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SYSTEMIC INFLAMMATION IN XENOGRAFT RECIPIENTS (SIXR): A NEW PARADIGM IN PIG-TO-PRIMATE XENOTRANSPLANTATION?

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Abstract

Inflammation is a complex response that involves interactions between multiple proteins in the human body. The interaction between inflammation and coagulation is well-recognized, but its role in the dysregulation of coagulation in xenograft recipients is not well-understood. Additionally, inflammation is known to prevent the development of T cell tolerance after transplantation.

Recent evidence indicates that systemic inflammation precedes and may be promoting activation of coagulation after pig-to-primate xenotransplantation. Activated recipient innate immune cells expressing tissue factor are increased after xenotransplantation, irrespective of immunosuppressive therapy. With immunosuppression, C-reactive protein (C-RP), fibrinogen, and interleukin-6 levels are significantly increased in pig artery patch recipients. In pig organ recipients, increased C-RP levels are observed prior to the development of features of consumptive coagulopathy. Systemic inflammation in xenograft recipients (SIXR) may be a key factor in the development of dysregulation of coagulation, as well as in resistance to immunosuppressive therapy.

While genetic modification of the donor pigs provides protection against humoral responses and the development of thrombotic microangiopathy, therapeutic prevention of SIXR may be essential in order to prevent systemic dysregulation of coagulation in xenograft recipients without the use of intensive immunosuppression.

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Author contribution

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Keywords

Coagulation; Innate immune cells; Inflammation; Pig; Primate; Xenotransplantation

Introduction

Inflammation is a natural and essential response in the human body to infection, cell stress, and tissue damage. It is frequently associated with the development of pathological conditions as diverse as diabetes, ischemia, and atherosclerosis. At sites of inflammation, a wide range of cytokines and chemokines are produced. Chemokines are required for recruiting innate immune cells to the site of inflammation. Cytokines modulate maturation, growth and responsiveness of various immune cells that are responsible for elimination of pathogens, removal of dead cells, and stimulation of tissue repair.

Following an inflammatory response, effective resolution of inflammation requires the cessation of pro-inflammatory signals, followed by the disappearance of inflammatory cellular infiltrates. However, under certain conditions, inflammation persists. Prolonged (or chronic) inflammatory responses are associated with various pathological conditions, such as rheumatoid arthritis and atherosclerosis. Similarly, intensified inflammatory responses can be associated adverse effects.

In the last two decades, a substantial network of interactions has been revealed between inflammation, the coagulation system, and the innate immune system [1]. Multiple cellular and molecular mechanisms have been identified linking inflammatory responses and homeostasis. The pro-inflammatory roles of coagulation proteins and the role of cytokines in promoting activation of coagulation have been well-characterized. Accordingly, the interaction between inflammation and coagulation results in an amplification circuit [2] promoting the production of both inflammatory mediators as well as coagulation factors.

Dysregulation of the coagulation system is considered a hallmark associated with failure of organ xenografts. Thrombotic microangiopathy (TM) in the graft and/or consumptive coagulopathy (CC) in the recipient are characteristic features associated with xenograft failure or rejection, and are major barriers to prolonged xenograft survival in nonhuman primates [3–6].

The roles of inflammatory signals in promoting T cell activation after allotransplantation have been recognized as key elements in the loss of T cell tolerance and prevention of long-term allograft survival [7], associated with B cell activation and antibody production. Recent studies suggest that the inflammatory response in pig-to-nonhuman primate xenotransplantation may have been underappreciated. Systemic inflammation in xenograft recipients (SIXR) may play a key role in the perpetuation of activation of coagulation (and therefore of coagulation dysregulation), as well as in increased resistance to the effects of immunosuppressive therapy (IS) after xenotransplantation [8].

Role of innate immune cells in inflammation and activation of coagulation

Innate immune cells are considered a barrier to prolonged xenograft survival in nonhuman primates [9,10]. In addition to their role in the process of inflammation and production of pro-inflammatory cytokines, innate immune cells, e.g., dendritic cells, are known to promote activation of coagulation through multiple mechanisms, providing an additional link between inflammation and activation of coagulation [11].

In response to pro-inflammatory signals, activated monocytes and dendritic cells upregulate tissue factor (TF) expression, and potentiate activation of coagulation [11–13]. Activated dendritic cells express thrombin receptors [14], while thrombin is also known to influence dendritic cell functions [15]. Furthermore, activated dendritic cells have been shown to express and release functional TF [13].

Expression of porcine TF by activated endothelial cells is an expected initial mechanism in the development of TM and CC. It is also likely that upregulation of recipient TF expression may further augment systemic activation of coagulation [16], particularly with prolonged xenograft survival. Pig aortic endothelial cells (pAECs) are known to stimulate human CD14⁺ monocytes associated with the production of pro-inflammatory cytokines [17] and the upregulation of TF expression [18]. Inflammatory CD11c⁺ dendritic cells have been shown to be monocyte-derived [19,20]. Our previous studies have shown that macrophages and peripheral blood mononuclear cells upregulate TF expression after pig organ xenotransplantation [10,21]. In pig artery patch recipient baboons, we have observed a significant increase in the percentage of peripheral blood CD14⁺CD11c⁺ cells, in the presence or absence of IS. This was associated with significant upregulation of TF expression on monocytes and dendritic cells in the recipient baboons [8].

While transgenic expression of human coagulation-regulatory proteins in pig xenografts provides protection against the development of TM, it will be essential to prevent long-term activation and upregulation of TF expression by innate immune cells in xenograft recipients in order to prevent dysregulation of coagulation.

Pro-inflammatory cytokines after xenotransplantation

Pro-inflammatory cytokines are essential for protection against infections. However, excessive cytokine production can promote inflammation [22]. Pro-inflammatory cytokines are known to promote activation of coagulation. Tumor necrosis factor-alpha (TNF- α) [23] and interleukin (IL)-6 [24] promote TF expression by various cell types, which can lead to activation of coagulation [23,25]. The potential roles of pro-inflammatory cytokines and chemokines in the development of dysregulation of coagulation in xenograft recipients are not well understood, despite their recognized contribution to xenograft rejection [26–29].

After organ xenotransplantation, IS may not efficiently reduce pro-inflammatory cytokines despite efficient blockade of adaptive immune responses. Detectable levels of interferon gamma (IFN- γ), IL-12 and IL-8 can still be observed after pig organ xenotransplantation in baboons. Significantly high levels of TNF- α , monocyte chemoattractant protein 1 (MCP1), and IL-6 are also detected [8]. In pig artery patch recipient baboons, costimulation-based IS

efficiently reduced pro-inflammatory cytokines except IL-6 [8], which has been linked to both inflammatory and thrombotic complications [30] and promotion of TF expression [31] on innate immune cells [25]. This observation suggests that blockade of IL-6 activity may be beneficial after xenotransplantation

C-reactive protein (C-RP) in xenograft recipients

C-RP is a long-studied acute-phase protein recognized to be produced in humans and nonhuman primates during episodes of inflammation, e.g. infection [32,33]. C-RP has been considered a sensitive, but not specific, marker for graft-related complications after organ allotransplantation in humans [34,35]. Notably, C-RP can also promote TF expression [36].

C-RP levels are increased in pig organ xenograft recipients within days after transplantation [8]. Importantly, this rise in C-RP levels occurred *prior to* signs of activation of coagulation and the development of CC in the recipients (Figure 1). Of note, higher C-RP levels were associated with rapid development of dysregulation of coagulation and earlier failure of kidney xenografts compared to heart xenografts [8]. The variable incidence of systemic complications in organ xenograft recipients may be attributed to organ xenograft heterogeneity and organ-specific vascular gene expression [37]. At the time of euthanasia, C-RP deposition was detected in the heart and kidney xenografts. Furthermore, C-RP-positive immune cells were observed in the native lungs of xenograft recipients, indicating a systemic inflammatory response.

In addition to its pro-inflammatory effects, IL-6 is known to induce C-RP production by hepatocytes [38] and smooth muscle cells [39]. High IL-6 levels are associated with high C-RP levels in humans [40]. In pig artery patch recipient baboons, we have documented a significant positive correlation between IL-6 and C-RP levels, as well as between C-RP and fibrinogen levels in the blood [8]. Both C-RP and fibrinogen are known to be acute-phase reactant proteins produced in response to acute inflammation.

Dysregulation of coagulation does not develop in pig artery patch recipients, where high levels of both C-RP and fibrinogen are maintained. In contrast, while C-RP levels continue to increase in organ xenograft recipients, fibrinogen levels gradually decline in concomitance with the onset of activation of coagulation due to consumption of coagulation factors and the development of CC. Hence, steps that to initially prevent the increase in C-RP and fibrinogen levels in pig organ recipients should be beneficial in preventing dysregulation of coagulation and in promoting long-term xenograft survival.

The effect of IS on the inflammatory responses and activation of coagulation in xenograft recipients

Activation of coagulation and fibrin deposition remain central to pig organ xenograft failure in nonhuman primates. Upregulation of pro-coagulant proteins by pig endothelial cells is thought to be critical for the development of TM in organ xenografts, where induction of a pro-coagulant phenotype by pig endothelial cells can be a result of increased binding of natural and elicited anti-pig antibodies [3]. Prevention of production of elicited antibodies is

thought to be essential for prevention of pig endothelial cell activation and upregulation of pro-coagulant proteins. Accordingly, efficient IS is critical to achieve prolonged xenograft survival, not only through the prevention of the adaptive immune response and elicited anti-pig antibodies, but also through the prevention of consequent activation of coagulation.

While dysregulation of the coagulation system in nonhuman primates is a characteristic feature of xenograft failure, anticoagulation has been shown to be relatively inefficient in prolonging pig kidney xenograft survival in monkeys receiving IS [41]. Furthermore, anticoagulation was shown to be inefficient in promoting pig heart xenograft survival in baboons [42]. These observations indicated that IS - but not anticoagulation - promotes long-term xenograft survival in nonhuman primates. It is important to note that these studies were performed using wild-type pigs expressing human complement-regulatory proteins CD55 (DAF) or CD46, respectively.

Therefore, while it might be expected that anticoagulants would ameliorate dysregulation of coagulation after xenotransplantation, effective IS may be more critical in preventing and/or delaying activation of coagulation. Prevention of T cell responses and depletion of T cells in the blood is associated with reduced pro-inflammatory cytokines [8]. Costimulation-based IS prevented the production of pro-inflammatory cytokines in pig artery patch recipients [8,43]. However, the level of IL-6 in the blood was higher when IS was administered, compared to the level seen in baboons not receiving IS. Similarly, C-RP levels were continuously elevated when IS was administered, but remained low in baboons without IS (Figure 2). Interestingly, there was a significant positive correlation between the levels of IL-6, fibrinogen, and C-RP in those recipients. However, when a similar IS regimen was administered to pig kidney and heart recipients, it did not have a comparable effect (i.e., it did not efficiently reduce the production of pro-inflammatory cytokines) indicating that more intensive IS and/or anti-inflammatory agents may be required to reduce SIXR after organ xenotransplantation.

These observations suggest that SIXR and the effect of IS on the inflammatory response may vary based on the type of xenograft, the extent of the antigenic load, and the intensity of IS. The IS regimens adopted by different groups may have variable effects on the inflammatory milieu in the xenograft recipient. The effect of IS on both pro- and anti-inflammatory cytokine levels in xenograft recipients is not clear and requires further investigation.

Observations reported in humans suggest that IS can on occasion stimulate an inflammatory response [44]. Also, a systemic inflammatory response syndrome in humans is well-described and involves dysregulation of coagulation [45]. In contrast, under certain conditions, prolonged inflammation may lead to *reduced* immune responses [46–48]. The long-term effects of IS on SIXR as well as the effects of SIXR on immune responses are yet to be determined. This may be critical in order to avoid complications associated with prolonged and intensive IS.

Conclusions

The underlying mechanisms of the inflammatory response in xenograft recipients are poorly understood. It is yet to be determined whether the inflammatory response varies dependent on the nature of the xenograft and the type of the administered IS. Recent studies suggest that SIXR may be playing a crucial role in the outcome of organ and cell xenotransplantation. Also, it will be important to determine the effect of SIXR on the adaptive immune response and on activation of coagulation. Defining both pro- and anti-inflammatory mechanisms may be critical to develop effective therapies after xenotransplantation. Clinically-relevant IS may be easier to achieve by also targeting pro-inflammatory factors in xenograft recipients. Anti-inflammatory therapy may enable a reduction in the intensity of the IS required to maintain pig xenograft survival.

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ABBREVIATIONS

| | |
|-------------|---|
| CC | Consumptive coagulopathy |
| C-RP | C-reactive protein |
| IS | immunosuppressive therapy |
| SIXR | systemic inflammation in xenograft recipients |
| TF | tissue factor |
| TM | Thrombotic microangiopathy |

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Highlights

- Systemic inflammation in xenograft recipients (SIXR) precedes activation of coagulation
- Increased inflammation may promote activation of coagulation in xenograft recipients
- Innate immune cells upregulate tissue factor expression after xeno-Tx
- Immunosuppression does not consistently reduce pro-inflammatory cytokines after xeno-Tx
- Prevention of SIXR may be essential to reduce dysregulation of coagulation after xeno-Tx

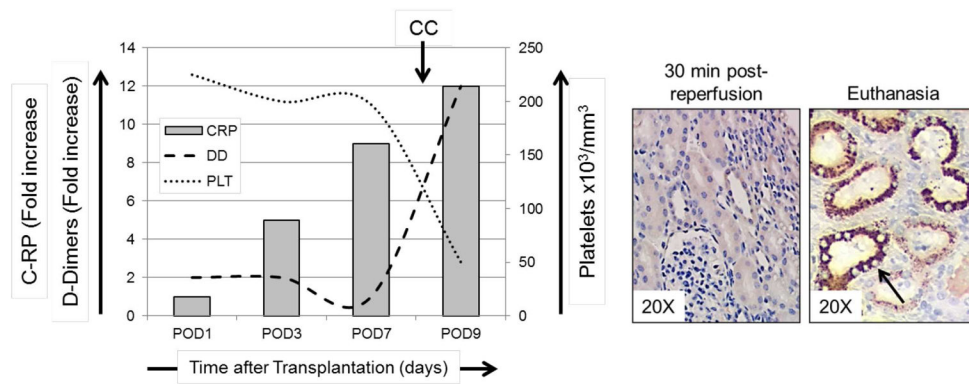


Figure 1. Increased C-RP levels in xenograft recipient baboon and deposition of C-RP in the pig xenografts

Left: Platelet counts, and fold increases in C-RP and D-Dimer were calculated in a pig kidney xenograft recipient. High levels of C-RP were detected as early as 3 days after kidney xenotransplantation, prior to the development of consumptive coagulopathy (CC), as indicated by reduced platelet counts and elevated D-Dimer levels. **Right:** At 30 min after reperfusion, no C-RP was detected, while at the time of euthanasia C-RP deposition was detected in the kidney tubules (arrow).

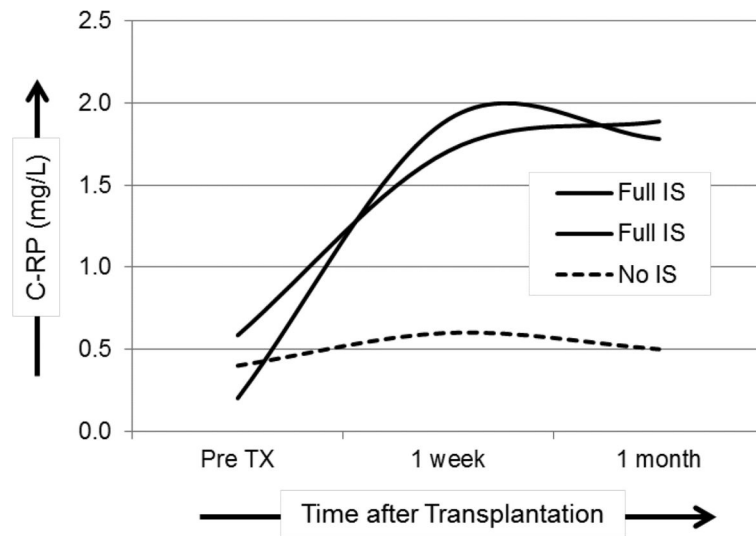


Figure 2. Immunosuppressive therapy (IS) is associated with increased C-RP levels in baboon recipients of pig artery patch grafts

C-RP levels were measured in baboons receiving full IS (n=2) or without IS (n=1), before, and one week and one month after pig artery patch xenotransplantation.