

ORIGINAL ARTICLE

Open Access



Molecular breeding of lignin-degrading brown-rot fungus *Gloeophyllum trabeum* by homologous expression of laccase gene

Misa Arimoto¹, Kenji Yamagishi², Jianqiao Wang¹, Kanade Tanaka³, Takanori Miyoshi⁴, Ichiro Kamei⁵, Ryuichiro Kondo⁶, Toshio Mori¹, Hirokazu Kawagishi^{1,7,8} and Hirofumi Hirai^{1,8*}

Abstract

The basidiomycete *Gloeophyllum trabeum* KU-41 can degrade Japanese cedar wood efficiently. To construct a strain better suited for biofuel production from Japanese cedar wood, we developed a gene transformation system for *G. trabeum* KU-41 using the hygromycin phosphotransferase-encoding gene (*hpt*) as a marker. The endogenous laccase candidate gene (*Gtlcc3*) was fused with the promoter of the *G. trabeum* glyceraldehyde-3-phosphate dehydrogenase-encoding gene and co-transformed with the *hpt*-bearing pAH marker plasmid. We obtained 44 co-transformants, and identified co-transformant L#61, which showed the highest laccase activity among all the transformants. Moreover, strain L#61 was able to degrade lignin in Japanese cedar wood-containing medium, in contrast to wild-type *G. trabeum* KU-41 and to a typical white-rot fungus *Phanerochaete chrysosporium*. By using strain L#61, direct ethanol production from Japanese cedar wood was improved compared to wild type. To our knowledge, this study is the first report of the molecular breeding of lignin-degrading brown-rot fungus and direct ethanol production from softwoods by co-transformation with laccase overproduction constructs.

Keywords: *Gloeophyllum trabeum* KU-41, Laccase gene, Homologous expression, Lignin degradation

Introduction

Gloeophyllum trabeum, along with *Postia placenta*, is a well-characterized brown-rot fungus used as a model laboratory organism. Brown-rot fungi are important recyclers in coniferous forest ecosystems and cause a highly destructive type of wood decay. Thus, elucidating the mechanisms employed by brown-rot fungi in the biodegradation of lignified plant cell walls is very important. A Fenton system is proposed to play an important role in cellulose decomposition by brown-rot fungi (Arantes et al. 2012). This system recently has been receiving increased attention because utilization of this process could facilitate the cost-effective transformation of lignocellulose biomass into biofuel or renewable chemicals (Kerem et al. 1999; Jensen et al. 2001). Therefore, many

studies have been conducted to understand the lignocellulose breakdown mechanisms in *G. trabeum*. In addition, decoding of the genomic sequences of brown-rot fungi, including those of *G. trabeum*, has facilitated the identification of key enzymes involved in lignocellulose metabolism (Floudas et al. 2012).

Ethanol is the most widely utilized liquid biofuel alternative to fossil fuels. Cellulosic biomass has been widely regarded as a readily available sugar source to replace starch materials in fermentation. Use of a single microorganism to produce ethanol from lignocellulosic biomass is expected to be especially cost-efficient; such an organism would have to possess the combined abilities to degrade lignin, hydrolyze cellulose, and ferment monosaccharides to ethanol (Lynd et al. 2005). Basidiomycetes play important roles in the carbon cycle as decomposers in forest ecosystems. Recently, we have proposed a new process of unified aerobic delignification and anaerobic saccharification and fermentation of hardwood by a single microorganism, the white-rot

*Correspondence: hirai.hirofumi@shizuoka.ac.jp

¹ Department of Applied Biological Chemistry, Faculty of Agriculture, Shizuoka University, 836 Ohya, Suruga-ku, Shizuoka 422-8529, Japan
Full list of author information is available at the end of the article

fungus *Phlebia* sp. MG-60 (Kamei et al. 2012). In contrast to hardwoods, softwoods such as Japanese cedar are more difficult to degrade by microbial activity (Katagiri et al. 1997). Brown-rot fungi have different mechanisms for the wood degradation with white-rot fungi, which are rapidly depolymerize the cellulose and hemicellulose in wood with modified lignin in the brown residue (Goodell et al. 1997; Hyde and Wood 1997). Brown-rot fungi are accessible to cellulose due to the high xylanase activity (Kim et al. 2014). The accessibility of cellulose increases in *G. trabeum* due to the removal of xylan (Gao et al. 2012). Furthermore, *G. trabeum* has the ability to produce ethanol from sugar under anaerobic or stressed conditions (Rasmussen et al. 2010). Thus, to address this challenge, we are investigating *G. trabeum* KU-41 as a model for the fungal degradation of Japanese cedar wood. Specifically, we hypothesized that it would be possible to develop this strain as a host basidiomycete capable of integrating the delignification, saccharification, and fermentation of softwoods. In this context, the construction of a novel genetic transformation system for *G. trabeum* was expected to permit the construction of strains with improved combinations of these properties. *T. versicolor*, *Pycnoporus coccineus*, *T. hirsute* were used as the laccase-producing wood rot fungi for the control in this study, because these fungi with high laccase production have been widely reported (Bourbonnais et al. 1995; Jaouani et al. 2005; Wu et al. 2011). *Phanerochaete chrysosporium* is a model white-rot fungus, used for comparing the lignin degradation in this study.

In the present study, we developed a novel genetic transformation system for *G. trabeum* KU-41, and demonstrated the molecular breeding of ligninolytic *G. trabeum* KU-41 by the homologous expression of an endogenous gene encoding a putative laccase activity.

Materials and methods

Strains and media

G. trabeum KU-41 (NBRC 111644) was described previously (Gao et al. 2012). The *G. trabeum* ATCC11539 strain was purchased from the American Type Culture Collection. The typical white-rot basidiomycetes *Trametes versicolor*, *Pycnoporus coccineus*, and *T. hirsuta* were purchased from NITE (National Institute of Technology and Evaluation). The strains used in this study are listed in Additional file 1: Table S1. All strains were maintained on semi-solid CYM medium (Yamagishi et al. 2007) at 28 °C.

Cloning of genomic DNA fragments encoding *Gtact* and *Gtgpd*

The reagents and protocols for manipulating DNA and RNA (DNA and RNA purification, cDNA synthesis,

3'-RACE, inverse PCR procedure, error-prone PCR, cloning of PCR products) were described previously (Yamagishi et al. 2013). The *G. trabeum* KU-41 genomic DNAs encoding cytosolic actin (*Gtact*) and glyceraldehyde-3-phosphate dehydrogenase (*Gtgpd*) were cloned by a series of PCR procedures (Additional file 1: Figs. S1, S2). The primers that were used are listed in Additional file 1: Table S2. These primers were designed based on nucleotide sequences and intron positions obtained from the DDBJ (<http://www.ddbj.nig.ac.jp/>). The accession numbers for *Gtact* and *Gtgpd* are AB856051 and AB856282, respectively.

Construction of transforming plasmids

The construction of transforming plasmids pAH and pGL are depicted in Additional file 1: Figs. S3, S4, S5. The plasmids used for the transformation of *G. trabeum* KU-41 were purified using Quantum Prep Midi kits (Bio-Rad Laboratories KK, Tokyo, Japan).

Preparation of *G. trabeum* protoplasts

G. trabeum protoplasts were prepared by slight modification of the previously described procedure (Yamagishi et al. 2007). Briefly, the KU-41 strain was pre-cultured for 10 days at 28 °C in liquid CYM medium (100 mL/flask in four 500-mL flasks). The mycelium was homogenized with a Waring blender (5000 rpm, 3 min) and cultured (for 4 days at 28 °C) in liquid CYM medium (100 mL/flask in twenty 500-mL flasks). Approximately 5 g of wet mycelium was collected from these cultures and treated with 20 mL of enzyme solution [Lyzing enzymes from Trichoderma (Sigma L1412-10G) 2.5 %, Cellulase Onozuka RS (Yakult) 2.5 %/0.5 M MgSO₄] at 30 °C for 6 h with gentle shaking. Protoplasts were separated from undigested mycelial debris by overlaying onto SorbOsm and centrifuging (4000×g) at 4 °C for 20 min. The viability of the resulting protoplasts was estimated as described previously (Yamagishi et al. 2007).

Genetic transformation procedure

Protoplasts of *G. trabeum* KU-41 (0.8×10^8 cells), marker plasmid pAH (2.5 to 20-μg), co-transformed plasmid pGL (20 μg), and 40 mM CaCl₂ (final concentration) were combined with SorbOsm to yield a total transformation volume of 1.6 mL. After incubation in ice for 30 min, 50 % PEG solution (1.6 mL) was added, and the combination was gently mixed before being diluted by addition of 20 mL SorbOsm. The transformation mix then was mixed with 150 mL of regeneration medium and poured into 20 Petri dishes (7.5 mL dish⁻¹). The composition of the regeneration medium was as described previously (Yamagishi et al. 2007), except that the pH of the medium used in the present study was adjusted to 6.5 (optimal

pH for hygromycin) with NaOH prior to use. Following solidification, the plates were incubated at 28 °C for 1 day, and the regenerating protoplasts then overlaid with 7.5 mL/plate of selection medium. (Selection medium consisted of regeneration medium, pH 6.5, supplemented with 40 $\mu\text{g mL}^{-1}$ hygromycin.) Regenerated clones, which appeared in 7–14 days, were picked and subcultured in the individual wells of a 24-well micro-plate containing 1 mL/well of semi-solid CYM medium (pH 6.5) supplemented with 20 $\mu\text{g mL}^{-1}$ hygromycin.

Amplification of transformed plasmids in regenerated clones

The direct PCR procedure for each regenerated clone was described previously (Yamagishi et al. 2013). Forward primers (hph-F1 and gpd-F12) and reverse primers (hph-R1 and lcc3-R1) were used to amplify the transformed plasmids pAH and pGL, respectively. The locations of the primers within the respective plasmids are depicted in Fig. 1.

Measurement of laccase activities of clones co-transformed with pGL

Initial screening was performed as follows. For each transformant harboring pGL (44 clones total), two 5-mm² pieces of mycelium were used to inoculate Kirk liquid medium (Tien and Kirk 1988) (2 mL/well in 12-well multititer plate), and plates were grown at 28 °C for 5 days. The laccase activity of each clone (each well) was measured using 3-ethylbenzothiazoline-6-sulfonic acid (ABTS) as the substrate (Nagai et al. 2002). Eleven clones showing appreciable laccase activity were further characterized for laccase activity. For each clone, three flasks were generated as follows: five 5-mm² pieces of

mycelium were used to inoculate Kirk liquid medium (10 mL/flask in 100-mL flask), and cultures were grown at 28 °C for 5 days. The laccase activity of each clone was calculated as the mean \pm standard deviation of the values from each trio of cultures. The laccase activity was also detected as zymogram. Extracellular proteins were separated by native polyacrylamide gel electrophoresis (PAGE), then visualized as zymogram by laccase active staining with 2,6-dimethoxyphenol.

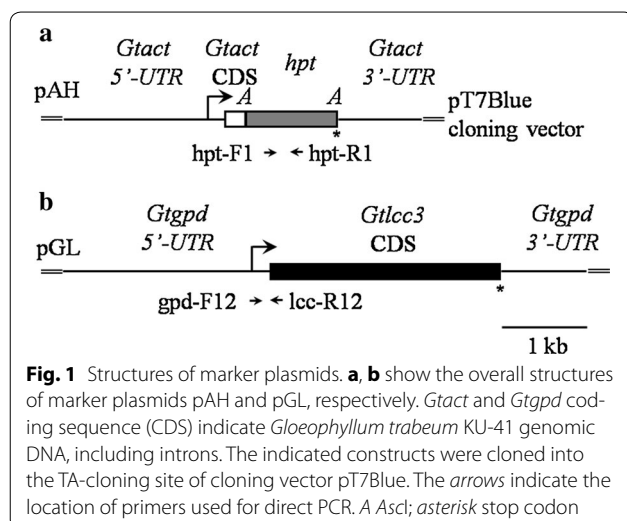
Assessment of genetic stability of clones co-transformed with pGL

Transformed clone L#61 was subcultured on semi-solid CYM medium without hygromycin until the border of colony reached the edge of the Petri dish (90 \times 15 mm). A piece of mycelium was punched out from the peripheral zone and subcultured on a new plate of semi-solid CYM medium without hygromycin. This process was repeated for a total of five times, and the sequential passages were denoted as generations 1–5. After subculturing, generations 1 and 5 of transformant L#61 were cultured in Kirk liquid medium (10-mL/flask in three 100-mL flasks) and their laccase activities were measured as described above. The laccase activity of each of the two tested generations was calculated as the mean \pm standard deviation of the values from each trio of cultures.

Determination of ligninolytic properties and ethanol production from wood

Erlenmeyer flasks (50-mL) containing 0.5 g extractive-free Japanese cedar wood meal (80–100 mesh) and 1.25 mL distilled water each were inoculated with mycelial discs of wild-type *G. trabeum* KU-41(WT) or transformant L#61 and the resulting cultures were incubated at 30 °C for 28 days. Weight loss, Klason lignin content, and acid-soluble lignin content in the fungal-treated wood meal were determined after the incubation period, as described previously (Hirai et al. 1994). Total lignin content was calculated as sum of Klason lignin and acid-soluble lignin contents.

Following the 28-day incubation, each wood meal culture was supplemented by the addition of 10 mL of basal liquid medium without saccharides (10 g L⁻¹ yeast extract, 10 g L⁻¹ KH₂SO₄, 2 g L⁻¹ (NH₄)₂SO₄, and 0.5 g L⁻¹ MgSO₄·7H₂O, pH 4.5) and then homogenized using a Polytron PT1200E. The cultures were further incubated with shaking (150 rpm) at 30 °C for 3, 6, 9, and 12 days in semi-aerobic conditions (obtained by sealing each culture flask with a silicon plug stopper). After fermentation, the wood meal cultures were centrifuged (10,000 \times g, 5 min). Ethanol levels in the resulting supernatants were measured by high-performance liquid chromatography (HPLC) using a Shodex SH1821 column



(8.0 mm × 300 mm, Showa Denko K.K., Tokyo, Japan) at 75 °C with 0.5 mM H₂SO₄ as the mobile phase at a flow rate of 0.6 mL min⁻¹. Detection was performed using an online Refractive Index Detector.

Results

Construction of a genetic transformation system for KU-41

We varied experimental conditions to investigate the appropriate conditions for preparing protoplasts of our strain. A sufficient number of protoplasts (0.2×10^8 protoplasts per gram of wet mycelium, 4 % cell viability) were obtained from the 4-day mycelial cultures. The marker plasmid pAH was constructed to express the hygromycin resistance-encoding gene (*hpt*) under the control of a constitutive promoter (for a gene encoding a cytosolic actin) (Fig. 1a). Genetic transformation of KU-41 protoplasts with 10 µg of the marker plasmid pAH yielded 26–35 transformants; no clones were obtained in the absence of pAH (Table 1). Direct PCR of clones transformed with pAH confirmed that all hygromycin-resistant clones contained the *hpt* gene (Fig. 2a).

Forced expression of the endogenous gene *Gtlcc3*

To confirm transgene expression in KU-41, the co-transforming plasmid pGL was constructed (Fig. 1b). pGL was designed to express an endogenous (*G. trabeum*) laccase candidate gene (*Gtlcc3*) under control of the *Gtgpd* promoter (Additional file 1: Figs. S4, S5). [*Gtlcc3* represents one of 4 putative laccase genes (*Gtlcc1–4*) identified in the *G. trabeum* genome database (http://genome.jgi-psf.org/Glotr1_1/Glotr1_1.home.html; protein ID: 127593)]. A total of 72 hygromycin-resistant clones were characterized following co-transformation with pAH and pGL; direct PCR revealed that 44 of the 72 clones harbored pGL (Fig. 2b; Table 1). All 72 clones were inoculated on semi-solid medium containing ABTS. After 2 days, green haloes were visible around 30 of the 44 pGL-positive clones. Haloes were not observed around any of the

Table 1 Transformed clones regenerated in uracil-free regenerating medium

Transformed plasmids		Exp. 1	Exp. 2
pAH	20 µg	75	77
pAH	10 µg	26	35
pAH	5 µg	20	24
pAH	2.5 µg	10	17
pAH + pGL	10 + 30 µg	100	106
Control	–	0	0

Transformation experiments were performed twice (experiment 1 and experiment 2)

Exp experiment

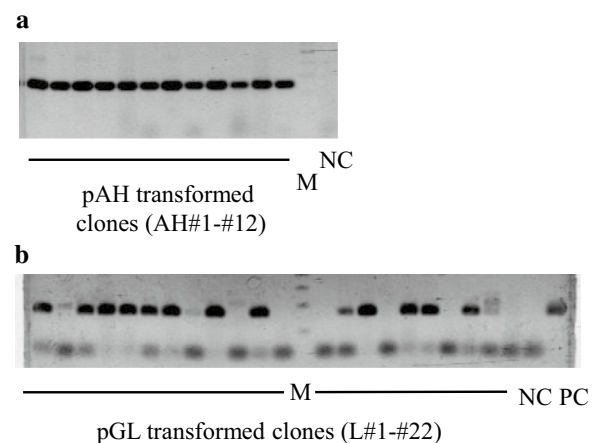


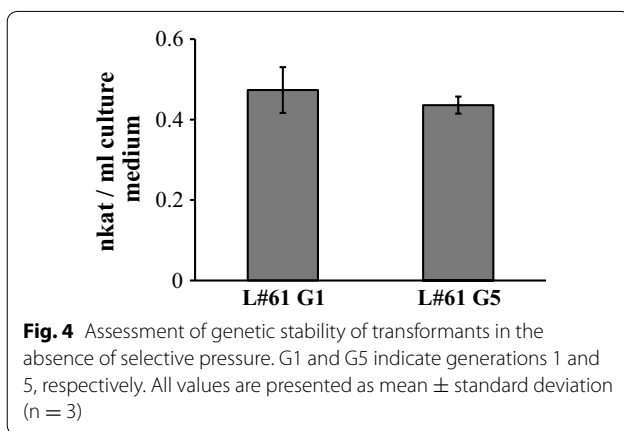
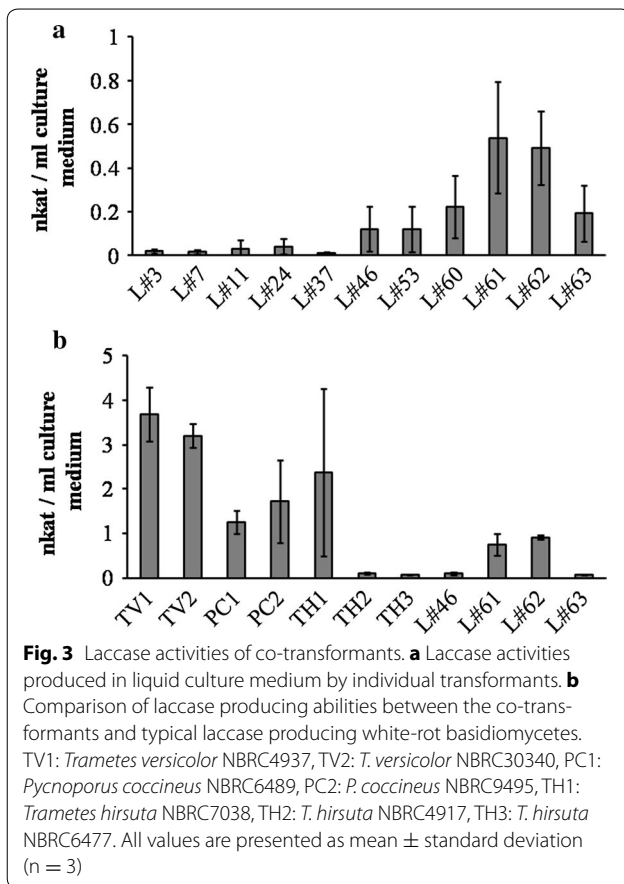
Fig. 2 Direct PCR analysis of genomic DNA recovered from individual transformants. Supernatants of homogenized mycelium of the transformants were used in PCR amplification with primer pair *hptF1-hptR1* (a) or *gpdF12-lccR12* (b). “M” indicates a 100 bp ladder size marker. “NC” indicates negative control (reactions with no PCR template). “PC” indicates positive control using pGL as PCR template. The PCR products were electrophoresed in 2 % agar and stained with SYBR Green (Takara)

negative control strains, including the KU-41 parent; 17 hygromycin-resistant strains recovered following transformation with pAH alone; and the separate *G. trabeum* ATCC 11539 strain. The production of laccase in Kirk liquid medium also was evaluated (Fig. 3). Co-transformant L#61 exhibited 0.73 nkat mL⁻¹ laccase activity. Although the laccase activity of co-transformant L#61 was lower than that of typical white-rot fungus *T. versicolor*, the activity was similar to that of *P. coccineus* (Fig. 3b).

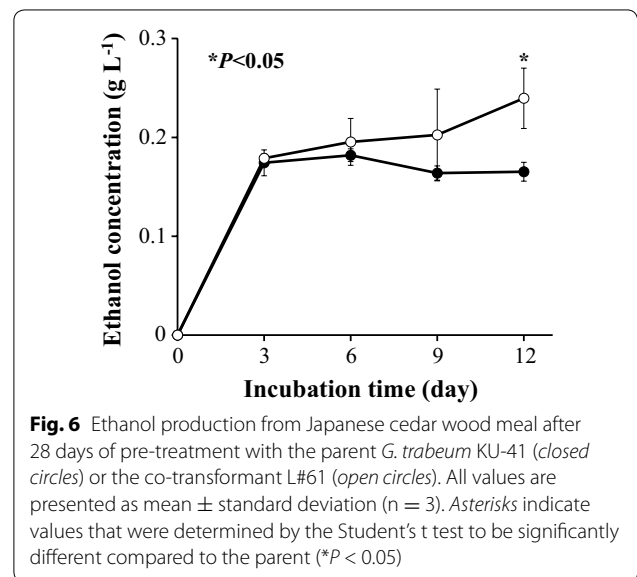
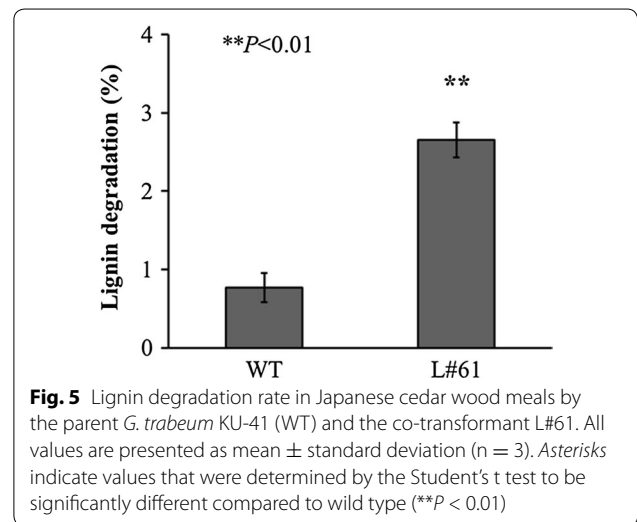
Co-transformant L#61 was repeatedly subcultured on semi-solid CYM medium without hygromycin to evaluate the genetic stability of the laccase construct in the clones co-transformed with pAH and pGL. Laccase production by the L#61 co-transformant did not appear to decrease even after 5 cycles of subculturing on hygromycin-free semisolid medium (Fig. 4; Additional file 1: Fig. S6).

Degradation of lignin in Japanese cedar wood meal by co-transformant L#61

We tested whether the L#61 co-transformant was capable of degrading lignin in Japanese cedar wood meal. The co-transformant L#61 exhibited significantly elevated ligninolytic activity (2.7 %) compared to the parent (0.8 %), as shown in Fig. 5. L#61 indicates values that were determined by the student's *t* test to be significantly different compared to wild type. In a parallel assay, the white-rot fungus *P. chrysosporium* also hardly degraded Japanese cedar wood meal (data not shown).



After a 28-day pretreatment period, the WT or transformant L#61 wood meal cultures were transferred to a basal liquid medium and incubated under semi-aerobic conditions to assess ethanol production. After 12 days of semi-aerobic incubation, co-transformant L#61 exhibited a significant increase (of 45 %) in ethanol production compared to WT (Fig. 6).



Discussion

In the present study, we investigated the appropriate conditions for preparing protoplasts of the KU-41 strain, because preparation of a sufficient number of protoplasts is essential for construction of a genetic transformation system. Among various parameters (e.g., the age of the mycelium, concentration of the osmotic stabilizer in the enzyme solution, and the composition of the regeneration medium), mycelial age showed the strongest effects on protoplast isolation (data not shown). Older mycelium produced a viscous substance that hindered the enzyme treatment. In addition, because some reports have indicated that native introns (i.e., introns derived from the host species) were essential for the expression

of exogenous genes in some basidiomycetes (Lugones et al. 1999; Ma et al. 2001; Burns et al. 2005; Yamazaki et al. 2006; Yamagishi et al. 2013), the first and second introns were retained as part of the *actin* promoter in pAH. However, the necessity of the introns was not confirmed in this study. Genetic transformation of KU-41 protoplasts with the marker plasmid pAH was successful, thus demonstrating our construction of a genetic transformation system for the KU-41 strain of the brown-rot fungus *G. trabeum*. The endogenous laccase candidate gene was fused with the *G. trabeum* *gpd* promoter and co-transformed with marker plasmid pAH. We obtained 44 co-transformants, and identified co-transformant L#61, which showed the highest laccase activity among all of the screened transformants. Our results indicated that forced expression of the introduced endogenous gene was successful and confirmed that *Gtlcc3* indeed encodes a laccase activity. Although the laccase activity of co-transformant L#61 is lower than that of typical white-rot fungus *T. versicolor*, the activity was similar to that of *P. coccineus*. Transformants with elevated laccase production might be generated by modifying the pGL promoter region and/or by screening among a larger number of co-transformants. Given that pGL does not contain a known origin of replication for *G. trabeum* and that the transformed plasmids were stably retained in the absence of selective pressure, we infer that the transformed plasmids were integrated into the chromosomes of the transformants.

Our primary concern was whether the co-transformant L#61 could degrade lignin; the parent *G. trabeum*, like all brown-rot fungi, does not exhibit appreciable lignin degradation activity (Gao et al. 2012). We assayed the ability of co-transformant L#61 to degrade lignin using wood meal derived from Japanese cedar. Laccase mediates one-electron oxidation of phenolic substrates to form many degradation products via various pathways, but the enzyme can not oxidize non-phenolic substrate (Kawai et al. 1989). Thus, co-transformant L#61 showed low delignification activity, because this fungus does not have the delignification systems for non-phenolic structure like white-rot fungi. Additionally, hardwood tree species contain both guaiacyl and syringyl lignin but softwood tree species contain only guaiacyl lignin. Although white rot fungi preferably degrade syringyl units (Dong et al. 2013), they are not contained in Japanese cedar wood (a softwood). Thus, it is suggested that *P. chrysosporium* hardly degraded Japanese cedar wood meal. From the above, it is estimated that co-transformant L#61 was only able to oxidize phenolic residues that were presented at surface of lignin molecules in Japanese cedar wood.

Our results indicated that KU-41 was invested with ligninolytic activity upon the expression of *Gtlcc3*;

delignification improved the direct yield of ethanol following fermentation of the wood meal. Autoclave sterilization, which could cause an effect similar to steam explosion pretreatment frequently used as pre-treatment for lignocellulosic substrates, was performed before fungal inoculation. Therefore, it is likely that the wood structure was changed into the structure that has better accessibility for fungal hydrolases to cellulose by the sterilization. In addition, due to relatively high xylanase activity, *G. trabeum* could make wood to accessible structure to cellulose. (Kim et al. 2014). From these reason, *G. trabeum* wild type might have been able to hydrolyze cellulose effectively and produce ethanol, in this experiment. Moreover, it is generally considered that both lignin and hemicellulose limit the accessibility of cellulase. And hemicellulose and lignin are chemically bind. Therefore, we estimated that recombinant laccase assists the hemicellulose removal due to polysaccharide hydrolases as a result of lignin surface oxidation, and that the improvement of lignin and hemicellulose removal leads to higher ethanol productivity of co-transformant L#61.

In conclusion, the homologous overexpression of a putative laccase-encoding gene provided *G. trabeum* with ligninolytic activity against Japanese cedar wood. To our knowledge, this is the first report that the molecular construction of a lignin-degrading brown-rot fungus. We expect that our transformation system and the resulting strain constructs will facilitate molecular analysis of the lignocellulose biodegrading system of *G. trabeum* and other brown-rot fungi.

Additional file

Additional file 1: Table S1. List of strains used in this study. **Table S2.** Oligonucleotides used as primers in this study. **Fig. S1.** Procedure for cloning *Gloeophyllum trabeum* actin-encoding genomic DNA. **Fig. S2.** Procedure for cloning *Gloeophyllum trabeum* glyceraldehyde-3-phosphate dehydrogenase-encoding genomic DNA. **Fig. S3.** Construction of marker plasmid pAH. **Fig. S4.** Addition of an *AscI* site in pGtgpd. **Fig. S5.** Construction of pGL. **Fig. S6.** Zymogram activity of the purified laccase enzyme with native PAGE.

Authors' contributions

MA, KY, JW, and TM participated in the experimental procedures of this study and drafting of the manuscript. KT, TM, IK, RK and HK involved in the data analysis and interpreted the results. HH participated in the design this study, data analysis, interpretation of results and drafting of the manuscript. All authors read and approved the final manuscript.

Author details

¹ Department of Applied Biological Chemistry, Faculty of Agriculture, Shizuoka University, 836 Ohya, Suruga-ku, Shizuoka 422-8529, Japan. ² NARO National Food Research Institute, 2-1-12 Kannondai, Tsukuba, Ibaraki 305-8642, Japan. ³ Integrative Technology Research Institute, Teijin Limited, Iwakuni 740-8511, Japan. ⁴ New Business Development Business Unit, Teijin Limited, Tokyo 100-8585, Japan. ⁵ Faculty of Agriculture, University of Miyazaki, Miyazaki 889-2192, Japan. ⁶ Faculty of Agriculture, Kyushu University, Fukuoka 812-8581, Japan. ⁷ Graduate School of Science and Technology,

Shizuoka University, Shizuoka 422-8529, Japan. ⁸ Research Institute of Green Science and Technology, Shizuoka University, Shizuoka 422-8529, Japan.

Acknowledgements

This work was supported in part by a project, titled "Research and Development of Catalytic Process for Efficient Conversion of Cellulosic Biomass into Biofuels and Chemicals", which was funded through the Special Funds for Education and Research (No. 21248023); and by a Grant-in-Aid for Scientific Research (A) from the Ministry of Education, Culture, Sports, Science and Technology of Japan (No. 24248030).

Competing interests

The authors declare that they have no competing interests.

Received: 16 September 2015 Accepted: 11 December 2015

Published online: 22 December 2015

References

- Arantes V, Jellison J, Goodell B. Peculiarities of brown-rot fungi and biochemical Fenton reaction with regard to their potential as a model for bioprocessing biomass. *Appl Microbiol Biotechnol*. 2012;94:323–38.
- Bourbonnais R, Paice MG, Reid ID, Lanthier P, Yaguchi M. Lignin oxidation by laccase isozymes from *Trametes versicolor* and role of the mediator 2,29-azinobis(3-ethylbenzthiazoline-6-sulfonate) in kraft lignin depolymerization. *Appl Environ Microbiol*. 1995;61:1876–80.
- Burns C, Gregory KE, Kirby M, Cheung MK, Riquelme M, Elliott TJ, Challen MP, Bailey A, Foster GD. Efficient GFP expression in the mushrooms *Agaricus bisporus* and *Coprinus cinereus* requires introns. *Fungal Genet Biol*. 2005;42:191–9.
- Dong XQ, Yang JS, Zhu N, Wang ET, Yuan HL. Sugarcane bagasse degradation and characterization of three white-rot fungi. *Bioresour Technol*. 2013;131:443–51.
- Floudas D, Binder M, Riley R, Barry K, Blanchette RA, Henrissat B, Martínez AT, Otiillar R, Spatafora JW, Yadav JS, Aerts A, Benoit I, Boyd A, Carlson A, Copeland A, Coutinho PM, de Vries RP, Ferreira P, Findley K, Foster B, Gaskell J, Glotzer D, Górecki P, Heitman J, Hesse C, Hori C, Igarashi K, Jurgens JA, Kallen N, Kersten P, Kohler A, Kües U, Kumar TK, Kuo A, LaButti K, Larrondo LF, Lindquist E, Ling A, Lombard V, Lucas S, Lundell T, Martin R, McLaughlin DJ, Morgenstern I, Morin E, Murat C, Nagy LG, Nolan M, Ohm RA, Patyshakuliyeva A, Rokas A, Ruiz-Dueñas FJ, Sabat G, Salamov A, Samejima M, Schmutz J, Slot JC, St John F, Stenlid J, Sun H, Sun S, Syed K, Tsang A, Wiebenga A, Young D, Pisabarro A, Eastwood DC, Martin F, Cullen D, Grigoriev IV, Hibbett DS. The Paleozoic origin of enzymatic lignin decomposition reconstructed from 31 fungal genomes. *Science*. 2012;336:1715–9.
- Gao Z, Mori T, Kondo R. The pretreatment of corn stover with *Gloeophyllum trabeum* KU-41 for enzymatic hydrolysis. *Biotechnol Biofuels*. 2012;5:28. doi:10.1186/1754-6834-5-28.
- Goodell B, Jellison J, Liu J, Daniel G, Paszczynski A, Fekete F, Krishnamurthy S, Jun L, Xu G. Low molecular weight chelators and phenolic compounds isolated from wood decay fungi and their role in the fungal biodegradation of wood. *J Biotechnol*. 1997;53:133–62.
- Hirai H, Kondo R, Sakai K. Screening of lignin degrading fungi and their ligninolytic enzyme activities during biological bleaching of kraft pulp. *Mokuzai Gakkaishi*. 1994;40:980–6.
- Hyde SM, Wood PM. A mechanism for production of hydroxyl radicals by the brown-rot fungus *Coniophora puteana*: Fe(III) reduction by cellobiose dehydrogenase and Fe(II) oxidation at a distance from the hyphae. *Microbiology*. 1997;143:259–66.
- Jaouani A, Guillén F, Penninckx MJ, Martínez AT, Martínez MJ. Role of *Pycnoporus coccineus* laccase in the degradation of aromatic compounds in olive oil mill wastewater. *Enzyme Microb Technol*. 2005;36:478–86.
- Jensen KA Jr, Houtman CJ, Ryan ZC, Hammel KE. Pathways for extracellular Fenton chemistry in the brown rot basidiomycete *Gloeophyllum trabeum*. *Appl Environ Microbiol*. 2001;67:2705–11.
- Kamei I, Hirota Y, Meguro S. Integrated delignification and simultaneous saccharification and fermentation of hard wood by a white-rot fungus, *Phlebia* sp. MG-60. *Bioresour Technol*. 2012;126:137–41.
- Katagiri N, Tsutsumi Y, Nishida T. Biobleaching of softwood kraft pulp by white-rot fungi and its related enzymes. *Mokuzai Gakkaishi*. 1997;43:678–85.
- Kawai S, Umezawa T, Higuchi T. Oxidation of methoxylated benzyl alcohol by *Coriolus versicolor* in the presence of syringaldehyde. *Wood Res*. 1989;76:10–6.
- Kerem Z, Kenneth AJ, Kenneth EH. Biodegradative mechanism of the brown rot basidiomycete *Gloeophyllum trabeum*: evidence for an extracellular hydroquinone-driven fenton reaction. *FEBS Lett*. 1999;446:49–54.
- Kim HM, Lee KH, Kim KH, Lee DS, Nguyen QA, Bae HJ. Efficient function and characterization of GH10 xylanase (Xyl110g) from *Gloeophyllum trabeum* in lignocellulose degradation. *J Biotechnol*. 2014;172:38–45.
- Lugones LG, Scholtmeijer K, Klootwijk R, Wessels JG. Introns are necessary for mRNA accumulation in *Schizophyllum commune*. *Mol Microbiol*. 1999;32:681–9.
- Lynd LR, van Zyl WH, McBride JE, Laser M. Consolidated bioprocessing of cellulosic biomass: an update. *Curr Opin Biotechnol*. 2005;16:577–83.
- Ma B, Mayfield MB, Gold MH. The green fluorescent protein gene functions as a reporter of gene expression in *Phanerochaete chrysosporium*. *Appl Environ Microbiol*. 2001;67:948–55.
- Nagai M, Sato T, Watanabe H, Saito K, Kawata M, Enei H. Purification and characterization of an extracellular laccase from the edible mushroom *Lentinula edodes*, and decolorization of chemically different dyes. *Appl Microbiol Biotechnol*. 2002;60:327–35.
- Rasmussen ML, Shrestha P, Khanal SK, Pometto AL 3rd, Hans van Leeuwen J. Sequential saccharification of corn fiber and ethanol production by the brown rot fungus *Gloeophyllum trabeum*. *Bioresour Technol*. 2010;101:3526–33.
- Tien M, Kirk TK. Lignin peroxidase of *Phanerochaete chrysosporium*. *Methods Enzymol*. 1988;161:238–48.
- Wu J, Zhang X, Wan J, Ma F, Tang Y, Zhang X. Production of fiberboard using corn stalk pretreated with white-rot fungus *Trametes hirsute* by hot pressing without adhesive. *Bioresour Technol*. 2011;102:11258–61.
- Yamagishi K, Kimura T, Oita S, Sugiura T, Hirai H. Transformation by complementation of a uracil auxotroph of the hyper lignin-degrading basidiomycete *Phanerochaete sordida* YK-624. *Appl Microbiol Biotechnol*. 2007;76:1079–91.
- Yamagishi K, Kimura T, Watanabe T. Construction of an endogenous selectable marker gene for the rice straw degrading white rot basidiomycete *Cyathus stercoreus*. *Biosci Biotechnol Biochem*. 2013;77:771–5.
- Yamazaki T, Okajima Y, Kawashima H, Tsukamoto A, Sugiura J, Shishido K. Intron-dependent accumulation of mRNA in *Coriolus hirsutus* of lignin peroxidase gene the product of which is involved in conversion/degradation of polychlorinated aromatic hydrocarbons. *Biosci Biotechnol Biochem*. 2006;70:1293–9.

Submit your manuscript to a SpringerOpen® journal and benefit from:

- Convenient online submission
- Rigorous peer review
- Immediate publication on acceptance
- Open access: articles freely available online
- High visibility within the field
- Retaining the copyright to your article

Submit your next manuscript at ► springeropen.com