

# Transretinoic acid inhibits rats gastric epithelial dysplasia induced by N-methyl-N-nitro-N-nitrosoguanidine: influences on cell apoptosis and expression of its regulatory genes

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## INTRODUCTION

Gastric epithelial dysplasia (GED) hypothetically is a straight-forward concept: dysplastic epithelium replacing the normal gastric epithelium of the stomach<sup>[1]</sup>. In the stomach, like any other segment of the gut, it is defined as an unequivocal non-invasive epithelial change<sup>[2,3]</sup>. The observation of gastric dysplasia as a cancerous lesion was recognized over a century ago, but it is only after the advent of gastroscopy that its clinical significance has been stressed<sup>[4-7]</sup>.

All-trans retinoic acid can effectively reverse dysplasia in gastric epithelial cells, thereby inhibiting its progression to gastric cancer<sup>[8-10]</sup>. However, its mechanism is not yet clear. We used N-methyl-N-nitro-N-nitrosoguanidine (MNNG) to establish a rat model of gastric epithelial cell dysplasia, and to study the influence of all-trans

retinoic acid. Changes in apoptosis and expression of Bcl-2, Fas and ICE were observed to investigate into insight of its mechanism.

## MATERIALS AND METHODS

### *Induction of gastric epithelial cell dysplasia and treatment with all-trans retinoic acid*

Forty-five 8-week-old male Wistar rats weighing 120 g-140 g were housed in individual cages at a controlled temperature of 22 °C, and a relative humidity of 50%. These animals were randomly divided into three groups each 15 rats. Group 1 served as blank control. Groups 2 and 3 were fed with MNNG to induce gastric epithelial cell dysplasia. Two grams MNNG (Fluka Co.) was dissolved in 2000 mL distilled water, placed in a brown bottle and kept at 4 °C. The preserved MNNG solution was further diluted to the concentration of 1 g/mL for use as drinking water ad lib<sup>[11,12]</sup>. In addition, at one, three, five and seven weeks, 2 mL absolute alcohol was infused into the stomach of each animal. After 24 weeks, when gastric epithelial cell dysplasia had been induced, animals in group 2 were given 40 µg/kg all-trans retinoic acid (Shanghai No.6 Pharmaceutical Co.) through infusion into the stomach every day. Group 3 served as treatment controls and a placebo (distilled water) was given instead of all-trans retinoic acid. The animals were killed at 36 weeks.

The stomachs of the rats were cut along the greater curvature. Specimens were taken from the pyloric area and five paraffin sections were made for each rat following the conventional method. These sections were used for routine pathological examinations, apoptosis determination and measurements for the expression of Bcl-2, Fas and ICE. Pathological diagnoses and gradings were carried out according to the criteria set by the National Gastric Cancer Research Group<sup>[13,14]</sup>.

### *Determination of apoptosis by TUNEL method*

After dewaxing a 4 µm thick section, the fundamental steps of the procedure were: addition of proteinase K (20 µL/mL); incubated at 37 °C; addition of Triton x-100 in 0.1% sodium citrate on ice, stood up; addition of 50 g TUNEL (Boehringer Co., Cat. no. 1684817); incubated at 37 °C in a humidified chamber; staining with diaminobenzidine (DAB); sealing the slide; and

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examined under a light microscope. Apoptotic cells presented as brownish staining in the nucleus, although part of the cytoplasm also can be stained because of leakage of some nuclear DNA fragments. At least 500 cells were examined and the number of apoptotic cells per 100 cells calculated to arrive at the apoptosis index (AI)<sup>[15,16]</sup>.

#### **Expression of Bcl-2, Fas and ICE measured by immunohistochemical staining**

**Bcl-2** After dewaxing, the 4 µm thick section was stained using the Avidin-Biotin Complex (ABC) method<sup>[16]</sup>. The primary antibody used was 2 mg/L rabbit antirat Bcl-2 polyclonal antibody (Santa Cruz Co. USA, Cat. No. Sc-578) diluted 1:50.

**Fas** To detect Fas activity, sections were stained by ABC method. The primary antibody used was 2 mg/L rabbit antirat Fas polyclonal antibody (Santa Cruz Co. USA, Cat. No. sc-716G) diluted 1:50.

**ICE** To detect ICE activity, sections were stained by the ABC method. The primary antibody used was 2 mg/L rabbit antirat ICE polyclonal antibody (Santa Cruz Co. USA, Cat. No. sc-514) diluted 1:50.

**Positivity control** Slides of multiple gene-protein-positive gastric cancer specimens were used as positive controls. If brown granules appeared on the nuclear membrane and in the cytoplasm of gastric epithelial cells, the specimen was considered to be Bcl-2 positive. If brown granules appeared on the cell membrane, the specimen was considered to be Fas positive; and if brown granules appeared in the cytoplasm, the specimen was considered to be ICE positive. The expression was graded according to the degree of staining: strongly positive (+++), moderately positive (++) and weakly positive (+). If more than 30% of cells in a slide were also considered positive. Those with strongly and moderately positive responses were considered to be overexpressing the protein in question<sup>[17-20]</sup>.

#### **Probes**

The following oligonucleotide probes were used in this study. Bcl-2, TGATACCAGCACTGGAGCAG, was synthesized by Shanghai Shengong Company. Fas, CAGCCAGGAAAGATCAAACAGAGAGC, was bought from Fuzuo Company.

#### **Northern-blot analysis**

Total RNA was isolated from the gastric epithelial tissue by extraction of guanidine isothiocyanate and centrifugation in cesium chloride<sup>[21,22]</sup>. Poly(A)<sup>+</sup>RNA was selected by oligo(dt)-cellular chromatography<sup>[23]</sup>. Six micograms of poly(A)<sup>+</sup>RNA from each sample was electrophoresed in 1% agarose gel containing 0.66 mol/L formaldehyde

and ethidium bromide (0.66 mg/L).

After electrophoresis, the gels were photographed under UV light to confirm that approximately equal amounts of RNA were loaded. The gels were pretreated with 0.05N NaOH for 30 minutes at room temperature<sup>[24,25]</sup> and RNA was transferred onto nitrocellulose. Then, appropriate probes were labeled with <sup>32</sup>P deoxycytidine triphosphate using a Random Prime DNA labeling kit (Boehringer Mannheim, Indianapolis, IN). The blots were hybridized overnight at 62 °C (bcl-2) or 52 °C (Fas) in 59% formamide, 10% dextran sulfate, 1% sodium dodecyl sulfate, 1 mol/L NaCl, and 100 g/L of sonicated salmon sperm. Then the blots were washed in 2 × standard saline citrate (1×SSC is 150 mmol/L NaCl, 15 mmol/L sodium citrate, pH 7.4), and 0.5% sodium dodecyl sulfate four times at room temperature for 5 minutes and also washed (3 × 10 minutes) in 0.2 × SSC and 0.5% sodium dodecyl sulfate at 60 °C. The blots were then exposed to Kodak XAR film (Eastman Kodak, Rochester, NY) at -70 °C. The nitrocellulose filter was boiled in 0.1% SSC and 0.1% SDS for 30 minutes to strip off the radioactivity probes and rehybridized with another <sup>32</sup>P labelled Cdna prob in a similar manner<sup>[26]</sup>. The quantity of specificity transcripts in different lanes was determined by densitometric analysis of autoradiographs.

## **RESULTS**

### **Histopathological changes of the gastric mucosa**

The incidence of dysplasia in group 3 was significantly higher than that in group 2 (73.3% vs 26.7%,  $P = 0.05$ , Table 1).

**Table 1 Histopathological changes and apoptosis of the gastric mucosa**

Group	n	Modest dysplasia (n)	Modest and severe dysplasia (n)	Percentage (%)	Apoptotic index
Normal	15	0	0	0	8.3±3.1
DIM	15	3	11	73.3%	2.2±0.4 <sup>a</sup>
DIMTR	15	4	4	26.7% <sup>ab</sup>	7.8±2.6 <sup>bc</sup>

<sup>a</sup> $P < 0.05$  vs normal; <sup>b</sup> $P > 0.05$  vs normal; <sup>c</sup> $P > 0.05$  vs DIM group.

### **Apoptosis**

The apoptosis index was not significantly different in groups 1 and 2 ( $P > 0.05$ ), but there was a significant difference between groups 1 and 3 ( $P < 0.05$ ) and also between groups 2 and 3 ( $P < 0.05$ ).

### **Expression of apoptosis-associated proteins**

**Bcl-2** In group 1, two rats (13.3%) expressed Bcl-2 and one rat (6.7%) overexpressed Bcl-2. Expression of Bcl-2 was found in 10 rats (66.7%) and overexpression of Bcl-2 in five rats (33.3%) in group 3. Both the expression and overexpression

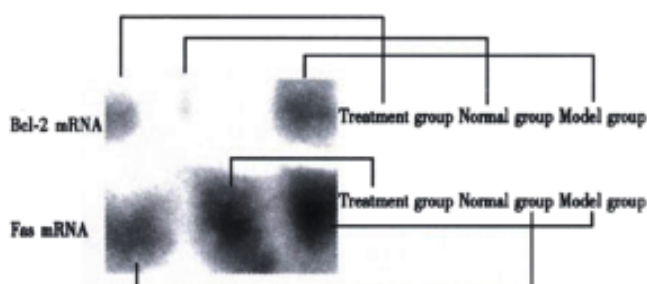
were significantly higher than that in group 1 ( $P < 0.05$ ). Group 2 had five rats (33.3%) expressing Bcl-2 and one rat (6.7%) overexpressing Bcl-2. Neither expression nor overexpression was significantly different from that of group 1 rats ( $P > 0.05$ ) but was significantly different from that of group 3 ( $P < 0.05$ ).

**Fas** In group 1, seven rats (46.7%) expressed Fas and two rats (13.3%) overexpressed of Fas. Group 3 had two rats (13.3%) with expression of Fas and two rats (13.3%) with overexpression of Fas. Expression of Fas was significantly different from that of group 1 ( $P < 0.05$ ), but overexpression was not ( $P > 0.05$ ). In group 2, six rats (40%) expressed Fas and four rats (26.7%) overexpressed Fas. Neither expression nor overexpression were significantly different from that of group 1 ( $P > 0.05$ ), but when compared with group 3, a significant difference was seen in the expression of Fas ( $P < 0.05$ ), yet not in overexpression ( $P > 0.05$ ).

**ICE** In group 1, three rats (20%) expressed ICE and no rats were found to be overexpressing this protein. Group 3 had two rats (13.3%) with expressing ICE and one rat (6.7%) with overexpressing ICE. Neither expression nor overexpression were significantly different from that in group 1 ( $P > 0.05$ ). In group 2, expression of ICE was seen in nine rats (60%) and overexpression was found in two rats (13.3%). Expression of ICE was significantly different from that in group 1 ( $P < 0.05$ ) but overexpression was not ( $P > 0.05$ ), and when compared with that of group 3, expression of ICE was significantly different ( $P < 0.05$ ) while overexpression was not ( $P > 0.05$ ).

#### **Bcl-2 and Fas mRNA expression**

The expression of bcl-2 mRNA increased and Fas mRNA was decreased in comparison of group 2 with group 1. The expression of bcl-2 mRNA was decreased and Fas mRNA was increased in comparison of group 2 with group 3 (Figure 1).



**Figure 1** Bcl-2 and Fas mRNA expression.

## **DISCUSSION**

With the introduction of fiberoptic endoscopy in the late 1960s and early 1970s, Nakamura and Nagayo in Japan were among the first to identify possible precancerous lesion on biopsical material and develop several categories for dysplasia<sup>[27,28]</sup>. In the West, Grundmann in 1975 quote for the first time the word dysplasia to describe exclusively precancerous gastric lesions<sup>[29]</sup>. Shortly after the WHO committee sanctioned this usage and detailed general diagnostic principles based on cellular atypia, abnormal differentiation and disorganised architecture<sup>[30,31]</sup>. There are differences between the Japanese and the Western criteria. A multicenter study is now under way to unify the diagnostic criteria<sup>[32-34]</sup>. So far there has been no unified criteria of dysplasia among Chinese. We used the Japanese criteria in this study.

**Is dysplasia reversible** Several investigators have addressed the issue, but the cumulative results remain inconclusive. There are still controversies in the interpretation of atrophic changes and intestinal metaplasia resulting in wide discrepancy in the conclusions reached by different authors. Controlled, long-term prospective studies conducted in different ethnic and geographic settings are needed to provide sound evidence-based answers to the question of reversibility of atrophy, intestinal metaplasia, and epithelial dysplasia<sup>[6,35]</sup>, trying to look for new drugs to reverse the dysplasia of great importance.

Normal gastric mucosal epithelial cells undergo apoptosis to clear up the senile cells and maintain the physiological balance of mucosal epithelial cells<sup>[36]</sup>. The homeostasis of gastric epithelial cells is maintained by the balance between cell proliferation and apoptosis. Alterations of these physiological cellular events in chronic pathological conditions of the stomach, as far as the proliferative pattern is concerned, an increase in the total number of epithelial proliferating cells and an abnormal distribution of the latter are frequently observed in chronic gastritis, gastric atrophy, intestinal metaplasia, gastric dysplasia and gastric cancer. Conversely, apoptosis has been found to be impaired in intestinal metaplasia, gastric dysplasia and cancer<sup>[37,38]</sup>. We consider the development of gastric cancer as a simple problem of balance, which is explained in the following formula: cumulative rate of epithelial cells' proliferation rate of epithelial cells' apoptotic rate of epithelial cells. When overaccumulation of gastric mucosal epithelial cells due to any reason, there is the possibility of development of gastric cancer. Thus, abnormal apoptosis might be one of the causes of gastric cancer development.

Apoptosis is modulated by regulatory genes. Bcl-2, Fas and ICE are the regulatory genes that have been predominantly studied. The statement of Bcl-2 can inhibit apoptosis, allowing the

proliferating cells to accumulate and inhibiting the removal of the malignant potential cells, thereby facilitating the development of cancerous change. Several scientists have studied the relation among bcl-2, gastric epithelial dysplasia and apoptosis and found that the expression abnormality leads to the apoptosis changed, the result in dysplasia and carcinoma<sup>[19,39-42]</sup>. The statement of Fas promotes apoptosis and its abnormality has relation with dysplasia too<sup>[43]</sup>. Interleukin-1-coverzyme can induce apoptosis in certain types of cells<sup>[44-46]</sup>.

A few investigation of bcl-2 expression and dysplasia found that bcl-2 expression did not correlate with the presence or degree of dysplasia in either benign gastric mucosa or gastric carcinoma (GC) patients. bcl-2 protein is frequently expressed in GC<sup>[39,47]</sup>. The reason perhaps is the difference of rat's model and human gastric epithelial dysplasia, model induced by MNNG caused mainly by chemistrial carcinogenesis, is not all the same with human.

Retinoic acid and its analog retinoid has a reverse effect on experimentally induced gastric mucosal precancerous lesions in rats<sup>[8-10,48]</sup>. The results of the present study further confirmed this. The results of the present study also revealed that retinoic acid can inhibit the overstatement of the Bcl-2 protein, promote the normal statement of the Fas protein and enhance the overstatement of the ICE protein, thereby promoting the apoptosis of gastric mucosal dysplastic epithelial cells. This may be one of the mechanisms by which retinoic acid reverses gastric mucosal precancerous lesions and the high statement of ICE may partly explain the side effects of retinoic acid.

The results of the present study reveal that in moderate and severe gastric mucosal dysplasia precancerous lesions, there is already abnormality in apoptosis and changes in associated genes. The increase in Bcl-2 statement, decrease in Fas statement and inhibition of apoptosis may be an important mechanisms in the progression of dysplasia to cancer.

## REFERENCES

- Goldstein NS, Lewin KJ. Gastric epithelial dysplasia and adenoma: historical review and histological criteria for grading. *Hum Pathol*, 1997;28:127-133
- Lewin KJ. Nomenclature problems of gastrointestinal epithelial neoplasia. *Am J Surg Pathol*, 1998;22:1043-1047
- Lauwers GT, Riddell RH. Gastric epithelial dysplasia. *Gut*, 1999; 45:784-790
- Talbot IC. Pathology and natural history of gastric carcinoma. In: Cancer of the stomach. Edited by Preece PE, Cuschieri A, Wellwood JM. London, 1985:73-85
- Fertitta AM, Comin U, Terruzzi V, Minoli G, Zambelli A, Cannatelli G, Bodini P, Bertoli G, Negri R, Brunati S, Fiocca R, Turpini F, Prada A, Ceretti E, Gullotta R, Cornaggia M. Clinical significance of gastric dysplasia: a multicenter follow-up study. *Hum Pathol*, 1993;25:265-268
- Fang DC. Status of researches in precancerous lesions of gastric mucosa. *Huaren Xiaohua Zazhi*, 1998;6:645-646
- You WC, Zhang L, Gail MH, Li JY, Chang YS, Blot WJ, Zhao CL, Liu WD, Li HQ, Ma JL, Hu YR, Bravo JC, Correa P, Xu GW, Fraumeni JF Jr. Precancerous lesions in two counties of China with contrasting gastric cancer risk. *Int J Epidemiol*, 1998;27:945-948
- Zhu ZH, Xia ZS, He SG. The effects of ATRA and 5-Fu on telomerase activity and cell growth of gastric cancer cells *in vitro*. *Shijie Huaren Xiaohua Zazhi*, 2000;8:669-673
- Jiang SY, Shyu RY, Chen HY, Lee MMS, Wu KL, Yeh MY. In vitro and in vivo growth inhibition of SC M1 gastric cancer cells by retinoic acid. *Oncology*, 1996;53:334-340
- Xia ZS, Zhu ZH, He SG. Effects of ATRA and 5-Fu on growth and telomerase activity of xenografts of gastric cancer in nude mice. *Shijie Huaren Xiaohua Zazhi*, 2000;8:674-677
- Campbell Thompson M, Lauwers GY, Reyher KK, Cromwell J, Shiverick KT. 17Beta-estradiol modulates gastroduodenal preneoplastic alterations in rats exposed to the carcinogen N-methyl-N'-nitro-nitrosoguanidine. *Endocrinology*, 1999;140: 4886-4894
- Chen ZY, Yan MX, Xiang BK. Effect of Welcome on experimental gastric precancerous lesions in rats. *World J Gastroenterol*, 2000;6(Suppl 3):22
- Farinati F, Rugge M, Mario FD, Valiante F, Baffa R. Early and advanced gastric cancer in the follow-up of moderate and severe gastric dysplasia patients. *A prospective study. Hum Pathol*, 1993; 25:261-264
- Rugge M, Farinati F, Baffa R, Sonogo F, Mario FD, Leandro G, Valiante F. Gastric epithelial-dysplasia in the natural history of gastric cancer: a multicenter prospective follow-up study. *Gastroenterology*, 1994;107:1288-1296
- Gu QL, Li NL, Zhu ZG, Yin HR, Lin YZ. A study on arsenic trioxide inducing in vitro apoptosis of gastric cancer cell lines. *World J Gastroenterol*, 2000;6:435-437
- Sun ZX, Ma QW, Zhao TD, Wei YL, Wang GS, Li JS. Apoptosis induced by norcantharidin in human tumor cells. *World J Gastroenterol*, 2000;6:263-265
- Lauwers GY, Scott GV, Karpeh MS. Immunohistochemical evaluation of bcl-2 protein expression in gastric adenocarcinomas. *Cancer*, 1995;75:2209-2213
- Lin J, Shen JK, Yu SX, Dai J, Bo AH, Yao XX. P21 and P53 protein expression in intestinal metaplasia and dysplasia of gastric mucosa. *Xin Xiaohuabingxue Zazhi*, 1997;5:711-712
- Dai J, Yu SX, Qi XL, Bo AH, Xu YL, Guo ZY. Expression of bcl-2 and c-myc protein in gastric carcinoma and precancerous lesions. *World J Gastroenterol*, 1998;4(Suppl 2):84-85
- Soslow RA, Remotti H, Baergen RN, Altorki NK. Suppression of apoptosis does not foster neoplastic growth in Barrett's esophagus. *Mod Pathol*, 1999;12:239-250
- Schweizer J, Goertler K. Synthesis in vivo of keratin polypeptides directed by mRNA isolation from newborn and adult mouse epidermis. *Eur J Biochem*, 1980;112:243-249
- Chomczynski P, Sacchi N. Single-step method of RNA isolation by acid guanidinium thiocyanate-phenol-chloroform extraction. *Anal Biochem*, 1987;162:156-159
- Sizemore N, Cox AD, Barnard JA, Oldham SM, Reynolds ER, Der CJ, Coffey RJ. Pharmacological inhibition of Ras-transformed epithelial cell growth is linked to down-regulation of epidermal growth factor-related peptides. *Gastroenterology*, 1999;117:567-576
- Aviv H, Leder P. Purification of biologically active globin messenger RNA by chromatography on oligothymidylic acid-cellulose. *Proc Natl Acad Sci USA*, 1972;69:1408-1412
- Sambrook J, Fritsch EF, Maniatis T. Transfer of denatured RNA to nitrocellulose filters. In: Rapley R, Manning DL. Vol.1. Humana Press, NY, 1989;7:46
- Takahashi M, Ota S, Shimada E. Hepatocyte growth factor is the most potent endogenous stimulant of rabbit gastric epithelia cell proliferation and migration in primary culture. *J Clin Invest*, 1995; 95:1994-2003
- Nakamura K, Sugano H, Takagi K, Fuchigami A. Histopathological study on early carcinoma of stomach: criteria for diagnosis of atypical epithelium. *Gann*, 1966;57:613-620
- Nagayo T. Histological diagnosis of biopsied gastric mucosae with special reference to that of borderline lesion. *Gann Monogr*, 1971; 11:245-256
- Grundmann E. Histologic types and possible initial stages in early gastric carcinoma. *Beitr Path Bd*, 1975;154:256-280
- Serck-Hansen A. Precancerous lesions of the stomach. *Scand J Gastroenterol*, 1979;14(Suppl 54):104-109
- Morson BC, Sobin LH, Grundmann E, Johansen A, Nagayo T, Serck-Hanssen A. Precancerous conditions and epithelial dysplasia in the stomach. *J Clin Pathol*, 1980;33:711-721
- Lauwers GY, Shimizu M, Correa P, Riddell RH, Kato Y, Lewin KJ, Yamabe H, Sheahan DG, Lewin D, Sipponen P, Kubilis PS, Watanabe H. Evaluation of gastric biopsies for neoplasia differences between Japanese and Western Pathologists. *Am J Surg*, 1999;23:

- 511-518
- 33 Genta RM, Rugge M. Gastric precancerous lesions: heading for an international consensus. *Gut*, 1999;45(Suppl 1):15-18
- 34 Koktysz R, Zielinski KW, Kulig A. Grading of gastric epithelial dysplasia. An interobserver study and analysis of diagnostic criteria. *Pol J Pathol*, 1997;48:5-14
- 35 Genta RM. Atrophy, metaplasia and dysplasia: are they reversible? *Ital J Gastroenterol Hepatol*, 1998;30(Suppl 3):S324-S325
- 36 Anti M, Armuzzi A, Gasbarrini G. Epithelial cell turnover and apoptosis. *Ital J Gastroenterol Hepatol*, 1998;30(Suppl 3):S276-S278
- 37 Xu AG, Li SG, Liu JH, Nong JG, Jiang P, Gan AH. Correlation between apoptosis and proliferation in gastric pre-carcinoma. *Zhonghua Yixue Zazhi*, 1999;79:185-186
- 38 Xu AG, Li SG, Liu JH, Gan AH. The function of apoptosis and protein expression of bcl-2, p53 and C-myc in the development of gastric cancer. *World J Gastroenterol*, 2000;6(Suppl 3):27
- 39 Clarke MR, Safatle AV, Ribeiro U, Sakai P, Reynolds JC. Bcl-2 protein expression in gastric remnant mucosa and gastric cancer 15 or more years after partial gastrectomy. *Mod Pathol*, 1997;8:114-119
- 40 Saegusa W, Takano Y, Okayasu I. Bcl-2 expression and its association with cell kinetic in human gastric carcinoma and intestinal metaplasia. *J Cancer Res Clin Oncol*, 1995;121:357-363
- 41 Lauwers GY, Scott GV, Hendricks J. Immunohistochemical evidence of aberrant bcl-2 protein expression in gastric epithelial dysplasia. *Cancer*, 1994;73:2900-2904
- 42 Liu HF, Liu WW, Fang DC, Men RP. Expression of bcl-2 protein in gastric carcinoma and its significance. *World J Gastroenterol*, 1998;4:228-230
- 43 Liu HF, Liu WW, Fang DC, Men RP. Relationship between Fas antigen expression and apoptosis in human gastric carcinoma and adjacent noncancerous tissues. *Huaren Xiaohua Zazhi*, 1998;6:321-322
- 44 Nett MA, Cerretti DP, Berson DR, Seavitt K, Gilbret DJ, Jenkins NA, Copeland NG, Black RA, Chaplin DD. Molecular cloning of the murine IL-1  $\beta$ -converting enzyme cDNA. *J Immunol*, 1992;149:3254-3259
- 45 Miura M, Zhu H, Rotello R. Induction of apoptosis in fibroblasts by IL-1  $\beta$ -converting enzyme, a mammalian homolog of the *C. elegans* cell death gene *ced-3*. *Cell*, 1993;75:6553
- 46 Cohen JJ. Exponential growth in apoptosis. *Immunol Today*, 1995;16:346-355
- 47 Feng YL, Zhang QX, Li SL. Combined expression of gastrointestinal hormone SP and anti-apoptosis gene Bcl-2 in gastric carcinoma. *World J Gastroenterol*, 2000;6(Suppl 3):22
- 48 Jiang SY, Shen SR, Shyu RY, Yu JC, Harn HJ, Yeh MY, Lee MM, Chang YC. Expression of nuclear retinoid receptors in normal, premalignant and malignant gastric tissues determined by *in situ* hybridization. *Br J Cancer*, 1999;80:206-214

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