

Chemopreventive effect of oltipraz on AFB₁-induced hepatocarcinogenesis in tree shrew model

Yuan Li¹, Jian Jia Su¹, Liu Liang Qin¹, Chun Yang¹, Dan Luo¹, Ke Chen Ban¹, TW Kensler² and BD Roebuck³

Subject headings hepatocellular carcinoma; tupajidae; aflatoxin B₁; hepatitis B virus; incidence; carcinogens, environmental

Li Y, Su JJ, Qin LL, Yang C, Luo D, Ban KC, Kensler TW, Roebuck BD. Chemopreventive effect of oltipraz on AFB₁-induced hepatocarcinogenesis in tree shrew model. *World J Gastroentero*, 2000; 6(5):647-650

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the major cancers in the world with a mortality of more than 250 000 cases yearly. More than 137 000 cases of HCC were diagnosed each year in China, which account approximately for more than 40 percent of the total number in the world. HCC has become the second major cause of death for cancer in China since 1990, and its annual mortality is expected to be 21.2 cases per 100 000 population in the year 2000. Even though progresses have been achieved for HCC diagnosis and treatment, its 5-year mortality is still higher than 95 percent^[1-3].

The prevalence of HCC is quite different among different areas around the world^[4,5]. It is considerably high in South-East Asia and sub-Saharan Africa, particularly in some southern and eastern regions inside China such as Fusui County in Guangxi Zhuang Autonomous Region and Qidong City in Jiangsu Province^[6-9]. The standardized incidence of HCC in these high-risk regions may exceed 100 cases per 100 000 of population^[10]. The obvious difference in geographic distribution of HCC indicates that there must be environmental factors for its pathogenesis.

Aflatoxin B₁ (AFB₁), which is produced by some strains of *Aspergillus flavus*, is a potent hepatotoxin and hepatocarcinogen^[11,12], and is considered as a major cause of HCC in some regions^[13-18]. It has also been postulated that

chronic infection with hepatitis B virus (HBV) in combination with exposure to AFB₁ in the diet may contribute to the extraordinary high risk of human HCC in some areas. Actually, two case-control studies in Shanghai have demonstrated a strong interaction between HBV and AFB₁ for risk of HCC^[14,15]. A similar chemical-viral interaction has been observed in Taiwan^[18-20]. The synergism between virus and mycotoxic carcinogen for the development of human HCC suggests that reduction in both risk factors may bring important public health consequences.

The concept of chemoprevention of cancer is over 40 years old and a number of works have been done in this field^[21,22]. Looking for effective and safe reagents against AFB₁ and/or other HCC related risk factors is one of the most important chemopreventive strategies for HCC^[23-26]. Green tea was identified years ago as one of the effective chemopreventive reagents against HCC through a series of animal experiments as well as a clinical trial^[27,28]. Recently oltipraz, another preventive agent which was previously described as a potent inhibitor of AFB₁ induced hepatocarcinogenesis in rat^[29-33], has been shown to inhibit the bioactivation of aflatoxin and enhance its detoxification in a clinical trial^[34-37] as well as in human hepatocytes in primary culture^[38]. Meanwhile, a universal vaccination program against HBV that started a decade ago now results in lower rates of HCC in children^[39]. An experimental model to test the synergistic effect of these two agents and their prevention, therefore, is needed.

RESEARCH ON TREE SHREW MODEL OF HEPATOCARCINOGENESIS

Tree shrew (*Tupaia spp.*) is a kind of small, squirrel-like mammals. Formerly it was considered to belong to the Primate order; currently it is classified into a separate order Scandentia and is supposed to be more closely related to human being than rodents^[40,41]. They have been used in biomedical researches since as early as the 1960s. Many researches have been done on its visual and nervous systems. In 1976, however, Reddy *et al*^[42] successfully induced liver cancer in tree shrew by AFB₁. Yan *et al*^[43,44] reported that tree shrews can be infected with HBV and they successfully used this HBV-infected tree shrew model for liver cancer

¹Department of Pathology, Guangxi Cancer Institute, Nanning 530027, China

²Department of Environmental Health Sciences, Johns Hopkins School of Hygiene and Public Health, Baltimore, MD 21205, USA

³Department of Pharmacology and Toxicology, Dartmouth Medical School, Hanover, NH 03755, USA

Dr. Yuan Li, received master degree in 1985 from Guangxi Medical University, associate professor of experimental pathology, majoring in liver cancer, having 20 papers published.

Project supported in part by USPHS grant CA-39416.

Correspondence to: Dr. Yuan Li, Department of Pathology, Guangxi Cancer Institute, Nanning, 530027, China

Tel. 0086-771-5331100

Email. li-yuan@public.nn.gx.cn

Received 2000-06-13 Accepted 2000-06-29

research. Recently Walter *et al*^[45] reported their *in vivo* and *in vitro* study results from tree shrews infected with HBV. Yan and Li reported a significantly higher incidence of HCC in tree shrews both infected with human HBV and exposed to AFB₁ than with either agent alone^[46,47]. Thus, this tree shrew model appears to closely mirror the most common causative factors of human HCC in some prevalent regions. Furthermore, with the exception of the chimpanzee, tree shrew is the only known animal that can be infected with human HBV. Therefore, the application of tree shrew in research related to liver cancer and hepatitis is receiving increasing attentions and a number of works have been published^[48-52].

Because of the difficulties in raising tree shrews artificially, most of the tree shrews used so far for research in China are captured individually from Yunnan Province. The drawback of using tree shrews captured in the wild for animal experiments is that their age, health status and reproductive history are unknown. In an attempt to avoid this drawback, we have conducted a preliminary experiment on rearing tree shrews and a promising result was obtained^[53].

RESEARCH ON THE PREVENTIVE EFFECT OF OLTIPRAZ IN TREE SHREW

In order to study the preventive effect of oltipraz on AFB₁ by animal models other than rodents, a short-term experiment was conducted on tree shrews.

Male and female adult tree shrews (*Tupaia belangeri chinensis*) were purchased from the Kunming Institute of Zoology (Yunnan Province, P.R. China). Their body weights ranged from 100g to 160g. Upon arrival, 1mL blood was collected from each animal and tested for HBV markers (HBsAg, anti-HBsAg, anti-HBcAg) and ALT. The tree shrews that were negative for these markers of HBV infection and that had ALT value below 55 units were divided into 4 groups with 6 or 7 animals for each group. Group A: normal control; group B: AFB₁ alone; group C: AFB₁+oltipraz daily; group D: AFB₁+oltipraz weekly. All the tree shrews were allowed 1 week to acclimatize to the facilities prior to the experiment. They were housed in separate, suspended, stainless steel wire cages under controlled environmental conditions with a 12-hour light/dark photoperiod. They had free access to tap water and a natural ingredient diet.

The experimental design is presented schematically in Figure 1. Tree shrews in groups B, C and D were given AFB₁ (400μg/kg b.w./day in liquid milk) daily beginning at one week after the experiment started and continued for 4 weeks. One week before giving AFB₁, tree shrews in group C and D were respectively given oltipraz (0.5mmol/kg b.w.) daily or weekly, by gavage in a saturated solution of sucrose for 5 weeks. Blood

samples and 24-hour urine samples were collected once a week from each animal throughout the experiment. At the termination of the 9-week experiment, tree shrews were killed by cervical dislocation. Three blocks of liver tissue were taken from each animal. Serial sections from each block were stained histochemically for r-glutamyl transpeptidase (γ-GT)^[54] and HE respectively. The γ-GT positive liver cells were counted with a nest-ruler under microscope. The results were analyzed by the medical statistics analyzing software PEMS that was designed by West China University of Medical Sciences. The levels of aflatoxin-albumin adducts in serum samples were determined by radioimmune assay^[55] and the levels of Aflatoxin-N⁷-guanine adducts in urine samples were assayed by HPLC^[29].

No γ-GT positive liver cell focus, a postulated precancerous marker^[54-56] was observed in any liver of the variously treated tree shrews in our study. However, different numbers of γ-GT positive liver cells, which scattered mainly around the portal spaces, were observed in each group. Even though the distribution patterns of these cells were similar among the 4 groups, the number was quite different. Groups B and D had obviously less γ-GT positive cells than groups A and C (Table 1).

γ-GT normally exists in embryonic liver cell in human being and rat. In adult rat, it exists only in some cells around portal spaces^[57] but can be re-expressed by mature hepatocytes during the recovery process after liver damage^[58]. In this study a number of γ-GT positive hepatocytes presented in periportal regions in the normal control group. On the contrary, the number of γ-GT positive hepatocytes of the same sites was markedly reduced in the AFB₁ treated B group. This phenomenon is fairly consistent with the findings on AFB₁ induced damage in rat, in which the periportal hepatocytes are the major targets of AFB₁. As shown in the same table, the number of γ-GT positive hepatocytes in group C was strikingly similar to the normal control group. This result indicates strongly the preventive action of oltipraz against AFB₁ toxicity. The apparent ineffectiveness of oltipraz in group D is most possibly due to its inadequate dose^[59]. These results might indicate that oltipraz has the preventive dose-related effect on AFB₁.

Insufficient duration and/or insufficient dosage of AFB₁ treatment may result in that no separate focus of γ-GT positive liver cell formed at the end of this 9-week experiment^[60]. However, the decrease of γ-GT positive hepatocytes in the periportal regions may be an early marker for the damage induced by AFB₁.

The levels of both aflatoxin-albumin adducts in serum samples and aflatoxin-N⁷-guanine adducts in urine samples of the tree shrews were also significantly affected by oltipraz. Following daily

exposures to AFB₁, the levels of serum aflatoxin-albumin adducts in group B increased rapidly over 2 weeks to reach a plateau that did not diminish until cessation of AFB₁ exposure. In group C however, oltipraz attenuated the aflatoxin-albumin adducts significantly ($P < 0.05$) with a median reduction of 80%. The mean levels of aflatoxin-N⁷-guanine (ng/mg creatinine) in the urine samples collected at week 5 were 6.34 ± 2.04 and 0.47 ± 0.13 in groups B and C respectively. This 93% decrease represented a statistically significant difference ($P < 0.05$). These results were reported in detail in another article^[61]. The major mechanism of oltipraz's chemopreventive effect is probably through inducing the activities of cytochrome P450 system and phase 2 enzymes such as glutathione transferases, epoxide hydrolase, etc, as reported by Langouet *et al.*^[38] and Fahey *et al.*^[62].

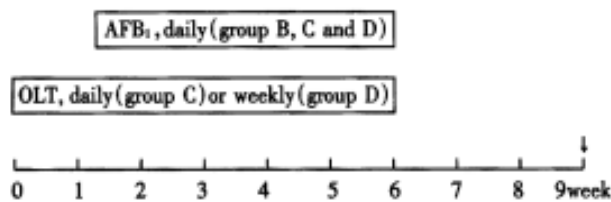


Figure 1 Experimental design for animal treatment. OLT: oltipraz; ↓: all the animals were sacrificed.

Table 1 The number and size of γ -GT positive hepatocyte-group ($\bar{x} \pm s_x$)

Group	Treatment	No./cm ²	mm ² /cm ²	mm ² /No.
A	Normal control	167.29±50.47	2.90±1.01	0.017±0.002
B	AFB ₁	71.92±42.19 ^a	1.14±0.69 ^a	0.015±0.002
C	AFB ₁ +OLT daily	167.10±45.94 ^{c,d}	2.73±0.87 ^{c,d}	0.016±0.001
D	AFB ₁ +OLT weekly	83.66±34.94 ^a	1.33±0.86 ^a	0.015±0.002 ^b

t test: ^a $P < 0.05$, ^b $P < 0.01$, vs group A; ^c $P < 0.01$, vs group B; ^d $P < 0.01$, vs group D.

OLT: oltipraz;

No./cm²: the number of γ -GT positive hepatocyte-group per cm² of liver tissue;

mm²/cm²: mm² of γ -GT positive hepatocyte-group per cm² of liver tissue;

mm²/No.: mm² of γ -GT positive hepatocyte-group per each one.

SUMMARY

Tree shrew is phylogenetically more closely related to human being than rodents. It is susceptible both to HBV infection and AFB₁ intoxication. It is a suitable experimental animal for hepatocarcinogenesis. Attempt for its rearing is promising.

Oltipraz is an effective reagent to protect AFB₁ intoxication. This effect is proved clearly not only by histological examination, but also by reduction of aflatoxin-albumin adducts in serum and aflatoxin-N⁷ guanine adducts in urine.

All of these studies mentioned above provide a foundation for further HCC chemoprevention study

by using tree shrews in the future.

ACKNOWLEDGEMENTS The authors express their appreciation to Drs. Guo-Hua Huang, Chao Ou, Xue-Lan Deng and Hua-Ping Huang for the contributions to the animal experiment. Preliminary accounts of this work were presented at the 1999 Annual Meeting of the American Association for Cancer Research^[63].

REFERENCES

- Skolnick AA. Armed with epidemiologic research, China launches programs to prevent liver cancer. *JAMA*, 1996;276:1458-1459
- Guyton KZ, Kensler TW. Prevention of liver cancer. *Curr Opin Oncol*, 1997;9:492-496
- Li LD, Lu FZ, Zhang SW, Mu R, Sun XD, Huangpu XM, Sun J, Zhou YS, Ouyang NH, Rao KQ, Chen YD, Sun AM, Xue ZF, Xia Y. The alterations of malignant tumors' mortality in China during the 20 years and the forecasting. *Zhonghua Zhong Liu Zazhi*, 1997;19:3-9
- Schafer DF, Sorrell MF. Hepatocellular carcinoma. *Lancet*, 1999;353:1253-1257
- Wild CP, Jiang YZ, Allen SJ, Jansen LAM, Hall AJ, Montesano R. Aflatoxin-albumin adducts in human sera from different regions of the world. *Carcinogenesis*, 1990;11:2271-2274
- Ruan CC, Chen YH, Zhang ZQ. Drinking water and liver cancer. *China Natl J New Gastroenterol*, 1997;3:47-49
- Deng ZL, Ma Y. Aflatoxin sufferer and p53 gene mutation in hepatocellular carcinoma. *World J Gastroenterol*, 1998;4:28-29
- Chen JG. The epidemiologic tendency and forecasting of liver cancer in Qidong (Abstract). *Zhonghua Yufang Yixue Zazhi*, 1996;30:180
- Groopman JD, Zhu JQ, Donahue PR, Pikul A, Zhang LS, Chen JS, Wogan GN. Molecular dosimetry of urinary aflatoxin DNA adducts in people living in Guangxi Autonomous Region, People's Republic of China. *Cancer Res*, 1992;52:45-52
- Yeh FS, Yu MC, Mo CC, Luo S, Tong MJ, Henderson BE. Hepatitis B virus, aflatoxins, and hepatocellular carcinoma in southern Guangxi, China. *Cancer Res*, 1989;49:2506-2509
- Wogan GN. Aflatoxin as a human carcinogen. *Hepatology*, 1999;30:573-575
- Choy WN. A review of the dose response induction of DNA adducts by aflatoxin B-1 and its implications to quantitative cancer risk assessment. *Mutation Res*, 1993;296:181-198
- Wang D, Shi JQ. Overexpression and mutations of tumor suppressor gene p53 in hepatocellular carcinoma. *China Natl J New Gastroenterol*, 1996;2:161-164
- Qian GS, Ross RK, Yu MC, Yuan JM, Gao YT, Henderson BE, Wogan GN, Groopman JD. A follow up study of urinary markers of aflatoxin exposure and liver cancer risk in Shanghai, People's Republic of China. *Cancer Epidemiol Biomark Prevent*, 1994;3:3-10
- Ross RK, Yuan JM, Yu MC, Wogan GN, Qian GS, Tu JT, Groopman JD, Gao YT, Henderson BE. Urinary aflatoxin biomarkers and risk of hepatocellular carcinoma. *Lancet*, 1992;339:943-946
- Chen CJ, Wang LY, Lu SN, Wu MH, You SL, Zhang YJ, Wang LW, Santella RM. Elevated aflatoxin exposure and increased risk of hepatocellular carcinoma. *Hepatology*, 1996;24:38-42
- Wogan GN. Biomarkers for molecular epidemiology of aflatoxin as a risk factor for hepatocellular carcinoma: the essential role of basic science. *CIIT Activities*, 1999;19:4-10
- Wang LY, Hatch M, Chen CJ, Levin B, You SL, Lu SN, Wu MH, Wu WP, Wang LW, Wang Q, Huang GT, Yang PM, Lee HS, Santella RM. Aflatoxin exposure and risk of hepatocellular carcinoma in Taiwan. *Int J Cancer*, 1996;67:620-625
- Lunn RM, Zhang YJ, Wang LY, Chen CJ, Lee PH, Lee CS, Tsai WY, Santella RM. p53 Mutations, chronic hepatitis B virus infection, and aflatoxin exposure in hepatocellular carcinoma in Taiwan. *Cancer Res*, 1997;57:3471-3477
- McGlynn KA, Rosvold EA, Lustbader ED, Hu Y, Clapper ML, Zhou T, Wild CP, Xia XL, Baffoe Bonnie A, Ofori Adjei D, Chen GC, London WT, Shen FM, Buetow KH. Susceptibility to hepatocellular carcinoma is associated with genetic variation in the enzymatic detoxification of aflatoxin B1. *Proc Natl Acad Sci USA*, 1995;92:2384-2387
- Hong WK, Sporn MB. Recent advances in chemoprevention of cancer. *Science*, 1997;278:1073-1077
- Mukhtar H, Ahmad N. Contemporary issues in toxicology: cancer chemoprevention: future holds in multiple agents. *Toxicol Appl Pharmacol*, 1999;158:207-210
- Gu GW, Zhou HG. Traditional Chinese Medicine in prevention of liver

- cancer. *Shijie Huaren Xiaohua Zazhi*, 1999;7:80-81
- 24 Zhou HG, Gu GW. Retinoids preventing liver cancer. *Shijie Huaren Xiaohua Zazhi*, 1999;7:82-83
- 25 Tan W, Lin DX, Xiao Y, Kadlubar FF, Chen JS. Chemoprevention of 2 amino 1 methyl 6 phenylimidazo [4,5-b] pyridine induced carcinogen DNA adducts by Chinese cabbage in rats. *World J Gastroentero*, 1999;5:138-142
- 26 Yuan JH, Zhang RP, Zhang RG, Guo LX, Wang XW, Luo D, Xie Y, Xie H. Growth inhibiting effects of taxol on human liver cancer *in vitro* and in nude mice. *World J Gastroentero*, 2000;6:210-215
- 27 Chen ZY, Yan RQ, Qin GZ, Qin LL. Effects of six edible plants on the development of AFB 1 induced γ GT positive hepatocyte foci in rat. *Zhonghua Zhongliu Zazhi*, 1987;9:109-111
- 28 Li Y, Qin GZ, Qin LL, Duan XX, Yan RQ. A series of experimental animal study of green tea on prevention of hepatic cancer. *Sichuan Zhongliu angzhi*, 1997;10:1-4
- 29 Kensler TW, Egner PA, Dolan PM, Groopman JD, Roebuck BD. Mechanism of protection against aflatoxin tumorigenicity in rats fed 5 (2 pyrazinyl) 4 methyl 1,2 dithiol 3 thione (oltipraz) and related 1,2 dithiol 3 thiones and 1,2 dithiol 3 ones. *Cancer Res*, 1987;47:4271-4277
- 30 Roebuck BD, Liu YL, Rogers AE, Groopman JD, Kensler TW. Protection against aflatoxin B-1 induced hepatocarcinogenesis in F344 rats by 5 (2 pyrazinyl) 4 methyl 1,2 dithiole 3 thione (oltipraz): predictive role for short term molecular dosimetry. *Cancer Res*, 1991;51:5501-5506
- 31 Kensler TW, Gange SJ, Egner PA, Dolan PM, Mućkoz A, Groopman JD, Rogers AE, Roebuck BD. Predictive value of molecular dosimetry: Individual versus group effects of oltipraz on aflatoxin albumin adducts and risk of liver cancer. *Cancer Epidemiol Biomark Prevent*, 1997;6:603-610
- 32 Maxuitenko YY, Curphey TJ, Kensler TW, Roebuck BD. Protection against aflatoxin B-1 induced hepatic toxicity as a short term screen of cancer chemo preventive dithiolethiones. *Fundament Appl Toxicol*, 1996;32:250-259
- 33 Maxuitenko YY, Libby AH, Joyner HH, Curphey TJ, MacMillan DL, Kensler TW, Roebuck BD. Identification of dithiolethiones with better chemopreventive properties than oltipraz. *Carcinogenesis*, 1998;19:1609-1615
- 34 Wang JS, Shen X, He X, Zhu YR, Zhang BC, Wang JB, Qian GS, Kuang SY, Zarba A, Egner PA, Jacobson LP, Muoz A, Helzlsouer KJ, Groopman JD, Kensler TW. Protective alterations in phase 1 and 2 metabolism of aflatoxin B-1 by oltipraz in residents of Qidong, People's Republic of China. *J Natl Cancer Inst*, 1999;91:347-354
- 35 Wang JS, Qian GS, Zarba A, He X, Zhu YR, Zhang BC, Jacobson L, Gange SJ, Muoz A, Kensler TW, Groopman JD. Temporal patterns of aflatoxin albumin adducts in hepatitis B surface antigen positive and antigen negative residents of Daxin, Qidong County, People's Republic of China. *Cancer Epidemiol Biomark Prevent*, 1996;5:253-261
- 36 Kensler TW, He X, Otieno M, Egner PA, Jacobson LP, Chen B, Wang JS, Zhu YR, Zhang BC, Wang JB, Wu Y, Zhang QN, Qian GS, Kuang SY, Fang X, Li YF, Yu LY, Prochaska HJ, Davidson NE, Gordon GB, Gorman MB, Zarba A, Enger C, Muñoz A, Helzlsouer KJ, Groopman JD. Oltipraz chemoprevention trial in Qidong, People's Republic of China: modulation of serum aflatoxin albumin adduct biomarkers. *Cancer Epidemiol Biomark Prevent*, 1998;7:127-134
- 37 Kensler TW, Groopman JD, Sutter TR, Curphey TJ, Roebuck BD. Development of cancer chemopreventive agents: Oltipraz as a paradigm. *Chem Res Toxicol*, 1999;12:113-126
- 38 Langou tS, Coles B, Morel F, Becquemont L, Beaune P, Guengerich FP, Ketterer B, Guillouzo A. Inhibition of CYP1A2 and CYP3A4 by oltipraz results in reduction of aflatoxin B-1 metabolism in human hepatocytes in primary culture. *Cancer Res*, 1995;55:5574-5579
- 39 Chang MH, Chen CJ, Lai MS, Hsu HM, Wu TC, Kong MS, Liang DC, Shau WY, Chen DS. Universal hepatitis B vaccination in Taiwan and the incidence of hepatocellular carcinoma in children. *N Engl J Med*, 1997;336:1855-1859
- 40 Wang YX, Li CY, Ma SL. Chapter 1. The classification and ecology of Chinese tree shrew. In: Peng YZ, Ye ZZ, Zou RJ, Wang YX, Tian BP, Ma YY, Shi LM, eds. *Biology of Chinese tree shrews*. Kunming, China: Yunnan Science and Technology Press, 1991:21-69
- 41 Bearder S, Pitts RS. Chapter 36, Prosimians and tree shrews. In: Trevor B. Poole eds. *The UFAW Handbook on the Care and Management of Laboratory Animals*. Sixth edition. Avon, Great Britain: Bath Press, 1987:551-567
- 42 Reddy JK, Svoboda DJ, Rao MS. Induction of liver tumors by aflatoxin B-1 in the tree shrew (*Tupaia glis*), a nonhuman primate. *Cancer Res*, 1976;36:151-160
- 43 Yan RQ, Su JJ, Huang DR, Gan YC, Yang C, Huang GH. Human hepatitis B virus and hepatocellular carcinoma I. Experimental infection of tree shrews with hepatitis B virus. *J Cancer Res Clin Oncol*, 1996;122:283-288
- 44 Yan RQ, Su JJ, Huang DR, Gan YC, Yang C, Huang GH. Human hepatitis B virus and hepatocellular carcinoma II. Experimental induction of hepatocellular carcinoma in tree shrews exposed to hepatitis B virus and aflatoxin B1. *J Cancer Res Clin Oncol*, 1996;122:289-295
- 45 Walter E, Keist R, Niederst B, Pult I, Blum HE. Hepatitis B virus infection of Tupaia hepatocytes *in vitro* and *in vivo*. *Hepatology*, 1996;24:1-5
- 46 Yan RQ, Su JJ, Huang DR, Yang C, Huang GH. A study on primary liver cancer in tree shrews induced by human hepatitis B virus and aflatoxin B1. *Zhonghua Binglixue Zazhi*, 1989;18:19-21
- 47 Li Y, Su JJ, Qin LL, Yang C, Ban KC, Yan RQ. Synergistic effect of hepatitis B virus and aflatoxin B1 in hepatocarcinogenesis in tree shrews. *Ann Acad Med Singapore*, 1999;28:67-71
- 48 Li Y, Su JJ, Yan RQ, Qin LL, Yang C, Ban KC, Duan XX, Huang GH. Expression of insulin like growth factor II (IGF-II) protein during tree shrews' hepatocarcinogenesis differently induced by AFB-1 and/or HBV. *Guangxi Yike Daxue Xuebao*, 1999;16:395-398
- 49 Su JJ, Qin GZ, Yan RQ, Huang DR, Yang C, Huang GH, Lotlikar PD. Expression of p53 gene in hepatocellular carcinomas induced by aflatoxin B1 with or without human hepatitis B virus in tree shrews. *Experiment Mole Med*, 1997;29:177-182
- 50 Su JJ, Qin GZ, Yan RQ, Huang DR, Yang C, Lotlikar PD. The expression of insulin like growth factor II, hepatitis B virus X antigen and p21 in experimental hepatocarcinogenesis in tree shrews. *Ann Acad Med Singapore*, 1999;28:62-66
- 51 Ban KC, Su JJ, Yang C, Qin LL, Li Y, Huang GH, Luo XL, Duan XX, Yan RQ. Expression of ras gene in experimental hepatocarcinogenesis in tree shrews. *Chin J Cancer Res*, 1999;11:23-25
- 52 Qin LL, Su JJ, Li Y, Yang C, Ban KC, Yan RQ. Expression of IGF-II, p53, p21 and HBxAg in precancerous events of hepatocarcinogenesis induced by AFB-1 and/or HBV in tree shrews. *World J Gastroentero*, 2000;6:138-139
- 53 Li Y, Baumgartner K, MacMillan D, Roebuck BD. Hand rearing of tree shrew. *Dongwuxue Zazhi*, in press
- 54 Li Y, Yan RQ, Qin GZ, Qin LL, Duan XX. Reliability of a short term test for hepatocarcinogenesis induced by aflatoxin B-1. *IARC Sci Publ*, 1991;105:431-433
- 55 Li Y, Su JJ, Qin LL, Yang C, Luo D, Ban KC, Huang GH, Ou C, Kensler T, Roebuck B. Chemopreventive effect of oltipraz (OLT) on AFB1 induced precancerous changes in liver of tree shrews. *Aizheng*, 1999;18:34-36
- 56 Zhu HZ, Zhang XL, Chen YS. Expression of glutathione S transferase placental mRNA in hepatic preneoplastic lesions in rats. *World J Gastroentero*, 1998;4:38-40
- 57 Kitagawa T, Imai F, Sato K. Re-elevation of γ -glutamyl transpeptidase activity in periportal hepatocytes of rats with age. *Gann*, 1980;71:362-366
- 58 Kitten O, Ferry N. Mature hepatocytes actively divide and express gamma glutamyl transpeptidase after D galactosamine liver injury. *Liver*, 1998;18:398-404
- 59 Butler WH. Acute toxicity of aflatoxin B-1 in rats. *Br J Cancer*, 1964;18:756-762
- 60 Kalengayi MMR, Ronchi G, Desmet VJ. Histochemistry of gamma-glutamyl transpeptidase in rat liver during aflatoxin B-1 induced carcinogenesis. *J Natl Cancer Inst*, 1975;55:579-588
- 61 Li Y, Su JJ, Qin LL, Egner PA, Wang JS, Groopman JD, Kensler TW, Roebuck BD. Reduction of aflatoxin B-1 adduct biomarkers by oltipraz in the tree shrew (*Tupaia belangeri chinensis*). *Cancer Letters*, 2000;154:79-83
- 62 Fahey JW, Zhang Y, Talalay P. Broccoli sprouts: An exceptionally rich source of inducers of enzymes that protect against chemical carcinogens. *Proc Natl Acad Sci USA*, 1997;94:10367-10372
- 63 Li Y, Su JJ, Qin LL, Egner PA, Wang JS, Groopman JD, Kensler TW, Roebuck BD. Modulation of aflatoxin B-1 adducts by oltipraz in the tree shrew. *Proc Am Assoc Cancer Res*, 1999;40:261