

# Draft Genome of the *Arthrobacter* sp. Strain Edens01

M. B. Couger,<sup>a</sup> Radwa A. Hanafy,<sup>a</sup> Curtis Edens,<sup>a</sup> Connie Budd,<sup>a</sup> Donald P. French,<sup>b</sup> Wouter D. Hoff,<sup>a</sup> Mostafa S. Elshahed,<sup>a</sup> Noha H. Youssef<sup>a</sup>

Department of Microbiology and Molecular Genetics, Oklahoma State University, Stillwater, Oklahoma, USA<sup>a</sup>; Department of Integrative Biology, Oklahoma State University, Stillwater, Oklahoma, USA<sup>b</sup>

**We report the draft genome sequence of *Arthrobacter* sp. strain Edens01, isolated from a leaf surface of a *Rosa* hybrid plant as part of the Howard Hughes Medical Institute-funded Student Initiated Microbial Discovery (SIMD) project. The genome has a total size of 3,639,179 bp and contig  $N_{50}$  of 454,897 bp.**

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Address correspondence to Noha H. Youssef, Noha@okstate.edu.

*Arthrobacter*, a bacterial genus frequently encountered in soil, has the capacity to metabolize numerous recalcitrant compounds (1), and is hence a valuable contributor to natural-attenuation-based, and engineered bioremediation schemes in multiple environments including groundwater (2, 3), crude-oil fields (4), and soil (5). Compounds that have been demonstrated to be metabolized by *Arthrobacter* spp. include chlorophenols (6), commonly found in pesticides, atrazine (7), a commonly used herbicide, nitrobenzoates (8), used in chemical and dye synthesis, and phthalates (9), compounds commonly used as a softening agent in plastics. Here, we present the genome of *Arthrobacter* sp. strain Edens01, which contains numerous genes important for bioremediation.

*Arthrobacter* sp. strain Edens01 was isolated from the leaf surface of a *Rosa* hybrid plant and sequenced at the University of Georgia Genomics Facility using the Illumina MiSeq sequencing platform, and 300 × 2 paired-end chemistry. Reads were quality-filtered with standard Illumina filtering settings, resulting in 753,618 read pairs, 452.2 MB of quality sequence data. All quality-filtered reads were assembled using the short read de Bruijn graph assembly (10) program Velvet (11) with set to a k-mer value of 101 bp and a minimum contig coverage value of 7×. Gene models were created using the prokaryotic gene calling software package Prodigal (12). The Velvet assembly had a total size of 3,639,179 bp, a G+C content of 64.73%, and 3,374 predicted proteins. Translated protein sequences were functionally annotated using a combination of NCBI Blast C++ homology search (13) and HMMER 3.0 hmmscan (14) against the PFAM 26.0 database (15).

Based on 16S rRNA gene-based comparisons to genomes publicly available in GenBank database ( $n = 302,955,543$ , October 2015), strain Edens01 was most closely related (97.0% sequence similarity) to *Arthrobacter* sp. 35W genomic scaffold K254DRAFT (GenBank accession number NZ\_AXVQ01000000). *Arthrobacter* sp. strain Edens01 16S rRNA gene also shared 97.0% sequence similarity with *Arthrobacter* sp. Rue61a, *Arthrobacter aurescens* TC1, *Arthrobacter* sp. M2012083, *Arthrobacter* sp. H41, *Arthrobacter* sp. 31Y, *Arthrobacter* sp. 135MFC05, *Arthrobacter* sp. Br18, *Arthrobacter* sp. CAL618, and *Arthrobacter* sp. TB 23. Whole-

genome comparison of *Arthrobacter* sp. Edens01 to protein coding gene models in the related genomes using BLASTp ( $e^{-5}$  cutoff) revealed a high proportion of shared genes (core genome) between *Arthrobacter* sp. Edens01 and closely related *Arthrobacter* spp. (2,703/3,374, 80.1% with strain Rue61a, 2,702/3,374, 80.1% with *Arthrobacter aurescens* TC1, 2,715/3,374, 81.9% with *Arthrobacter* sp. M2012083, 2,402/3,374, 71.2% with *Arthrobacter* sp. H41, 2,645/3,374, 78.4% with *Arthrobacter* sp. 135MFC05, 2,482/3,374, 73.6% with *Arthrobacter* sp. Br18, 2,498/3,374, 74.0% with *Arthrobacter* sp. CAL618, and 2,536/3,374, 75.2% with *Arthrobacter* sp. TB 23). Genomic analysis of *Arthrobacter* sp. Edens01 revealed numerous genes putatively involved in the degradation of monoaromatics and xenobiotics, including protocatechuate 3,4-dioxygenase (16), phenol 2-monooxygenase (17), 4-hydroxybenzoate 3-monooxygenase (18), ethyl tert-butyl ether degradation protein (19), and pentachlorophenol 4-monooxygenase (20).

In conclusion, this initial genomic analysis of strain Edens01 reveals the presence of many genes involved in bioremediation and contributes to the study and pangenomic repertoire of the metabolically versatile genus *Arthrobacter*.

**Nucleotide sequence accession number.** The GenBank accession number for the genome is [LKIU000000000](https://genbank.ncbi.nlm.nih.gov/GenBank/FASTA/seqview.fcgi?acc=KLU000000000).

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## REFERENCES

- Shapir N, Mongodin EF, Sadowsky MJ, Daugherty SC, Nelson KE, Wackett LP. 2007. Evolution of catabolic pathways: genomic insights into microbial s-triazine metabolism. *J Bacteriol* 189:674–682. <http://dx.doi.org/10.1128/JB.01257-06>.
- Lu Q, Zhu RL, Yang J, Li H, Liu YD, Lu SG, Luo QS, Lin KF. 2015. Natural attenuation model and biodegradation for 1,1,1-trichloroethane contaminant in shallow groundwater. *Front Microbiol* 6:839. <http://dx.doi.org/10.3389/fmicb.2015.00839>.
- Nousiainen AO, Björklöf K, Sagarkar S, Nielsen JL, Kapley A, Jørgensen

- KS. 4 August 2015. Bioremediation strategies for removal of residual atrazine in the boreal groundwater zone. *Appl Microbiol Biotechnol*. <http://dx.doi.org/10.1007/s00253-015-6828-2>.
4. Sun W, Li J, Jiang L, Sun Z, Fu M, Peng X. 2015. Profiling microbial community structures across six large oilfields in China and the potential role of dominant microorganisms in bioremediation. *Appl Microbiol Biotechnol* 99:8751–8764. <http://dx.doi.org/10.1007/s00253-015-6748-1>.
  5. Sagarkar S, Mukherjee S, Nousiainen A, Björklöf K, Purohit HJ, Jørgensen KS, Kapley A. 2013. Monitoring bioremediation of atrazine in soil microcosms using molecular tools. *Environ Pollut* 172:108–115. <http://dx.doi.org/10.1016/j.envpol.2012.07.048>.
  6. Arora PK, Mohanta TK, Srivastava A, Bae H, Singh VP. 2014. Metabolic pathway for degradation of 2-chloro-4-aminophenol by *Arthrobacter* sp. SPG. *Microb Cell Fact* 13:164. <http://dx.doi.org/10.1186/s12934-014-0164-6>.
  7. Sagarkar S, Bhardwaj P, Storck V, Devers-Lamrani M, Martin-Laurent F, Kapley A. 25 September 2015. s-triazine degrading bacterial isolate *Arthrobacter* sp. AK-YN10, a candidate for bioaugmentation of atrazine contaminated soil. *Appl Microbiol Biotechnol*. <http://dx.doi.org/10.1007/s00253-015-6975-5>.
  8. Arora PK, Sharma A. 2015. New metabolic pathway for degradation of 2-nitrobenzoate by *Arthrobacter* sp. SPG. *Front Microbiol* 6:551. <http://dx.doi.org/10.3389/fmicb.2015.00551>.
  9. Wen ZD, Gao DW, Wu WM. 2014. Biodegradation and kinetic analysis of phthalates by an *Arthrobacter* strain isolated from constructed wetland soil. *Appl Microbiol Biotechnol* 98:4683–4690. <http://dx.doi.org/10.1007/s00253-014-5568-z>.
  10. Compeau PE, Pevzner PA, Tesler G. 2011. How to apply de Bruijn graphs to genome assembly. *Nat Biotechnol* 29:987–991. <http://dx.doi.org/10.1038/nbt.2023>.
  11. Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 18:821–829. <http://dx.doi.org/10.1101/gr.074492.107>.
  12. Hyatt D, Chen GL, Locascio PF, Land ML, Larimer FW, Hauser LJ. 2010. Prodigal: prokaryotic gene recognition and translation initiation site identification. *BMC Bioinformatics* 11:119. <http://dx.doi.org/10.1186/1471-2105-11-119>.
  13. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <http://dx.doi.org/10.1186/1471-2105-10-421>.
  14. Johnson LS, Eddy SR, Portugaly E. 2010. Hidden Markov model speed heuristic and iterative HMM search procedure. *BMC Bioinformatics* 11:431. <http://dx.doi.org/10.1186/1471-2105-11-431>.
  15. Finn RD, Bateman A, Clements J, Coggill P, Eberhardt RY, Eddy SR, Heger A, Hetherington K, Holm L, Mistry J, Sonnhammer EL, Tate J, Punta M. 2014. Pfam: the protein families database. *Nucleic Acids Res* 42:D222–D230. <http://dx.doi.org/10.1093/nar/gkt1223>.
  16. Guzik U, Hupert-Kocurek K, Krysiak M, Wojcieszynska D. 2014. Degradation potential of protocatechuate 3,4-dioxygenase from crude extract of *Stenotrophomonas maltophilia* strain kb2 immobilized in calcium alginate hydrogels and on glyoxyl agarose. *BioMed Res Int* 2014:138768. <http://dx.doi.org/10.1155/2014/138768>.
  17. Ma F, Shi SN, Sun TH, Li A, Zhou JT, Qu YY. 2013. Biotransformation of benzene and toluene to catechols by phenol hydroxylase from *Arthrobacter* sp. W1. *Appl Microbiol Biotechnol* 97:5097–5103. <http://dx.doi.org/10.1007/s00253-012-4301-z>.
  18. Chen K, Huang L, Xu C, Liu X, He J, Zinder SH, Li S, Jiang J. 2013. Molecular characterization of the enzymes involved in the degradation of a brominated aromatic herbicide. *Mol Microbiol* 89:1121–1139. <http://dx.doi.org/10.1111/mmi.12332>.
  19. Li S-s, Zhang D, Yan W. 2014. Enhanced biodegradation of methyl tert-butyl-ether by a microbial consortium. *Curr Microbiol* 68:317–323. <http://dx.doi.org/10.1007/s00284-013-0480-9>.
  20. Hlouchova K, Rudolph J, Pietari JM, Behlen LS, Copley SD. 2012. Pentachlorophenol hydroxylase, a poorly functioning enzyme required for degradation of pentachlorophenol by *Sphingobium chlorophenolicum*. *Biochemistry* 51:3848–3860. <http://dx.doi.org/10.1021/bi300261p>.