

Draft Genome Sequences of Six *Pseudoalteromonas* Strains, P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26, Which Induce Larval Settlement and Metamorphosis in *Hydractinia echinata*

Jonathan L. Klassen,^a Thomas Wolf,^b Maja Rischer,^b Huijuan Guo,^b Ekaterina Shelest,^b Jon Clardy,^c Christine Beemelmans^b

University of Connecticut, Department of Molecular & Cell Biology, Storrs, Connecticut, USA^a; Leibniz Institute for Natural Product Research and Infection Biology eV, Jena, Germany^b; Harvard Medical School, Department of Biological Chemistry and Molecular Pharmacology, Boston, Massachusetts, USA^c

To gain a broader understanding of the importance of a surface-associated lifestyle and morphogenic capability, we have assembled and annotated the genome sequences of *Pseudoalteromonas* strains P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26, isolated from *Hydractinia echinata*. These genomes will allow detailed studies on bacterial factors mediating interkingdom communication.

Received 27 October 2015 Accepted 30 October 2015 Published 17 December 2015

Citation Klassen JL, Wolf T, Rischer M, Guo H, Shelest E, Clardy J, Beemelmans C. 2015. Draft genome sequences of six *Pseudoalteromonas* strains, P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26, which induce larval settlement and metamorphosis in *Hydractinia echinata*. *Genome Announc* 3(6):e01477-15. doi:10.1128/genomeA.01477-15.

Copyright © 2015 Klassen et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to Christine Beemelmans, christine.beemelmans@hki-jena.de.

Pseudoalteromonas strains P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26 were isolated from the tissue of a feeding polyp of the marine hydroid *Hydractinia echinata* (1) purchased from the Marine Biological Laboratory in Woods Hole, MA, USA. *Pseudoalteromonas* are commonly isolated from biofilms of marine surfaces and host tissue of marine invertebrates (2, 3). Their effects on the settlement and metamorphosis of biofouling invertebrates (4–6) and the production of pharmacologically active compounds (7) have been extensively studied. Six *Pseudoalteromonas* strains were isolated from *H. echinata* and screened for their effects on its larval settlement and metamorphosis using a colony-based assay (1). Genomes from the most inductive strains P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26 were sequenced to identify candidate genes responsible for larval settlement. Genomic DNA was extracted using the GenElute Blood Genomic DNA kit (Sigma-Aldrich) according to the manufacturer's protocol. Sequencing performed at the Harvard Medical School Biopolymers Facility used Illumina TruSeq 50 bp single-read libraries and a HiSeq2000 instrument (Illumina CASAVA 1.8.2). After subsampling reads to achieve ~50× coverage, genomes were assembled using the A5 pipeline v20120518 (8) and screened for contamination using blobology (9). Genomes were annotated using Prokka v1.10 (10) and assembly statistics were calculated using scripts from the Assemblathon2 project (11).

The draft genome sequence of strain P1-7a was sequenced to 52× coverage, and comprises 189 contigs totaling 4,374,565 bases in length and having a G+C content of 40.8%. Its annotation includes 3,853 coding sequences (CDSs), 96 tRNAs, and 4 rRNAs.

The draft genome of strain P1-9 was sequenced to 47× coverage, and comprises 211 contigs totaling 4,808,111 bases in length and having a G+C content of 40.7%. Its annotation includes 4,321 CDSs, 84 tRNAs, and 3 rRNAs.

The draft genome sequence of strain P1-13-1a was sequenced

to 51× coverage, and comprises 174 contigs totaling 4,442,776 bases in length and having a G+C content of 40.7%. Its annotation includes 3,930 CDSs, 93 tRNAs, and 3 rRNAs.

The draft genome of strain P1-16-1b was sequenced to 57× coverage, and comprises 90 contigs totaling 3,977,637 bases in length and having a G+C content of 40.1%. Its annotation includes 3,562 CDSs, 90 tRNAs, and 4 rRNAs.

The draft genome sequence of strain P1-25 was sequenced to 51× coverage, and comprises 163 contigs totaling 4,399,610 bases in length and having a G+C content of 40.7%. Its annotation includes 3,855 CDSs, 97 tRNAs, and 3 rRNAs.

The draft genome sequence of strain P1-26 was sequenced to 48× coverage, and comprises 219 contigs totaling 4,715,935 bases in length and having a G+C content of 41.2%. Its annotation includes 4,183 CDSs, 96 tRNAs, and 4 rRNAs.

Genes associated with secretion (e.g., type II secretion system), biofilm formation (e.g., curli, extracellular polymers) (12), secondary metabolite production (e.g., NRPS), siderophore (e.g., desferrioxamine) (13, 14), and bacteriocin biosynthesis were detected in all genomes indicating the successful adaptation to persistence and competition on marine surfaces. These genome sequences will help elucidate the mechanisms involved in *H. echinata* settlement and metamorphosis (1), and help identify novel biotechnologically important molecules.

Nucleotide sequence accession numbers. These whole-genome shotgun projects for strains P1-7a, P1-9, P1-13-1a, P1-16-1b, P1-25, and P1-26 have been deposited in DDBJ/EMBL/GenBank under the accession numbers [LKDU000000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDU000000000), [LKDV000000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDV000000000), [LKGQ000000000](https://www.ncbi.nlm.nih.gov/nuccore/LKGQ000000000), [LKDW000000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDW000000000), and [LKDX000000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDX000000000), respectively. The versions described in this paper are the first versions, [LKDU010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDU010000000), [LKBD010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKBD010000000), [LKDV010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDV010000000), [LKGQ010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKGQ010000000), [LKDW010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDW010000000), and [LKDX010000000](https://www.ncbi.nlm.nih.gov/nuccore/LKDX010000000).

ACKNOWLEDGMENTS

We are grateful for financial support from the NIH to J.C. (GM086258), and the German National Academy of Sciences Leopoldina for a postdoctoral fellowship to C.B. (LPDS 2011-2). J.L.K. was supported by funds from the University of Connecticut. M.R. was supported by the graduate school Jena School for Microbial Communication (JSMC) financed by the Deutsche Forschungsgemeinschaft, and T.W. was supported by the International Leibniz Research School for Microbial and Molecular Interactions (ILRS), as part of the JSMC.

REFERENCES

- Frank U, Leitz T, Müller WA. 2001. The hydroid *Hydractinia*: a versatile, informative cnidarian representative. *Bioessays* 23:963–971. <http://dx.doi.org/10.1002/bies.1137>.
- Gauthier G, Gauthier M, Christen R. 1995. Phylogenetic analysis of the genera *Alteromonas*, *Shewanella*, and *Moritella* using genes coding for small-subunit rRNA sequences and division of the genus *Alteromonas* into two genera, *Alteromonas* (emended) and *Pseudoalteromonas* gen. nov., and proposal of twelve new species combinations. *Int J Syst Bacteriol* 45:755–761. <http://dx.doi.org/10.1099/00207713-45-4-755>.
- Holmstrom C, Kjelleberg S. 1999. Marine *Pseudoalteromonas* species are associated with higher organisms and produce biologically active extracellular agents. *FEMS Microbiol Ecol* 30:285–293. [http://dx.doi.org/10.1016/S0168-6496\(99\)00063-X](http://dx.doi.org/10.1016/S0168-6496(99)00063-X).
- Bowman JP. 2007. Bioactive compound synthetic capacity and ecological significance of marine bacterial genus *Pseudoalteromonas*. *Mar Drugs* 5:220–241. <http://dx.doi.org/10.3390/md504220>.
- Qian P-, Lau SCK, Dahms H-, Dobretsov S, Harder T. 2007. Marine biofilms as mediators of colonization by marine macroorganisms: implications for antifouling and aquaculture. *Mar Biotechnol* 9:399–410. <http://dx.doi.org/10.1007/s10126-007-9001-9>.
- Hadfield MG. 2011. Biofilms and marine invertebrate larvae: what bacteria produce that larvae use to choose settlement sites. *Annu Rev Marine Sci* 3:453–470. <http://dx.doi.org/10.1146/annurev-marine-120709-142753>.
- Machado H, Sonnenschein EC, Melchiorson J, Gram L. 2015. Genome mining reveals unlocked bioactive potential of marine gram-negative bacteria. *BMC Genomics* 16:158–170. <http://dx.doi.org/10.1186/s12864-015-1365-z>.
- Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for *de novo* assembly of microbial genomes. *PLoS One* 7:e42304. <http://dx.doi.org/10.1371/journal.pone.0042304>.
- Kumar S, Jones M, Koutsovoulos G, Clarke M, Blaxter M. 2013. Blobology: exploring raw genome data for contaminants, symbionts and parasites using taxon-annotated GC-coverage plots. *Front Genet* 4:237. <http://dx.doi.org/10.3389/fgene.2013.00237>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <http://dx.doi.org/10.1093/bioinformatics/btu153>.
- Bradnam KR, Fass JN, Alexandrov A, Baranay P, Bechner M, Birol I, Boisvert S, Chapman JA, Chapuis G, Chikhi R, Chitsaz H, Chou W, Corbeil J, Del Fabbro C, Docking T, Durbin R, Earl D, Emrich S, Fedotov P, Fonseca NA, Ganapathy G, Gibbs RA, Gnerre S, Godzaridis E, Goldstein S, Haimel M, Hall G, Haussler D, Hiatt JB, Ho IY, Howard J, Hunt M, Jackman SD, Jaffe DB, Jarvis ED, Jiang H, Kazakov S, Kersey PJ, Kitzman JO, Knight JR, Koren S, Lam T-W, Lavenier D, Laviolette F, Li Y, Li Z, Lio B, Liu Y, Luo R, MacCallum I, MacManes MD, Maillat N, Melnikov S, Naquin D, Ning Z, Otto TD, Paten B, Paulo OS, Phillippy AM, Pina-Martins F, Place M, Przybylski D, Qin X, Qu C, Ribeiro FJ, Richards S, Rokhsar DS, Ruby JG, Scalabrin S, Schatz MC, Schwartz DC, Sergushichev A, Sharpe T, Shaw TI, Shendure J, Shi Y, Simpson JT, Song H, Tsarev F, Vezzi F, Vicedomini R, Vieira BM, Wang J, Worley KC, Yin S, Yiu S-M, Yuan J, Zhang G, Zhang H, Zhou S, Korf IF. 2013. Assemblathon 2: Evaluating *de novo* methods of genome assembly in three vertebrate species. *GigaScience* 2:10. <http://dx.doi.org/10.1186/2047-217X-2-10>.
- Thomas T, Evans FF, Schleheck D, Mai-Prochnow A, Burke C, Penevyan A, Dalisay DS, Stelzer-Braid S, Saunders N, Johnson J, Ferreira S, Kjelleberg S, Egan S. 2008. Analysis of the *Pseudoalteromonas tumidica* genome reveals properties of a surface-associated life style in the Marine environment. *PLoS One* 3:e3252. <http://dx.doi.org/10.1371/journal.pone.0003252>.
- Wolf T, Shelest V, Nath N, Shelest E. 2015. CASSIS and SMIPS—promoter-based prediction of secondary metabolite gene clusters in eukaryotic genomes. *Bioinformatics*, in press.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Brucoleri R, Lee SY, Fischbach MA, Müller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0—a comprehensive resource for the genome mining of biosynthetic gene clusters. *Nucleic Acids Res* 43:W237–W243. <http://dx.doi.org/10.1093/nar/gkv437>.