



Published in final edited form as:

*Hypertension*. 2015 September ; 66(3): 582–589. doi:10.1161/HYPERTENSIONAHA.115.05627.

## Profibrotic Role for Interleukin-4 in Cardiac Remodeling and Dysfunction

Hongmei Peng, Zeyd Sarwar, Xiao-Ping Yang, Edward L. Peterson, Jiang Xu, Branislava Janic, Nadia Rhaleb, Oscar A. Carretero, and Nour-Eddine Rhaleb

Hypertension and Vascular Research Division, Department of Internal Medicine (H.P., Z.S., X.-P.Y., J.X., B.J., N.R., O.A.C., N.-E.R.) and Department of Public Health Sciences (E.L.P.), Henry Ford Hospital, Detroit, MI.

### Abstract

Elevated interleukin-4 (IL-4) levels are associated with cardiac fibrosis in hypertension and heart failure in both patients and experimental animals. We hypothesized that chronically elevated IL-4 induces cardiac fibrosis, resulting in a predisposition of the heart to angiotensin II-induced damage. Wild-type Balb/c (WT, high circulating IL-4) and IL-4-deficient Balb/c mice (IL-4<sup>-/-</sup>) were used. WT mice exhibited cardiac fibrosis (evidenced by an increase in expression of procollagen genes/interstitial collagen fraction), enlarged left ventricle chamber, and declined cardiac function associated with a greater number of mast cells and macrophages in the heart compared with IL-4<sup>-/-</sup>. In contrast, IL-4<sup>-/-</sup> mice had normal cardiac architecture/function while showing a 57.9% reduction in heart interstitial collagen compared with WT, despite elevated proinflammatory cytokines in heart tissue. In response to angiotensin II administration, IL-4<sup>-/-</sup> had reduced interstitial myocardial fibrosis and were protected from developing dilated cardiomyopathy, which was seen in WT mice. This was associated with increased macrophage infiltration into the hearts of WT mice, despite a similar degree of hypertension and increased cardiac transforming growth factor- $\beta$ 1 in both groups. In vitro data demonstrated that IL-4 upregulates procollagen genes and stimulates collagen production in mouse cardiac fibroblasts. This process is mediated by signal transducer and activator of transcription 6 signaling pathway via IL-4 receptor alpha. This study not only establishes a causal relationship between IL-4 and cardiac fibrosis/dysfunction, but also reveals a critical role for IL-4 in angiotensin II-induced cardiac damage. IL-4 could serve as an additional target for the treatment of cardiac fibrosis.

### Keywords

angiotensin II; balb/c mice; fibrosis; heart; interleukin-4

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Correspondence to Hongmei Peng, Hypertension and Vascular Research Division, Henry Ford Hospital, 2799 West Grand Blvd, Detroit, MI 48202. hpeng1@hfhs.org or Nour-Eddine Rhaleb, Hypertension and Vascular Research Division, Henry Ford Hospital, 2799 West Grand Blvd, Detroit, MI 48202. nrhaleb1@hfhs.org.

The online-only Data Supplement is available with this article at <http://hyper.ahajournals.org/lookup/suppl/doi:10.1161/HYPERTENSIONAHA.115.05627/-/DC1>

Disclosures  
None.

Progressive cardiac fibrosis is a major maladaptive response to hemodynamic stress and various profibrotic stimuli. It leads to a reduction in myocardial compliance and eventually to cardiac failure.<sup>1</sup> Cytokine interleukin (IL)-4 has been implicated in this process.<sup>2-5</sup>

IL-4 is produced by immune cells, including CD4<sup>+</sup> T-helper (Th) lymphocytes<sup>6</sup> and mast cells.<sup>7</sup> Among its multiple biological effects, IL-4 promotes tissue fibrotic remodeling in diseases involving lung,<sup>8</sup> skin,<sup>9</sup> and liver.<sup>10</sup> Recent studies have shown a positive correlation between systemic IL-4 levels and cardiac fibrotic remodeling in both patients<sup>2,3</sup> and experimental animals.<sup>4,5</sup> Furthermore, previous studies have demonstrated that Balb/c mice, which are characterized by high levels of circulating IL-4, exhibited increased cardiac collagen deposition, left ventricular enlargement, and depressed cardiac function (fibrotic cardiomyopathy).<sup>11,12</sup> When these mice were challenged with angiotensin II (Ang II), severe fibrosis and dilated cardiomyopathy (DCM) developed.<sup>12</sup> Although these observations demonstrate a putative profibrotic role for IL-4 in the heart, the direct evidence linking high levels of IL-4 to cardiac fibrosis is lacking.

The role of IL-4 in cardiac fibrosis has recently been determined in a loss-of-function study showing that administration of an anti-IL-4 neutralizing antibody significantly blunted cardiac fibrotic remodeling in C57BL/6 mice with aortic coarctation.<sup>13</sup> This study established a causal relationship between IL-4 and cardiac fibrosis in hypertension. However, it remains unknown whether (1) chronically elevated IL-4, as seen in primary hypertension<sup>5</sup> and aging,<sup>4</sup> is sufficient to initiate fibrotic response, leading to cardiac fibrosis and dysfunction and (2) stress challenges, such as Ang II–induced hypertension, exacerbate IL-4–induced fibrotic cardiomyopathy, leading to heart failure.

In this report, we used wild-type (WT) and IL-4<sup>-/-</sup> Balb/c mice to delineate the functional significance of chronic elevations in IL-4 levels during the development of cardiac fibrosis and dysfunction, further clarifying their causal relationship. We also demonstrated that IL-4 represents a critical determinant of progression to DCM (evidenced by severe cardiac fibrosis, dilated left ventricular chamber, and declined cardiac function) in Ang II–induced hypertension. Furthermore, we examined the effects of IL-4 on cardiac fibroblasts, the primary cellular source of collagens in the heart, and found that IL-4 not only significantly increased procollagen mRNA but also promoted collagen production through the signal transducer and activator of transcription 6 (STAT6) signaling pathway via IL-4 receptor alpha (IL-4R $\alpha$ ).

## Methods

Detailed description of methods is available in the online-only Data Supplement.

## Results

### Systolic Blood Pressure, Cardiac Remodeling, and Function

Basal systolic blood pressure (SBP) was similar between WT and IL-4<sup>-/-</sup> mice (Figure 1A). Ang II administration resulted in a significant increase in SBP within 2 weeks of exposure to Ang II and stayed elevated until the end of the experiment (8 weeks). There were no

differences in the degree of induced hypertension between the groups at any time point measured (Figure 1A). Ang II also induced comparable cardiac hypertrophy in WT and IL-4<sup>-/-</sup> mice as indicated by significantly increased left ventricle (LV) weights after 8 weeks of Ang II infusion (Figure 1B).

Cardiac geometry and function are shown in Figures 1C–1F. At 16 to 20 weeks of age, WT mice showed significantly enlarged LV chamber and declined cardiac function compared with IL-4<sup>-/-</sup>. WT mice exhibited increased LV weight, severe LV chamber dilatation, wall thinning, and dramatically decreased ejection fraction and shortening fraction after 8 weeks of Ang II administration. In contrast, IL-4<sup>-/-</sup> mice had significantly increased LV weight to tibia length ratio and diastolic posterior wall thickness in response to Ang II and increased SBP. In Ang II–treated IL-4<sup>-/-</sup> mice, ejection fraction and shortening fraction were decreased compared with vehicle-treated mice.

### Cardiac Fibrotic Remodeling

As detected by immunoblotting, IL-4 was expressed in the hearts of WT but was absent in IL-4<sup>-/-</sup> mice (Figure 2A). The fibrosis shown in WT mice with Ang II–induced hypertension was characterized as a dense crisscrossing meshwork of collagen fibers encircling muscle fibers and accumulated collagen extending outward from perivascular/pericapillary space (Figure 2B). Cardiac fibrosis seen in WT mice showed a significantly higher interstitial collagen fraction in the LV myocardium when compared with mice with genetic IL-4 deletion (Figure 2C). IL-4 deletion attenuated cardiac fibrosis by 57.9% (3.85% ± 0.53% in WT versus 1.62% ± 0.22% in IL-4<sup>-/-</sup>,  $P < 0.005$ ). Ang II infusion for 8 weeks resulted in a significant increase in interstitial collagen fraction in both WT and IL-4<sup>-/-</sup> mice with significantly higher interstitial collagen fraction in WT compared with IL-4<sup>-/-</sup> mice (Figure 2C). In addition, significantly higher mRNA levels of procollagen type-I alpha 1 (Col1 $\alpha$ 1) and procollagen type-III alpha 1 (Col3 $\alpha$ 1) were found in the hearts of WT mice compared with that in IL-4<sup>-/-</sup> (Figure 2D and 2E). Ang II induced significant upregulation of these 2 procollagen genes in the hearts of both WT and IL-4<sup>-/-</sup> mice with significantly higher mRNA level of Col3 $\alpha$ 1 in WT+Ang II than in the IL-4<sup>-/-</sup> mice+Ang II group (Figure 2D and 2E).

### Mast Cells and CD68<sup>+</sup> Macrophages in the Heart

Mast cells were primarily observed surrounding the vessels and the pericardium (Figure 3A). A more than 2-fold increase in total mast cells was seen in the hearts of WT mice compared with IL-4<sup>-/-</sup>; however, Ang II infusion for 8 weeks did not change the numbers of mast cells in the hearts (Figure 3B). A similar pattern was observed with CD68<sup>+</sup> macrophages in the hearts of vehicle-treated WT and IL-4<sup>-/-</sup> mice. Chronic Ang II administration significantly increased the numbers of macrophages in the hearts of WT but not in IL-4<sup>-/-</sup> mice (Figure 3C and 3D). Noticeable accumulation of macrophages was also seen around the vessels.

### Cardiac Cytokines and Monocyte Chemoattractant Protein-1 in the Heart

There were no significant differences in cardiac transforming growth factor- $\beta$ 1 (TGF- $\beta$ 1) levels between WT and IL-4<sup>-/-</sup> mice. Ang II administration for 8 weeks significantly

increased TGF- $\beta$ 1 levels in the hearts of both groups (Figure 4A). Values of IL-4 and IL-5 in the mouse hearts were below the lowest levels of detection. Interferon gamma (IFN $\gamma$ ) in the hearts of WT mice was too low to detect with the used kit; however, IL-4 deletion induced robust IFN $\gamma$  production (Figure 4B). Significant increases in IL-10, IL-1 $\beta$ , IL-2, and tumor necrosis factor- $\alpha$  were observed in the hearts of IL-4<sup>-/-</sup> mice compared with WT mice (Figure 4C–4F). Interestingly, chronic Ang II administration for 4 weeks resulted in a significant reduction in cardiac IL-1 $\beta$ , IL-2, and tumor necrosis factor- $\alpha$  only in IL-4-deficient mice. Cardiac monocyte chemoattractant protein-1 (MCP-1) levels in WT mice were >2 $\times$  greater than those in IL-4<sup>-/-</sup> mice. Ang II infusion for 4 weeks induced increases in MCP-1 in the hearts, which was significant in WT mice (Figure 4G).

### IL-4 Signaling and Collagen Production in Cardiac Fibroblasts

Immunocytochemistry results showed constitutive expression of functional IL-4R $\alpha$  on the cell-surface of mouse cardiac fibroblasts (Figure 5A). Western blot analysis revealed that IL-4R $\alpha$  expression was doubled after cells were incubated with IL-4 for 24 hours in a STAT6-dependent fashion (Figure 5B). IL-4 rapidly induced phosphorylation of STAT6 (within minutes), with the phosphorylation lasting >24 hours (Figure 5C). Importantly, IL-4 markedly increased collagen production (Figure 5D) and significantly induced Col1 $\alpha$ 1 and Col3 $\alpha$ 1 mRNA expression by cardiac fibroblasts (Figure 5E and 5F); these effects were STAT6-dependent.

## Discussion

This is the first study to establish a causal relationship between chronically elevated IL-4 and cardiac fibrotic remodeling and dysfunction. Not only does high IL-4 induce LV fibrosis and dysfunction, but its deletion markedly attenuates this effect as demonstrated in Balb/c hearts and IL-4<sup>-/-</sup> Balb/c hearts, respectively. The present study also showed that in contrast to WT, IL-4<sup>-/-</sup> mice were protected from fibrotic lesions and did not develop DCM in Ang II-induced hypertension.

The role for IL-4 in the regulation of blood pressure has not been adequately addressed previously. We did not observe significant differences in SBP between WT and IL-4<sup>-/-</sup> mice at both basal condition and Ang II-induced hypertension. This finding is in line with a previous study showing that systemic administration of an anti-IL-4 neutralizing antibody did not affect the increased blood pressure in C57BL/6 mice with aortic coarctation.<sup>13</sup> Vascular endothelial cells express IL-4R $\alpha$  and respond to IL-4.<sup>14</sup> In the lungs, IL-4 activates pulmonary endothelial cells and induces endothelin-1 production, resulting in the development of pulmonary hypertension.<sup>15</sup> Therefore, we cannot exclude the possibility that IL-4 may play a role in the development of hypertension over time, especially under the conditions combined with other activated signaling pathways, such as Ang II signaling.

An increase in content or a transformed structure of cardiac fibrillar collagen can impede muscular contraction and relaxation.<sup>16</sup> Cardiac fibrosis may cause systolic dysfunction through several distinct mechanisms, including impaired force generation by myocytes,<sup>17</sup> disrupted normal coordination of myocardial excitation–contraction coupling,<sup>18</sup> and an asynchronous contraction of the myocardium.<sup>19</sup> These proposed mechanisms may be

involved in cardiac fibrosis–induced decline in the cardiac function observed in Balb/c mice. The fact that this decline in cardiac function disappeared when cardiac fibrosis was absent in IL-4<sup>-/-</sup> further supports the notion of cardiac fibrosis–induced systolic dysfunction.

In the present study, we found positive correlation between IL-4 and Col1 $\alpha$ 1/Col3 $\alpha$ 1 gene expression/collagen production in the hearts of mice, as well as in cultured cardiac fibroblasts. In cultured cells, upregulation of Col1 $\alpha$ 1/Col3 $\alpha$ 1 gene was mediated through STAT6 signaling via IL-4R $\alpha$ . These findings are consistent with the results of *in vivo*<sup>20</sup> and *in vitro*<sup>21–23</sup> studies using noncardiac tissue and fibroblasts, respectively. The important finding of our study was an increase in IL-4R $\alpha$  expression on cardiac fibroblasts in response to IL-4, which was mediated via STAT6 signaling. This phenomenon was also observed in T and B cells from Balb/c mice.<sup>24</sup> Increased IL-4 receptor expression by IL-4 provides an important mechanism for amplification of IL-4–dependent activation of STAT6 signaling in cardiac fibroblasts. Therefore, IL-4–induced collagen production in fibroblasts significantly contributes to cardiac fibrosis and dysfunction.

We observed that IL-4 was associated with an increase in the number of mast cells in the heart. This finding could have been because of IL-4–induced mast cell proliferation because mast cells express IL-4R $\alpha$  *in vivo*.<sup>25</sup> In disease states, mast cells can be activated by cytokines, such as IL-4, leading to either secretion of mediators by degranulation or release of distinct mediators without overt degranulation.<sup>13,26</sup> The positive association between total mast cells and cardiac fibrosis shown in our study is consistent with reported results from clinical studies<sup>27</sup> and in experimental animal models.<sup>28</sup> Notably, a study using mast cell–deficient mice<sup>29</sup> confirms a profibrotic role for mast cells in the evolution of congestive heart failure. Here, we provide evidence for the first time that mast cells, which are known to secrete IL-4,<sup>30</sup> might participate in mediating IL-4–induced fibrotic cardiomyopathy. Thus, IL-4 produced by mast cell may function in an autocrine manner, leading to further mast cell proliferation and IL-4 production. We propose that mast cells could be an important cellular source of cardiac IL-4, which is significantly elevated in Balb/c mice.

Our data indicate that IL-4 contributes to an increased number of macrophages in the fibrotic heart. This finding may be as a result of IL-4–induced upregulation of MCP-1 expression because deletion of IL-4 was associated with a markedly decreased number of macrophages and MCP-1 in the myocardium. The mechanisms underlying IL-4–induced accumulation of macrophages in the heart could be because of (1) IL-4–induced recruitment of the cells via upregulation of expressions of adhesion molecules and MCP-1 in endothelial cells<sup>31–33</sup> and (2) IL-4–stimulated proliferation of resident and recruited macrophages.<sup>34</sup> In an IL-4 rich environment, macrophages polarize toward an M2 phenotype (alternative activation),<sup>35</sup> which has been shown to significantly contribute to cardiac fibrosis.<sup>36</sup>

Although it has been reported that IL-4 might mediate fibrosis in part by increasing the expression of the TGF- $\beta$  mRNA in fibroblasts,<sup>37</sup> we observed neither a significant difference in TGF- $\beta$ 1 levels between WT and IL-4–deficient mouse hearts nor an induction of TGF- $\beta$ 1 production in cultured rat cardiac fibroblasts stimulated with IL-4 (Figure S3 in the online-only Data Supplement). Our data are consistent with reported findings showing that IL-4–induced cardiac fibrosis in hypertension was independent of TGF- $\beta$ 1.<sup>13</sup> IL-4 not

only promotes Th2 differentiation, but at the same time suppresses Th1 differentiation.<sup>38</sup> Not surprisingly, genetic deletion of IL-4 in Balb/c mice results in the skew from Th2 differentiation to Th1 as evidenced by markedly increased IFN $\gamma$  in the myocardium of IL-4<sup>-/-</sup> mice. Our finding that cardiac fibrosis was abrogated along with a marked induction of IFN $\gamma$  and IL-10 in IL-4-deficient Balb/c mice is consistent with reports indicating antifibrotic role of IFN $\gamma$  in the heart<sup>39</sup> and IL-10's efficacy in the treatment of fibrosis in disease models.<sup>40,41</sup>

Enhanced Th1 responses, as a result of a striking induction of IFN $\gamma$ , were observed in the hearts of IL-4<sup>-/-</sup> mice as evidenced by significantly increased cardiac IL-1 $\beta$ , IL-2, and tumor necrosis factor- $\alpha$ . These cytokines have been shown to have negative inotropic effects on cardiac myocytes.<sup>42,43</sup> Despite the inhibitory effects of these cytokines on cardiomyocytes, cardiac functions in IL-4<sup>-/-</sup> mice were preserved up to at least 16 to 20 weeks of age. However, it is possible that cardiac function of IL-4<sup>-/-</sup> mice could gradually decline with age.

Ang II significantly increased TGF- $\beta$ 1 levels in the hearts of both WT and IL-4<sup>-/-</sup> mice. In fibroblasts, TGF- $\beta$ 1 induces a sustained increase in Col1 $\alpha$ 1 mRNA expression and collagen type-I secretion, but does not do the same with Col3 mRNA and collagen type-III.<sup>44,45</sup> This phenomenon may explain why only Col3 $\alpha$ 1 mRNA levels, but not Col1 $\alpha$ 1, were significantly different between Ang II-treated WT and IL-4<sup>-/-</sup> mice in our study. This finding stems from the fact that significant increases in Col1 $\alpha$ 1 mRNA in both WT and IL-4<sup>-/-</sup> mice diminish the difference in Col1 $\alpha$ 1 mRNA expression induced by IL-4. TGF- $\beta$ 1 also induces other profibrotic factors, including connective tissue growth factor,<sup>44</sup> which might upregulate both Col1 $\alpha$ 1 and Col3 $\alpha$ 1 mRNAs in both WT and IL-4<sup>-/-</sup> mice treated with Ang II. This results in significantly higher Col1 $\alpha$ 1/Col3 $\alpha$ 1 mRNA levels compared with its vehicle-treated group. An interesting finding of the current study was the reduction of IL-1 $\beta$ , IL-2, and tumor necrosis factor- $\alpha$  in the hearts of IL-4<sup>-/-</sup> mice treated with Ang II. The mechanisms by which these cytokines were reduced in mice with Ang II exposure are unknown. Recent studies have provided evidence of an important role for MCP-1 in the recruitment of bone marrow-derived fibroblast precursors in Ang II-induced cardiac fibrosis.<sup>46,47</sup> In addition, it has been shown that IL-4 induces MCP-1 expression in endothelial cells.<sup>33,48</sup> The cardiac MCP-1 levels in our WT mice were significantly higher than that in IL-4<sup>-/-</sup>. Therefore, robust MCP-1 induction by IL-4 and Ang II results in a strong recruitment of monocytes (then M2 macrophage differentiation) and bone marrow-derived fibroblast precursors. This may explain the dense collagen extending outward from the perivascular/pericapillary space to the myocardium in our Ang II-treated WT mice. The pathophysiologic effects of Ang II administration over imbalanced Th1/Th2 cytokine expression on the heart are complex and involve many variables. Our data suggest that in the presence of high levels of IL-4 and Ang II-induced high systolic afterload and stimulation, cardiac fibrosis is a major cause for impaired cardiac function in mice. This could be the outcomes of synergic effects of IL-4 and Ang II on cardiac resident cells and infiltrated immune cells.



## Perspectives

The present study establishes the causal relationship between chronically elevated IL-4 and fibrotic cardiomyopathy by using WT and IL-4<sup>-/-</sup> Balb/c mice. In addition, by infusing Ang II, we develop a hypertension-induced DCM mouse model in Balb/c mice. This is likely the result of IL-4 accumulation over time because IL-4-deficient mice were protected from exacerbated fibrotic cardiac remodeling and DCM. Our in vitro study proposes a mechanism by which IL-4-enhanced collagen production in cardiac fibroblasts is mediated through the STAT6 signaling pathway. Of note, fibrogenesis has been recognized as a major cause of morbidity and mortality in many chronic diseases,<sup>49</sup> but for which no specific therapies are available yet. The discovery of IL-4 upregulating fibrogenesis would be of great clinical interest and would allow IL-4 to serve as a potential target for future therapeutic intervention.

## Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

## Acknowledgments

We thank Gulser Gurocak for her excellent technical assistance.

### Sources of Funding

This study was supported by American Heart Association 14GRNT20460148 (H. Peng) and Henry Ford Hospital Institutional Fund (N.E. Rhaleb).

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## Novelty and Significance

### What Is New?

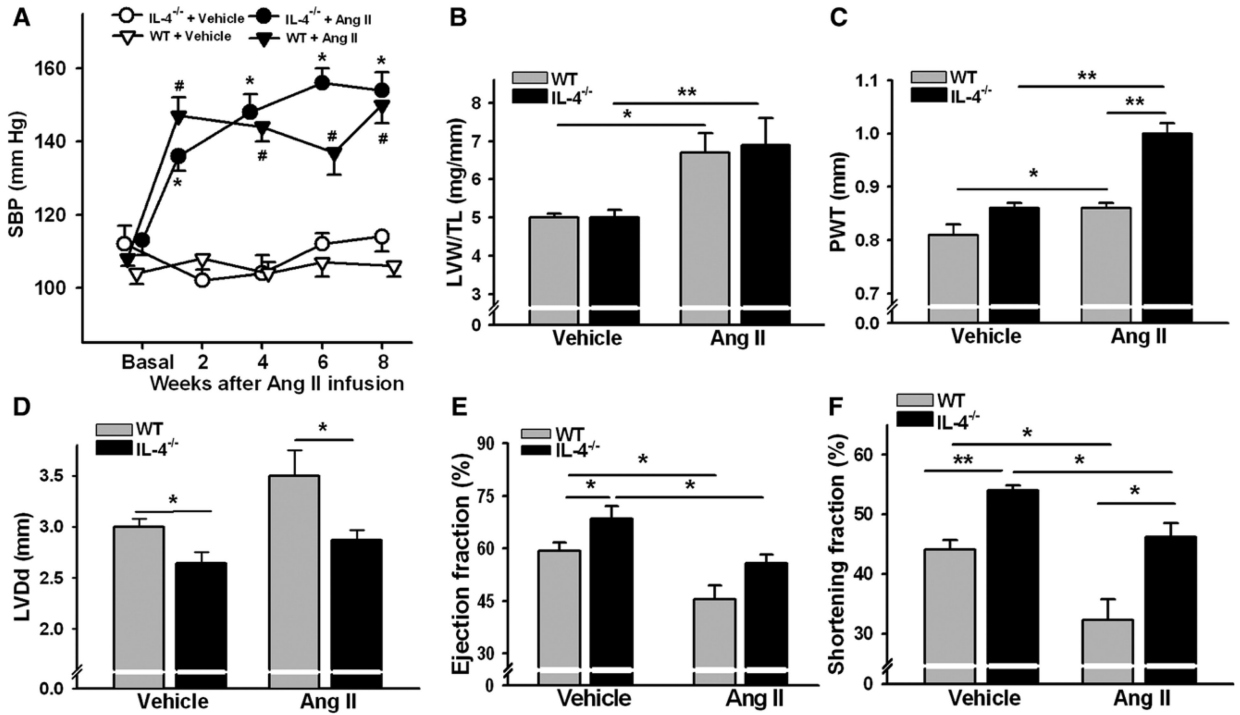
- This study shows that chronically elevated interleukin-4, a cytokine produced by immune cells, is sufficient to induce cardiac fibrosis/dysfunction, resulting in the heart being susceptible to angiotensin II–induced cardiac damage.

### What Is Relevant?

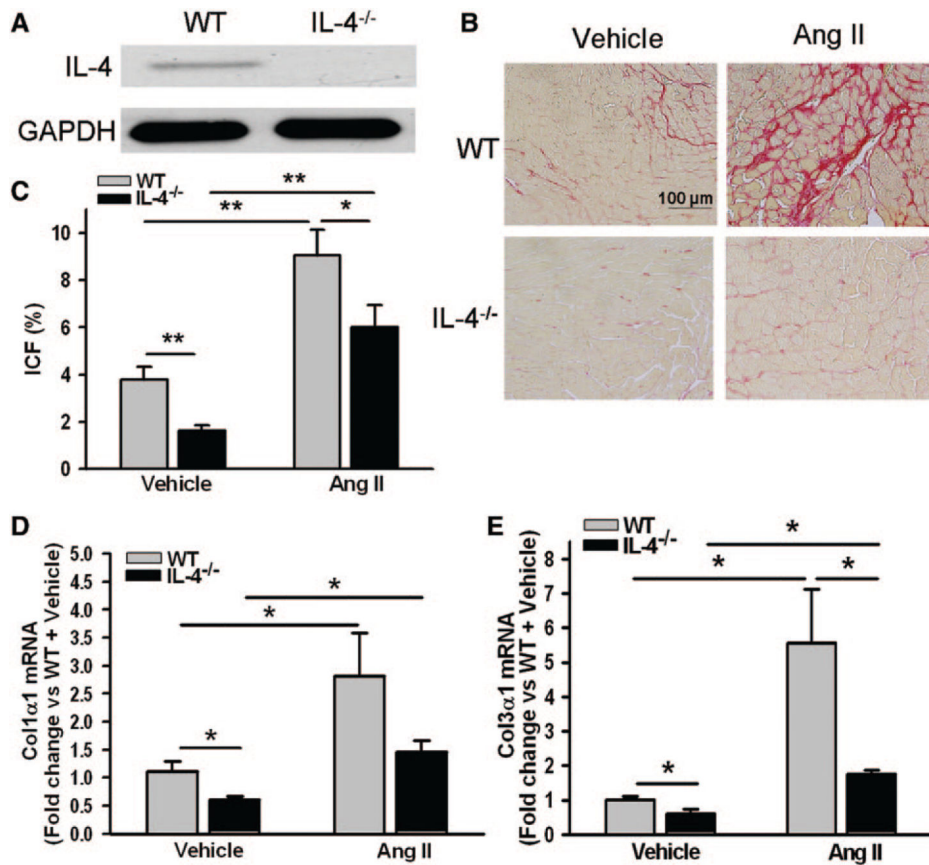
- Cardiac fibrosis represents a critical component of chronic heart diseases and hypertension-related end-organ damage.
- Fibrogenesis represents a common pathophysiological change in many chronic diseases, becoming a major cause of morbidity and mortality, but no specific therapies are currently available to halt or reverse fibrosis.

### Summary

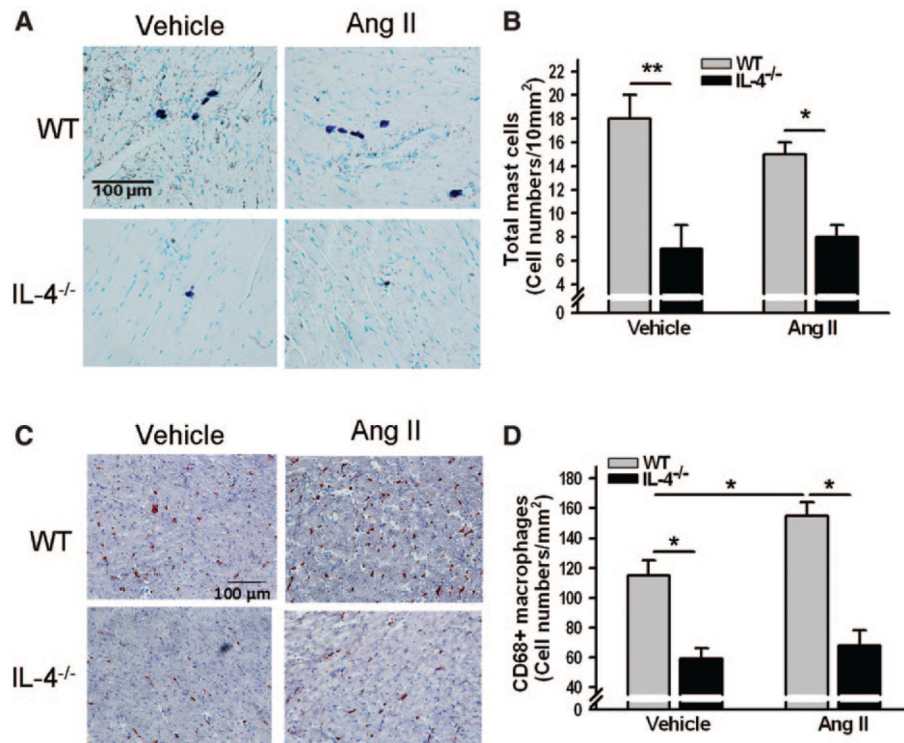
Our findings reveal that interleukin-4 is a key profibrotic factor in the heart, which implicates interleukin-4 as an additional diagnostic test for the diseases and a promising new therapeutic target.



**Figure 1.** Systolic blood pressure (SBP) and cardiac remodeling in wild-type (WT) and interleukin-4 (IL-4)<sup>-/-</sup> mice at the steady-state condition and angiotensin (Ang) II–induced hypertension. **A**, SBP data. Results represent mean±SEM. #*P*<0.005 vs WT+vehicle; \**P*<0.005 vs IL-4<sup>-/-</sup>+vehicle, a Student's *t* test with a Hochberg correction for multiple testing. Cardiac hypertrophy/remodeling and function assessed by left ventricle weight (LVW) to tibia length (TL) ratio (**B**) or echocardiography as the sum of diastolic posterior wall thickness (PWT; **C**), left ventricular diastolic dimension (LVDd; **D**), ejection fraction (**E**), and shortening fraction (**F**) after 8 weeks of Ang II treatment in WT and IL-4<sup>-/-</sup> mice. The bars represent mean±SEM. \**P*<0.05, \*\**P*<0.005, a 2-sample 2-sided Wilcoxon test with a Hochberg correction for multiple testing, n=6 to 14 per group.

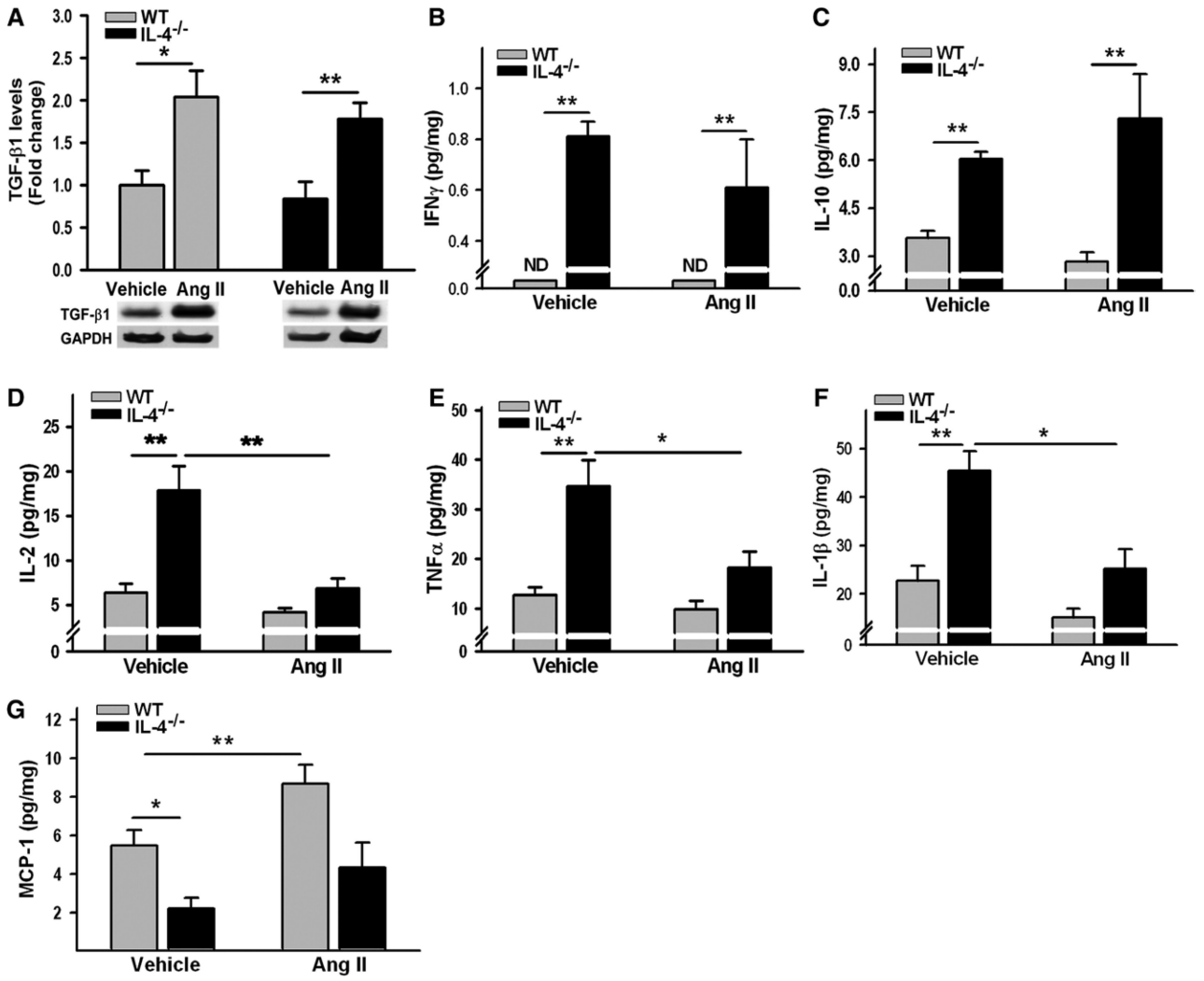


**Figure 2.** Cardiac fibrotic remodeling in the hearts of wild-type (WT) and interleukin-4 (IL-4)<sup>-/-</sup> mice. **A**, Left ventricle lysates were analyzed for IL-4 by immunoblotting with an antibody against mouse IL-4. Representative images of interstitial fibrillar collagen (red) in the hearts of mice treated with either vehicle or angiotensin (Ang II) for 8 weeks (**B**) and quantification of interstitial collagen fraction (ICF) of the study animals (**C**). Procollagen type-I alpha 1 (Col1α1; **D**) mRNA and procollagen type-III alpha 1 (Col3α1; **E**) mRNA in the myocardium of mice treated with either vehicle or Ang II for 8 weeks. The bars represent mean±SEM, \**P*<0.05 and \*\**P*<0.005, a 2-sample 2-sided Wilcoxon test with a Hochberg correction for multiple testing, n=6 to 14 per group.



**Figure 3.** Mast cells and CD68<sup>+</sup> macrophages in the hearts of wild-type (WT) and interleukin-4 (IL-4)<sup>-/-</sup> mice. Representative images of mast cells (purple; **A**) and CD68<sup>+</sup> macrophages (red-brown; **C**) in the myocardium of mice treated with either vehicle or angiotensin (Ang) II for 8 weeks and quantification of the numbers of mast cells (**B**) and CD68<sup>+</sup> macrophages (**D**). The bars represent mean±SEM, \* $P < 0.05$ , \*\* $P < 0.005$ , a 2-sample 2-sided Wilcoxon test with a Hochberg correction for multiple testing,  $n = 5$  to 9 per group.





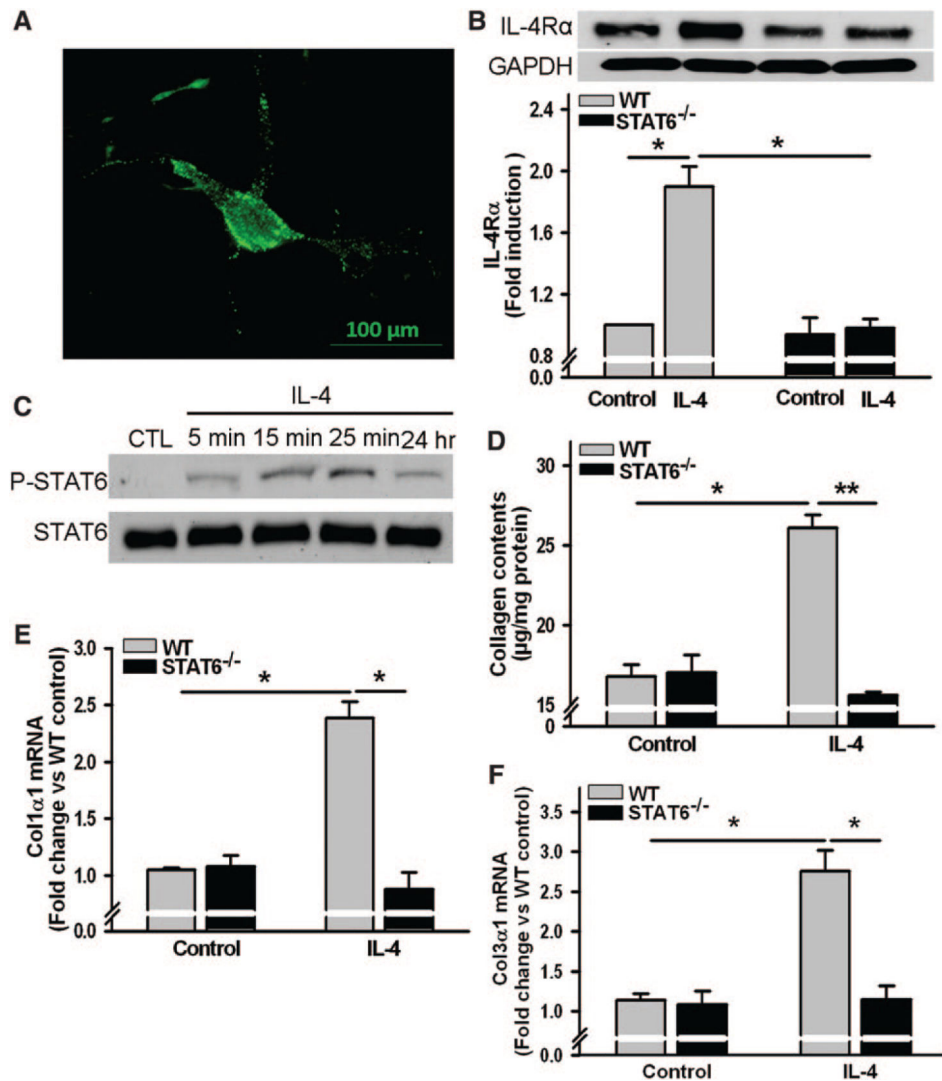
**Figure 4.** Cytokine and monocyte chemoattractant protein-1 (MCP-1) expression in the hearts of wild-type (WT) and interleukin (IL)-4<sup>-/-</sup> mice. Left ventricle lysates from WT and IL-4<sup>-/-</sup> mice treated with either vehicle or angiotensin (Ang) II were processed by Western blot for transforming growth factor-β1 (TGF-β1; **A**) by a cytokine bead array for interferon gamma (IFN<sub>γ</sub>), IL-10, IL-2, IL-1<sub>β</sub>, and tumor necrosis factor-α (TNF<sub>α</sub>; **B-F**) and by ELISA for MCP-1 (**G**). The bars represent mean±SEM, \**P*<0.05, \*\**P*<0.005, a 2-sample 2-sided Wilcoxon test with a Hochberg correction for multiple testing, n=4 to 5 per group.

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**Figure 5.** Effects of interleukin-4 (IL-4) on cultured mouse cardiac fibroblasts. **A**, Representative immunofluorescent image of cell-surface IL-4 receptor alpha (IL-4R $\alpha$ ) on mouse cardiac fibroblasts (green). Primary cardiac fibroblasts were prepared from wild-type (WT) or signal transducer, and activator of transcription 6 (STAT6)<sup>-/-</sup> mice, IL-4R $\alpha$  expression (**B**), and IL-4–induced phosphorylated STAT6 (P-STAT6; **C**) were analyzed by Western blot. **A** and **C**, Results from 3 independent experiments. **D**, Collagen contents in conditioned media of cardiac fibroblasts treated with IL-4 (10 ng/mL) for 48 hours. mRNA abundance of procollagen type-I  $\alpha$ 1 (Col1 $\alpha$ 1; **E**) and procollagen type-III  $\alpha$ 1 (Col3 $\alpha$ 1; **F**) after cells were incubated with IL-4 (10 ng/mL) for 6 hours. The bars represent mean $\pm$ SEM, \* $P$ <0.05, \*\* $P$ <0.005, a 2-sample 2-sided Wilcoxon test with a Hochberg correction for multiple testing,  $n$ =4 per group.