Original Article

Correlation between polymorphism of endothelial nitric oxide synthase and avascular necrosis of femoral head

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Abstract: Objective: We analyzed the correlation between mutation in intron 4 and exon 7 of endothelial nitric oxide synthase (eNOS) and avascular necrosis of femoral head (ANFH). Method: A total of 260 ANFH cases without history of hip joint injuries were diagnosed and subject to staging according to Ficat standard, with 262 health subjects as control. Venous blood was collected to extract genome DNA, which was then amplified by PCR. The polymorphism of 27 bp repeat sequence in intron 4 and G894T polymorphism in exon 7 of eNOS gene was detected. Results: The b/b, b/a and a/a genotype frequency of intron 4 was 77.7%, 19.2% and 3.1% in ANFH group, respectively, and that in the control group was 58.0%, 32.8% and 9.2%, respectively. The b allele frequency in ANFH group was obviously higher than that in the control (P<0.0001). The frequency of 894 G/G wild type, G/T heterozygote and T/T homozygote in eNOS exon 7 was analyzed by PCR-RLFP: 65.4%, 26.5% and 8.1% in ANFH group, and 46.2%, 37.8% and 16% in normal control, respectively. The frequency of TT genotype in ANFH was obviously higher than that in the control group (P<0.001). Conclusion: Polymorphism of eNOS was correlated with ANFH.

Keywords: Polymorphism/genetics, endothelial cells/enzymology, nitric oxide synthase (NOS)/genetics, ischemia, necrosis of femoral head/genetics

Introduction

Avascular necrosis of femoral head (ANFH) is a common joint disease in China which affects millions of people. Pathogenic factors of ANFH include genetics, injuries, hormone abuse, alcohol abuse and coagulation factors [1-3]. These factors can lead to reduced blood supply to femoral head, osteonecrosis and finally the collapse of bone cortex and osteochondral destruction. The future direction of AHFH treatment is early discovery and early prevention, so as to reverse or slow down the progression.

Endothelial nitric oxide synthase (eNOS) is responsible for the production of nitric oxide (NO) and reduces vascular tension and thrombosis risk. Recent studies have shown that two polymorphic loci of eNOS gene, namely, 27 bp repeat sequence in intron 4 and G894y poly-

morphism in exon 7, are related to coronary heart disease, hypertension and nephropathy [4-6]. Though the connections between eNOS polymorphism and vascular diseases are established, very few reports are published concerning the relationship between eNOS gene and ANFH [7]. We observed the effects of 27 bp repeat sequence in intron 4 and G894y polymorphism in exon 7 of eNOS gene on the incidence of ANFH. The polymorphism of the two loci can be used for the screening of high-risk groups.

Materials and methods

General data

ANFH patients treated at Department of Orthopedics of our hospital from January 2008 to December 2014 were selected. Excluding

those with a history of hip joint injuries, 260 cases were finally recruited (157 males and 103 females, aged 55.4±10.2 years old). By pathogenic factors, the cases were divided into idiopathic ANFH, hormone-induced ANFH and alcohol-induced ANFH. All cases were diagnosed and subject to staging according to Ficat standard and relevant staging criteria. For control group, 262 healthy people receiving physical examination at our hospital were included (161 males and 101 females, aged 55.9±10.5 years old). All cases signed informed consent. The tube containing EDTA as anticoagulant was used to collect 5 mL of venous blood from each case. Peripheral blood monocular cells (PBMC) were isolated by Ficoll method and cryopreserved.

Equipment and reagents

Milli-Q Ultrapure Water Systems (Millipore Corporation, USA), BD202 Balance (Metfler-Toledo International Inc., Germany), UV spectro-photometer (Eppendorf, Germany), ABI PRISM 310 Genetic Analyzer (PE Corporation, USA), 1640 culture medium (GIBCO, USA); DNeasy Tissue Kit (QIAGEN, Germany), RNase (Roche, Switzerland), Gel Extraction Kit (MACHER-EY-NAGEL, USA), BigDye Sequencing Kit (Applied Biosystems, USA).

Experimental methods

DNA extraction and purification detection: Frozen PBMC specimens were thawed and used for genome DNA extraction with DNeasy Tissue Kit (QIAGEN, Germany). Cell lysis buffer was added and cell membrane was disrupted by oscillation. For each specimen 100 μL of RNaseA was added. The specimen was passed through CB6 column and TB buffer was used for elution. Genome DAN extraction was carried out according to the kit instruction. The recovered DNA fragments were detected for concentration and purity by 0.8% agarose gel electrophoresis and ultraviolet spectroscopy. Images were taken using FR-200 UV-Vis detector for quantification by referring to DNA markers.

Synthesis of specific primers and PCR amplification: Genbank was searched for base sequences of 27 bp repeat sequence in intron 4 and G894T polymorphism in exon 7 of eNOS gene. Primers were designed using Primer 5.0 (see **Table 1**). Conditions of PCR amplification

were as follows: denaturation at 95°C for 15 min, at 95°C for 30 s, annealing at 55°C for 30 s, extension at 72°C for 1 min, 33 cycles, final extension at 72°C for 5 min. The PCR products were separated by agarose gel electrophoresis and the fragments were recovered using Gel Extraction Kit (MACHEREY-NAGEL).

Agarose gel electrophoresis of DNA: The 1.5% agarose gel was filled into the tank and solidified, and the specimen was loaded. Electrophoresis was carried out for 2 h at the voltage of 3-5 V/cm, and then the gel was removed and stained with ethidium bromide. Electrophoresis patterns were obtained at the wave length of 254 nm.

Restriction enzyme digestion for exon 7: The system of restriction enzyme digestion consisted of the followings: 1 μ L Ban II, 2 μ L 10 X Buffer, DNA \leq 1 μ g, and sterile water up to 20 μ L. The reaction temperature was 37°C. After digestion at 37°C for 1 h in the above 20 μ L reaction system, 10 μ L of product of digestion was collected and mixed with 6 X loading buffer. Agarose gel electrophoresis was carried out at the 100 V for 30-60 min.

Sequencing of intron 4: PCR products were purified by using sodium acetate/ethanol method and pretreated before electrophoresis. Capillary tube was mounted according to the instruction and the position of the tube was calibrated. Gel filling was performed manually and the sequential files were established. Then the gel was automatically filled into the capillary tube. After pre-electrophoresis at 1.2 kV for 5 min, sample loading was performed automatically according to the programmed sequence. This was followed by pre-electrophoresis (1.2 kV, 20 min) and electrophoresis at 7.5 kV for 2 h. When the electrophoresis was over, the equipment would be cleaned automatically. The next round of gel filling, pre-electrophoresis and electrophoresis began for the new specimen. The total electrophoresis time for each specimen was 2.5 h. The color sequencing chromatograms were printed automatically.

Statistical processes: All statistical analyses were performed using SPSS 18.0. Data were compared with X² test or t test, and P<0.05 was considered as statistically significant.

Table 1. The primer sequences of the two loci

Locus	Sense primer	Antisense primer			
27 bp repeat polymorphism	5'AGGCCCTATGGTAGTGCCTT3'	5'TCTCTTAGTGCTGTGGTCAC3'			
G894T polymorphism	5'AAGGCAGGAGACAGTGGAGGT3'	5'CCCAGTCAATCCCTTTGGTGCTCA3'			

Table 2. Characteristics of the participants

Groups	N	Age (years)	Gender	BMI Kg/m ²	GLU	TG	TC	HDL-C	LDL-C
			(M/F)	DIVII Ng/III	(mmol/L)	(mmol/L)	(mmol/L)	(mmol/L)	(mmol/L)
ANFH group	260	55.4±10.2	157/103	24.1±3.3	5.9±1.6	2.1±1.3	5.5±2.7	1.7±0.9	2.7±1.5
Control group	262	55.9±10.5	161/101	24.4±3.4	5.6±1.3	1.7±1.4	4.9±2.6	1.6±0.8	2.6±1.7
Р		0.432	0.417	0.112	0.652	0.079	0.088	0.184	0.559

Note: BMI=Body mass index; GLU=Glucose; TC=Total cholesterol; LDL-C=Low-density lipoprotein-cholesterol; LDL-C=High-density lipoprotein-cholesterol; TG=Triglycerides.

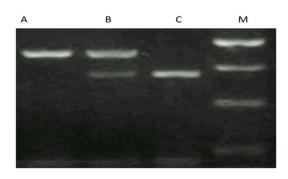


Figure 1. Genotyping results of G894T polymorphism in exon 7. A: TT genotype; B: GT genotype; C: GG genotype; M: DNA marker.

Results

Clinical data of subjects

The course of disease in 260 ANFH cases was 3 months to 23 years, with an average of 110.2±60.1 months. Staging results showed that 38 cases belonged to stage II, 75 to stage III (including transitional cases) and 147 to stage IV. According to etiological classification, 65, 132 and 63 cases were alcohol-induced ANFH, idiopathic ANFH and hormone-induced ANFH, respectively. The characteristics of the patients and the control were shown in the **Table 2**.

Detection of G894T polymorphism in exon 7

The amplified products were hydrolyzed by specific endonucleases. Wild-type DNA was hydrolyzed by Banll into two fragments of unequal length, corresponding to 163 bp and 85 bp bands on the electrophoretograms, respective-

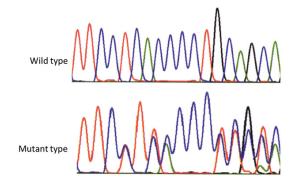


Figure 2. The wild type and mutant type sequences 27 bp repeat polymorphism of NOS gene.

ly. However, mutant DNA could not be hydrolyzed with a length of 248 bp, corresponding to the third band on the electrophoretogram (**Figure 1**).

Polymorphism detection of 27 bp repeat sequence in intron 4 of eNOS gene

Sequencing of Intron 4 of eNOS gene polymorphism indicated that three polymorphisms, which were b/b, b/a and a/a (Figure 2).

Data analysis

Distribution of eNOS alleles in normal people and ANFH cases: The frequencies of intron 4 b allele and 894T allele in exon 7 in ANFH group were obviously higher than those of the control (P<0.001, Table 3).

The frequency of eNOS intron 4 b/a genotype in ANFH group was significantly higher than that of the normal control (P<0.0001). The frequen-

Table 3. Distribution of genotype and alleles between two groups

CND	Cround	N -	Genotype				Allele		
SNP	Groups		a/a	a/b	b/b	Р	а	b	Р
27 bp repeat polymorphism	ANFH group	260	8 (3.1)	50 (19.2)	202 (77.7)	<0.001	66 (12.7)	454 (87.3)	<0.001
	Control group	262	24 (9.2)	86 (32.8)	152 (58.0)		134 (25.7)	390 (74.3)	
			TT	GT	GG		G	Т	
G894T	ANFH group	260	21 (8.1)	69 (26.5)	170 (65.4)	<0.001	111 (21.3)	409 (78.7)	<0.001
	Control group	262	42 (16.0)	99 (37.8)	121 (46.2)		183 (34.9)	341 (65.1)	

cies of G894T polymorphism in exon 7 and G/T genotype in ANFH group were considerably higher than those of the control (P<0.001, **Table 3**).

Discussion

In the present study, we found genetic polymorphisms of eNOS were associated with ANFA in a Chinese population. The risk factors of ANFH include use of steroid hormones, alcohol abuse, trauma and genetics [2, 3, 8]. The cases of ANFH may present no typical symptoms during early onset and there is a lack of practical screening tool. Along with progress of medical sciences, the correlation between ANFH and gene polymorphism has drawn increasing attention. New molecular methods are proposed for treating ANFH. We aimed to study the correlation between the polymorphism of eNOS gene and the risk of ANFH based on the molecular mechanism of ANFH, so as to offer new technique for screening, early diagnosis and intervention.

NO produced by eNOS has a wide range of bioactivity, including anti-oxidative, anti-inflammatory, anticoagulation, antiplatelet aggregation and adhesion [9-12]. eNOS in bones is involved in the regulation of bone turnover. The normal functioning of some hormones such as IGF and estrogen also relies on eNOS [13]. Directly acting on platelets. NO can inhibit platelet aggregation and adhesion and reduce platelet thrombosis formation. NO can facilitate wound healing by promoting angiogenesis [14]. Given these factors, the author believes that eNOS is involved in ANFH pathogenesis. Two pathogenic mutation loci have been found in eNOS gene. One is G894T polymorphism in exon 7 of eNOS gene, leading to the change from glutamic acid to aspartic acid at site 298 of peptide. The other is the mutation of five 27 bp repeat sequences (b allele, wild type) in intron 4 into

four 27 bp repeat sequences and one inverted repeat sequence (a allele, mutant type). It is generally believed that intronic mutation affects mRNA transcription, while exonic mutation affects the function and activity of enzymes [15]. Recently, Tesauro et al. [16] showed that eNOS mutant gene was less stable than that by wild-type gene. Although mutation had no impact on the substrate binding ability and activity of enzyme, the peptide encoded by mutant eNOS gene was more easily hydrolyzed than that by wild-type eNOS gene. As a result, eNOS content was reduced and NO production declined accordingly. Besides, the plasma level of NO in individual with eNOS intron 4a mutation was obviously lower than that with wildtype eNOS gene [17]. That is to say, the mutation of two loci reduced NO content.

The correlations of polymorphism of eNOS with coronary heart disease, hypertension and strokes have been established in many studies [18-21]. We observed that a significantly higher number of ANFH cases carried mutant intron 4 of eNOS gene than normal controls, and this difference was especially prominent in idiopathic ANFH. Since the mutation of intron 4 can reduce eNOS synthesis, it is inferred that intron 4a mutation is one risk factor of ANFH. NO produced by eNOS has a protective effect on femoral bones and is therefore considered as a beneficial factor.

Alcohol and steroid hormone abuse is the non-traumatic risk factor of ANFH. Some reports showed that glucocorticoids inhibit eNOS gene expression, but we did not find obvious differences in allele frequency between hormone-induced ANFH and normal controls. The possible reason may be the limited sample size. However, few data are available with respect to the influence of alcohol on eNOS gene expression, so no conclusive findings have been reached. We did not find obvious differences in

allele frequency between alcohol-induced ANFH and normal controls, which may be also due to the limited sample size.

Frequencies of intron 4 b/a genotype and G894T polymorphism in exon 7 were calculated and compared with those of normal controls. PBMC isolation, cell whole-genome extraction, specific PCR amplification, determination of nucleotide sequencing of target genes and polymorphism analysis of DNA fragments by restriction enzyme digestion were performed. The results showed that ANFH group and normal control group differed in the polymorphism of eNOS gene. It was discovered that intron 4 b/a genotype of eNOS gene among Chinese population caused the increase of ANFH risk. G894T polymorphism in exon 7 also led to an obvious rise of ANFH incidence. In order to confirm the correlation between the polymorphism of eNOS gene and idiopathic ANFH, the sample size should be enlarged. Therefore, polymorphism of eNOS gene provides a candidate target for the screening of high-risk population and early intervention of ANFH.

Disclosure of conflict of interest

None.

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