONE 10(12): e0145105. doi:10.1371/journal.pone.0145105

Editor: Chen-Hua Liu, National Taiwan University Hospital, TAIWAN

Received: September 7, 2015

Accepted: November 29, 2015

Published: December 15, 2015

Copyright: © 2015 About et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](http://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its Supporting Information files.

Funding: This work was supported by INSERM-ANRS (France REcherche Nord & sud Sida-HIV Hépatites-FRENSH, Grant n° 2010-A01273-36, <http://www.anrs.fr>), Institut National de la Santé et de la Recherche Médicale (INSERM, <http://www.inserm.fr>), University Paris Descartes (<http://www.parisdescartes.fr>), the French National Research Agency (ANR, <http://www.agence-nationale-recherche.fr>).

1 Laboratory of Human Genetics of Infectious Diseases, Necker Branch, Institut National de la Santé et de la Recherche Médicale (INSERM) U1163, Paris, France, **2** Paris Descartes University, Imagine Institute, Paris, France, **3** Laboratory of Immunity and Infection, Centre d'Immunologie et des Maladies Infectieuses de Paris (CIMI), INSERM U1135, Groupe Hospitalier Pitié Salpêtrière, Assistance Publique-Hôpitaux de Paris (AP-HP), Paris, France, **4** Plateforme Génomique Inserm-ANRS, Groupe Hospitalier Pitié Salpêtrière, AP-HP, UPMC Université Paris 6, Paris, France, **5** Sorbonne Universités, UPMC Université Paris 06, INSERM, Institut Pierre Louis d'épidémiologie et de Santé Publique (IPLESP UMRS 1136), Paris, France, **6** Wellcome Trust Centre for Human Genetics, University of Oxford, Oxford, United Kingdom, **7** Centre for Human Immunology, Department of Immunology, Institut Pasteur, Paris, France, **8** The Laboratory of Dendritic Cell Biology, Department of Immunology, Institut Pasteur, INSERM U818, Paris, France, **9** Département d'Hépatologie, Hôpital Cochin, AP-HP, Université Paris Descartes, Paris, France, **10** INSERM UMS20, Institut Pasteur, Paris, France, **11** Unit for Basic and Clinical research on Viral Hepatitis, Inserm-ANRS (France REcherche Nord & sud Sida-HIV Hépatites-FRENSH), Paris, France, **12** Service de Santé Publique, Hôpital Saint Antoine, AP-HP, Paris, France, **13** Service d'Hépatologie, Hôpital Beaujon, Clichy, France, **14** Centre de recherche en cancérologie de Lyon (CRCL), INSERM UMR I 1052/CNRS 5286, Lyon cedex 03, France, **15** Université Claude-Bernard Lyon 1, Villeurbanne, France, **16** Hospices civils de Lyon, Hôpital de la Croix-Rousse, service d'hépatologie et de gastroentérologie, Lyon, France, **17** CHU St Eloi Hospital, Liver Unit, Montpellier, France, **18** Department of Hepatology and Gastroenterology, Hôpital Henri Mondor, AP-HP, Université Paris-Est Créteil (UPEC), Créteil, France, **19** Institut Mondor de Recherche Biomédicale (IMRB), INSERM U955, UPEC, Créteil, France, **20** Department of Hepatogastroenterology, INSERM U954, CHU de Nancy, Université de Lorraine, Vandoeuvre-Lès-Nancy, France, **21** St. Giles Laboratory of Human Genetics of Infectious Diseases, Rockefeller Branch, The Rockefeller University, New York, NY, United States of America

☯ These authors contributed equally to this work.

† Membership of the ANRS CO20-CUPIC study group is listed in the Acknowledgments.

* aurelie.cobat@inserm.fr (AC); laurent.abel@inserm.fr (LA)

Abstract

Background

Human genetic factors influence the outcome of pegylated interferon and ribavirin hepatitis C therapy. We explored the role of *IL28B*, *APOH* and *ITPA* SNPs on the outcomes of triple therapy including telaprevir or boceprevir in patients with compensated cirrhosis chronically infected with HCV-1.

recherche.fr) under the "Investments for the future" program (grant n°ANR-10-IAHU-01), and in part by the Association Française pour l'Etude du Foie (AFEF, <http://www.afef.asso.fr>). FA is the recipient of a fellowship from Fondation pour la Recherche Médicale (FRM, Grant n° FDM20140630671, <http://www.frm.org>). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

Patients and Methods

A total of 256 HCV-1 Caucasian treatment-experienced patients with compensated cirrhosis from the ANRS CO20-CUPIC cohort were genotyped for a total of 10 candidate SNPs in *IL28B* (rs12979860 and rs368234815), *APOH* (rs8178822, rs12944940, rs10048158, rs52797880, rs1801689 and rs1801690) and *ITPA* (rs1127354 and rs7270101). We tested the association of *IL28B* and *APOH* SNPs with sustained virological response and of *ITPA* SNPs with anemia related phenotypes by means of logistic regression assuming an additive genetic model.

Results

None of the six *APOH* SNPs were associated with sustained virological response. The favorable alleles of the *IL28B* SNPs rs12979860 and rs368234815 were associated with sustained virological response (rs12979860: OR = 2.35[1.50–3.70], $P = 2 \times 10^{-4}$). Refined analysis showed that the effect of *IL28B* SNPs on sustained virological response was restricted to prior PegIFN/RBV relapse (OR = 3.80[1.82–8.92], $P = 8 \times 10^{-4}$). We also confirmed the association between *ITPA* low activity alleles and protection against early hemoglobin decline in triple therapy ($P = 2 \times 10^{-5}$).

Conclusion

Our results suggest that the screening of rs12979860 may remain interesting for decision making in prior relapse HCV-1 Caucasian patients with compensated cirrhosis eligible for a telaprevir- or boceprevir-based therapy.

Introduction

HCV infection is a major public health issue with ~80 million people chronically infected worldwide [1]. Up to 2011, standard of care treatment was based on pegylated interferon and ribavirin (PegIFN/RBV) which leads to viral clearance in ~50% of the patients [2]. Well-established baseline predictors of sustained virological response (SVR) to PegIFN/RBV include viral load, HCV genotype, age, ethnicity, body weight, insulin resistance, steatosis, fibrosis stage, and *IL28B* single nucleotide polymorphism (SNP) rs12979860 [2–5]. A dinucleotide frameshift variant (rs368234815) creating a novel gene encoding *IFN-λ-4*, in strong linkage disequilibrium (LD) with rs12979860, was recently identified as a stronger predictor than rs12979860 of treatment-induced clearance [6]. One of the most common side effects of ribavirin therapy is anemia that mainly appears at the beginning of treatment. Two variants, rs1127354 and rs7270101, from the inosine triphosphate (*ITPA*) gene, encoding a protein that hydrolyses inosine triphosphate, are independent predictors of RBV-induced anemia [7].

Since 2011, first generation direct acting antiviral drugs (DAAs) targeting the HCV NS3/4A protease, such as telaprevir (TVR) and boceprevir (BOC), are available in several countries. These drugs combined with a PegIFN/RBV backbone significantly improved the SVR as compared to PegIFN/RBV alone in both treatment-naïve and previous treatment-failure patients with chronic HCV genotype 1 (HCV-1) infection [8,9]. However, side effects such as anemia are more frequent, particularly in patients with cirrhosis [10]. More recently, new IFN-free therapies with second generation DAAs have emerged and provide SVR rates over 90% [11].

However, these therapies are still very expensive and not yet widely used in real life settings. Therefore, triple therapy combining PegIFN/RBV with first generation protease inhibitors (PIs) remains the standard of care for HCV-1 infected patients in most countries. In the present study we aimed to explore the role of *IL28B*, *APOH* and *ITPA* SNPs on the outcomes of triple therapy including telaprevir or boceprevir in patients with compensated cirrhosis chronically infected with HCV-1.

Patients and Methods

Study population

The ANRS CO20-CUPIC (Compassionate Use of Protease Inhibitors in viral C Cirrhosis) study is a French multicenter cohort study that enrolled 660 HCV genotype 1 (HCV-1) treatment-experienced cirrhotic patients to assess safety and efficacy of triple therapy with TVR or BOC for difficult to treat patients in real-life settings [10,12]. Briefly, patients with compensated cirrhosis chronically infected with HCV-1, and who failed a prior course of IFN alone or IFN/RBV started a triple combination therapy including PegIFN/RBV and TVR or BOC for a total course of 48 weeks [10]. The choice between TVR and BOC was at the investigator's discretion. Results showed a substantial benefit of triple therapy in difficult to treat patients with SVR rates of 43–52% but with an increased frequency and severity of side effects [12]. Interestingly, a recent study conducted in 189 patients from the CUPIC cohort identified baseline levels of apolipoprotein H (apoH), encoded by *APOH* gene, as a surrogate marker for SVR to triple therapy [13]. *APOH* polymorphisms have previously been associated with triglyceride levels, which itself is an independent correlate of HCV clearance [14].

Written informed consent was obtained from each patient before enrolment. The study was conducted in accordance with the Declaration of Helsinki and French law for biomedical research and was approved by the "Ile de France IX" Ethics Committee (Créteil, France).

Outcomes and statistical analysis

In the present study we took advantage of the well characterized CUPIC cohort study to assess the role of candidate SNPs in *IL28B*, *APOH* and *ITPA* on efficacy and safety of TVR- or BOC-based triple therapy. Only Caucasian patients who gave their consent for genetic testing were included (n = 256). Efficacy was assessed by SVR, defined as an undetectable HCV-RNA level 12 weeks after the end of therapy. For safety analysis, we focused on anemia and first considered a broad definition of clinically relevant anemia corresponding to patients with grade 2, 3 or 4 anemia (i.e. $Hb < 9.5 \text{ g.dl}^{-1}$) and/or blood transfusion and/or use of erythropoietin (EPO) occurring during the 48 weeks of treatment. We also focused on early significant hemoglobin decline, defined as a decrease of hemoglobin level of at least 3 g.dl^{-1} between baseline and week 4 as proposed in [7]. For early significant hemoglobin decline analysis, patients for whom EPO therapy (N = 22) or RBV dose reduction (N = 4) was instituted before week 4 were excluded and 209 patients were included in this analysis.

For the SVR binary phenotype, all statistical analyses were conducted by means of logistic regression. Association with *IL28B* and *APOH* SNPs was tested by assuming an additive genetic model (i.e. the coding of the genotype represents the number of reference alleles 0, 1 or 2). We performed both univariate and multivariate analysis, including covariates previously identified as independent predictors of SVR in the CUPIC cohort (i.e.: prior treatment response, lead-in phase, platelet count and HCV-1 subtypes) [12]. Interaction between the *IL28B* SNP rs12979860 and binary covariates such as prior response to treatment (non-response versus relapse including breakthrough) and treatment group (TVR versus BOC), was modeled in the

logistic regression framework by adding an interaction multiplicative term between the two main effects, e.g. *IL28B* SNP and the prior response to treatment.

The logistic regression framework was also used for the statistical analyses of the anemia related binary phenotypes (i.e. clinically relevant anemia and early significant hemoglobin decline). We performed both univariate and multivariate analysis, including *ITPA* SNPs (assuming an additive model), and other predictors of anemia (i.e.: age, sex, lead-in phase, hemoglobin at baseline, albumin at baseline <35g/L). To measure the joint effect of the two *ITPA* SNPs on anemia we considered a combined variable which estimates the severity of *ITPA* deficiency from rs1127354 and rs7270101 genotypes, as previously done in [7]. The severity of *ITPA* deficiency was defined as follows (S4 Table): Full *ITPA* activity (100%) was considered for rs1127354 C/C and rs7270101 A/A genotypes combination; 60% *ITPA* activity was considered for rs1127354 C/C and rs7270101 A/C genotypes combination; 30% *ITPA* activity was considered for rs1127354 C/C and rs7270101 C/C genotypes combination or rs1127354 A/C and rs7270101 A/A genotypes combination; and very low *ITPA* activity (0%) was considered for combined heterozygosity or rs1127354 A/A and rs7270101 A/A genotype combination [15]. Predicted *ITPA* activity was then considered as a quantitative covariate with four possible values (0; 0.3; 0.6; 1) in our logistic regression model. For the early hemoglobin decline phenotype, interaction between *ITPA* activity and the binary lead-in covariate was model in the logistic regression framework by adding a multiplicative interaction term between the two main effects, i.e. *ITPA* activity and lead-in.

All analyses were performed using the R software version 3.1.2 (<http://cran.r-project>), and p-values lower than 0.05 were considered as significant.

Genotyping

We genotyped 10 SNPs (S1 Fig) using TaqMan SNP genotyping assays (Applied Biosystems Inc., Foster City, CA) a total of 10 SNPs, two *ITPA* SNPs (rs1127354 and rs7270101), two *IL28B* variants (rs12979860 and rs368234815) and six *APOH* SNPs (rs8178822 [16,17], rs12944940, rs10048158 [18], rs52797880, rs1801689 [16,19] and rs1801690 [16,20]) selected based on their potential impact on apoH plasma levels via a search on NCBI Pubmed and regu-
lomeDB (score $\geq 2b$, <http://regulomedb.org/>).

Results

Patient characteristics

A total of 256 Caucasian patients were genotyped, 162 receiving TVR and 94 receiving BOC with comparable baseline characteristics (S1 Table). A total of 172 (67%) individuals were men. The mean (Standard deviation, SD) age at inclusion was 58.1y (9.8y). Prior treatment response was null in 31 (12%), partial in 108 (42%), breakthrough in 10 (4%) and relapse in 92 (36%) patients. As previously proposed [21,22], null and partial responders were grouped in a “prior non-response” category, and breakthrough and relapse were grouped in a “prior relapse” category for further analyses. Eighty six individuals (33%) were infected with genotype 1a, 147 (57%) with 1b and 21 (8%) with 1c. The mean (SD) hemoglobin level at baseline was 14.6g.dl⁻¹ (1.7g.dl⁻¹), and the mean (SD) platelet count at baseline was 150,000mm⁻³ (66,000mm⁻³). As expected by the protocol, patients treated with BOC were more likely to receive 4 weeks lead-in with PegIFN/RBV (95.7% vs 26.5%, $P < 0.001$).

Association of SVR status with APOH and IL28B SNPs

SVR was achieved for 119 (46.5%) patients and no significant difference ($P = 0.48$) was observed between the two treatment groups. Results of univariate analysis of *IL28B* and *APOH*

Table 1. Effects of *IL28B* and *APOH* variants on SVR and of *ITPA* variants on anemia in univariate analysis.

SNP	chr:position	Closest gene (variant type)	Reference/ alternative (aaf*)	n	call rate	HWE p-value	OR (95%CI)	P-value
SVR phenotype								
rs12979860	19:39248147	<i>IL28B</i> (intron)	C/T (0.52)	254	99.2	9.9 10 ⁻⁶	2.35 [1.50–3.70]	2.0x10 ⁻⁴
rs368234815	19:39248514	<i>IL28B</i> (splice)	TT/dG (0.52)	242	94.5	3.8 10 ⁻⁷	2.35 [1.46–3.79]	4.7x10 ⁻⁴
rs1801690	17:66212167	<i>APOH</i> (missense)	C/G (0.07)	251	98.0	0.62	1.47 [0.73–2.96]	0.28
rs1801689	17:66214462	<i>APOH</i> (missense)	A/C (0.04)	251	98.0	1	0.65 [0.26–1.61]	0.35
rs52797880	17:66220736	<i>APOH</i> (missense)	A/G (0.08)	242	94.5	1	1.40 [0.71–2.74]	0.33
rs8178822	17:66229411	<i>APOH</i> (5'UTR)	G/T (0.08)	253	98.8	1	1.31 [0.67–2.57]	0.43
rs12944940	17:66235598	<i>APOH</i> (intron)	T/C (0.21)	252	98.4	0.33	1.01 [0.64–1.57]	0.98
rs10048158	17:66240200	<i>APOH</i> (intron)	C/T (0.47)	251	98.0	0.42	0.96 [0.68–1.36]	0.84
Clinically relevant anemia								
rs1127354	20:3213196	<i>ITPA</i> (missense)	C/A (0.06)	255	99.6	1	1.36 [0.65–2.84]	0.42
rs7270101	20:3213247	<i>ITPA</i> (intron)	A/C (0.13)	255	99.6	0.83	1.31 [0.78–2.19]	0.31
Early Hb decline								
rs1127354	20:3213196	<i>ITPA</i> (missense)	C/A (0.06)	209	99.6	1	4.20 [1.38–12.8]	0.01
rs7270101	20:3213247	<i>ITPA</i> (intron)	A/C (0.13)	209	99.6	0.83	2.27 [1.20–4.29]	0.01

* aaf, alternative allele frequency.

doi:10.1371/journal.pone.0145105.t001

SNPs are presented in Table 1. All *APOH* SNPs were in Hardy Weinberg equilibrium (HWE), and some of them were in LD (S1 Fig). No significant association was observed between *APOH* SNPs and SVR. As previously observed in Caucasian individuals, *IL28B* variants, rs12979860 and rs368234815, were in almost complete LD ($r^2 = 0.94$; S1 Fig). As the results for both SNPs were very similar and the call rate of rs12979860 was slightly higher than that of rs368234815, results are presented only for rs12979860. As expected by the selection bias of the sample including only treatment-experienced patients, rs12979860 was not in HWE ($P = 9.9 \times 10^{-6}$) due to an enrichment of unfavorable allele T for clearance (52% in CUPIC patients vs 32% in the 1000 genomes European population (www.ensembl.org). Despite this skewed distribution, the favorable allele C of rs12979860 was significantly associated with SVR to triple therapy in univariate analysis ($P = 2 \times 10^{-4}$) with an Odds ratio (OR) of achieving SVR per increase of one copy of the favorable allele C (i.e. $OR_{C/CvsC/T}$ or $OR_{C/TvsT/T}$) of 2.35 (95% confidence interval: 1.50–3.70) (Table 1 and Fig 1A). The effect of rs12979860 genotype on SVR did not differ significantly between the two treatment groups ($P_{interaction} = 0.3$).

We performed further multivariate analysis including covariates previously identified as independent predictors of SVR (i.e. prior treatment response, no lead-in phase, platelet count $\geq 100,000 \text{mm}^{-3}$ and HCV-1b subtype [12]) (S2 Table), and showed that rs12979860 was independently associated with SVR ($OR = 2.05 [1.24–3.48]$, $p = 5.9 \times 10^{-3}$). The best predictor of SVR in the multivariate analysis was previous relapse to PegIFN/RBV ($OR = 2.69 [1.50–4.88]$, $p = 9.6 \times 10^{-4}$). Hence, we further explored the combined effect of rs12979860 and prior treatment response on SVR, and found a significant interaction ($p = 0.03$) between these two factors. As shown in Fig 1B, the effect of rs12979860 on SVR was observed only in prior relapse ($P = 8.2 \times 10^{-4}$) with a stronger OR of 3.80 [1.82–8.92], while this effect is no more significant ($P = 0.6$) in prior non-responders with an OR of 1.20 [0.63–2.33]. Multivariate analysis restricted to previous relapse patients (S2 Table) showed that rs1279860 has the most significant effect on SVR among other known predictors ($P = 4.4 \times 10^{-4}$) with an OR of 5.01 [2.16–13.3]. Overall, our results suggest that the effect of rs12979860 on triple therapy-induced clearance in treatment-experienced patients is restricted to those who experienced prior PegIFN/RBV relapse.

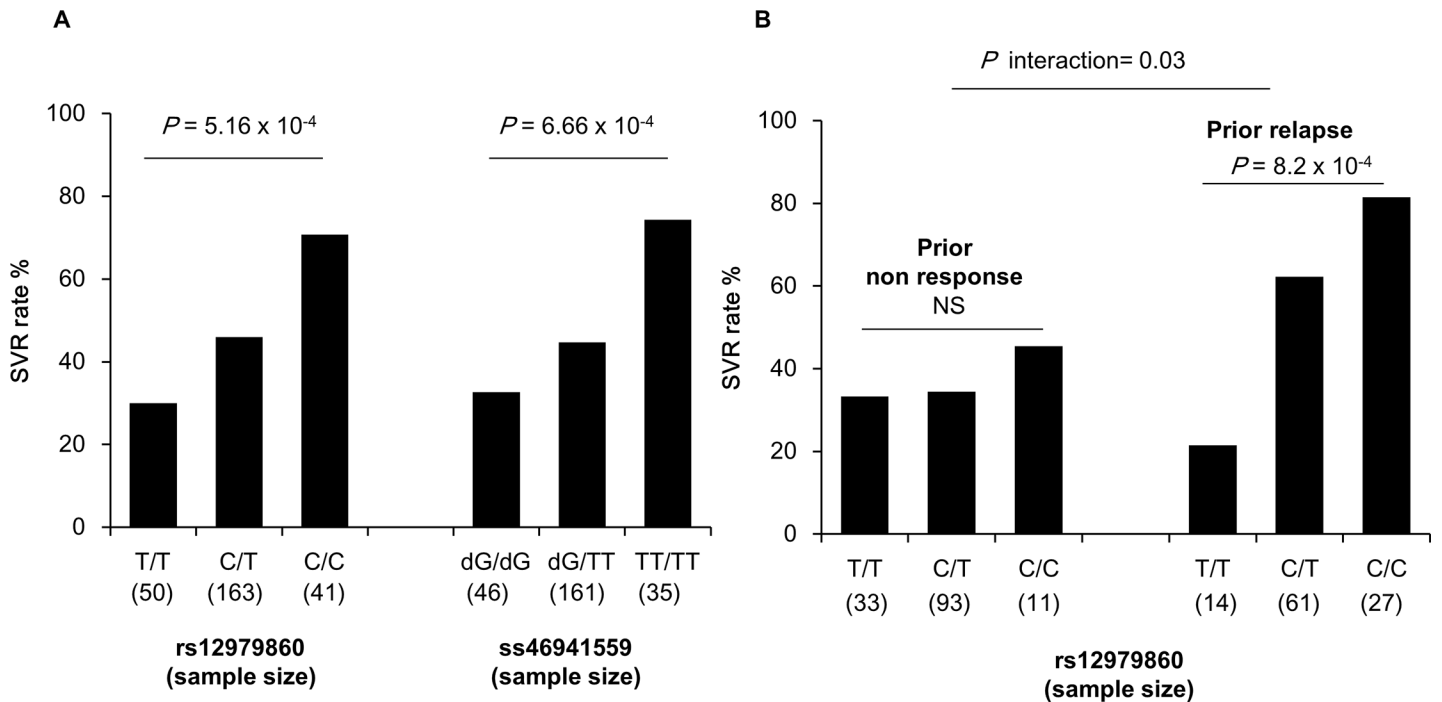


Fig 1. Rate of SVR by genotypes for *IL28B* SNPs. (A) SVR rates by genotypes for rs12979860 (left panel) and rs368234815 (right panel). (B) SVR rates by genotypes for rs12979860 according to prior treatment response.

doi:10.1371/journal.pone.0145105.g001

Association of anemia with ITPA SNPs

Clinically relevant anemia and early significant Hb decline were observed in 155 (60.5%) and 89 (42.6%) patients, respectively. Both *ITPA* SNPs were in HWE (Table 1), and were not in LD (S1 Fig). The frequencies of major alleles of rs1127354 (C) and rs7270101 (A), associated with normal ITPA activity, were 0.94 and 0.87, respectively. In univariate analysis, clinically relevant anemia was associated neither with rs1127354 ($p = 0.42$), nor with rs7270101 ($p = 0.31$). In contrast, early significant Hb decline was significantly associated with rs1127354 ($OR_{C/CvsC/A} = OR_{C/AvsA/A} = 4.20[1.38-12.8]$, $p = 0.01$), and with rs7270101 ($OR_{A/AvsA/C} = OR_{A/CvsC/C} = 2.27[1.20-4.29]$, $P = 0.01$) (Table 1 and Fig 2A). In multivariate analysis including other predictors of anemia (i.e. age in years, sex, lead-in, Hb at baseline and albumin at baseline [10,12]), rs1127654 and rs7270101 major alleles were strongly and independently associated with early significant Hb decline (rs1127354, $OR = 7.83[2.64-29.2]$, $P = 6.0 \times 10^{-4}$; rs7270101, $OR = 3.28[1.65-6.95]$, $P = 1.2 \times 10^{-3}$).

We further explored the joint effect of rs1127354 and rs7270101 by considering as a quantitative covariate the severity of ITPA deficiency estimated from the genotypes at rs1127354 and rs7270101, as shown in S4 Table [7]. Consistent with the SNP effects, clinically relevant anemia was not associated ($p = 0.12$) with ITPA activity while there was a strong effect of ITPA activity ($P = 1.1 \times 10^{-4}$) on the rate of early Hb decline ranging from 15% for 30% activity to 52.3% for full activity (Fig 2B). Early significant Hb decline was defined at week 4 of therapy and was the consequence of either PegIFN/RBV alone for patients having had a lead-in or PegIFN/RBV plus TVR or BOC for individuals who started triple therapy without a lead-in. As already shown [8,9], the absence of lead-in (and consequently the triple therapy) was significantly associated ($P = 6.4 \times 10^{-3}$ in multivariate analysis) with early Hb decline (S3 Table). However, refined analysis showed that the effect of ITPA activity on early Hb decline did not differ significantly

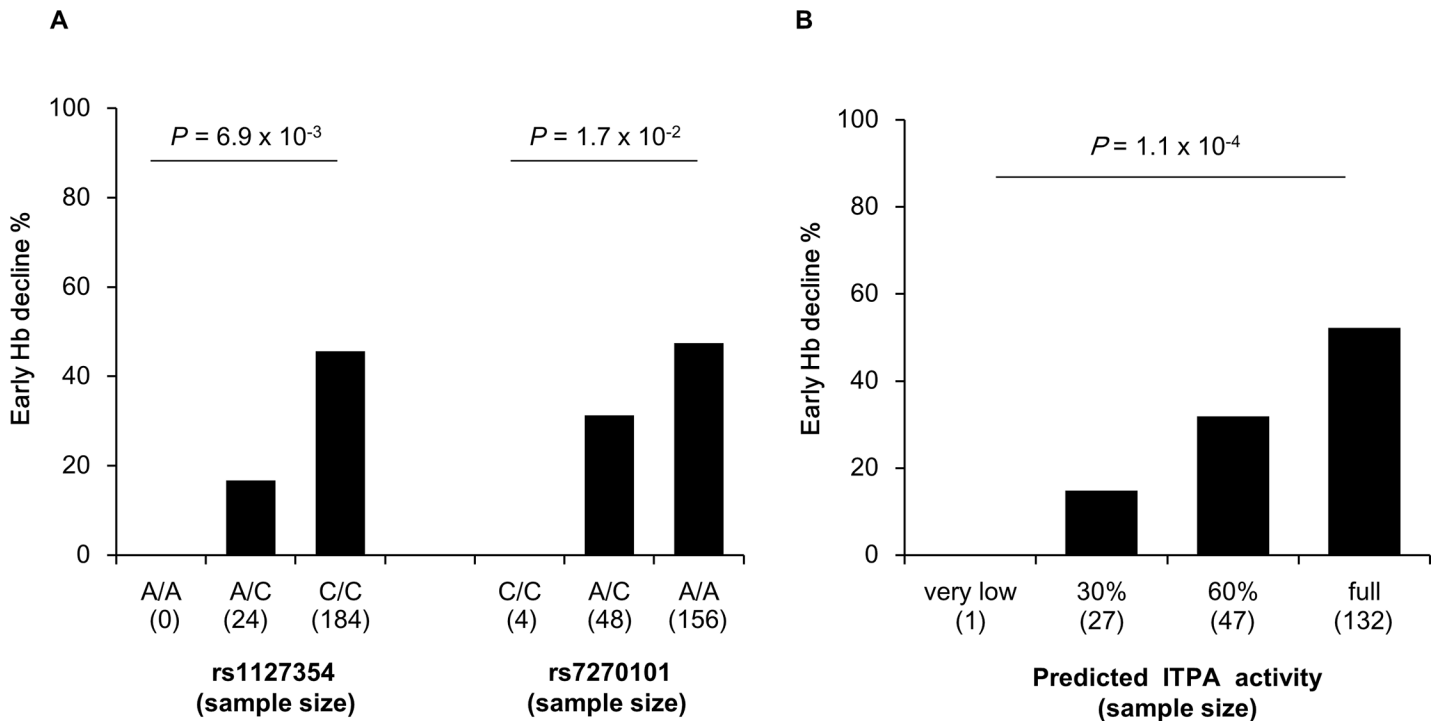


Fig 2. Rate of early hemoglobin decline by genotypes for rs1127354 and rs7270101 (A) and by predicted ITPA activity (B). Severity of ITPA deficiency was defined as in *Fellay et al [7]* considering rs1127354 C and rs7270101 A equivalent low activity variants.

doi:10.1371/journal.pone.0145105.g002

($P_{\text{interaction}} = 0.68$) between the groups of patients with or without lead-in, with similar OR values observed in these two groups of 2.45[1.28–5.47] and 2.99[1.62–5.97], respectively. These results indicate that PIs increase the risk of early Hb decline, but do not have a significant influence on the relationship between early Hb decline and ITPA activity.

Discussion

Triple therapy using BOC or TVR remains the reference treatment for chronically HCV-1 infected patients in a large number of countries, and is of major importance for patients with cirrhosis who are at risk to develop severe complications as liver failure or hepatocellular carcinoma [23]. However, the overall chance of success with triple therapy is around 50% in those cirrhotic patients and the risk of adverse effects remains high [12]. Therefore, providing information that could help to the prediction of achieving SVR for cirrhotic patients under triple therapy is of major interest [23]. In this study, the unique CUPIC cohort of well characterized treatment experienced cirrhotic patients allowed us refining the association between *IL28B* SNPs and SVR to triple therapy, and between *ITPA* SNPs and anemia. *IL28B* genotype is the strongest predictor of SVR to the standard PegIFN/RBV therapy in patients chronically infected with HCV-1. In treatment-naïve HCV-1 patients receiving TVR or BOC in combination with PegIFN/RBV, the association between *IL28B* genotype and SVR remains clinically relevant [21,23,24]. We report here that *IL28B* alleles favorable for clearance were associated with a twofold increase of SVR rate in a cohort of treatment-experienced patients of Caucasian origin, chronically infected by HCV-1, with compensated cirrhosis and receiving either TVR or BOC in triple therapy. Refined analysis showed that the effect of *IL28B* SNPs was restricted to individuals who previously relapsed (i.e. breakthrough or relapse) to PegIFN alone or PegIFN/RBV therapy, with a stronger effect of these SNPs on SVR in this population. This

result suggests that TVR and BOC may potentiate *IL28B*-dependent clearance transiently induced by PegIFN/RBV therapy, and that *IL28B*-independent mechanisms are involved in the non-response to PegIFN/RBV therapy.

Several studies including both treatment-naïve and treatment experienced HCV patients receiving TVR- or BOC-based therapy have also consistently identified *IL28B* genotype as a predictor of SVR independent of treatment history [25–28]. However, results of the few studies focusing only on treatment-experienced patients were less conclusive. In a Japanese cohort of 103 treatment-experienced patients, mono-infected with HCV-1b and receiving TVR, the *IL28B* variant, rs8099917, was an independent predictor of SVR [29]. In the RESPOND-2 study of BOC-based therapy in treatment-experienced patients (N = 207), rs12979860 C/C genotype was predictive of a good interferon response at week 4 but only a non-statistically significant trend was observed with SVR [21]. In the REALIZE study of TVR-based therapy in 422 treatment-experienced patients, SVR rates were slightly higher among patients with rs12979860 C/C genotype compared with C/T and T/T genotypes but the difference was not statistically significant [22]. In these two latter studies, patients included variable proportions of prior responders (slightly higher in the RESPOND-2 than in the REALIZE study), and a rather low proportion of cirrhotic patients (<30%). Further studies are needed to identify the factors, like prior response, ethnic origin, liver fibrosis status, or HCV-1 genotype, which could explain the differences observed in the strength of association between SVR and *IL28B* genotype in treatment-experienced patients receiving either TVR or BOC.

No significant association was observed between *APOH* SNPs and SVR. One explanation could be that the impact of each individual variant on apoH levels is not large enough to further impact on SVR. Moreover, our study was underpowered to detect an association signal between SVR and variant with minor allele frequency below 0.1, which is the case for four of the six tested *APOH* SNPs. As an example, the power to detect an association between SVR and the missense SNP rs1801690 was 55% at the 0.05 type I error level for an additive OR of 2. Finally, despite a careful literature search and public database screening, we may have missed some variants impacting on apoH levels that are not identified yet.

Anemia is a well-established adverse event with PegIFN/RBV treatment of chronic HCV infection and the addition of PIs, such as TVR and BOC, has significantly increased its incidence [8,9]. Our data are consistent with this observation as the patients without lead-in had a higher rate of early Hb decline. The role of *ITPA* polymorphisms on early Hb decline and/or early anemia was identified by GWAS [7] and further replicated in different ethnic groups treated both by PegIFN/RBV [30–35], and by a triple combination therapy with TVR [31,36]. Here, we confirmed the protective effect of low *ITPA* activity variants on early Hb decline in treatment-experienced patients having received pegIFN/RBV alone or triple therapy combination for 4 weeks. We could also show that the relationship between *ITPA* SNPs and early Hb decline was not influenced by the presence of PIs, indicating that the effect of PIs on Hb decline is probably independent of *ITPA* activity. In our study, *ITPA* deficiency did not protect against clinically relevant anemia, which has a broader definition based on both Hb decline and anemia management during the whole period, a finding consistent with the previous study of Aghemo et al. [36].

Overall, our results suggest that the screening of rs12979860 remains interesting for decision making in Caucasian difficult-to-treat HCV-1 patients (in particular if they presented a prior PegIFN/RBV relapse) with compensated cirrhosis eligible for a PI-based triple therapy. In those patients, the genotyping of *ITPA* SNPs are very useful to predict the development of early severe Hb decline.

Supporting Information

S1 Fig. LD plot of *APOH*, *IL28B* and *ITPA* SNPs (chromosome 17, 19 and 20 respectively).
(DOCX)

S1 Table. Characteristics of the patients
(DOCX)

S2 Table. Factors related to SVR: multivariate analysis.
(DOCX)

S3 Table. Factors related to early hemoglobin decline: univariate and multivariate analysis.
(DOCX)

S4 Table. Predicted ITPA activity according to genotypes at the two ITPA SNPs and corresponding number of observed patients for early hemoglobin decline analysis.
(DOCX)

Acknowledgments

We are grateful to all the study participants. We thank Jean-Laurent Casanova, all members of both branches of the laboratory of Human Genetics of Infectious Diseases, and Estelle Mottez from INSERM UMS20. We thank the INSERM-ANRS CO20 CUPIC Study Group, led by Christophe Hézode (christophe.hezode@hmn.aphp.fr) and H el ene Fontaine (helene.fontaine@cch.aphp.fr):

Thierry Poynard, Service d'H epato-gastro-ent erologie, H opital La Piti e Salp etri re, Paris, France; Val erie Canva, Service des Maladies de l'appareil Digestif, Centre Hospitalier R egional Universitaire de Lille-H opital Huriez, Lille, France; Patrice Cacoub, Service de M edecine Interne, H opital La Piti e Salp etri re, Paris, France; Didier Samuel, Service d'H epato-gastro-ent erologie, H opital Paul Brousse, Villejuif, France; Patrick Marcellin, Service d'H epato-gastro-ent erologie, H opital Beaujon, Clichy-la Garenne, France; Laurent Alric, P ole Digestif-Service de M edecine Interne, Centre Hospitalier Universitaire de Purpan, Toulouse, France; Victor De L edinghen, Service d'H epato-gastro-ent erologie, H opital Haut-L ev eque, Pessac, France; Marc Bourli ere, Service d'H epato-gastro-ent erologie, fondation H opital Saint Joseph, Marseille, France; Jean-Pierre Zarski, Clinique Universitaire d'H epato-gastro-ent erologie, Centre Hospitalier R egional Universitaire Saint Antoine, Grenoble, France; Jean-Jacques Raabe, Unit e de Recherche Clinique, Centre Hospitalier de Metz, Thionville-H opital Mercy, Metz, France; Sophie M etivier, p ole Digestif, Gastro-ent erologie-H epato-gastro-ent erologie, Centre hospitalier universitaire Purpan, Toulouse, France; Lawrence Serfaty, Service d'H epato-gastro-ent erologie, H opital Saint-Antoine, AP-HP, Paris, France; Ghasan Riachi, Service d'H epato-gastro-ent erologie et nutrition, Centre Hospitalier Universitaire de Rouen, Rouen, France; Armand Abergel, P ole Digestif, Centre Hospitalier Estaing, Clermont Ferrand, France; V eronique Loustaud-Ratti, Service d'H epato-gastro-ent erologie, Centre Hospitalier Universitaire Dupuyten, Limoges, France; Albert Tran, P ole Digestif, Centre hospitalier Universitaire de Nice, Nice, France; Xavier Causse, Service d'H epato-gastro-ent erologie et de Gastroent erologie et oncologie digestive, Centre Hospitalier R egional La Source, Orl eans, France; Dominique Guyader, Service des Maladies du Foie, Centre Hospitalier Universitaire de Pontchaillou, Rennes, France; Pierre-Henri Bernard, Service d'H epato-gastro-ent erologie, H opital Saint Andr e, Bordeaux, France; Pierre Attali, Service d'H epato-gastro-ent erologie, H opital de Bic etre, AP-HP, Le Kremlin Bic etre, France; Vincent Di-Martino, Service d'H epato-gastro-ent erologie, Centre hospitalier de Jean Minjoz, Besan on, France; Paul Cal es, Service d'H epato-gastro-ent erologie, Centre Hospitalier Universitaire, Angers, France; V eronique Grando-Lemaire, Service d'H epato-gastro-ent erologie, H opital Jean Verdier,

AP-HP, Bondy, France; Isabelle Rosa, Service d'Hépatologie et de Gastroentérologie, Centre hospitalier Intercommunal de Créteil, Créteil, France; Danièle Botta-Friedland, Service d'Hépatologie et Gastroentérologie, Hôpital de la Conception, Marseille, France; Thong Dao, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier Universitaire, Caen, France; Damien Lucidarne, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier de l'institut Catholique Lillois, Lille, France; Patrick Hillon, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier Universitaire de Dijon, Dijon, France; Cyrille Feray, Service d'Hépatologie et Gastroentérologie, Hôpital Hôtel Dieu, Nantes, France; Thierry Fontanges, Service d'Hépatologie et Gastroentérologie, Hôpital P Oudot, Bourgoin-Jallieu, France; Jean-Didier Grange, Service d'Hépatogastroentérologie, Hôpital Tenon, Paris, France; Gilles Gatineau-Sailliant, Centre Hospitalier, Meaux, France; Eric Poncin, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier de Dax, Dax, France; Jean-Pierre Arpurt, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier, Avignon, France; Yannick Bacq, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier Universitaire Trousseau, Tours, France; Patrick Delasalle, Service d'Hépatologie et Gastroentérologie, Clinique du Palais, Grasse, France; Denis Ouzan, Service d'Hépatologie et Gastroentérologie, Institut Arnaud Tzanck, Saint-Laurent du Var, France; Jean-Baptiste Nousbaum, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier Universitaire de la Cavale Blanche, Brest, France; Christine Sylvain, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier; Universitaire, Poitiers, France; Didier Ribard, Service d'Hépatologie et Gastroentérologie, Centre Hospitalier Universitaire Caremeau, Nîmes, France; Philippe Renard, Centre Hospitalier Victor Dupouy, Argenteuil, France; Caroline Lasoux-Combe, Service de Maladies Infectieuses, Hôpital Saint-Louis, APHP, Paris, France; Stéphanie de Montigny-Lenhardt, Centre Hospitalier Edmond Garcin, Aubagne, France; Christophe Pilette, Centre Hospitalier, Le Mans, France; Jacques Denis, Centre Hospitalier Sud Francilien, Corbeil-Essonnes, France; Thierry Allègre, Service d'Hématologie-Oncologie Médecine Interne, centre Hospitalier du Pays d'Aix, Aix en Provence, France; Matthieu Schnee, service d'Hépatogastroentérologie, Centre Hospitalier Départemental Les Oudairies, aA Roche Sur Yon, France; Gaëtan Franck, Cabinet de GastroEntéroHépatologie- Appareil Digestif, La Rochelle, France; Jean-Marc Combis, Clinique Ambroise Paré, Toulouse, France; Pierre Bedossa, Service d'Anatomo-Pathologie, Hôpital Beaujon, Clichy, France; Jean-Michel Pawlotsky, Centre National de Référence pour les Hépatites Virales B, C et delta, Service de Virologie, Hôpital Henri Mondor, AP-HP, Créteil, France; Marianne L'Henaff, Association de patients, ARCAT, TRT-5, Paris France; Michelle Sizorn, Association de patients, SOS Hépatites, Paris France; Ventzislava Petrov-Sanchez, Inserm-ANRS, Service Hépatites, Paris, France; Olivier Chazouillères, Service d'Hépatologie, Hôpital Saint-Antoine, AP-HP, Paris, France; Jean Dubuisson, Unité de Virologie, INSERM U1019, CNRS UMR8204, Lille, France; Francesco Negro, Hôpital Cantonal, Genève, Suisse; Georges-Philippe Pageaux, Hôpital Saint Eloi, Montpellier, France; Valérie Paradis, Service d'anatomopathologie, Hôpital Beaujon, AP-HP, Clichy, France; Bruno Spire, Santé Publique, Sciences Humaines et Sociales, INSERM, Marseille, France; Anne-Marie Taburet, Pharmacologie, Hôpital Bicêtre, AP-HP, Le Kremlin-Bicêtre, France; Jean-Claude Trinchet, Service d'Hépatologie, Hôpital Jean Verdier, AP-HP, Bondy, France; Yazdan Yazdanpanah, Economie de la Santé, Hôpital Bichat, AP-HP, Paris, France; Céline Dorival-Mouly, INSERM UMR-S 1136 Université Pierre et Marie Curie Paris 6, Paris, France; Cécilie Dufour, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France; Céline Fréhaut, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France; Aurélie Lesel, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France; Nathalie Zahraa, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France; Marion Pirot, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France; Yoann Barthe, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6,

Paris, France; Frédéric Chau, INSERM UMR-S 1136, Université Pierre et Marie Curie Paris 6, Paris, France.

Author Contributions

Conceived and designed the experiments: AC LA IT. Performed the experiments: TOM JN FA. Analyzed the data: FA AC VP. Contributed reagents/materials/analysis tools: PR DD CC PS FC PM FZ DL CH HF JPB SP MLA IT. Wrote the paper: FA AC LA.

References

1. Gower E, Estes C, Blach S, Razavi-Shearer K, Razavi H. Global epidemiology and genotype distribution of the hepatitis C virus infection. *J Hepatol*. 2014 Nov; 61(1, Supplement):S45–57.
2. Fried MW, Shiffman ML, Reddy KR, Smith C, Marinos G, Gonçales FL, et al. Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. *N Engl J Med*. 2002 Sep 26; 347(13):975–82. PMID: [12324553](#)
3. Manns MP, McHutchison JG, Gordon SC, Rustgi VK, Shiffman M, Reindollar R, et al. Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: a randomised trial. *Lancet*. 2001 Sep 22; 358(9286):958–65. PMID: [11583749](#)
4. Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, et al. Genetic variation in IL28B predicts hepatitis C treatment-induced viral clearance. *Nature*. 2009 Sep 17; 461(7262):399–401. doi: [10.1038/nature08309](#) PMID: [19684573](#)
5. Casanova J-L, Abel L. The genetic theory of infectious diseases: a brief history and selected illustrations. *Annu Rev Genomics Hum Genet*. 2013; 14:215–43. doi: [10.1146/annurev-genom-091212-153448](#) PMID: [23724903](#)
6. Prokunina-Olsson L, Muchmore B, Tang W, Pfeiffer RM, Park H, Dickensheets H, et al. A variant upstream of IFNL3 (IL28B) creating a new interferon gene IFNL4 is associated with impaired clearance of hepatitis C virus. *Nat Genet*. 2013 Feb; 45(2):164–71. doi: [10.1038/ng.2521](#) PMID: [23291588](#)
7. Fellay J, Thompson AJ, Ge D, Gumbs CE, Urban TJ, Shianna KV, et al. ITPA gene variants protect against anaemia in patients treated for chronic hepatitis C. *Nature*. 2010 Mar 18; 464(7287):405–8. doi: [10.1038/nature08825](#) PMID: [20173735](#)
8. Bacon BR, Gordon SC, Lawitz E, Marcellin P, Vierling JM, Zeuzem S, et al. Boceprevir for previously treated chronic HCV genotype 1 infection. *N Engl J Med*. 2011 Mar 31; 364(13):1207–17. doi: [10.1056/NEJMoa1009482](#) PMID: [21449784](#)
9. Zeuzem S, Andreone P, Pol S, Lawitz E, Diago M, Roberts S, et al. Telaprevir for Retreatment of HCV Infection. *N Engl J Med*. 2011 Jun 23; 364(25):2417–28. doi: [10.1056/NEJMoa1013086](#) PMID: [21696308](#)
10. Hézode C, Fontaine H, Dorival C, Larrey D, Zoulim F, Canva V, et al. Triple therapy in treatment-experienced patients with HCV-cirrhosis in a multicentre cohort of the French Early Access Programme (ANRS CO20-CUPIC)—NCT01514890. *J Hepatol*. 2013 Sep; 59(3):434–41. doi: [10.1016/j.jhep.2013.04.035](#) PMID: [23669289](#)
11. Zeuzem S, Jacobson IM, Baykal T, Marinho RT, Poordad F, Bourlière M, et al. Retreatment of HCV with ABT-450/r-ombitasvir and dasabuvir with ribavirin. *N Engl J Med*. 2014 Apr 24; 370(17):1604–14. doi: [10.1056/NEJMoa1401561](#) PMID: [24720679](#)
12. Hézode C, Fontaine H, Dorival C, Zoulim F, Larrey D, Canva V, et al. Effectiveness of telaprevir or boceprevir in treatment-experienced patients with HCV genotype 1 infection and cirrhosis. *Gastroenterology*. 2014 Jul; 147(1):132–42.e4. doi: [10.1053/j.gastro.2014.03.051](#) PMID: [24704719](#)
13. Sultanik P, Mallet V, Lagaye S, Casrouge A, Dorival C, Barthe Y, et al. Plasma apolipoprotein H limits HCV replication and associates with response to NS3 protease inhibitors-based therapy. *Liver Int*. 2015;n/a–n/a.
14. Cassader M, Ruiu G, Gambino R, Guzzon F, Pagano A, Veglia F, et al. Influence of apolipoprotein H polymorphism on levels of triglycerides. *Atherosclerosis*. 1994 Sep 30; 110(1):45–51. PMID: [7857369](#)
15. Shipkova M, Lorenz K, Oellerich M, Wieland E, Ahsen N von. Measurement of erythrocyte inosine triphosphate pyrophosphohydrolase (ITPA) activity by HPLC and correlation of ITPA genotype-phenotype in a Caucasian population. *Clin Chem*. 2006 Feb; 52(2):240–7. PMID: [16384889](#)
16. Mehdi H, Manzi S, Desai P, Chen Q, Nestlerode C, Bontempo F, et al. A functional polymorphism at the transcriptional initiation site in beta2-glycoprotein I (apolipoprotein H) associated with reduced gene expression and lower plasma levels of beta2-glycoprotein I. *Eur J Biochem FEBS*. 2003 Jan; 270(2):230–8.

17. Suresh S, Demirci FYK, Jacobs E, Kao AH, Rhew EY, Sanghera DK, et al. Apolipoprotein H promoter polymorphisms in relation to lupus and lupus-related phenotypes. *J Rheumatol*. 2009 Feb; 36(2):315–22. doi: [10.3899/jrheum.080482](https://doi.org/10.3899/jrheum.080482) PMID: [19132787](https://pubmed.ncbi.nlm.nih.gov/19132787/)
18. Athanasiadis G, Sabater-Lleal M, Buil A, Souto JC, Borrell M, Lathrop M, et al. Genetic determinants of plasma β_2 -glycoprotein I levels: a genome-wide association study in extended pedigrees from Spain. *J Thromb Haemost JTH*. 2013 Mar; 11(3):521–8. doi: [10.1111/jth.12120](https://doi.org/10.1111/jth.12120) PMID: [23279374](https://pubmed.ncbi.nlm.nih.gov/23279374/)
19. Leduc MS, Shimmin LC, Klos KLE, Hanis C, Boerwinkle E, Hixson JE. Comprehensive evaluation of apolipoprotein H gene (APOH) variation identifies novel associations with measures of lipid metabolism in GENOA. *J Lipid Res*. 2008 Dec; 49(12):2648–56. doi: [10.1194/jlr.M800155-JLR200](https://doi.org/10.1194/jlr.M800155-JLR200) PMID: [18676959](https://pubmed.ncbi.nlm.nih.gov/18676959/)
20. Mehdi H, Aston CE, Sanghera DK, Hamman RF, Kamboh MI. Genetic variation in the apolipoprotein H (beta2-glycoprotein I) gene affects plasma apolipoprotein H concentrations. *Hum Genet*. 1999 Aug; 105(1–2):63–71. PMID: [10480357](https://pubmed.ncbi.nlm.nih.gov/10480357/)
21. Poordad F, Bronowicki J-P, Gordon SC, Zeuzem S, Jacobson IM, Sulkowski MS, et al. Factors that predict response of patients with hepatitis C virus infection to boceprevir. *Gastroenterology*. 2012 Sep; 143(3):608–18.e1–5. doi: [10.1053/j.gastro.2012.05.011](https://doi.org/10.1053/j.gastro.2012.05.011) PMID: [22626609](https://pubmed.ncbi.nlm.nih.gov/22626609/)
22. Pol S, Aerssens J, Zeuzem S, Andreone P, Lawitz EJ, Roberts S, et al. Limited impact of IL28B genotype on response rates in telaprevir-treated patients with prior treatment failure. *J Hepatol*. 2013 May; 58(5):883–9. doi: [10.1016/j.jhep.2012.12.023](https://doi.org/10.1016/j.jhep.2012.12.023) PMID: [23321318](https://pubmed.ncbi.nlm.nih.gov/23321318/)
23. Jacobson IM, Catlett I, Marcellin P, Bzowej NH, Muir AJ, Adda N, et al. 1369 TELAPREVIR SUBSTANTIALLY IMPROVED SVR RATES ACROSS ALL IL28B GENOTYPES IN THE ADVANCE TRIAL. *J Hepatol*. 2011 Mar 1; 54:S542–3.
24. Holmes JA, Desmond PV, Thompson AJ. Does IL28B genotyping still have a role in the era of direct-acting antiviral therapy for chronic hepatitis C infection? *J Viral Hepat*. 2012 Oct; 19(10):677–84. doi: [10.1111/jvh.12003](https://doi.org/10.1111/jvh.12003) PMID: [22967098](https://pubmed.ncbi.nlm.nih.gov/22967098/)
25. Chayama K, Hayes CN, Abe H, Miki D, Ochi H, Karino Y, et al. IL28B but not ITPA polymorphism is predictive of response to pegylated interferon, ribavirin, and telaprevir triple therapy in patients with genotype 1 hepatitis C. *J Infect Dis*. 2011 Jul 1; 204(1):84–93. doi: [10.1093/infdis/jir210](https://doi.org/10.1093/infdis/jir210) PMID: [21628662](https://pubmed.ncbi.nlm.nih.gov/21628662/)
26. Akuta N, Suzuki F, Fukushima T, Kawamura Y, Sezaki H, Suzuki Y, et al. Prediction of treatment efficacy and telaprevir-resistant variants after triple therapy in patients infected with hepatitis C virus genotype 1. *J Clin Microbiol*. 2013 Sep; 51(9):2862–8. doi: [10.1128/JCM.01129-13](https://doi.org/10.1128/JCM.01129-13) PMID: [23784126](https://pubmed.ncbi.nlm.nih.gov/23784126/)
27. Tsubota A, Shimada N, Atsukawa M, Abe H, Kato K, Ika M, et al. Impact of IL28B polymorphisms on 24-week telaprevir-based combination therapy for Asian chronic hepatitis C patients with hepatitis C virus genotype 1b. *J Gastroenterol Hepatol*. 2014 Jan; 29(1):144–50. doi: [10.1111/jgh.12402](https://doi.org/10.1111/jgh.12402) PMID: [24117654](https://pubmed.ncbi.nlm.nih.gov/24117654/)
28. Sterling RK, Kuo A, Rustgi VK, Sulkowski MS, Stewart TG, Fenkel JM, et al. Virological outcomes and treatment algorithms utilisation in observational study of patients with chronic hepatitis C treated with boceprevir or telaprevir. *Aliment Pharmacol Ther*. 2015 Apr; 41(7):671–85. doi: [10.1111/apt.13095](https://doi.org/10.1111/apt.13095) PMID: [25627020](https://pubmed.ncbi.nlm.nih.gov/25627020/)
29. Shimada N, Tsubota A, Atsukawa M, Abe H, Ide T, Takaguchi K, et al. A 48-week telaprevir-based triple combination therapy improves sustained virological response rate in previous non-responders to peginterferon and ribavirin with genotype 1b chronic hepatitis C: A multicenter study. *Hepatol Res Off J Jpn Soc Hepatol*. 2014 Mar 10;
30. Thompson AJ, Fellay J, Patel K, Tillmann HL, Naggie S, Ge D, et al. Variants in the ITPA gene protect against ribavirin-induced hemolytic anemia and decrease the need for ribavirin dose reduction. *Gastroenterology*. 2010 Oct; 139(4):1181–9. doi: [10.1053/j.gastro.2010.06.016](https://doi.org/10.1053/j.gastro.2010.06.016) PMID: [20547162](https://pubmed.ncbi.nlm.nih.gov/20547162/)
31. Suzuki F, Suzuki Y, Akuta N, Sezaki H, Hirakawa M, Kawamura Y, et al. Influence of ITPA polymorphisms on decreases of hemoglobin during treatment with pegylated interferon, ribavirin, and telaprevir. *Hepatol Baltim Md*. 2011 Feb; 53(2):415–21.
32. Eskesen AN, Melum E, Moghaddam A, Bjørø K, Verbaan H, Ring-Larsen H, et al. Genetic variants at the ITPA locus protect against ribavirin-induced hemolytic anemia and dose reduction in an HCV G2/G3 cohort. *Eur J Gastroenterol Hepatol*. 2012 Aug; 24(8):890–6. doi: [10.1097/MEG.0b013e3283546efd](https://doi.org/10.1097/MEG.0b013e3283546efd) PMID: [22584257](https://pubmed.ncbi.nlm.nih.gov/22584257/)
33. Rau M, Stickel F, Russmann S, Manser CN, Becker PP, Weisskopf M, et al. Impact of genetic SLC28 transporter and ITPA variants on ribavirin serum level, hemoglobin drop and therapeutic response in patients with HCV infection. *J Hepatol*. 2013 Apr; 58(4):669–75. doi: [10.1016/j.jhep.2012.11.027](https://doi.org/10.1016/j.jhep.2012.11.027) PMID: [23195617](https://pubmed.ncbi.nlm.nih.gov/23195617/)
34. Ahmed WH, Furusyo N, Zaky S, Eldin AS, Aboalam H, Ogawa E, et al. Pre-treatment role of inosine triphosphate pyrophosphatase polymorphism for predicting anemia in Egyptian hepatitis C virus patients.

World J Gastroenterol WJG. 2013 Mar 7; 19(9):1387–95. doi: [10.3748/wjg.v19.i9.1387](https://doi.org/10.3748/wjg.v19.i9.1387) PMID: [23538996](https://pubmed.ncbi.nlm.nih.gov/23538996/)

35. Clark PJ, Aghemo A, Degaspero E, Galmozzi E, Urban TJ, Vock DM, et al. Inosine triphosphatase deficiency helps predict anaemia, anaemia management and response in chronic hepatitis C therapy. *J Viral Hepat*. 2013 Dec; 20(12):858–66. doi: [10.1111/jvh.12113](https://doi.org/10.1111/jvh.12113) PMID: [24304455](https://pubmed.ncbi.nlm.nih.gov/24304455/)
36. Aghemo A, Grassi E, Rumi MG, Ambrosio R D', Galmozzi E, Degaspero E, et al. Limited utility of ITPA deficiency to predict early anemia in HCV patients with advanced fibrosis receiving Telaprevir. *PloS One*. 2014; 9(4):e95881. doi: [10.1371/journal.pone.0095881](https://doi.org/10.1371/journal.pone.0095881) PMID: [24760000](https://pubmed.ncbi.nlm.nih.gov/24760000/)