



2016 Alcoholic Liver Disease: Global view

Metabolic derivatives of alcohol and the molecular culprits of fibro-hepatocarcinogenesis: Allies or enemies?

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Abstract

Chronic intake of alcohol undoubtedly overwhelms the structural and functional capacity of the liver by initiating complex pathological events characterized by steatosis, steatohepatitis, hepatic fibrosis and cirrhosis. Subsequently, these initial pathological events are sustained and ushered into a more complex and progressive liver disease, increasing the risk of fibro-hepatocarcinogenesis. These coordinated pathological events mainly result from buildup of toxic metabolic derivatives of alcohol including but not limited to acetaldehyde (AA), malondialdehyde (MDA), CYP2E1-generated reactive oxygen species, alcohol-induced gut-derived lipopolysaccharide, AA/MDA protein and DNA adducts. The metabolic derivatives of alcohol together with other comorbidity factors, including hepatitis B and C viral infections, dysregulated iron metabolism, abuse of antibiotics, schistosomiasis, toxic drug metabolites, autoimmune disease and other non-specific factors, have been shown to underlie liver diseases. In view of the multiple etiology of liver diseases, attempts to delineate the mechanism by which each etiological factor causes liver disease has always proved cumbersome if not impossible. In the case of alcoholic liver disease (ALD), it is even more cumbersome and complicated as a result of the many toxic metabolic derivatives of alcohol with their varying liver-specific toxicities. In spite of all these hurdles, researchers and experts in hepatology have strived to expand knowledge and scientific discourse, particularly on ALD and its associated complications through the medium of scientific research, reviews and commentaries. Nonetheless, the molecular

mechanisms underpinning ALD, particularly those underlying toxic effects of metabolic derivatives of alcohol on parenchymal and non-parenchymal hepatic cells leading to increased risk of alcohol-induced fibro-hepatocarcinogenesis, are still incompletely elucidated. In this review, we examined published scientific findings on how alcohol and its metabolic derivatives mount cellular attack on each hepatic cell and the underlying molecular mechanisms leading to disruption of core hepatic homeostatic functions which probably set the stage for the initiation and progression of ALD to fibro-hepatocarcinogenesis. We also brought to sharp focus, the complex and integrative role of transforming growth factor beta/small mothers against decapentaplegic/plasminogen activator inhibitor-1 and the mitogen activated protein kinase signaling nexus as well as their cross-signaling with toll-like receptor-mediated gut-dependent signaling pathways implicated in ALD and fibro-hepatocarcinogenesis. Looking into the future, it is hoped that these deliberations may stimulate new research directions on this topic and shape not only therapeutic approaches but also models for studying ALD and fibro-hepatocarcinogenesis.

Key words: Alcoholic hepatitis; Lipopolysaccharide; Fibro-hepatocarcinogenesis; Mitogen activated protein kinase; Transforming growth factor beta; Small mother against decapentaplegic

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Core tip: Alcoholic liver disease (ALD) leading to fibro-hepatocarcinogenesis may show a bidirectional origin within the gut-liver axis. We bring to light the subtle reprogramming of the gut epithelium, gut microbiome and hepatic cells by both metabolic derivatives and unstable chemical species secondary to chronic alcohol intake, and their concerted role in ALD. We specifically highlight the integrative role of transforming growth factor- β /Smad, which synchronizes inflammatory and fibrogenic signals within the gut-liver axis. The gut may provide a less invasive option not only for prognosis and treatment of ALD but also for future research. We suggest that therapies for ALD and fibro-hepatocarcinogenesis should focus on restoring the gut microbiome.

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INTRODUCTION

It is a common knowledge that continuous heavy alcohol intake (80 g/d by men and 40 g/d by

women) spanning several years (10-20 years)^[1-3] may ultimately lead to chronic liver injury most often characterized by steatosis, steatohepatitis, liver fibrosis and cirrhosis leading to increased risk of fibro-hepatocarcinogenesis^[1,4-6]. Many reviews^[6-9] and research reports^[10-12], just to mention but a few, have all emphasized the pathological role of alcohol and its metabolic derivatives in ALD as well as efforts to identify some of the signaling pathways crucial in alcohol-induced liver disease. These important expert inputs have provided new insights in our understanding of ALD and fibro-hepatocarcinogenesis and have also provided new research directions about these diseases. Nevertheless, the pathological and molecular signaling pathways which underpin the initiation and progression of alcohol-induced liver injury leading to fibro-hepatocarcinogenesis still remain incompletely elucidated. For instance, signaling pathways that integrate gut-dependent alcohol-induced dysbiosis, inflammation and liver-specific alcohol-related inflammation, immune regulation and fibrogenic signals have so far remained elusive. The current difficulty in elucidating the molecular pathogenesis of alcohol-induced liver disease is multifaceted. (1) The anatomical position of the liver coupled with the diversity of agents in terms of number, biochemical properties, physicochemical properties, toxicity potential, their duration/frequency of exposure to the liver have obscured well designed attempts to delineate and characterize agent-specific effects on the liver much less the signaling pathways involved; (2) There is accumulating evidence, which seems to indicate that buildup of mutations in hepatic alcohol metabolizing enzymes (alcohol dehydrogenase, aldehyde dehydrogenase, CYP2E1) and genetic alterations induced by alcohol in hepatic cells^[13,14] may have further obscured attempts to elucidate the signaling pathways in ALD and fibro-hepatocarcinogenesis; and (3) Perhaps the most major difficulty is the bidirectional origin of ALD and fibro-hepatocarcinogenesis (gut to liver or liver to gut) and the dysregulation of key homeostatic functions (inflammation, immune regulation and regulation of fibrogenic signals). Notably, hepatic metabolism of alcohol as well as effect of alcohol on the gut generates many toxic chemical species with different mechanisms of hepatic toxicity which makes it difficult to distinctly identify their individual effects and signaling pathways involved.

This review takes a close look at current perspectives and scientific investigations on the effect of alcohol and its metabolic derivatives on hepatic parenchymal and non-parenchymal cells, bidirectional origin of ALD as well as the subtle conspiracy at the molecular level involving inflammatory, immune and fibrogenic signaling pathways underpinning ALD and fibro-hepatocarcinogenesis. Specifically, we put into perspective the complex and integrative roles of TGF- β (a key fibrogenic cytokine), Smad proteins, and

MAPK signaling pathways which pathologically suffer complicity in ALD and fibro-hepatocarcinogenesis mainly due to up-regulation of PAI-1 gene (a key downstream target gene of dysregulated TGF- β /Smad signaling in fibro-hepatocarcinogenesis) as well as recruitment of inflammatory and immune signaling pathways to promote ALD and fibro-hepatocarcinogenesis. Of note, the pathological role of PAI-1 in liver fibrosis, cirrhosis and cancer in general has been reported^[15]. And these pathological roles of PAI-1 may be linked to dysregulated TGF- β /Smad and MAPK pathways.

Transforming growth factor beta is a prototype of a superfamily of multi-functional cytokines including bone morphogenetic protein (BMPs), activin, inhibin, growth and differentiation factors, nodal, and anti-Mullerian hormone^[16]. The TGF- β_1 subtype has been extensively studied, mainly due to its physiological and pathological roles in the regulation of metazoan development, differentiation and homeostasis. It is in the light of these that the TGF- β class of cytokines is seen as a necessary evil in metazoan biology. In fact, TGF- β signaling pathway plays an important role during embryonic development, normal physiological processes and disease states by regulating several cellular processes, including cell growth and differentiation, cell migration, apoptosis, extracellular matrix formation^[16] and inhibition of cell proliferation in the early stages of carcinogenesis mainly by blocking uncontrolled proliferation of epithelial, endothelial and hematopoietic cells^[17]. However, genetic and epigenetic alterations of the TGF- β ligand, TGF- β -specific membrane receptors (T β R I, T β R II and T β R III), and mediators (Smad proteins) may switch its tumor suppressor effects into tumor promotion. The susceptibility of TGF- β to loss of function mutations in various cancers has been reported^[18]. For example, loss or gain of function mutations in T β R I^[19] T β R II^[20-22], Smad2^[23,24], Smad3^[25] and Smad4^[23,26] have all been implicated in various human cancers. Therefore, it is not surprising that dysregulated TGF- β signaling pathway suffer complicity in almost all known human cancers^[27-30]. It is maintained that genetic and epigenetic factors conspire to mastermind switching of TGF- β function by rendering tumor cells resistant or unresponsive to TGF- β -mediated growth arrest, and other homeostatic functions. TGF- β has been branded as the key factor regulating the acquisition of all the phenotypic hallmarks of cancer (cell proliferation, induction of epithelial to mesenchymal transition (EMT), induction of tissue invasion and migration, induction of tumor angiogenesis, inhibition of immune surveillance, induction of cancer cell survival, cancer cell immortality and resistance to TGF- β -mediated cytostasis)^[27,30]. Recent evidence shows that mitogen activated protein kinase (MAPK) pathway regulate linker-dependent phosphorylation of receptor mediated Smads (Smad2 and Smad3) to promote pathological roles of dysregulated TGF- β /Smad signaling in liver fibrosis and hepatocellular carcinoma (HCC)^[31,32]. The question arises as to how

these signaling pathways act in synchrony to modulate alcohol-dependent activation of the hepatic cells to promote ALD and fibro-hepatocarcinogenesis from the perspective of the gut and the liver. Does chronic alcohol exposure alter TGF- β /Smad and MAPK signaling pathways? If it does, how and which component of the TGF- β /Smad signaling mediators is/are altered and how? Finally, how do deliberations on these questions inform us of future research directions and therapeutic strategies against ALD and fibro-hepatocarcinogenesis? The above questions are the preoccupation of the present review.

HEPATIC ALCOHOL METABOLISM

The liver metabolizes alcohol by employing two mechanisms, either through cytosol degradation by alcohol dehydrogenase to acetaldehyde (AA), then to acetic acid by aldehyde dehydrogenase in the mitochondria or *via* the cytochrome P450 (CYP) isoenzyme system where CYP2E1 actively metabolizes alcohol in cases of heavy alcohol ingestion^[33-35]. Efficient functioning of these two hepatic alcohol metabolic processes ensure that toxic metabolites of alcohol, mainly AA (a hepatotoxin as well as a neurotoxin), MDA (a hepatotoxin) and some other unstable derivatives of the metabolites including CYP2E1-generated free radicals, protein adducts of AA and MDA, are rendered inactive or cleared from the system long before they cause any cellular damage. Indeed, buildup of AA and MDA, an inevitable phenomenon in chronic alcohol intake, is implicated for most of the toxic effects associated with chronic alcohol use^[34].

Interestingly, it was reported that CYP2E1 activity may be induced about two to tenfold after chronic alcohol exposure and the underlying mechanism was linked to oxidative stress^[36]. It was also reported that CYP2E1-dependent alcohol metabolism causes oxidative stress through increased output of reactive oxygen species (ROS)^[37-39], which has already been implicated in lipid peroxidation and liver injury^[40].

It must be noted that both cytosolic and mitochondrial alcohol metabolic pathways reduce NAD⁺ to NADH (addition of a hydrogen atom to NAD⁺ to convert it to NADH), however, impairment of any of the two metabolic pathways as a result of chronic alcohol intake may lead to a high NADH/NAD⁺ ratio which by extension affects cytosolic and mitochondrial metabolism of carbohydrate and lipid substrates leading to impaired gluconeogenesis^[4]. It was reported that alcohol exposure induces fatty liver disease by increasing NADH/NAD⁺ ratio^[41]. It remains to be established whether alcohol-induced NADH/NAD⁺ turnover underlies reprogramming and switching energy metabolism of pre-neoplastic hepatic cells from efficient mitochondria oxidative phosphorylation to that of inefficient but protective aerobic glycolysis (so-called Warburg effect). The net effect is that there is

diminished substrate flow through the Krebs' cycle, giving rise to diversion of acetyl CoA to fatty acid synthesis and this possibly underlies NADH-induced inhibition of mitochondria fatty acid β -oxidation and elevated fatty acid synthesis leading to the onset of alcoholic liver disease^[42-44].

Currently, it has been proposed that the pathogenesis of a healthy liver to one of alcohol-induced liver damage may involve a two-hit progression with steatosis being considered as the "first hit", followed by cellular insults such as oxidative stress, lipid peroxidation, direct lipid toxicity, mitochondrial dysfunction and/or infection to cause hepatic inflammation leading to alcoholic steatohepatitis^[4-6]. As useful as this current "two hit" proposal may be, it remains to be clarified whether the pathological sequence of ALD leading to fibro-hepatocarcinogenesis lend itself to any particular set pattern, in view of the fact that diverse toxic agents of non-alcoholic origin may also influence ALD progression. The effect of co-morbidity factors such as hepatitis B and C infections has been shown to increase the progression of ALD. However, it is still difficult to clarify the question of which toxic agent first initiates liver damage and which toxic agent takes over at what cellular time scale and how? Is it alcohol or the co-morbidity factors that first initiate liver damage? It appears that alcohol-induced liver damage leading to ALD and fibro-hepatocarcinogenesis may not follow any specific temporal sequence, in view of the presence of other non-alcohol toxic agents. It is possible that the underlying non-alcohol liver-specific toxic agents may be the determinants of the temporal sequence of alcohol-induced liver disease.

Alcohol-induced liver damage displays bidirectional origin in view of the significant nauseating role of lipopolysaccharide (LPS) derived from progressive alteration of the gut microenvironment by chronic alcohol intake.

ALCOHOL AND ALTERATION OF GUT MICROBIOME

The invention of the microscope^[45] provided the impetus to uncover the co-existence of micro-organisms and humans^[46]. It is now a common knowledge that the human gut, (a prominent barrier organ) harbors a metagenomic community of some 10^{14} micro-organisms^[47] mainly dominated by bacteria. The gut microbiome does play many regulatory functions spanning regular modulation of the innate and adaptive immune systems^[48], synthesis and release of nutrients, vitamins, and preservation of the structural and functional integrity of the gut wall^[46]. For instance, in the course of evolution of the gut microbiome, the diverse gut microorganisms have progressively managed to adapt as commensals, producing nutrients, such as vitamins of the B and K subclasses^[49], and

short-chain fatty acids (SCFAs). About 60%-90% of SCFAs in the gut lumen are absorbed by enterocytes to regulate energy supply, control gut pH, and resist pathogenic growth^[50] probably *via* inflammasome^[51]. The gut microbiome also plays a role in bile acid regulation^[52,53], exchange of phenolic and aromatic acids^[54], cholines, fatty acids and phospholipids^[55,56]. Liver-specific biosynthesis of primary bile acids are reported to be dehydroxylated by some of these gut microbiome giving rise to secondary bile acids, which may be absorbed by the enterocytes to promote lipid absorption and energy homeostasis^[52,53]. In view of the above, the importance of the gut microbiome in anti-oxidant, inflammatory, immune and energy homeostasis cannot be underestimated and therefore it represents a crucial determinant of the body's susceptibility to irritants including alcohol and its metabolic derivatives.

It is not surprising that alterations in the number and species diversity of the gut microbiome culminating from host-behaviors including but not limited to chronic alcohol intake derail the essential benefits of the gut microbiome^[57] and may provide avenue for the onset of various inflammatory diseases of the gastrointestinal system and its accessory organs, of which the liver is the most affected. From hindsight, change in gut microbial diversity has long been implicated in Crohn's disease (CD)^[46], ulcerative colitis (UC)^[46] and irritable bowel disease (IBD)^[46,58]. The case is not different with chronic alcohol exposure and the possible increase in Gram negative/Gram positive bacteria ratio (Figure 1). Nakayama *et al.*^[59] have shown that increased translocation of *Streptococcus suis* and its degraded products across the gut wall secondary to alcohol exposure correlated with progression of ALD. Accumulating evidence show that chronic alcohol intake may switch the aforementioned essential regulatory functions of the gut microbiome into a rather deleterious one. For example, alcohol-induced gut dysbiosis increases endotoxin turnover^[60,61], particularly LPS^[6,62], which leads to increased leakage of endotoxins into portal circulation and chronic stimulation of the liver. By diverse mechanisms, alcohol and its metabolic derivatives have been implicated in dysbiosis of the gut mucosal layer^[63-66].

Lipopolysaccharides (LPS) are breakdown products of bacterial cell walls, specifically pathogenic Gram negative bacteria strains^[67] and it is reported that it can activate hepatic cells^[6,67] and initiate overt inflammatory responses *via* TNF- α mediation^[68,69]. Under normal physiological conditions, release of LPS from breakdown of pathogenic Gram negative bacteria into portal circulation is rendered harmless by endothelial cells lining blood vessels, sinusoidal endothelial cells (SECs) of the liver as well as liver-resident macrophages (KCs) or its cellular concentration is reduced to levels well below physiological concentrations insufficient to elicit any inflammatory response. But perturbations of the

Alcohol-induced perturbation of intestinal epithelium and derangement of liver sinusoidal micro-structure in ALD and fibro-hepatocarcinogenesis

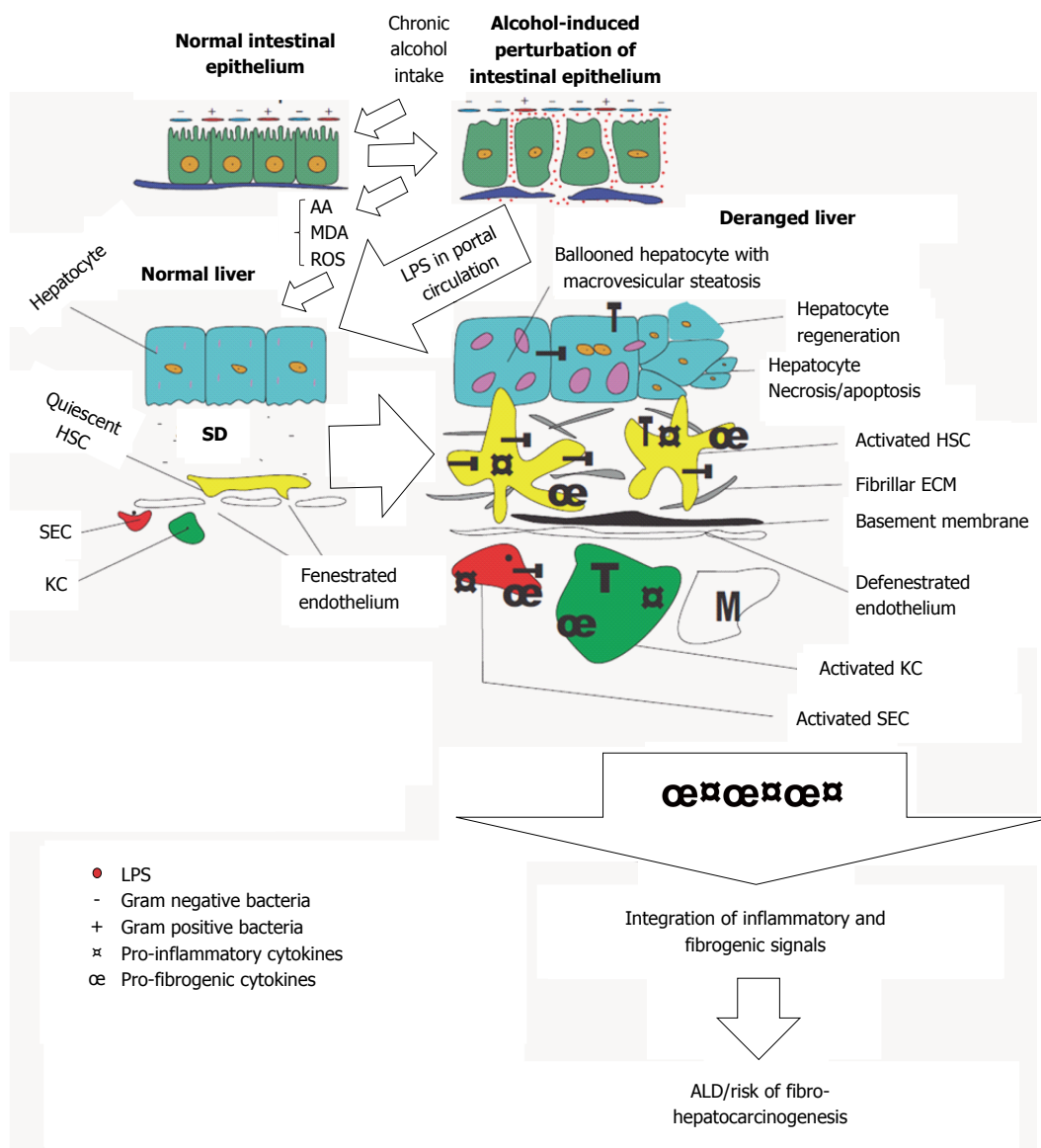


Figure 1 An illustration of the bidirectional origin of alcoholic liver disease and fibro-hepatocarcinogenesis within the gut-liver axis. Chronic alcohol use induces derangement of the gut epithelium, increases Gram-/Gram+ bacteria ratio, increases endotoxin turnover, increases permeability of gut epithelium to endotoxins including lipopolysaccharide (LPS). Subsequently leakage of LPS into portal circulation gain access to liver to initiate activation of hepatic cells. LPS-dependent activation of hepatic cells is further augmented by metabolic derivatives of alcohol to promote alcoholic liver disease (ALD) and fibro-hepatocarcinogenesis. T: Toll-like receptor 4; HSC: Hepatic stellate cell; KC: Kupffer cell; SD: Space of disse; SEC: Sinusoidal endothelial cell; M: Monocyte.

gut wall as a result of chronic alcohol intake (Figure 1), increases gut permeability to LPS derived from degraded bacterial cells^[62] and this certainly leads to leakage of high concentrations of LPS into portal circulation. The high levels of LPS in portal circulation overwhelms the regulatory capacity of SECs and endothelial cells leading to chronic liver injury^[41]. Continual exposure of the liver to gut-derived LPS serves as an inflammatory nosae, first by disrupting the balance between inflammatory and anti-inflammatory homeostatic regulation. This balance is shifted to favor heightened or sustained inflammatory response. To sustain the exaggerated inflammatory response, LPS first activates hepatic

parenchymal cells, precisely SECs KCs and hepatic stellate cells (HSCs) leading to re-programming of their core functions. It is not surprising that a correlation was reported between increased intestinal LPS permeability and alcoholic hepatitis^[60,61]. Similarly, LPS derived from alcohol-induced increase in gut permeability was shown in alcoholics to correlate with the pattern and the amount of alcohol consumed^[70,71] while high levels of LPS were detected in the sera and livers of patients with alcohol-induced liver disease^[62]. The hepatotoxic effect of pathogen associated molecular patterns (PAMPs), e.g., LPS^[72] has been shown to be mediated through toll-like receptors (TLRs)^[73]. LPS is specifically reported

to be the ligand for TLR4 subtype^[67]. Importantly, TLR4 as well as other TLR subtypes have been shown to be expressed on KCs, HSCs and hepatocytes under inflammatory conditions^[9,74]. TLRs are crucial in the regulation of innate immune responses, sensing of damage associated molecular patterns (DAMPs) as well as PAMPs, of which LPS is an integral component. Also, it was reported that LPS/TLR4 signaling involves LPS-binding protein (LBP), CD14 and MD-2^[75,76]. LBP facilitates the transfer of LPS from the outer membrane of bacterial cells to CD14, which in turn ensures the formation of TLR4-MD-2^[77] to trigger LPS/TLR4 signaling, but downstream of this TLR4-mediated LPS-induced liver inflammation is myeloid differentiation factor protein 88 (MyD88). LPS-induced release of pro-inflammatory cytokines (TNF- α , IL-1 β , IL-6, IL-8, IL-12), chemokines (INF- γ , MCP-1)^[78] as well as cell adhesion molecules (ICAM-1, VCAM-1) *via* activation of NF- κ B and I κ B in injured hepatic cells was MyD88-dependent^[78]. LPS-induced activation of the MAPK pathway leading to the expression of activator protein (AP)-1 was also reported to be MyD88-dependent^[79-81].

In short, chronic alcohol intake may generate an inflammatory arsenal comprising LPS, ROS, AA and MDA together with their respective protein adducts, which launches continuous attack on hepatic cells (Figure 1). The pathological manifestation of these alcohol-mediated attacks on the liver may depend in part on the synergistic interaction between alcohol and non-alcohol co-morbidity factors. Consequently, this sets the stage for the onset of liver fibrosis and its progression to cirrhosis, thus increasing the risk of fibro-hepatocarcinogenesis. To appreciate how the alcohol generated inflammatory arsenal attack the hepatic cells, we take a cursory look at each of the hepatic cells in the light of their normal structure and function vs their alteration by alcohol and its metabolic derivatives.

ALCOHOL, METABOLIC DERIVATIVES OF ALCOHOL AND ACTIVATION OF HEPATIC CELLS

Sinusoidal endothelial cells

Sinusoidal endothelial cells (SECs) are hepatic non-parenchymal cells characterized by flattened and highly fenestrated features^[6]. SECs have no basement membrane and their microstructure endows them with the ability to selectively filter blood components from portal circulation into the space of Disse, for subsequent presentation to the hepatocytes and lipid storage cells^[6].

Additionally, SECs are endowed with scavenger receptors (SRs) and higher permeability properties, and these factors make it possible for SECs to engage in phagocytosis, clearing blood of harmful toxins and ensuring bidirectional exchange of substances between the hepatic parenchyma cells and portal blood. The

SECs, by logic provides the first line of defense for the liver, essentially scavenging and clearing potential harmful products from attacking the liver. Many immunological functions, including but not limited to the following, removal of small molecules (< 200 nm) from the blood using innate immune mechanisms such as scavenger and mannose receptors^[82,83], expressions of MHC class II and co-stimulatory molecules (CD54 and CD106), and antigen processing, presentation, and leukocyte recruitment^[83], have been attributed to SECs.

The afore-mentioned homeostatic functions of SECs in the liver of an alcoholic become derailed due to continuous and sustained activation of SECs by gut-derived LPS leakage into portal circulation. For example, exposure of SECs to LPS was shown to have induced basement membrane formation^[84], and this observation was said to have preceded fibrogenesis. Some results from *in vitro* studies have also shown that SECs can reverse activated HSCs back to their quiescent state, but SECs lose this property following capillarization due to LPS activation^[85]. Serious is the fact that continuous LPS-activation of SECs induces decreased responsiveness of SECs to LPS and also diminishes SEC-dependent scavenger functions. Eventually, the liver may be exposed to potentially damaging insults including metabolic derivatives of alcohol.

Accordingly, AA and MDA were shown to have formed protein adducts, which in turn stimulated SECs to produce more fibrogenic cytokines^[6]. A clear attestation to this observation was demonstrated by fibronectin. Fibronectin was shown to be overexpressed following alcohol-induced liver damage and it was implicated in the activation of HSCs leading to liver fibrosis^[6]. MDA-derived protein adducts were reported to have increased the expression of soluble fibronectin, cellular fibronectin and EIII A fibronectin variant (the variant form of fibronectin, most implicated in HSC activation)^[86,87]. Pro-inflammatory cytokines including TNF- α , MCP-1, and MIP-2 were similarly shown to have increased following treatment of isolated SECs with MDA-derived protein adducts^[87,88]. Also, AA/MDA modified proteins were reported to have induced SECs to release both pro-inflammatory and pro-fibrogenic signals^[88], while LPS was similarly reported to induce AA/MDA modified protein adduct-dependent release of chemokines and cytokines by SECs^[89]. Evidently, LPS, AA, MDA, and their protein adducts act in concert to potentially induce apoptosis of SECs which leads to weakening of SEC-dependent defense mechanisms of the liver, further, compounding an already compromised liver. LPS-activated SECs may in turn trans-activate KCs and HSCs in addition to their direct stimulation by LPS.

Kupffer cells

As part of the reticuloendothelial system are cells called Kupffer cells (KCs). Kupffer cells are monocyte-

derived cells resident in the liver as specialized hepatic macrophages^[6,90]. These cells were first observed by Karl Wilherm von Kupffer in 1876^[91] to whom is credited the name KCs, but it was not until 1898 that KCs were correctly identified as macrophages^[92]. Their origin can be traced to the bone marrow, where promonocytes and monoblasts cells differentiate into monocytes, which then enter circulation and finally transform into KCs^[93].

Functionally, KCs form a major part of the reticuloendothelial system within the liver sinusoidal compartment, phagocytizing senescent red blood cells as well as phagogenic presentations. As a result, they are widely scattered within the sinusoids. In support of the phagocytizing capacity of KCs, Helmy *et al.*^[94] have reported a receptor of the immunoglobulin family (CRIg), and they further showed that CRIg null mice could not clear complement system-coated pathogens, and that CRIg is well conserved in mice and humans, emphasizing the relevance of the CRIg as a component of the innate immune system and that of the role of KCs in the innate and complement systems.

In normal physiological state, KCs perform their immuno-regulatory functions without overt release of pro-inflammatory cytokines and chemokines; however, alcohol and its associated metabolic derivatives have the potential to reprogram KCs through repeated or continuous activation. This undue activation of KCs renders them more pro-inflammatory and pro-fibrogenic due to KC-dependent release of inflammatory and fibrogenic cytokines. For instance, it was reported that under stress conditions, KCs and other hepatic cells release cytokines (IL-1, IL-6, IL-8, TNF-) and chemokines (MIP-2, IP-109, KC/GRO, MIP-1 α , and RANTES)^[90,95]. It is the unregulated release of these inflammatory and fibrogenic cytokines that induces liver injury. It was further shown that each of the pro-inflammatory cytokines and chemokines could directly cause liver injury by targeting hepatic cells or indirectly through chemo-attraction of immune cells including neutrophils and lymphocytes^[95]. Also, it was reported that the expression of adhesion molecules changes during LPS/alcohol-dependent liver injuries. Notably the enhanced expression of PECAM-1 and down-regulation of ICAM-1 characteristic of normal liver were reported to be reversed by TGF- β under inflammatory conditions^[96].

Chronic exposure of the liver to alcohol succeeds in changing the sensitivity of KCs to LPS stimulation^[97-99]. In a study to clarify alcohol-induced sensitization of KCs to LPS, Watanabe *et al.*^[100] have suggested that it could be due to the effect of alcohol on calcium channels, which is indispensable for TNF- α release. Exposure of KCs to alcohol does not only increase sensitivity of KCs to metabolites of alcohol but also increases intracellular calcium channels in KCs. It was shown that KCs exposed to alcohol for two hours lacked elevated intracellular calcium, but prolongation

of alcohol exposure time to 24 h showed increased intracellular calcium, which manifested as TNF- α production and expression of LPS binding receptor (CD14), and this perhaps explains the increased sensitivity of KCs to LPS. Chronic alcohol exposure was also shown to have increased the expression of α 2A-adrenoceptors in activated KCs and it was linked to the release of TNF- α and TNF- α -induced liver injury^[101]. LPS was shown to augment AA/MDA protein adduct-mediated release of pro-inflammatory and pro-fibrogenic cytokines and chemokines by KCs^[89]. LPS-induced acute and chronic liver injury was linked to activation of KC^[102,103].

The activation of KCs is not limited to only LPS-derived from gut microbiome, and proteins modified by AA and MDA may also activate KCs leading to overt inflammatory and immunological responses injurious to the liver (Figure 2). Many alcohol modified protein adducts have been widely implicated in alcoholic liver disease^[89,104,105]. Alcohol and its metabolic derivatives have also been implicated in KC/prostaglandin E2-induced liver injury^[106] and this is in part attributed to endotoxin-dependent release of nitric oxide (NO) in KCs^[107]. It appears that, chronic alcohol exposure in addition to AA and MDA dependent liver injury may also induce increase in gut-derived LPS, increase in LPS leakage into portal blood, increase in blood concentration of LPS, increase in LPS binding receptor expression and increased sensitivity of KCs to LPS and these may collectively sustain liver injury.

LPS activation of hepatic cells shows a snowballing effect, in that apart from LPS directly activating each hepatic cell, each activated hepatic cell may in turn influence the activation of its neighboring hepatic cells in a manner akin to paracrine or hormone-like cell to cell communication. For example, LPS-induced activation of SECs leads to activation of KCs *via* release of pro-inflammatory and pro-fibrogenic factors, similar activation of KCs also in turn activate quiescent HSCs through the release of TNF- α and TGF- β ^[6]. Typically, LPS-activated KCs were shown to have activated HSCs *in vitro* leading to HSC proliferation and increased ECM production^[108]. Next, we look at how alcohol and its metabolic derivatives act in concert to activate HSCs to usher in liver fibrosis.

Hepatic stellate cells

Hepatic stellate cells (HSCs) are non-parenchymal hepatic cells located within the space of Disse between endothelial cells and hepatocytes^[109,110]. HSCs can exist in two distinct forms depending on whether they are activated by an external inflammatory nosae or otherwise. In their normal inactivated state, also known as quiescent stellate (Ito) cells, they function normally by storing vitamin A, and may play modulatory roles during inflammation by expressing ICAM-1 and VCAM-1. It has been shown that HSC activation (tran-differentiation of quiescent vitamin

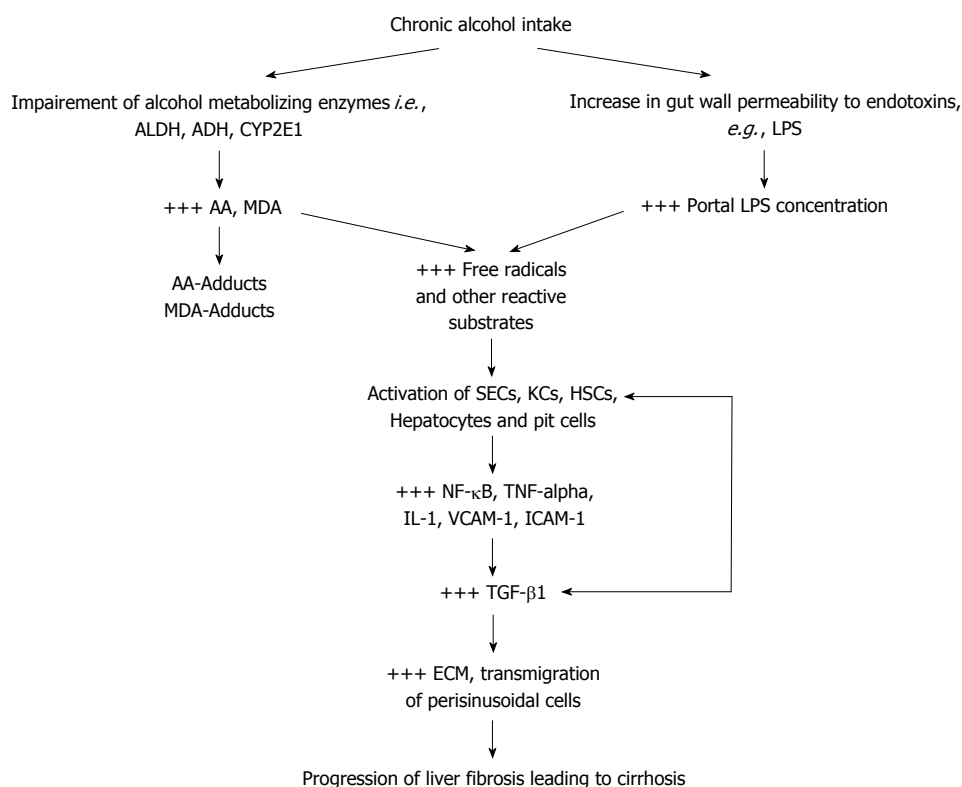


Figure 2 A schematic illustration of the two-way attack of hepatic cells in alcoholic liver disease and fibro-hepatocarcinogenesis. By multiple mechanisms alcohol metabolizing enzymes and gut-derived LPS induce production of free radicals which stimulate the release of pro-inflammatory and pro-fibrogenic cytokines. Free radical-dependent activation of hepatic cells leads to the release of pro-inflammatory transcription factor (NF- κ B) and inflammatory cytokines (TNF- α , IL-1 β) which provide the signal for injurious overt hepatic inflammatory response. Secondary to the injurious hepatic inflammatory response, is the activation of hepatic cells (mainly HSCs and KCs) to further release pro-fibrogenic factors, mainly the key fibrogenic cytokine (TGF- β) which mediates a high ECM turnover (increased fibrogenesis : fibrinolysis ratio) in HSCs and the space of Disse, and trans-migration of hepatic non-parenchymal cells. These pathological events initiate liver fibrosis and cirrhosis leading to fibro-hepatocarcinogenesis. AA: Acetaldehyde; ADH: Aldehyde dehydrogenase; ALDH: Alcohol dehydrogenase; ECM: Extracellular matrix; HSCs: Hepatic stellate cells; IL-1 β : Interleukin-1 beta; KCs: Kupffer cells; LPS: Lipopolysaccharide; MDA: Malondialdehyde; NF- κ B: Nuclear factor kappa B; SECs: Sinusoidal endothelial cells; TGF- β : Transforming growth factor beta; TNF- α : Tumor necrosis factor alpha; VCAM: Vascular cell adhesion molecule; +++: Up-regulated expression or overproduction.

A-storing cells to proliferative myofibroblast cells) by hepatotoxins plays a crucial role in liver fibrogenesis. To substantiate this claim, it was shown that inactivation of HSCs attenuates liver fibrosis^[111]. Activated HSCs, mostly release excess TGF- β ₁ which has been shown to down-regulate ICAM-1 and VCAM-1 while increasing N-CAM expression in HSCs. In addition, HSCs in their activated states may express TNF- α , which reduces N-CAM-coding mRNAs and also induces ICAM-I-and-VCAM-1 specific transcripts by many folds^[90]. Under inflammatory conditions, HSCs trans-differentiate into myofibroblast and concomitantly increase ECM production. There is dysregulation of ECM metabolism which promotes liver fibrosis. This underlies the initiation of fibrogenesis and the complicity of HSCs in liver fibrosis^[112]. Coherent with this, Knittel *et al.*^[113] have intimated the importance of HSCs during hepatic injury through the recruitment and migration of mononuclear cells in the peri-sinusoidal space. This perhaps sets the stage for the secretion of TGF- β and TNF- α and their pathological role in liver fibrosis^[114,115] and several other cytokines and chemokines including MCP-1, RANTE-1, and IL-8^[116-118].

The effect of alcohol and its metabolic derivatives on HSC activation and their roles in ALD progression have been extensively investigated. For example, alcohol-induced activation of HSCs was linked to the release of TGF- β ₁, matrix proteins, and initiation of fibrogenic response^[112,119]. Karaa *et al.*^[120] have also shown that alcohol exposure to mice produced HSC activation in addition to increased hepatic collagen output and neutrophil infiltration.

Evidence implicating alcohol-specific metabolites in liver injury was clearly demonstrated by using 4-methylpyrazole (4-MP), an inhibitor of alcohol metabolism. It was shown that sustained alcohol exposure to precision cut liver slices (PCLS) produced increased levels of IL-6, depletion of GSH stores, increased lipid peroxidation, increased expression of smooth muscle actin (α -SMA) and increased deposition of collagen in liver sinusoidal space^[6]. But all these aforementioned alcohol exposure-specific phenotypic hallmarks were reversed by 4-MP treatment^[6,121], emphasizing the involvement of alcohol and its metabolic derivatives in liver fibrosis.

Furthermore, LPS was reported to exert direct

effect on HSCs^[122] and also indirectly *via* LPS activated KCs^[108,123]. LPS was shown to enhance the effect of alcohol and AA on HSC activation^[124,125] and the expression of collagen 1 and IL-6^[126], while AA alone induced activation and proliferation of HSCs^[127,128] and expression of α -SMA^[127,129].

At the mechanistic level, AA-induced activation of HSCs was linked partly to ERK1/2/PI3K pathway^[130,131], JNK/ α 1-collagen pathway^[132] and TGF- β ₁/T β R II signaling^[133,134]. Transcriptional elevation of TGF- β and its membrane receptors (T β R I and T β R II) have been considered as key hallmarks of activated HSC^[135]. And this certainly increases the sensitivity of activated HSCs to TGF- β -mediated fibrogenic signals. Siegmund *et al.*^[9] have posited that the signal for ECM production by HSCs in liver-related diseases of all etiologies including alcohol emanates from TGF- β . Corroboratively, it was shown that TGF- β protein levels increased in both experimental and human liver fibrosis^[136]. Thus, TGF- β is the key contributor of irregular ECM accumulation in liver sinusoidal space by forming basal membranes leading to defenestration of the liver sinusoids^[137] as well as decreasing MMPs to further halt ECM degradation^[138].

Hepatocytes

Hepatocytes are the main hepatic cells in the liver accounting for 80% of the cytoplasmic mass of the liver^[90]. Characteristically, hepatocytes exhibit eosinophilic cytoplasm (a cytoplasm with abundant mitochondria) and basophilic stippling (abundance of endoplasmic reticulum and ribosomes)^[90]. Within the liver, hepatocytes are organized into cell-thick plates separated by vascular channels^[139,140]. When these structural and functional characteristics of hepatocytes are not disturbed or altered by any nosae, hepatocytes can attain an average life span of 5 months while retaining the capacity to regenerate^[90].

Functionally, hepatocytes are involved in protein synthesis, protein storage, and transformation of carbohydrates, synthesis of cholesterol, bile salts, phospholipids, detoxification, modification and excretion of exogenous and endogenous substances^[90]. Additionally, hepatocytes can biosynthesize hormones including insulin-like growth factor (IGF-1)^[90,141], thrombopoietin^[142], erythropoietin^[143] and cytokines such as IL-8^[144,145] needed for normal hepatic homeostatic processes.

In the event of liver injury due to alcohol and its metabolic derivatives, the degree of injury overwhelms the hepatocyte defense mechanisms, especially when the nosae is continuous and sustained (Figure 1). As a means of defense, hepatocytes may respond to acute phase inflammatory mediators such as IL-6 by releasing acute phase proteins including C-reactive protein (CRP)^[146], serum amyloid A (SAA)^[147] and also some intracellular defense proteins like heme-oxygenase-1 (HO-1)^[148]. In a desperate

attempt to fight back, hepatocytes employ various mechanisms including release of chemokines such as MIG^[149], IP-10^[149], cytokine-induced neutrophil chemo-attractant (KC), MIP-1, MIP-2, and MIP-3 which act in concert to recruit and activate pro-inflammatory cells (mononuclear phagocytes) and KCs respectively. But because the inbuilt intracellular hepatocyte defense mechanisms *i.e.*, CRP, SAA and HO-1, and the anti-oxidant system have already been weakened, by the continuous exposure of the damaging nosae, the hepatocyte defense response leads to mass hepatocyte apoptosis at a rate that further compromise the structural and functional integrity of the liver as a whole. Hepatocyte cytosol and microsomal compartments are the initial sites for alcohol metabolism before mitochondrial-dependent breakdown. The number and integrity of hepatocytes are rate-limiting factors in alcohol metabolism. Therefore, increased apoptosis of hepatocytes leads to poor alcohol metabolism which further increases the buildup of alcohol metabolites (AA and MDA), meanwhile KCs, SECs, and NKs which could clear the debris and the resulting buildup of toxins have been disabled by LPS, AA, and MDA. This situation exposes the liver to potentially damaging nosae which may set the stage for the initiation of cirrhosis leading to HCC.

Pit cells

Pit cells, lymphocyte-derived non-parenchymal cells, form about 1% of the non-parenchymal cell mass^[150]. Pit cells are liver representatives of natural killer cells (NKs) in other organs. Pit cells are suspected to have originated from the bone marrow transported by blood to finally settle in the liver, where they transmogrify into their current state by lowering their density and increasing their granular content. The very existence of the pit cells are dependent on KCs^[151], which suggest that whatever that happens to KCs will in turn affect the fate of pit cells.

Functionally, pit cells biosynthesize interferon gamma (IFN- γ) in response to damaging inflammatory nosae, but they can also partake in the destruction of virus-infected malignant cells. Pit cells are versatile migratory cells and they are normally activated by interleukin-2^[90]. Alcohol, AA, MDA, and LPS may directly damage pit cells through continuous activation or indirectly by activated KCs and HSCs, leading to functional impairment and consequences thereof on the liver, in view of the fact that pit cells *via* the perforin/granzyme-dependent mechanism are indispensable in the removal and apoptosis of splenic/blood-NK-resistant tumor cells^[152]. Importantly, TGF- β -induced repression of NKs, a phenomenon characteristic of chronic alcohol consumption^[153], has been linked to failure of NK cell-mediated apoptosis of HSCs^[154,155].

Evidently, chronic alcohol intake affects the structural and functional capacity of all the hepatic

cells and this possibly may set the cellular stage for the actions of pro-inflammatory and pro-fibrogenic cytokines acting in concert to promote ALD leading to fibro-hepatocarcinogenesis. At the center of this complex cellular conspiracy against the liver are the TGF- β /Smad and MAPK pathways and their downstream target genes.

TGF- β : A VERSATILE SIGNALING MODULATOR WITH COMPLEX FUNCTIONALITY IN ALD AND FIBRO-HEPATOGENESIS

To appreciate how alcohol and its metabolic derivatives alter TGF- β /Smad signaling to promote ALD and fibro-hepatocarcinogenesis, we should first take a panoramic view of TGF- β /Smad signaling and also the cell/context-specific functions of TGF- β which perhaps may explain its complex and integrative roles in ALD. TGF- β is considered to exert growth restraints on various cancer cells at the initial stages of fibro-hepatocarcinogenesis by mechanisms including cell cycle arrest at critical check points, induction of apoptosis and restoration of cellular structure^[156-158], an observation which highlights TGF- β as a potent anti-tumor cytokine^[31]. However, by a sudden twist of events, in a different cellular context such as in ALD, fibro-hepatocarcinogenesis as well as in other disease pathologies, it tends to promote disruption of cell adhesion, induces migration and invasion, and mediate immune suppression and angiogenesis, to become a crucial tumor promoter^[158]. For example, in ALD it was reported that AA does not only increase the steady levels of TGF- β mRNA transcripts^[159] but also promote activation of latent TGF- β and elevate the expression of T β R II^[134]. Furthermore, it was shown that a decrease in TGF- β protein levels correlated with a decrease in AA-induced α 2 collagen (I) gene^[160].

Basically, TGF- β signaling involves two major signaling modes, canonical and non-canonical. The former is mediated by Smad proteins while the latter involves cross signaling between TGF- β and other cell signaling pathways implicated in cancer such as Wnt/ β -catenin pathway^[161,162], VEGF pathway^[163,164], aberrant FGF/FGFR signaling pathways^[165,166], MAPK pathway^[167,168], PI3k/AKT/ mTOR pathway^[169], HGF signaling^[164], aberrant EGF/EGFR signaling pathway^[170], and deregulation of IGF pathway^[171]. To proceed, we highlight the Smad proteins and the MAPK pathway, which have so far been shown to work closely with TGF- β signaling in both cell and animal models of ALD (Table 1) to promote liver fibrosis and HCC^[31].

Smad proteins

Genetic studies spanning more than a decade ago using *Caenorhabditis elegans* (a nematode), and *Drosophila melanogaster* (a fruitfly) led to the dis-

covery of a group of genes, which were later named Smads from their original sources. Smads are the central mediators that carry signals from receptors of TGF- β , BMP, and activin cytokines to the nucleus^[172]. Smads which are now identified as substrate transcriptional factors play integral functions in the intracellular signaling responses to TGF- β and its related signaling complex^[173]. Basically, the Smad proteins have three forms, namely receptor mediated Smads (Smad 2 and Smad3 specific for TGF- β), inhibitory Smad (Smad7) and a common Smad (Smad4). It must be stated that mutations in these Smad types due to genetic and epigenetic causes such as alcohol exposure are linked to dysregulated TGF- β signaling in ALD and cancer in general. For example, Smad7 inhibits TGF- β /T β R I-dependent phosphorylation of Smad2 and Smad3^[174-176] to abrogate dysregulated TGF- β /Smad signaling transduction in many disease pathologies including but not limited to ALD and fibro-hepatocarcinogenesis. But, alcohol and LPS were shown to down-regulate Smad7 expression to induce liver fibrosis in a Smad3-dependent fashion^[120]. Similarly, a number of reports from cell and animal studies (Table 1) have implicated receptor-mediated Smad proteins (Smad2 and Smad3) in ALD as well as in HCC patients. Smad4 deletions have been mapped in HCC^[18]. Smad2^[23,24], Smad3^[25] and Smad4^[177] mutations have been detected in various cancer subtypes. But it remains to be elucidated how alcohol and its metabolic derivatives modulate the Smad proteins and the signaling pathways they mediate, particularly the upstream (TGF- β ligand, T β R I, T β R II, Smad2, Smad3, Smad4, Smad7) and downstream (Imp7/8, PAI-1) signaling mediators of TGF- β and how such findings may inform future research directions and precise therapeutic strategies against ALD and fibro-hepatocarcinogenesis. It is worth suggesting that Smad2, Smad3, and Smad7 deletions or/and mutations should be mapped in ALD as well as in HCC to facilitate early screening, diagnosis and treatment of ALD and its related complications.

Importantly, it must be mentioned that the switch of TGF- β function from tumor suppression in early stages of cancer as well as in early stages of ALD to tumor promotion in late HCC reflects an imbalance between canonical and non-canonical TGF- β signaling and recruitment of other oncogenic signaling pathways (Figure 3). What actually causes this switch has so far remained elusive. Refreshingly, in HCC it was reported that binding of Gas6 ligand to Axl induces activation of Axl/14-3-3 ζ to switch TGF- β signaling from tumor suppression to tumor promotion in a JNK/Smad3L-dependent fashion^[178]. It was also mentioned that suppression of Axl succeeded in blocking oncogenic TGF- β signaling in HCC, and this has raised hopes about indirect inhibition of oncogenic TGF- β signaling using Axl-specific inhibitors. A number of Axl-specific inhibitors are under various stages of preclinical studies including SGI-7079, BGB324, DP3975 and NA80x^[179].

Table 1 Involvement of Transforming growth factor- β , Smad, plasminogen activator inhibitor-1 and mitogen activated protein kinase signaling pathways in alcoholic liver disease and fibro-hepatocarcinogenesis

Alcohol/ metabolic derivative	Mechanism/pathway	Cellular context	Ref.
Alcohol, LPS, SAME	Inhibition of TGF- β /Smad signaling abrogates alcohol-induced liver injury alcohol and LPS induce liver fibrosis via activation of TGF- β signaling in a Smad3-dependent fashion and down-regulation of Smad7, but SAME could abrogate it and also restore Smad7 expression	Cultured HSCs, male rats	[120,221]
AA	Up-regulation of Smad3 and Smad4, increase in nuclear translocation of Smad3/4 complex, decrease in Smad7 expression, all leading to enhanced expression of COL1A2 Increase in TGF- β 1 secretion and up-regulated expression of T β RII in HSCs was linked to AA AA increased COL1 α 1 expression in HSCs in a Smad3-dependent manner	Human and mouse HSCs	[159,219,222,223]
Alcohol	Alcohol-induced increase in endotoxemia linked to up-regulated protein expression of TGF- β 1, IL-6, NF- κ B, TNF- α , I κ B α	Guinea pig liver	[224]
Alcohol, LPS	Alcohol potentiates LPS-induced pancreatic fibrosis <i>via</i> increased production of TGF- β 1	Human pancreatic tissue sample, pancreatic acinar-like cells (AR42J)	[225]
Alcohol	Alcohol-induced translocation of <i>S. suis</i> across gut wall and also up-regulated TGF- β 1 and COL 1 to promote liver disease	Alcoholics, mice, Caco-2 cells	[59]
Alcohol	While alcohol exposure impairs nuclear import of growth hormone-induced STAT5B and IL-6-induced STAT3, it had no effect on TGF- β 1-induced nuclear import of Smad2/3 TGF- β 1 mediates liver fibrosis in experimental rats in a Smad4-dependent fashion Alcohol-induces hepatic iron overload leading to liver damage <i>via</i> modulation of hepcidin through BMP6/Smad4 signaling pathway	Rat liver, adult male C57BL/6J mice	[11,184,226,227]
Alcohol	Alcohol modulates iron-induced liver injury <i>via</i> increased expression of TGF- β 1, BMP2, phosphorylated Smad2	Mice	[228]
Alcohol	Alcohol-induced steatosis and liver injury in Smad7 null mice is enhanced by TGF- β 1 signaling and TGF- β 1-induced EMT in hepatocytes	Alb-Cre mice, Smad7 (loxP/loxP) mice	[229]
Alcohol	Alcohol exposure induces TGF- β 1 release and activation of TGF- β 1-induced down-regulation of alcohol dehydrogenase 1 (ADH1) mRNA transcripts in part through TGF- β /ALK5/Smad2/3 signaling	Mice	[230]
Alcohol, AA	Alcohol and AA-induced activation of TGF- β 1, JNK and p38 signaling pathways were inhibited by butein	HSCs, HepG2 cells	[231]
Alcohol	TGF- β 1 mediates alcohol-induced activation of HSCs <i>via</i> activation of p38/JNK MAPK pathway and overexpression of α -SMA, procollagen1, betulin and betulinic acid can reverse these pathways to restore liver integrity	Rat HSCs	[232]
Alcohol, LPS	LPS and CYP2E1-dependent oxidative stress synergistically activate p38/JNK pathway <i>via</i> LPS/TNF- α signaling pathway	Hepatic cells	[233]
Alcohol	Alcohol induces cytotoxicity <i>via</i> activation of p38, JNK and ERK MAPK pathway, but COS reversed this by inhibiting the MAPK pathway and activation of Nrf2	Human L02 normal liver cells	[234]
Alcohol	Alcohol induces hepatotoxicity by activating p38/JNK MAPK pathway in addition to NF- κ B, IL-6, TNF- α , but these effects were reversed by MA	Mice	[235]
Alcohol	Activation of JNK and ERK MAPK pathway mediates alcohol-induced oxidative stress, but HO-1-derived CO reversed these effects by activating p38 MAPK pathway just as CORM-2, which suppressed TNF- α and IL-6	Adult male Balb/c mice, primary rat hepatocytes	[236]
Alcohol	TLR2/4, p38/ERK MAPK pathway, IL-1 β , TNF- α , COX-2 mediate alcohol-induced liver injury, but noni juice (NJ) effectively reverses alcohol-induced liver injury by modulating the above factors	Mice	[237]
Alcohol	Alcohol-induced hepatocyte apoptosis is mediated through activation of p38/JNK MAPK pathway and also involve Fas	Human liver adenocarcinoma (SK-Hep1) cells	[238]
LPS	LPS induces liver inflammation <i>via</i> multiple pathways including activation of p38 MAPK/Nrf2/HO-1, ICAM-1, VCAM-1, TNF- α	RAW264.7 cells, CLP-induced septic mice	[239]
Alcohol	ERK MAPK activation, increase in mRNA transcripts of egr-1 and PAI-1 are associated with alcohol-induced steatosis and hepatic necrosis	Rats	[240]
LPS	Activation of p38 MAPK pathway and COX-2 mediate LPS-induced liver injury, however, ES attenuates liver injury by modulating the above pathways	Sprague-Dawley (SD) rats	[241]
Alcohol	Activation of p38, JNK and ERK MAPK pathways and histone modification (acetylation, methylation and phosphorylation) mediates alcohol-induced hepatic cellular injury	Male SD rats	[242]
Alcohol	Alcohol enhances Fas-induced liver injury by activating p38/JNK MAPK pathway, increase caspase-3 and -8 and TNF- α	Mice	[243]

LPS	Alcohol induces CYP2E1 and LPS overproduction, and CYP2E1 sensitizes hepatocytes to LPS/TNF- α -dependent injury and this is mediated through activation of p38/JNK MAPK pathway		[244]
Alcohol	Inhibition of liver regeneration in partial hepatectomized rats is associated with alcohol-induced p38 activation and cyclin D1 expression	Male Wistar rats	[245]
Alcohol	Alcohol-induced inhibition of HO-1 is mediated through blockade of p38/ERK MAPK-dependent nuclear import of Nrf2, but quercetin can reverse this blockade to restore hepatoprotection against alcohol-induced oxidative stress	Human hepatocytes	[246]
Alcohol	Increased gastric mRNA transcripts was reported as a response to alcohol-dependent noxae on the gut wall, suggesting the protective role of PAI-1 in the gut	C57BL/6 mice, PAI-1-H/K β mice	[247]
Alcohol, LPS	Increase in PAI-1 correlated with progression of ALD PAI-1 was implicated in hepatic inflammation and fibrosis in a two-hit model of ALD involving alcohol and LPS Alcohol-induced increase in hepatic lipids was linked to up-regulation of PAI-1, but this was reversed by metformin	<i>In vitro</i> and <i>in vivo</i> models of ALD, mice	[199-201]

ALD: Alcoholic liver disease; HGF: Hepatocyte growth factor; ECM: Extracellular matrix; PAI: Plasminogen activator inhibitor; PI3K: Phosphatidylinositol 3-kinase; EGF: Epidermal growth factor; MAPK: Mitogen activated protein kinase; TGF- β : Transforming growth factor beta; Smad: Small mother against decapentaplegic; SAMe: S-adenosyl-L-methionine.

Canonical TGF- β signaling

The canonical TGF- β signaling is mediated primarily by Smad proteins (Smad2, Smad3, Smad4) *via* TGF- β -specific receptors (T β R I, T β R II and T β R III). Trans-membrane TGF- β signaling begins with ligand binding of T β R III, which then presents TGF- β to T β R II^[180], this is peculiar to TGF- β 2 which only interacts with T β R III before it becomes bound to T β R II. However, the other TGF- β family members, precisely TGF- β 1 and TGF- β 3 readily bind to T β R II without the need of binding to T β R III; therefore they can transduce intracellular signals either in the presence or absence of T β R III^[17]. TGF- β activated T β R II subsequently trans-phosphorylate T β R I. Activated T β R-I in turn trans-phosphorylate latent transcriptional factors (Smad2 and Smad3) at their C-terminal SXS motif^[17]. The phosphorylated Smad2/3 undergoes a rapid conformational change which facilitates their oligomerization with a common Smad4. The formation of Smad2/3/4 complex enhances preferential nuclear relocation and accumulation of the complex^[181-183]. The cellular responses to TGF- β are fine-tuned by continuous nucleocytoplasmic shuttling of Smad2/3, which permits continuous sensing and responds to changes in TGF- β receptor activity^[184,185]. The nucleocytoplasmic shuttling of Smad2/3/4 complex is crucial for TGF- β signaling and this is aided by karyopherins, particularly Imp7/8. In the nucleus, the Smad2/3/4 complex interacts with transcriptional co-activators or repressors to determine the transcription of TGF- β target genes, such as PAI-1 gene, therefore deciding the fate of cells^[186].

Though alcohol and its metabolic derivatives in many studies (Table 1) have been shown to stimulate increased expression of PAI-1, it remains to be explored how alcohol and its metabolic derivatives modulate Imp7/8 which facilitates Smad2/3/4 nucleocytoplasmic shuttling. It must be noted that TGF- β /Smad signaling mediated through C-terminal phosphorylation of Smad2/3 corresponds to all its tumor suppressor and cytostatic functions. The exact

modulation of canonical TGF- β /Smad, particularly Smad2/3/4 and their regulation by alcohol and its metabolic derivatives need to be clarified, in the light of gut-dependent and liver-dependent alcohol-induced inflammatory and fibrogenic signals. It is currently held that ALD may begin from the gut^[187-190]. A paradigm that does not only highlight the therapeutic potential of the gut^[191] but also the prognostic and diagnostic value of the gut. The possible crosstalk between LPS/TLR4/TNF- α /TNF- α RI/MLCK in gut dysbiosis/inflammation and alcohol-induced liver inflammation/fibrogenesis driven by dysregulated TGF- β /Smad and the MAPK pathways remain veiled. Unveiling the link between the gut-liver axis in the light of TGF- β /Smad/MAPK and TLR4/TNF- α RI signaling will undoubtedly not only help to explain the switch of TGF- β signaling from tumor suppression to tumor promotion in ALD, but will also open a new therapeutic avenue to advance target specific therapies against ALD and fibro-hepatocarcinogenesis.

Non-canonical TGF- β signaling

Aside the canonical TGF- β signaling *via* TGF- β -specific membrane receptors (T β R II and T β R I) and latent Smad proteins (Smad2, Smad3 and Smad4), TGF- β also activates other signaling pathways quiet independently. Among the signaling pathways activated by TGF- β are the MAPK pathway^[192-194], the growth and survival kinases (P13K, AKT/PKB, mTOR) and the small GTP-binding proteins (Ras, RhoA, Racl, and Cdc42)^[195-197]. It is worth notice that oncogenic non-canonical TGF- β signaling in its full activation as is the case in ALD and fibro-hepatocarcinogenesis far outweigh the canonical tumor suppressor TGF- β /Smad signaling and could therefore explain the connivance between alcohol and its metabolic derivatives and the oncogenic non-canonical TGF- β signaling in ALD, increasing the risk of fibro-hepatocarcinogenesis. This is evident in the elevated expression of PAI-1 in some cell and animal models of ALD and HCC. Importantly, PAI-1 is a key target gene of TGF- β signaling and plays

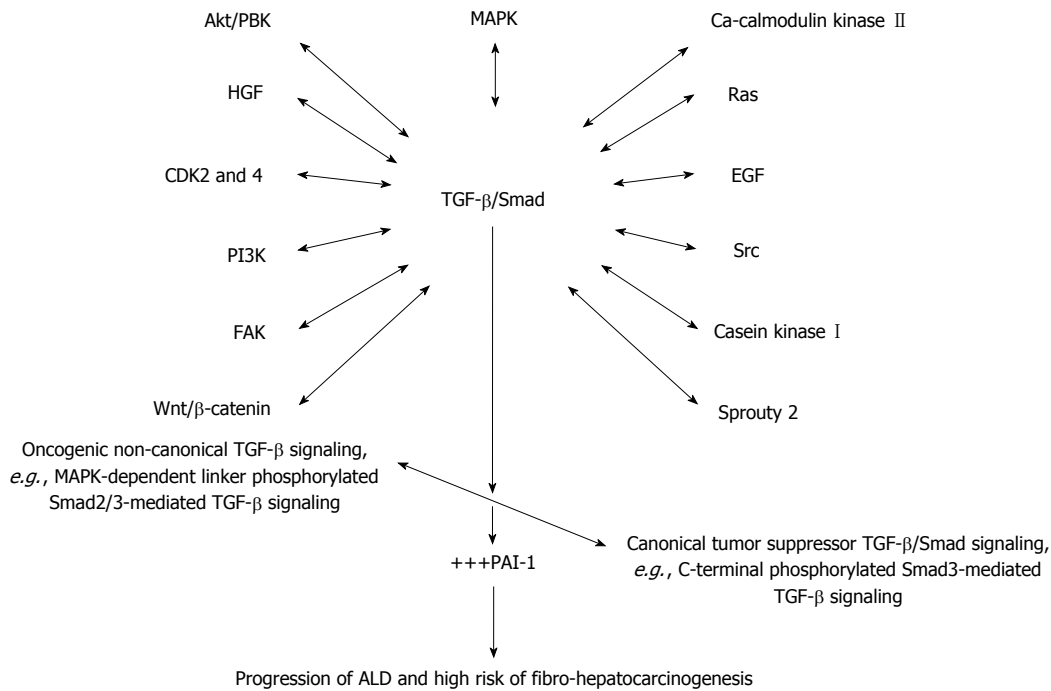


Figure 3 A proposed schematic illustration of the complex and integrative role of TGF-β/Smad signaling in alcoholic liver disease and alcohol-induced fibro-hepatocarcinogenesis. Alcohol and its metabolic derivatives induce the release and activation of TGF-β/Smad signaling through NF-κB/TNF-α mediation (Figure 1). The NF-κB/TNF-α-mediated activation of TGF-β/Smad signaling switches canonical tumor suppressor (C-terminal phosphorylated Smad3/Smad4-dependent TGF-β signaling) into oncogenic (MAPK-dependent linker phosphorylated Smad2/3-dependent TGF-β signaling) and also non-canonical TGF-β signaling involving cross-signaling with other signaling pathways implicated in hepatic malignancies. Key cross-signaling pathways which team up with TGF-β signaling includes but not limited to CDK 2 and 4, Ca-calmodulin kinase II, EGF, HGF, PI3K/AKT, FAK, Src, Sprouty2, casein kinase I, Wnt/β-catenin. This leads to imbalance between canonical and non-canonical TGF-β signaling. Increase in oncogenic TGF-β/Smad signaling leads to up-regulation of PAI-1 gene expression and PAI-1-mediated pathologies thereof. The key pathological effects of PAI-1 include dysregulated ECM regulation, cell proliferation and invasion, and dysregulated apoptosis and these underlie initiation and progression of alcohol-induced fibro-hepatocarcinogenesis. HGF: Hepatocyte growth factor; ECM: Extracellular matrix; PAI: Plasminogen activator inhibitor; PI3K: Phosphatidylinositol 3-kinase; EGF: Epidermal growth factor; MAPK: Mitogen activated protein kinase; TGF-β: Transforming growth factor beta; Smad: Small mother against decapentaplegic; +++: Up-regulated expression or overproduction.

crucial roles in all disease pathologies in which TGF-β is implicated.

Plasminogen activator inhibitor

Plasminogen activator inhibitor (*PAI*)-1 gene is an important inhibitor of both tissue and urokinase type plasminogen activators; as a result it inhibits fibrin degradation *via* inactivation of plasminogen^[6] and also mediates some inflammatory responses^[198]. Many reports have implicated PAI-I in alcohol-induced liver damage^[199-201]. PAI-I levels were shown to increase following acute and chronic alcohol exposure^[200], and a decrease in PAI-1 expression perhaps through RNA interference significantly reduced alcohol-induced steatosis and lipid peroxidation^[200].

Fibrinolysis has been shown to be a common feature in alcohol-induced liver disease; meanwhile PAI-1 gene is a crucial modulator of fibrinolysis through its inhibitory action on plasminogen activator^[201]. The involvement of PAI-1 in alcohol-induced liver damage could be traced to the link between TNF-α and TGF-β/Smad signaling and could also explain the integrative role of TGF-β in the gut-liver axis in alcohol-induced fibro-hepatocarcinogenesis, since PAI-1 is a major target gene of TGF-β/Smad signaling pathway^[31,32,202].

The complicity of TNF-α in alcohol-induced fatty liver disease, where it was purported to increase the expression of PAI-1,^[6] emphasizes a more complex interaction between pro-inflammatory and fibrogenic factors in ALD. TNF-α mediates the pro-inflammatory signaling while TGF-β₁ controls the fibrogenic signals. To confirm the TNF-α/PAI-1 link, alcohol-induced expression of PAI-1 in TNFR1 -/- mice was investigated and found inhibited^[200], indicating that TNF-α might induce PAI-1 through the MAPK pathway^[203]. But the crosstalk between the MAPK pathway and TGF-β/Smad signaling pathway is a common dysregulated signaling pathway implicated in many cancers, particularly MAPK-dependent linker phosphorylation of Smad2 and Smad3 leading to increased PAI-1 expression and occurrence of phenotypic hallmarks of fibro-hepatocarcinogenesis^[32].

MAPK pathway

Recent developments in the understanding of TGF-β/Smad signaling has revealed non-Smad TGF-β signaling^[204] and a growing indication of a crosstalk between MAPK pathway and Smad signaling downstream of TGF-β^[205]. The MAPKs represent a large class of serine/threonine protein kinases crucial in the initial

responses to a diversity of extracellular signals involved in cell growth, cell differentiation and apoptosis^[206] and activation of nuclear transcription factors by allowing nuclear sensing of extracellular signals^[207]. They are grouped into three sub-classes: the extracellular signal-regulated kinases (ERK1, ERK2), the stress-activated protein (SAP) kinases, also known as c-jun N-terminal kinases (JNK1, JNK2, and JNK3), and the p38 MAPKs (α , β , γ , and δ)^[193]. In several reports, TGF- β was shown to activate ERK, p38, and JNK in different cell types^[208,209]. As a result, TGF- β signaling can be regulated *via* linker-dependent phosphorylation of Smad2/3. A number of protein kinases including ERK1/2^[193], JNK^[210] and p38^[211] activated by TGF- β in turn phosphorylate Smad2/3 at the linker region. It is a common knowledge that linker-dependent phosphorylation of Smad2/3 in part switches the TGF- β /Smad signaling from tumor suppression to tumor promotion.

Notably, alcohol and its metabolic derivatives have been shown to activate the MAPK pathway, and this was reported to be crucial for collagen formation in HSCs^[212]. Alcohol-dependent activation of the MAPK pathway depended on the type of hepatic cell and their physiological state, duration of alcohol exposure, and the type of agonist^[212]. With specific reference to KCs, TNF- α production and release was linked to chronic alcohol exposure and LPS-induced activation of p38 MAPK^[212]. Similarly, LPS-induced activation of ERK and p38 MAPKs was shown to be responsible for liver injury^[213]. While elucidating how alcohol and its metabolic derivatives activate the MAPK pathway, Yao *et al.*^[214] showed that CD14-mediated LPS recognition of TLR4/MD-2 complex mediates TNF- α release secondary to MAPK activation. To stoke more arguments and perhaps stimulate search for explanation, it was reported that activation of p38 MAPK is indispensable for hepatocyte proliferation, while sustained activation of the MAPKs reverses this effect^[215,216]. The TGF- β /Smad and the MAPK signaling pathways may be crucial in the initiation and progression of ALD and fibro-hepatocarcinogenesis. It is apparent that a signaling loop possibly involves LPS/TLR4/MD-2/TNF- α -MAPK and TGF- β /Smad crosstalk at multiple levels and plays key roles in ALD as well as recruitment of other signaling pathways implicated in ALD (Figure 3). Some of the signaling pathways recruited into the TGF- β /Smad/MAPK signaling nexus may include but not limited to Spry2^[217], EGF and FGF^[218], Ras and Wnt/ β -catenin^[219]. For instance, TGF- β ₁ was shown to down-regulate Spry2 in a Smad-dependent fashion^[217], meanwhile, Spry2 aids phosphorylation of PTEN and its nuclear accumulation to induce p53-mediated growth arrest^[220] which perhaps underlies Spry2-dependent inhibition of HCC cell growth and inhibition of c-Met-induced proliferation and angiogenesis in fibro-hepatocarcinogenesis^[218]. But PTEN, which negatively regulates PI3K/AKT signaling, is in turn down-regulated by TGF- β ₁^[31]. However, it is yet to be determined how alcohol and its metabolic

derivatives modulate the tumor suppressor PTEN *via* the TGF- β /Smad/MAPK pathways. Will selective inhibition of TGF- β specific receptors, MAPK-dependent phosphorylation of Smad2/3, TNF- α /TNF- α RI in ALD enhance PTEN expression to halt ALD and the risk of fibro-hepatocarcinogenesis?

CONCLUSION

The structural and functional integrity of the liver is anchored on three pillars: effective modulation of inflammation, oxidative/nitrosative stress, and the innate and adaptive immune systems. From the foregoing, it appears that alcohol and its metabolic derivatives disrupt these three cardinal hepatic functions through reprogramming the functions of hepatic cells to favor ALD and fibro-hepatocarcinogenesis progression through concerted interplay of LPS/TLR4/MD-2/TNF- α -MAPK and TGF- β /Smad signaling. Consequently, alcohol primes the liver to diverse irritants and also increases the sensitivity of hepatic cells to inflammatory nosae derived from non-alcohol sources such as those from co-morbidity factors. This may underlie the spectrum of the pathological features of ALD and other liver disorders, in view of the fact that alcoholics and non-alcoholics alike have in one way or the other been exposed to alcohol and its metabolic derivatives once in their life time either through *de novo* biosynthesis of alcohol from food or from some endogenous substrates under conditions of low oxygen tension such as hypoxia.

Etiologically, metabolic derivatives of alcohol as well as alcohol-dependent alteration of gut microbiome, derangements of gut wall, increased production of PAMPs such as LPS and their subsequent leakage into portal circulation, certainly are the significant alcohol-induced disrupters of inflammatory, innate and adaptive immune regulation within the gut-liver axis. Alcohol-induced liver inflammation, liver fibrosis, and cirrhosis leading to increased risk of HCC may not follow a specific temporal sequence but could follow a multi-dimensional pattern determined in part by the existence of non-alcohol co-morbidity factors.

Mechanistically, alcohol and its metabolic derivatives provide a pathological platform for concerted interaction between pro-inflammatory factors (NF- κ B, TNF- α , and IL-1 β), pro-fibrogenic factors (TGF- β , Smad, MAPK and PAI-1) and recruitment of other signaling pathways such as the PI3K, Sprouty2 in a TGF- β /Smad-dependent fashion to promote ALD and fibro-hepatocarcinogenesis. TGF- β /Smad/MAPK and their associated cross-signaling nexus must be seen as indispensable in ALD and fibro-hepatocarcinogenesis.

At the research front, it is important that future studies on ALD and fibro-hepatocarcinogenesis focus on experimental models that will permit study of each of the distinct alcohol-derived inflammatory nosae, *i.e.*, AA, MDA, ROS, LPS, AA/MDA-derived protein/DNA adducts and also delineation of nosae-specific pathways, while at the same time excluding overlap

from co-morbidity factors. This will certainly further expand our understanding and discourse on ALD and fibro-hepatocarcinogenesis to guarantee informed prognosis, diagnosis and treatment.

Therapeutically, TGF- β receptor inhibitors, Smad3L-specific inhibitors, MAPK-specific inhibitors, TNF- α /TNF- α RI-specific inhibitors as well as gut-specific therapeutic strategies must feature prominently in therapies designed for ALD and fibro-hepatocarcinogenesis.

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REFERENCES

- Liu JD, Leung KW, Wang CK, Liao LY, Wang CS, Chen PH, Chen CC, Yeh EK. Alcohol-related problems in Taiwan with particular emphasis on alcoholic liver diseases. *Alcohol Clin Exp Res* 1998; **22**: 164S-169S [PMID: 9622397]
- Mandayam S, Jamal MM, Morgan TR. Epidemiology of alcoholic liver disease. *Semin Liver Dis* 2004; **24**: 217-232 [PMID: 15349801 DOI: 10.1055/s-2004-832936]
- Teli MR, Day CP, Burt AD, Bennett MK, James OF. Determinants of progression to cirrhosis or fibrosis in pure alcoholic fatty liver. *Lancet* 1995; **346**: 987-990 [PMID: 7475591]
- Gyamfi MA, Wan YJ. Pathogenesis of alcoholic liver disease: the role of nuclear receptors. *Exp Biol Med* (Maywood) 2010; **235**: 547-560 [PMID: 20463294 DOI: 10.1258/ebm.2009.009249]
- Stewart S, Jones D, Day CP. Alcoholic liver disease: new insights into mechanisms and preventative strategies. *Trends Mol Med* 2001; **7**: 408-413 [PMID: 11530336]
- Schaffert CS, Duryee MJ, Hunter CD, Hamilton BC, DeVeny AL, Huerter MM, Klassen LW, Thiele GM. Alcohol metabolites and lipopolysaccharide: roles in the development and/or progression of alcoholic liver disease. *World J Gastroenterol* 2009; **15**: 1209-1218 [PMID: 19291821 DOI: 10.3748/wjg.15.1209]
- Ceni E, Mello T, Galli A. Pathogenesis of alcoholic liver disease: role of oxidative metabolism. *World J Gastroenterol* 2014; **20**: 17756-17772 [PMID: 25548474 DOI: 10.3748/wjg.v20.i47.17756]
- Shepard BD, Fernandez DJ, Tuma PL. Alcohol consumption impairs hepatic protein trafficking: mechanisms and consequences. *Genes Nutr* 2010; **5**: 129-140 [PMID: 19890673 DOI: 10.1007/s12263-009-0156-z]
- Siegmund SV, Dooley S, Brenner DA. Molecular mechanisms of alcohol-induced hepatic fibrosis. *Dig Dis* 2005; **23**: 264-274 [PMID: 16508291 DOI: 10.1159/000090174]
- Jin M, Ande A, Kumar A, Kumar S. Regulation of cytochrome P450 2e1 expression by ethanol: role of oxidative stress-mediated pkc/jnk/sp1 pathway. *Cell Death Dis* 2013; **4**: e554 [PMID: 23519123 DOI: 10.1038/cddis.2013.78]
- Fernandez DJ, Tuma DJ, Tuma PL. Hepatic microtubule acetylation and stability induced by chronic alcohol exposure impair nuclear translocation of STAT3 and STAT5B, but not Smad2/3. *Am J Physiol Gastrointest Liver Physiol* 2012; **303**: G1402-G1415 [PMID: 23064763 DOI: 10.1152/ajpgi.00071.2012]
- Fritz KS, Green MF, Petersen DR, Hirschey MD. Ethanol metabolism modifies hepatic protein acylation in mice. *PLoS One* 2013; **8**: e75868 [PMID: 24073283 DOI: 10.1371/journal.pone.0075868]
- Farazi PA, DePinho RA. Hepatocellular carcinoma pathogenesis: from genes to environment. *Nat Rev Cancer* 2006; **6**: 674-687 [PMID: 16929323 DOI: 10.1038/nrc1934]
- El-Serag HB. Epidemiology of hepatocellular carcinoma in USA. *Hepatol Res* 2007; **37** Suppl 2: S88-S94 [PMID: 17877502 DOI: 10.1111/j.1872-034X.2007.00168.x]
- Van De Craen B, Declercq PJ, Gils A. The Biochemistry, Physiology and Pathological roles of PAI-1 and the requirements for PAI-1 inhibition in vivo. *Thromb Res* 2012; **130**: 576-585 [PMID: 22801256 DOI: 10.1016/j.thromres.2012.06.023]
- Derynck R, Akhurst RJ. Differentiation plasticity regulated by TGF-beta family proteins in development and disease. *Nat Cell Biol* 2007; **9**: 1000-1004 [PMID: 17762890 DOI: 10.1038/ncb434]
- Tian M, Neil JR, Schiemann WP. Transforming growth factor- β and the hallmarks of cancer. *Cell Signal* 2011; **23**: 951-962 [PMID: 20940046 DOI: 10.1016/j.cellsig.2010.10.015]
- Levy L, Hill CS. Alterations in components of the TGF-beta superfamily signaling pathways in human cancer. *Cytokine Growth Factor Rev* 2006; **17**: 41-58 [PMID: 16310402 DOI: 10.1016/j.cytogfr.2005.09.009]
- Goggins M, Shekher M, Turnacioglu K, Yeo CJ, Hruban RH, Kern SE. Genetic alterations of the transforming growth factor beta receptor genes in pancreatic and biliary adenocarcinomas. *Cancer Res* 1998; **58**: 5329-5332 [PMID: 9850059]
- Markowitz S, Wang J, Myeroff L, Parsons R, Sun L, Lutterbaugh J, Fan RS, Zborowska E, Kinzler KW, Vogelstein B. Inactivation of the type II TGF-beta receptor in colon cancer cells with microsatellite instability. *Science* 1995; **268**: 1336-1338 [PMID: 7761852]
- Parsons R, Myeroff LL, Liu B, Willson JK, Markowitz SD, Kinzler KW, Vogelstein B. Microsatellite instability and mutations of the transforming growth factor beta type II receptor gene in colorectal cancer. *Cancer Res* 1995; **55**: 5548-5550 [PMID: 7585632]
- Myeroff LL, Parsons R, Kim SJ, Hedrick L, Cho KR, Orth K, Mathis M, Kinzler KW, Lutterbaugh J, Park K. A transforming growth factor beta receptor type II gene mutation common in colon and gastric but rare in endometrial cancers with microsatellite instability. *Cancer Res* 1995; **55**: 5545-5547 [PMID: 7585631]
- Maliekal TT, Antony ML, Nair A, Paulmurugan R, Karunakaran D. Loss of expression, and mutations of Smad 2 and Smad 4 in human cervical cancer. *Oncogene* 2003; **22**: 4889-4897 [PMID: 12894231 DOI: 10.1038/sj.onc.1206806]
- Prunier C, Ferrand N, Frottier B, Pessah M, Atfi A. Mechanism for mutational inactivation of the tumor suppressor Smad2. *Mol Cell Biol* 2001; **21**: 3302-3313 [PMID: 11313456 DOI: 10.1128/mcb.21.10.3302-3313.2001]
- Han SU, Kim HT, Seong DH, Kim YS, Park YS, Bang YJ, Yang HK, Kim SJ. Loss of the Smad3 expression increases susceptibility to tumorigenicity in human gastric cancer. *Oncogene* 2004; **23**: 1333-1341 [PMID: 14647420 DOI: 10.1038/sj.onc.1207259]
- Burger B, Uhlhaas S, Mangold E, Propping P, Friedl W, Jenne D, Dockter G, Back W. Novel de novo mutation of MADH4/SMAD4 in a patient with juvenile polyposis. *Am J Med Genet* 2002; **110**: 289-291 [PMID: 12116240 DOI: 10.1002/ajmg.10411]
- Blobe GC, Schiemann WP, Lodish HF. Role of transforming growth factor beta in human disease. *N Engl J Med* 2000; **342**: 1350-1358 [PMID: 10793168 DOI: 10.1056/nejm200005043421807]
- Pasche B. Role of transforming growth factor beta in cancer. *J Cell Physiol* 2001; **186**: 153-168 [PMID: 11169452 DOI: 10.1002/1097-4652(200002)186:2]
- Jakowlew SB. Transforming growth factor-beta in cancer and metastasis. *Cancer Metastasis Rev* 2006; **25**: 435-457 [PMID: 16951986 DOI: 10.1007/s10555-006-9006-2]
- Shi Y, Massagué J. Mechanisms of TGF-beta signaling from cell membrane to the nucleus. *Cell* 2003; **113**: 685-700 [PMID: 12809600 DOI: 10.1016/S0092-8674(03)00432-X]
- Boye A, Yang Y. Hepatic microRNA orchestra: a new diagnostic, prognostic and therapeutic tool for hepatocarcinogenesis. *Mini Rev Med Chem* 2014; **14**: 837-852 [PMID: 25342194]
- Boye A, Kan H, Wu C, Jiang Y, Yang X, He S, Yang Y. MAPK inhibitors differently modulate TGF- β /Smad signaling in HepG2

- cells. *Tumour Biol* 2015; **36**: 3643-3651 [PMID: 25560488 DOI: 10.1007/s13277-014-3002-x]
- 33 **Zakhari S**, Li TK. Determinants of alcohol use and abuse: Impact of quantity and frequency patterns on liver disease. *Hepatology* 2007; **46**: 2032-2039 [PMID: 18046720 DOI: 10.1002/hep.22010]
- 34 **Niemelä O**, Parkkila S, Ylä-Herttua S, Villanueva J, Ruebner B, Halsted CH. Sequential acetaldehyde production, lipid peroxidation, and fibrogenesis in micropig model of alcohol-induced liver disease. *Hepatology* 1995; **22**: 1208-1214 [PMID: 7557872]
- 35 **Sladek NE**, Manthey CL, Maki PA, Zhang Z, Landkamer GJ. Xenobiotic oxidation catalyzed by aldehyde dehydrogenases. *Drug Metab Rev* 1989; **20**: 697-720 [PMID: 2680404 DOI: 10.3109/03602538909103572]
- 36 **Gouillon Z**, Lucas D, Li J, Hagbjork AL, French BA, Fu P, Fang C, Ingelman-Sundberg M, Donohue TM, French SW. Inhibition of ethanol-induced liver disease in the intragastric feeding rat model by chlormethiazole. *Proc Soc Exp Biol Med* 2000; **224**: 302-308 [PMID: 10964266]
- 37 **Lieber CS**. CYP2E1: from ASH to NASH. *Hepatol Res* 2004; **28**: 1-11 [PMID: 14734144]
- 38 **Lieber CS**. The discovery of the microsomal ethanol oxidizing system and its physiologic and pathologic role. *Drug Metab Rev* 2004; **36**: 511-529 [PMID: 15554233 DOI: 10.1081/dmr-200033441]
- 39 **Parola M**, Robino G. Oxidative stress-related molecules and liver fibrosis. *J Hepatol* 2001; **35**: 297-306 [PMID: 11580156]
- 40 **Seth D**, Haber PS, Syn WK, Diehl AM, Day CP. Pathogenesis of alcohol-induced liver disease: classical concepts and recent advances. *J Gastroenterol Hepatol* 2011; **26**: 1089-1105 [PMID: 21545524 DOI: 10.1111/j.1440-1746.2011.06756.x]
- 41 **Liu J**. Ethanol and liver: recent insights into the mechanisms of ethanol-induced fatty liver. *World J Gastroenterol* 2014; **20**: 14672-14685 [PMID: 25356030 DOI: 10.3748/wjg.v20.i40.14672]
- 42 **Crabb DW**, Dipple KM, Thomasson HR. Alcohol sensitivity, alcohol metabolism, risk of alcoholism, and the role of alcohol and aldehyde dehydrogenase genotypes. *J Lab Clin Med* 1993; **122**: 234-240 [PMID: 8409698]
- 43 **Crabb DW**, Galli A, Fischer M, You M. Molecular mechanisms of alcoholic fatty liver: role of peroxisome proliferator-activated receptor alpha. *Alcohol* 2004; **34**: 35-38 [PMID: 15670663 DOI: 10.1016/j.alcohol.2004.07.005]
- 44 **Lieber CS**. Alcoholic fatty liver: its pathogenesis and mechanism of progression to inflammation and fibrosis. *Alcohol* 2004; **34**: 9-19 [PMID: 15670660 DOI: 10.1016/j.alcohol.2004.07.008]
- 45 **Scher JU**, Abramson SB. The microbiome and rheumatoid arthritis. *Nat Rev Rheumatol* 2011; **7**: 569-578 [PMID: 21862983 DOI: 10.1038/nrrheum.2011.121]
- 46 **Actis GC**. The gut microbiome. *Inflamm Allergy Drug Targets* 2014; **13**: 217-223 [PMID: 24953716]
- 47 **Seksik P**. Gut microbiota and IBD. *Gastroenterol Clin Biol* 2010; **34** Suppl 1: S44-S51 [PMID: 20889004 DOI: 10.1016/s0399-8320(10)70020-8]
- 48 **Rosenstiel P**. Stories of love and hate: innate immunity and host-microbe crosstalk in the intestine. *Curr Opin Gastroenterol* 2013; **29**: 125-132 [PMID: 23337934 DOI: 10.1097/MOG.0b013e32835da2e7]
- 49 **Ramakrishna BS**. Role of the gut microbiota in human nutrition and metabolism. *J Gastroenterol Hepatol* 2013; **28** Suppl 4: 9-17 [PMID: 24251697 DOI: 10.1111/jgh.12294]
- 50 **Probert HM**, Apajalahti JH, Rautonen N, Stowell J, Gibson GR. Polydextrose, lactitol, and fructo-oligosaccharide fermentation by colonic bacteria in a three-stage continuous culture system. *Appl Environ Microbiol* 2004; **70**: 4505-4511 [PMID: 15294779 DOI: 10.1128/aem.70.8.4505-4511.2004]
- 51 **Elinav E**, Strowig T, Kau AL, Henao-Mejia J, Thaiss CA, Booth CJ, Peaper DR, Bertin J, Eisenbarth SC, Gordon JI, Flavell RA. NLRP6 inflammasome regulates colonic microbial ecology and risk for colitis. *Cell* 2011; **145**: 745-757 [PMID: 21565393 DOI: 10.1016/j.cell.2011.04.022]
- 52 **Ridlon JM**, Kang DJ, Hylemon PB. Bile salt biotransformations by human intestinal bacteria. *J Lipid Res* 2006; **47**: 241-259 [PMID: 16299351 DOI: 10.1194/jlr.R500013-JLR200]
- 53 **Dawson PA**, Lan T, Rao A. Bile acid transporters. *J Lipid Res* 2009; **50**: 2340-2357 [PMID: 19498215 DOI: 10.1194/jlr.R900012-JLR200]
- 54 **Lord RS**, Bralley JA. Clinical applications of urinary organic acids. Part 2. Dysbiosis markers. *Altern Med Rev* 2008; **13**: 292-306 [PMID: 19152477]
- 55 **Serino M**, Blasco-Baque V, Burcelin R. Microbes on-air: gut and tissue microbiota as targets in type 2 diabetes. *J Clin Gastroenterol* 2012; **46** Suppl: S27-S28 [PMID: 22955352 DOI: 10.1097/MCG.0b013e318264e844]
- 56 **Serino M**, Fernández-Real JM, García-Fuentes E, Queipo-Ortuño M, Moreno-Navarrete JM, Sánchez A, Burcelin R, Tinahones F. The gut microbiota profile is associated with insulin action in humans. *Acta Diabetol* 2013; **50**: 753-761 [PMID: 22711164 DOI: 10.1007/s00592-012-0410-5]
- 57 **Leone V**, Chang EB, Devkota S. Diet, microbes, and host genetics: the perfect storm in inflammatory bowel diseases. *J Gastroenterol* 2013; **48**: 315-321 [PMID: 23475322 DOI: 10.1007/s00535-013-0777-2]
- 58 **David LA**, Maurice CF, Carmody RN, Gootenberg DB, Button JE, Wolfe BE, Ling AV, Devlin AS, Varma Y, Fischbach MA, Biddinger SB, Dutton RJ, Turnbaugh PJ. Diet rapidly and reproducibly alters the human gut microbiome. *Nature* 2014; **505**: 559-563 [PMID: 24336217 DOI: 10.1038/nature12820]
- 59 **Nakayama T**, Takeuchi D, Matsumura T, Akeda Y, Fujinaga Y, Oishi K. Alcohol consumption promotes the intestinal translocation of *Streptococcus suis* infections. *Microb Pathog* 2013; **65**: 14-20 [PMID: 24036179 DOI: 10.1016/j.micpath.2013.08.006]
- 60 **Parlesak A**, Schäfer C, Schütz T, Bode JC, Bode C. Increased intestinal permeability to macromolecules and endotoxemia in patients with chronic alcohol abuse in different stages of alcohol-induced liver disease. *J Hepatol* 2000; **32**: 742-747 [PMID: 10845660]
- 61 **Choudhry MA**, Fazal N, Goto M, Gamelli RL, Sayeed MM. Gut-associated lymphoid T cell suppression enhances bacterial translocation in alcohol and burn injury. *Am J Physiol Gastrointest Liver Physiol* 2002; **282**: G937-G947 [PMID: 12016118 DOI: 10.1152/ajpgi.00235.2001]
- 62 **Schenker S**, Bay MK. Alcohol and endotoxin: another path to alcoholic liver injury? *Alcohol Clin Exp Res* 1995; **19**: 1364-1366 [PMID: 8561316]
- 63 **Elamin E**, Masclee A, Troost F, Pieters HJ, Keszthelyi D, Aleksa K, Dekker J, Jonkers D. Ethanol impairs intestinal barrier function in humans through mitogen activated protein kinase signaling: a combined in vivo and in vitro approach. *PLoS One* 2014; **9**: e107421 [PMID: 25226407 DOI: 10.1371/journal.pone.0107421]
- 64 **Yan AW**, Fouts DE, Brandl J, Stärkel P, Torralba M, Schott E, Tsukamoto H, Nelson KE, Brenner DA, Schnabl B. Enteric dysbiosis associated with a mouse model of alcoholic liver disease. *Hepatology* 2011; **53**: 96-105 [PMID: 21254165 DOI: 10.1002/hep.24018]
- 65 **Rao R**. Endotoxemia and gut barrier dysfunction in alcoholic liver disease. *Hepatology* 2009; **50**: 638-644 [PMID: 19575462 DOI: 10.1002/hep.23009]
- 66 **Purohit V**, Bode JC, Bode C, Brenner DA, Choudhry MA, Hamilton F, Kang YJ, Keshavarzian A, Rao R, Sartor RB, Swanson C, Turner JR. Alcohol, intestinal bacterial growth, intestinal permeability to endotoxin, and medical consequences: summary of a symposium. *Alcohol* 2008; **42**: 349-361 [PMID: 18504085 DOI: 10.1016/j.alcohol.2008.03.131]
- 67 **Frasinariu OE**, Ceccarelli S, Alisi A, Moraru E, Nobili V. Gut-liver axis and fibrosis in nonalcoholic fatty liver disease: an input for novel therapies. *Dig Liver Dis* 2013; **45**: 543-551 [PMID: 23280158 DOI: 10.1016/j.dld.2012.11.010]
- 68 **McClain CJ**, Song Z, Barve SS, Hill DB, Deaciuc I. Recent advances in alcoholic liver disease. IV. Dysregulated cytokine

- metabolism in alcoholic liver disease. *Am J Physiol Gastrointest Liver Physiol* 2004; **287**: G497-G502 [PMID: 15331349 DOI: 10.1152/ajpgi.00171.2004]
- 69 **Enomoto N**, Ikejima K, Kitamura T, Oide H, Takei Y, Sato N, Thurman RG. Alcohol enhances lipopolysaccharide-induced increases in nitric oxide production by Kupffer cells via mechanisms dependent on endotoxin. *Alcohol Clin Exp Res* 2000; **24**: 55S-58S [PMID: 10803781]
- 70 **Fukui H**, Brauner B, Bode JC, Bode C. Plasma endotoxin concentrations in patients with alcoholic and non-alcoholic liver disease: reevaluation with an improved chromogenic assay. *J Hepatol* 1991; **12**: 162-169 [PMID: 2050995]
- 71 **Nanji AA**, Khettry U, Sadrzadeh SM, Yamanaka T. Severity of liver injury in experimental alcoholic liver disease. Correlation with plasma endotoxin, prostaglandin E2, leukotriene B4, and thromboxane B2. *Am J Pathol* 1993; **142**: 367-373 [PMID: 8382006]
- 72 **Seki E**, Schnabl B. Role of innate immunity and the microbiota in liver fibrosis: crosstalk between the liver and gut. *J Physiol* 2012; **590**: 447-458 [PMID: 22124143 DOI: 10.1113/jphysiol.2011.219691]
- 73 **Yang L**, Seki E. Toll-like receptors in liver fibrosis: cellular crosstalk and mechanisms. *Front Physiol* 2012; **3**: 138 [PMID: 22661952 DOI: 10.3389/fphys.2012.00138]
- 74 **Schwabe RF**, Seki E, Brenner DA. Toll-like receptor signaling in the liver. *Gastroenterology* 2006; **130**: 1886-1900 [PMID: 16697751 DOI: 10.1053/j.gastro.2006.01.038]
- 75 **Miyake K**. Endotoxin recognition molecules MD-2 and toll-like receptor 4 as potential targets for therapeutic intervention of endotoxin shock. *Curr Drug Targets Inflamm Allergy* 2004; **3**: 291-297 [PMID: 15379597]
- 76 **Miyake K**. Innate recognition of lipopolysaccharide by Toll-like receptor 4-MD-2. *Trends Microbiol* 2004; **12**: 186-192 [PMID: 15051069 DOI: 10.1016/j.tim.2004.02.009]
- 77 **Freudenberg MA**, Tchaptchet S, Keck S, Fejer G, Huber M, Schütze N, Beutler B, Galanos C. Lipopolysaccharide sensing an important factor in the innate immune response to Gram-negative bacterial infections: benefits and hazards of LPS hypersensitivity. *Immunobiology* 2008; **213**: 193-203 [PMID: 18406367 DOI: 10.1016/j.imbio.2007.11.008]
- 78 **Guo J**, Friedman SL. Toll-like receptor 4 signaling in liver injury and hepatic fibrogenesis. *Fibrogenesis Tissue Repair* 2010; **3**: 21 [PMID: 20964825 DOI: 10.1186/1755-1536-3-21]
- 79 **Akira S**, Takeda K. Toll-like receptor signalling. *Nat Rev Immunol* 2004; **4**: 499-511 [PMID: 15229469 DOI: 10.1038/nri1391]
- 80 **Akira S**, Takeda K. Functions of toll-like receptors: lessons from KO mice. *C R Biol* 2004; **327**: 581-589 [PMID: 15330257]
- 81 **Takeda K**, Akira S. TLR signaling pathways. *Semin Immunol* 2004; **16**: 3-9 [PMID: 14751757]
- 82 **Braet F**, Wisse E. Structural and functional aspects of liver sinusoidal endothelial cell fenestrae: a review. *Comp Hepatol* 2002; **1**: 1 [PMID: 12437787]
- 83 **Knolle PA**, Limmer A. Control of immune responses by scavenger liver endothelial cells. *Swiss Med Wkly* 2003; **133**: 501-506 [PMID: 14652798]
- 84 **Wang BY**, Ju XH, Fu BY, Zhang J, Cao YX. Effects of ethanol on liver sinusoidal endothelial cells-fenestrae of rats. *Hepatobiliary Pancreat Dis Int* 2005; **4**: 422-426 [PMID: 16109529]
- 85 **Deleve LD**, Wang X, Guo Y. Sinusoidal endothelial cells prevent rat stellate cell activation and promote reversion to quiescence. *Hepatology* 2008; **48**: 920-930 [PMID: 18613151 DOI: 10.1002/hep.22351]
- 86 **Thiele GM**, Duryee MJ, Freeman TL, Sorrell MF, Willis MS, Tuma DJ, Klassen LW. Rat sinusoidal liver endothelial cells (SECs) produce pro-fibrotic factors in response to adducts formed from the metabolites of ethanol. *Biochem Pharmacol* 2005; **70**: 1593-1600 [PMID: 16202982 DOI: 10.1016/j.bcp.2005.08.014]
- 87 **Duryee MJ**, Freeman TL, Willis MS, Hunter CD, Hamilton BC, Suzuki H, Tuma DJ, Klassen LW, Thiele GM. Scavenger receptors on sinusoidal liver endothelial cells are involved in the uptake of aldehyde-modified proteins. *Mol Pharmacol* 2005; **68**: 1423-1430 [PMID: 16105988 DOI: 10.1124/mol.105.016121]
- 88 **Thiele GM**, Duryee MJ, Willis MS, Sorrell MF, Freeman TL, Tuma DJ, Klassen LW. Malondialdehyde-acetaldehyde (MAA) modified proteins induce pro-inflammatory and pro-fibrotic responses by liver endothelial cells. *Comp Hepatol* 2004; **3** Suppl 1: S25 [PMID: 14960177 DOI: 10.1186/1476-5926-2-s1-s25]
- 89 **Duryee MJ**, Klassen LW, Freeman TL, Willis MS, Tuma DJ, Thiele GM. Lipopolysaccharide is a cofactor for malondialdehyde-acetaldehyde adduct-mediated cytokine/chemokine release by rat sinusoidal liver endothelial and Kupffer cells. *Alcohol Clin Exp Res* 2004; **28**: 1931-1938 [PMID: 15608611]
- 90 **Ramadori G**, Moriconi F, Malik I, Dudas J. Physiology and pathophysiology of liver inflammation, damage and repair. *J Physiol Pharmacol* 2008; **59** Suppl 1: 107-117 [PMID: 18802219]
- 91 **Haubrich WS**. Kupffer of Kupffer cells. *Gastroenterology* 2004; **127**: 16 [PMID: 15236167]
- 92 **Stachura J**, Gałazka K. History and current status of Polish gastroenterological pathology. *J Physiol Pharmacol* 2003; **54** Suppl 3: 183-192 [PMID: 15075472]
- 93 **Naito M**, Hasegawa G, Takahashi K. Development, differentiation, and maturation of Kupffer cells. *Microsc Res Tech* 1997; **39**: 350-364 [PMID: 9407545 DOI: 10.1002/(sici)1097-0029(19971115)39]
- 94 **Helmy KY**, Katschke KJ, Gorgani NN, Kljavin NM, Elliott JM, Diehl L, Scales SJ, Ghilardi N, van Lookeren Campagne M. CRIG: a macrophage complement receptor required for phagocytosis of circulating pathogens. *Cell* 2006; **124**: 915-927 [PMID: 16530040 DOI: 10.1016/j.cell.2005.12.039]
- 95 **Ramadori G**, Saile B. Inflammation, damage repair, immune cells, and liver fibrosis: specific or nonspecific, this is the question. *Gastroenterology* 2004; **127**: 997-1000 [PMID: 15362057]
- 96 **Neubauer K**, Lindhorst A, Tron K, Ramadori G, Saile B. Decrease of PECAM-1-gene-expression induced by proinflammatory cytokines IFN-gamma and IFN-alpha is reversed by TGF-beta in sinusoidal endothelial cells and hepatic mononuclear phagocytes. *BMC Physiol* 2008; **8**: 9 [PMID: 18466611 DOI: 10.1186/1472-6793-8-9]
- 97 **Bautista AP**, Spitzer JJ. Cross-tolerance between acute alcohol intoxication and endotoxemia. *Alcohol Clin Exp Res* 1996; **20**: 1395-1400 [PMID: 8947315]
- 98 **D'Souza NB**, Nelson S, Summer WR, Deaciuc IV. Alcohol modulates alveolar macrophage tumor necrosis factor-alpha, superoxide anion, and nitric oxide secretion in the rat. *Alcohol Clin Exp Res* 1996; **20**: 156-163 [PMID: 8651446]
- 99 **Sarphie TG**, D'Souza NB, Deaciuc IV. Kupffer cell inactivation prevents lipopolysaccharide-induced structural changes in the rat liver sinusoid: an electron-microscopic study. *Hepatology* 1996; **23**: 788-796 [PMID: 8666333 DOI: 10.1002/hep.510230420]
- 100 **Watanabe N**, Suzuki J, Kobayashi Y. Role of calcium in tumor necrosis factor-alpha production by activated macrophages. *J Biochem* 1996; **120**: 1190-1195 [PMID: 9010769]
- 101 **Ajakaiye MA**, Jacob A, Wu R, Zhou M, Ji Y, Dong W, Wang Z, Qiang X, Chaung WW, Nicastro J, Coppa GF, Wang P. Upregulation of Kupffer cell $\alpha 2A$ -Adrenoceptors and downregulation of MKP-1 mediate hepatic injury in chronic alcohol exposure. *Biochem Biophys Res Commun* 2011; **409**: 406-411 [PMID: 21575605 DOI: 10.1016/j.bbrc.2011.05.007]
- 102 **Thurman RG**. II. Alcoholic liver injury involves activation of Kupffer cells by endotoxin. *Am J Physiol* 1998; **275**: G605-G611 [PMID: 9756487]
- 103 **Enomoto N**, Ikejima K, Bradford B, Rivera C, Kono H, Brenner DA, Thurman RG. Alcohol causes both tolerance and sensitization of rat Kupffer cells via mechanisms dependent on endotoxin. *Gastroenterology* 1998; **115**: 443-451 [PMID: 9679050]
- 104 **Niemelä O**. Distribution of ethanol-induced protein adducts in vivo: relationship to tissue injury. *Free Radic Biol Med* 2001; **31**: 1533-1538 [PMID: 11744326]

- 105 **Tuma DJ**. Role of malondialdehyde-acetaldehyde adducts in liver injury. *Free Radic Biol Med* 2002; **32**: 303-308 [PMID: 11841919]
- 106 **Enomoto N**, Ikejima K, Yamashina S, Enomoto A, Nishiura T, Nishimura T, Brenner DA, Schemmer P, Bradford BU, Rivera CA, Zhong Z, Thurman RG. Kupffer cell-derived prostaglandin E(2) is involved in alcohol-induced fat accumulation in rat liver. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G100-G106 [PMID: 10898751]
- 107 **Enomoto N**, Ikejima K, Bradford BU, Rivera CA, Kono H, Goto M, Yamashina S, Schemmer P, Kitamura T, Oide H, Takei Y, Hirose M, Shimizu H, Miyazaki A, Brenner DA, Sato N, Thurman RG. Role of Kupffer cells and gut-derived endotoxins in alcoholic liver injury. *J Gastroenterol Hepatol* 2000; **15** Suppl: D20-D25 [PMID: 10759216]
- 108 **Zhang X**, Yu WP, Gao L, Wei KB, Ju JL, Xu JZ. Effects of lipopolysaccharides stimulated Kupffer cells on activation of rat hepatic stellate cells. *World J Gastroenterol* 2004; **10**: 610-613 [PMID: 14966928 DOI: 10.3748/wjg.v10.i4.610]
- 109 **Ramadori G**, Saile B. Portal tract fibrogenesis in the liver. *Lab Invest* 2004; **84**: 153-159 [PMID: 14688800 DOI: 10.1038/labinvest.3700030]
- 110 **Saile B**, Ramadori G. Inflammation, damage repair and liver fibrosis--role of cytokines and different cell types. *Z Gastroenterol* 2007; **45**: 77-86 [PMID: 17236124 DOI: 10.1055/s-2006-927395]
- 111 **Day SA**, Lakner AM, Moore CC, Yen MH, Clemens MG, Wu ES, Schrum LW. Opioid-like compound exerts anti-fibrotic activity via decreased hepatic stellate cell activation and inflammation. *Biochem Pharmacol* 2011; **81**: 996-1003 [PMID: 21291870 DOI: 10.1016/j.bcp.2011.01.015]
- 112 **Friedman SL**. Preface. Hepatic fibrosis: pathogenesis, diagnosis, and emerging therapies. *Clin Liver Dis* 2008; **12**: xiii-xxiv [PMID: 18984462 DOI: 10.1016/j.cld.2008.07.009]
- 113 **Knittel T**, Dinter C, Kobold D, Neubauer K, Mehde M, Eichhorst S, Ramadori G. Expression and regulation of cell adhesion molecules by hepatic stellate cells (HSC) of rat liver: involvement of HSC in recruitment of inflammatory cells during hepatic tissue repair. *Am J Pathol* 1999; **154**: 153-167 [PMID: 9916930]
- 114 **Knittel T**, Mehde M, Kobold D, Saile B, Dinter C, Ramadori G. Expression patterns of matrix metalloproteinases and their inhibitors in parenchymal and non-parenchymal cells of rat liver: regulation by TNF-alpha and TGF-beta1. *J Hepatol* 1999; **30**: 48-60 [PMID: 9927150]
- 115 **Saile B**, Matthes N, Knittel T, Ramadori G. Transforming growth factor beta and tumor necrosis factor alpha inhibit both apoptosis and proliferation of activated rat hepatic stellate cells. *Hepatology* 1999; **30**: 196-202 [PMID: 10385656 DOI: 10.1002/hep.510300144]
- 116 **Marra F**, Delogu W, Petrai I, Pastacaldi S, Bonacchi A, Efsen E, Aleffi S, Bertolani C, Pinzani M, Gentilini P. Differential requirement of members of the MAPK family for CCL2 expression by hepatic stellate cells. *Am J Physiol Gastrointest Liver Physiol* 2004; **287**: G18-G26 [PMID: 15016614 DOI: 10.1152/ajpgi.00336.2003]
- 117 **Schwabe RF**, Bataller R, Brenner DA. Human hepatic stellate cells express CCR5 and RANTES to induce proliferation and migration. *Am J Physiol Gastrointest Liver Physiol* 2003; **285**: G949-G958 [PMID: 12829440 DOI: 10.1152/ajpgi.00215.2003]
- 118 **Paik YH**, Schwabe RF, Bataller R, Russo MP, Jobin C, Brenner DA. Toll-like receptor 4 mediates inflammatory signaling by bacterial lipopolysaccharide in human hepatic stellate cells. *Hepatology* 2003; **37**: 1043-1055 [PMID: 12717385 DOI: 10.1053/jhep.2003.50182]
- 119 **Wang JH**, Batey RG, George J. Role of ethanol in the regulation of hepatic stellate cell function. *World J Gastroenterol* 2006; **12**: 6926-6932 [PMID: 17109512 DOI: 10.3748/wjg.v12.i43.6926]
- 120 **Karaa A**, Thompson KJ, McKillop IH, Clemens MG, Schrum LW. S-adenosyl-L-methionine attenuates oxidative stress and hepatic stellate cell activation in an ethanol-LPS-induced fibrotic rat model. *Shock* 2008; **30**: 197-205 [PMID: 18180699 DOI: 10.1097/shk.0b013e318160f417]
- 121 **Klassen LW**, Thiele GM, Duryee MJ, Schaffert CS, DeVeeney AL, Hunter CD, Olinga P, Tuma DJ. An in vitro method of alcoholic liver injury using precision-cut liver slices from rats. *Biochem Pharmacol* 2008; **76**: 426-436 [PMID: 18599023 DOI: 10.1016/j.bcp.2008.05.012]
- 122 **Shi H**, Dong L, Dang X, Liu Y, Jiang J, Wang Y, Lu X, Guo X. Effect of chlorogenic acid on LPS-induced proinflammatory signaling in hepatic stellate cells. *Inflamm Res* 2013; **62**: 581-587 [PMID: 23483217 DOI: 10.1007/s00011-013-0610-7]
- 123 **Purohit V**, Brenner DA. Mechanisms of alcohol-induced hepatic fibrosis: a summary of the Ron Thurman Symposium. *Hepatology* 2006; **43**: 872-878 [PMID: 16502397 DOI: 10.1002/hep.21107]
- 124 **Ha MH**, Wei L, Rao HY, Liu F, Wang XY, Feng B, Fei R, Chen HS, Cong X. Effect of interferon-gamma on hepatic stellate cells stimulated by acetaldehyde. *Hepatogastroenterology* 2008; **55**: 1059-1065 [PMID: 18705328]
- 125 **Szuster-Ciesielska A**, Plewka K, Kandefer-Szerszeń M. Betulin, betulinic acid and butein are inhibitors of acetaldehyde-induced activation of liver stellate cells. *Pharmacol Rep* 2011; **63**: 1109-1123 [PMID: 22180353]
- 126 **Quiroz SC**, Bucio L, Souza V, Hernández E, González E, Gómez-Quiroz L, Kershenovich D, Vargas-Vorackova F, Gutiérrez-Ruiz MC. Effect of endotoxin pretreatment on hepatic stellate cell response to ethanol and acetaldehyde. *J Gastroenterol Hepatol* 2001; **16**: 1267-1273 [PMID: 11903746]
- 127 **Guo Y**, Wu XQ, Zhang C, Liao ZX, Wu Y, Xia ZY, Wang H. Effect of indole-3-carbinol on ethanol-induced liver injury and acetaldehyde-stimulated hepatic stellate cells activation using precision-cut rat liver slices. *Clin Exp Pharmacol Physiol* 2010; **37**: 1107-1113 [PMID: 20880187 DOI: 10.1111/j.1440-1681.2010.05450.x]
- 128 **Zhang L**, Wu T, Chen JM, Yang LL, Song HY, Ji G. Danshensu inhibits acetaldehyde-induced proliferation and activation of hepatic stellate cell-T6. *Zhong Xi Yi Jie He Xue Bao* 2012; **10**: 1155-1161 [PMID: 23073200]
- 129 **Liu Y**, Brymora J, Zhang H, Smith B, Ramezani-Moghadam M, George J, Wang J. Leptin and acetaldehyde synergistically promotes α SMA expression in hepatic stellate cells by an interleukin 6-dependent mechanism. *Alcohol Clin Exp Res* 2011; **35**: 921-928 [PMID: 21294755 DOI: 10.1111/j.1530-0277.2010.01422.x]
- 130 **Jiang MD**, Ma HD, Zhong XF, Xie FW, Zeng WZ. [Effects of Erk signal transduction on the cell cycle of rat hepatic stellate cells stimulated by acetaldehyde]. *Zhonghua Gan Zang Bing Za Zhi* 2003; **11**: 650-653 [PMID: 14636436]
- 131 **Svegliati-Baroni G**, Ridolfi F, Di Sario A, Saccomanno S, Bendia E, Benedetti A, Greenwel P. Intracellular signaling pathways involved in acetaldehyde-induced collagen and fibronectin gene expression in human hepatic stellate cells. *Hepatology* 2001; **33**: 1130-1140 [PMID: 11343241 DOI: 10.1053/jhep.2001.23788]
- 132 **Chen A**, Davis BH. The DNA binding protein BTEB mediates acetaldehyde-induced, jun N-terminal kinase-dependent α (I) collagen gene expression in rat hepatic stellate cells. *Mol Cell Biol* 2000; **20**: 2818-2826 [PMID: 10733585]
- 133 **Anania FA**, Potter JJ, Rennie-Tankersley L, Mezey E. Activation by acetaldehyde of the promoter of the mouse α 2(I) collagen gene when transfected into rat activated stellate cells. *Arch Biochem Biophys* 1996; **331**: 187-193 [PMID: 8660697 DOI: 10.1006/abbi.1996.0297]
- 134 **Chen A**. Acetaldehyde stimulates the activation of latent transforming growth factor-beta1 and induces expression of the type II receptor of the cytokine in rat cultured hepatic stellate cells. *Biochem J* 2002; **368**: 683-693 [PMID: 12223100 DOI: 10.1042/bj20020949]
- 135 **Kim Y**, Ratzu V, Choi SG, Lalazar A, Theiss G, Dang Q, Kim SJ, Friedman SL. Transcriptional activation of transforming growth factor beta1 and its receptors by the Kruppel-like factor ZF9/core promoter-binding protein and Sp1. Potential mechanisms for autocrine fibrogenesis in response to injury. *J Biol Chem* 1998;

- 273: 33750-33758 [PMID: 9837963]
- 136 **Friedman SL.** Molecular regulation of hepatic fibrosis, an integrated cellular response to tissue injury. *J Biol Chem* 2000; **275**: 2247-2250 [PMID: 10644669]
- 137 **Siegmund SV,** Brenner DA. Molecular pathogenesis of alcohol-induced hepatic fibrosis. *Alcohol Clin Exp Res* 2005; **29**: 102S-109S [PMID: 16344593]
- 138 **Border WA,** Noble NA. Transforming growth factor beta in tissue fibrosis. *N Engl J Med* 1994; **331**: 1286-1292 [PMID: 7935686 DOI: 10.1056/nejm199411103311907]
- 139 **Grisham JW,** Nopanitaya W, Compagno J, Nägel AE. Scanning electron microscopy of normal rat liver: the surface structure of its cells and tissue components. *Am J Anat* 1975; **144**: 295-321 [PMID: 1211369 DOI: 10.1002/aja.1001440304]
- 140 **Macchiarelli G,** Motta PM. The three-dimensional microstructure of the liver. A review by scanning electron microscopy. *Scan Electron Microsc* 1986; (**Pt 3**): 1019-1038 [PMID: 3541157]
- 141 **Scharf J,** Ramadori G, Bräulke T, Hartmann H. Synthesis of insulinlike growth factor binding proteins and of the acid-labile subunit in primary cultures of rat hepatocytes, of Kupffer cells, and in cocultures: regulation by insulin, insulinlike growth factor, and growth hormone. *Hepatology* 1996; **23**: 818-827 [PMID: 8666337 DOI: 10.1053/jhep.1996.v23.pm0008666337]
- 142 **Shimada Y,** Kato T, Ogami K, Horie K, Kokubo A, Kudo Y, Maeda E, Sohma Y, Akahori H, Kawamura K. Production of thrombopoietin (TPO) by rat hepatocytes and hepatoma cell lines. *Exp Hematol* 1995; **23**: 1388-1396 [PMID: 7498368]
- 143 **Eckardt KU,** Pugh CW, Ratcliffe PJ, Kurtz A. Oxygen-dependent expression of the erythropoietin gene in rat hepatocytes in vitro. *Pflugers Arch* 1993; **423**: 356-364 [PMID: 8351193]
- 144 **Ramadori G,** Christ B. Cytokines and the hepatic acute-phase response. *Semin Liver Dis* 1999; **19**: 141-155 [PMID: 10422197 DOI: 10.1055/s-2007-1007106]
- 145 **Sheikh N,** Tron K, Dudas J, Ramadori G. Cytokine-induced neutrophil chemoattractant-1 is released by the noninjured liver in a rat acute-phase model. *Lab Invest* 2006; **86**: 800-814 [PMID: 16715102 DOI: 10.1038/labinvest.3700435]
- 146 **Sambasivam H,** Rassouli M, Murray RK, Nagpurkar A, Mookerjee S, Azadi P, Dell A, Morris HR. Studies on the carbohydrate moiety and on the biosynthesis of rat C-reactive protein. *J Biol Chem* 1993; **268**: 10007-10016 [PMID: 8486673]
- 147 **Ramadori G,** Rieder H, Sipe J, Shirahama T, Meyer zum Büschenfelde KH. Murine tissue macrophages synthesize and secrete amyloid proteins different to amyloid A (AA). *Eur J Clin Invest* 1989; **19**: 316-322 [PMID: 2478371]
- 148 **Tron K,** Samoilenko A, Musikowski G, Kobe F, Immenschuh S, Schaper F, Ramadori G, Kietzmann T. Regulation of rat heme oxygenase-1 expression by interleukin-6 via the Jak/STAT pathway in hepatocytes. *J Hepatol* 2006; **45**: 72-80 [PMID: 16510205 DOI: 10.1016/j.jhep.2005.12.019]
- 149 **Ren X,** Kennedy A, Colletti LM. CXC chemokine expression after stimulation with interferon-gamma in primary rat hepatocytes in culture. *Shock* 2002; **17**: 513-520 [PMID: 12069190]
- 150 **Bioulac-Sage P,** Kuiper J, Van Berkel TJ, Balabaud C. Lymphocyte and macrophage populations in the liver. *Hepato gastroenterology* 1996; **43**: 4-14 [PMID: 8682487]
- 151 **Vanderkerken K,** Bouwens L, Van Rooijen N, Van den Berg K, Baekeland M, Wisse E. The role of Kupffer cells in the differentiation process of hepatic natural killer cells. *Hepatology* 1995; **22**: 283-290 [PMID: 7601422]
- 152 **Vermijlen D,** Luo D, Froelich CJ, Medema JP, Kummer JA, Willems E, Braet F, Wisse E. Hepatic natural killer cells exclusively kill splenic/blood natural killer-resistant tumor cells by the perforin/granzyme pathway. *J Leukoc Biol* 2002; **72**: 668-676 [PMID: 12377935]
- 153 **Jeong WI,** Park O, Gao B. Abrogation of the antifibrotic effects of natural killer cells/interferon-gamma contributes to alcohol acceleration of liver fibrosis. *Gastroenterology* 2008; **134**: 248-258 [PMID: 18166357 DOI: 10.1053/j.gastro.2007.09.034]
- 154 **Jeong WI,** Park O, Suh YG, Byun JS, Park SY, Choi E, Kim JK, Ko H, Wang H, Miller AM, Gao B. Suppression of innate immunity (natural killer cell/interferon- γ) in the advanced stages of liver fibrosis in mice. *Hepatology* 2011; **53**: 1342-1351 [PMID: 21480338 DOI: 10.1002/hep.24190]
- 155 **Radaeva S,** Sun R, Jaruga B, Nguyen VT, Tian Z, Gao B. Natural killer cells ameliorate liver fibrosis by killing activated stellate cells in NKG2D-dependent and tumor necrosis factor-related apoptosis-inducing ligand-dependent manners. *Gastroenterology* 2006; **130**: 435-452 [PMID: 16472598 DOI: 10.1053/j.gastro.2005.10.055]
- 156 **Derynck R,** Akhurst RJ, Balmain A. TGF-beta signaling in tumor suppression and cancer progression. *Nat Genet* 2001; **29**: 117-129 [PMID: 11586292]
- 157 **Heldin CH,** Landström M, Moustakas A. Mechanism of TGF-beta signaling to growth arrest, apoptosis, and epithelial-mesenchymal transition. *Curr Opin Cell Biol* 2009; **21**: 166-176 [PMID: 19237272]
- 158 **Massagué J.** TGF β signalling in context. *Nat Rev Mol Cell Biol* 2012; **13**: 616-630 [PMID: 22992590 DOI: 10.1038/nrm3434]
- 159 **Casini A,** Cunningham M, Rojkind M, Lieber CS. Acetaldehyde increases procollagen type I and fibronectin gene transcription in cultured rat fat-storing cells through a protein synthesis-dependent mechanism. *Hepatology* 1991; **13**: 758-765 [PMID: 2010171]
- 160 **Greenwel P.** Acetaldehyde-mediated collagen regulation in hepatic stellate cells. *Alcohol Clin Exp Res* 1999; **23**: 930-933 [PMID: 10371417]
- 161 **Behari J.** The Wnt/ β -catenin signaling pathway in liver biology and disease. *Expert Rev Gastroenterol Hepatol* 2010; **4**: 745-756 [PMID: 21108594 DOI: 10.1586/egh.10.74]
- 162 **Giles RH,** van Es JH, Clevers H. Caught up in a Wnt storm: Wnt signaling in cancer. *Biochim Biophys Acta* 2003; **1653**: 1-24 [PMID: 12781368 DOI: 10.1016/S0304-419X(03)00005-2]
- 163 **Shen YC,** Hsu C, Cheng AL. Molecular targeted therapy for advanced hepatocellular carcinoma: current status and future perspectives. *J Gastroenterol* 2010; **45**: 794-807 [PMID: 20567987 DOI: 10.1007/s00535-010-0270-0]
- 164 **Whittaker S,** Marais R, Zhu AX. The role of signaling pathways in the development and treatment of hepatocellular carcinoma. *Oncogene* 2010; **29**: 4989-5005 [PMID: 20639898 DOI: 10.1038/onc.2010.236]
- 165 **Turner N,** Grose R. Fibroblast growth factor signalling: from development to cancer. *Nature Reviews Cancer* 2010; **10**: 116-129 [DOI: 10.1038/nrc2780]
- 166 **Knights V,** Cook SJ. De-regulated FGF receptors as therapeutic targets in cancer. *Pharmacol Ther* 2010; **125**: 105-117 [PMID: 19874848 DOI: 10.1016/j.pharmthera.2009.10.001]
- 167 **Schmitz KJ,** Wohlschlaeger J, Lang H, Sotiropoulos GC, Malago M, Steveling K, Reis H, Cicinnati VR, Schmid KW, Baba HA. Activation of the ERK and AKT signalling pathway predicts poor prognosis in hepatocellular carcinoma and ERK activation in cancer tissue is associated with hepatitis C virus infection. *J Hepatol* 2008; **48**: 83-90 [PMID: 17998146 DOI: 10.1016/j.jhep.2007.08.018]
- 168 **Chang Q,** Zhang Y, Beezhold KJ, Bhatia D, Zhao H, Chen J, Castranova V, Shi X, Chen F. Sustained JNK1 activation is associated with altered histone H3 methylations in human liver cancer. *J Hepatol* 2009; **50**: 323-333 [PMID: 19041150 DOI: 10.1016/j.jhep.2008.07.037]
- 169 **Villanueva A,** Chiang DY, Newell P, Peix J, Thung S, Alsinet C, Tovar V, Roayaie S, Minguez B, Sole M, Battiston C, Van Laarhoven S, Fiel MI, Di Feo A, Hoshida Y, Yea S, Toffanin S, Ramos A, Martignetti JA, Mazzaferro V, Bruix J, Waxman S, Schwartz M, Meyerson M, Friedman SL, Llovet JM. Pivotal role of mTOR signaling in hepatocellular carcinoma. *Gastroenterology* 2008; **135**: 1972-1983.e1911 [PMID: 18929564 DOI: 10.1053/j.gastro.2008.08.008]
- 170 **Zhang H,** Berezov A, Wang Q, Zhang G, Drebin J, Murali R, Greene MI. ErbB receptors: from oncogenes to targeted cancer therapies. *J Clin Invest* 2007; **117**: 2051-2058 [PMID: 17671639]

- DOI: 10.1172/JCI32278]
- 171 **Lund P**, Schubert D, Niketeghad F, Schirmacher P. Autocrine inhibition of chemotherapy response in human liver tumor cells by insulin-like growth factor-II. *Cancer Lett* 2004; **206**: 85-96 [PMID: 15019164 DOI: 10.1016/j.canlet.2003.10.018]
 - 172 **Matsuzaki K**. Smad phosphoisoform signals in acute and chronic liver injury: similarities and differences between epithelial and mesenchymal cells. *Cell Tissue Res* 2012; **347**: 225-243 [PMID: 21626291 DOI: 10.1007/s00441-011-1178-6]
 - 173 **Derynck R**, Miyazono K. The TGF- family: Cold Spring Harbor Laboratory. NY: Press Cold Spring Harbor, 2008
 - 174 **Hayashi Y**, Hirose F, Nishimoto Y, Shiraki M, Yamagishi M, Matsukage A, Yamaguchi M. Identification of CFDD (common regulatory factor for DNA replication and DREF genes) and role of its binding site in regulation of the proliferating cell nuclear antigen gene promoter. *J Biol Chem* 1997; **272**: 22848-22858 [PMID: 9278447]
 - 175 **Nakao A**, Afrakhte M, Morén A, Nakayama T, Christian JL, Heuchel R, Itoh S, Kawabata M, Heldin NE, Heldin CH, ten Dijke P. Identification of Smad7, a TGFbeta-inducible antagonist of TGF-beta signalling. *Nature* 1997; **389**: 631-635 [PMID: 9335507 DOI: 10.1038/39369]
 - 176 **Souchelnytskyi S**, Nakayama T, Nakao A, Morén A, Heldin CH, Christian JL, ten Dijke P. Physical and functional interaction of murine and Xenopus Smad7 with bone morphogenetic protein receptors and transforming growth factor-beta receptors. *J Biol Chem* 1998; **273**: 25364-25370 [PMID: 9738003]
 - 177 **Friedl W**, Uhlhaas S, Schulmann K, Stolte M, Loff S, Back W, Mangold E, Stern M, Knaebel HP, Sutter C, Weber RG, Pistorius S, Burger B, Propping P. Juvenile polyposis: massive gastric polyposis is more common in MADH4 mutation carriers than in BMPR1A mutation carriers. *Hum Genet* 2002; **111**: 108-111 [PMID: 12136244 DOI: 10.1007/s00439-002-0748-9]
 - 178 **Reichl P**, Dengler M, van Zijl F, Huber H, Führlinger G, Reichel C, Sieghart W, Peck-Radosavljevic M, Grubinger M, Mikulits W. Axl activates autocrine transforming growth factor- β signaling in hepatocellular carcinoma. *Hepatology* 2015; **61**: 930-941 [PMID: 25251599 DOI: 10.1002/hep.27492]
 - 179 **Wu X**, Liu X, Koul S, Lee CY, Zhang Z, Halmos B. AXL kinase as a novel target for cancer therapy. *Oncotarget* 2014; **5**: 9546-9563 [PMID: 25337673]
 - 180 **López-Casillas F**, Wrana JL, Massagué J. Betaglycan presents ligand to the TGF beta signaling receptor. *Cell* 1993; **73**: 1435-1444 [PMID: 8391934]
 - 181 **Massagué J**, Gomis RR. The logic of TGFbeta signaling. *FEBS Lett* 2006; **580**: 2811-2820 [PMID: 16678165]
 - 182 **Moustakas A**, Heldin CH. Non-Smad TGF-beta signals. *J Cell Sci* 2005; **118**: 3573-3584 [PMID: 16105881 DOI: 10.1242/jcs.02554]
 - 183 **Gallihier AJ**, Neil JR, Schieman WP. Role of transforming growth factor-beta in cancer progression. *Future Oncol* 2006; **2**: 743-763 [PMID: 17155901 DOI: 10.2217/14796694.2.6.743]
 - 184 **Xu XB**, He ZP, Liang ZQ, Leng XS. [Obstruction of TGF-beta1 signal transduction by anti-Smad4 gene can therapy experimental liver fibrosis in the rat]. *Zhonghua Gan Zang Bing Za Zhi* 2004; **12**: 263-266 [PMID: 15161498]
 - 185 **Inman GJ**, Nicolás FJ, Hill CS. Nucleocytoplasmic shuttling of Smads 2, 3, and 4 permits sensing of TGF-beta receptor activity. *Mol Cell* 2002; **10**: 283-294 [PMID: 12191474]
 - 186 **Feng XH**, Derynck R. Specificity and versatility in tgf-beta signaling through Smads. *Annu Rev Cell Dev Biol* 2005; **21**: 659-693 [PMID: 16212511 DOI: 10.1146/annurev.cellbio.21.022404.142018]
 - 187 **Chen P**, Schnabl B. Host-microbiome interactions in alcoholic liver disease. *Gut Liver* 2014; **8**: 237-241 [PMID: 24827618 DOI: 10.5009/gnl.2014.8.3.237]
 - 188 **Szabo G**. Gut-liver axis in alcoholic liver disease. *Gastroenterology* 2015; **148**: 30-36 [PMID: 25447847 DOI: 10.1053/j.gastro.2014.10.042]
 - 189 **Hartmann P**, Seebauer CT, Schnabl B. Alcoholic liver disease: the gut microbiome and liver cross talk. *Alcohol Clin Exp Res* 2015; **39**: 763-775 [PMID: 25872593 DOI: 10.1111/acer.12704]
 - 190 **Chen P**, Stärkel P, Turner JR, Ho SB, Schnabl B. Dysbiosis-induced intestinal inflammation activates tumor necrosis factor receptor I and mediates alcoholic liver disease in mice. *Hepatology* 2015; **61**: 883-894 [PMID: 25251280 DOI: 10.1002/hep.27489]
 - 191 **Hartmann P**, Chen WC, Schnabl B. The intestinal microbiome and the leaky gut as therapeutic targets in alcoholic liver disease. *Front Physiol* 2012; **3**: 402 [PMID: 23087650 DOI: 10.3389/fphys.2012.00402]
 - 192 **Bakin AV**, Rinehart C, Tomlinson AK, Arteaga CL. p38 mitogen-activated protein kinase is required for TGFbeta-mediated fibroblastic transdifferentiation and cell migration. *J Cell Sci* 2002; **115**: 3193-3206 [PMID: 12118074]
 - 193 **Watanabe H**, de Caestecker MP, Yamada Y. Transcriptional cross-talk between Smad, ERK1/2, and p38 mitogen-activated protein kinase pathways regulates transforming growth factor-beta-induced aggrecan gene expression in chondrogenic ATDC5 cells. *J Biol Chem* 2001; **276**: 14466-14473 [PMID: 11278290 DOI: 10.1074/jbc.M005724200]
 - 194 **Zavadil J**, Bitzer M, Liang D, Yang YC, Massimi A, Kneitz S, Piek E, Bottinger EP. Genetic programs of epithelial cell plasticity directed by transforming growth factor-beta. *Proc Natl Acad Sci USA* 2001; **98**: 6686-6691 [PMID: 11390996 DOI: 10.1073/pnas.111614398]
 - 195 **Dumont N**, Bakin AV, Arteaga CL. Autocrine transforming growth factor-beta signaling mediates Smad-independent motility in human cancer cells. *J Biol Chem* 2003; **278**: 3275-3285 [PMID: 12421823 DOI: 10.1074/jbc.M204623200]
 - 196 **Bhowmick NA**, Ghiassi M, Bakin A, Aakre M, Lundquist CA, Engel ME, Arteaga CL, Moses HL. Transforming growth factor-beta1 mediates epithelial to mesenchymal transdifferentiation through a RhoA-dependent mechanism. *Mol Biol Cell* 2001; **12**: 27-36 [PMID: 11160820]
 - 197 **Perلمان R**, Schieman WP, Brooks MW, Lodish HF, Weinberg RA. TGF-beta-induced apoptosis is mediated by the adapter protein Daxx that facilitates JNK activation. *Nat Cell Biol* 2001; **3**: 708-714 [PMID: 11483955 DOI: 10.1038/35087019]
 - 198 **Haas SL**, Fitzner B, Jaster R, Wiercinska E, Gaitantzi H, Jesnowski R, Löhr JM, Singer MV, Dooley S, Breitkopf K. Transforming growth factor-beta induces nerve growth factor expression in pancreatic stellate cells by activation of the ALK-5 pathway. *Growth Factors* 2009; **27**: 289-299 [PMID: 19639490 DOI: 10.1080/08977190903132273]
 - 199 **Arteel GE**. New role of plasminogen activator inhibitor-1 in alcohol-induced liver injury. *J Gastroenterol Hepatol* 2008; **23** Suppl 1: S54-S59 [PMID: 18336665 DOI: 10.1111/j.1440-1746.2007.05285.x]
 - 200 **Berghheim I**, Guo L, Davis MA, Lambert JC, Beier JI, Duveau I, Luyendyk JP, Roth RA, Arteel GE. Metformin prevents alcohol-induced liver injury in the mouse: Critical role of plasminogen activator inhibitor-1. *Gastroenterology* 2006; **130**: 2099-2112 [PMID: 16762632 DOI: 10.1053/j.gastro.2006.03.020]
 - 201 **Beier JI**, Arteel GE. Alcoholic liver disease and the potential role of plasminogen activator inhibitor-1 and fibrin metabolism. *Exp Biol Med* (Maywood) 2012; **237**: 1-9 [PMID: 22238286 DOI: 10.1258/ebm.2011.011255]
 - 202 **Yang Y**, Yang S, Chen M, Zhang X, Zou Y, Zhang X. Compound Astragalus and Salvia miltiorrhiza Extract exerts anti-fibrosis by mediating TGF-beta/Smad signaling in myofibroblasts. *J Ethnopharmacol* 2008; **118**: 264-270 [PMID: 18502066 DOI: 10.1016/j.jep.2008.04.012]
 - 203 **Fearns C**, Loskutoff DJ. Induction of plasminogen activator inhibitor 1 gene expression in murine liver by lipopolysaccharide. Cellular localization and role of endogenous tumor necrosis factor-alpha. *Am J Pathol* 1997; **150**: 579-590 [PMID: 9033272]
 - 204 **Zhang YE**. Non-Smad pathways in TGF-beta signaling. *Cell Res* 2009; **19**: 128-139 [PMID: 19114990 DOI: 10.1038/cr.2008.328]
 - 205 **Javelaud D**, Mauviel A. Crosstalk mechanisms between the mitogen-activated protein kinase pathways and Smad signaling downstream of TGF-beta: implications for carcinogenesis.

- Oncogene* 2005; **24**: 5742-5750 [PMID: 16123807]
- 206 **Anderson P.** Kinase cascades regulating entry into apoptosis. *Microbiol Mol Biol Rev* 1997; **61**: 33-46 [PMID: 9106363]
- 207 **Whitmarsh AJ, Davis RJ.** Signal transduction by MAP kinases: regulation by phosphorylation-dependent switches. *Sci STKE* 1999; **1999**: PE1 [PMID: 11865181 DOI: 10.1126/stke.1999.1.pe1]
- 208 **Derynck R, Zhang YE.** Smad-dependent and Smad-independent pathways in TGF-beta family signalling. *Nature* 2003; **425**: 577-584 [PMID: 14534577]
- 209 **Wakefield LM, Roberts AB.** TGF-beta signaling: positive and negative effects on tumorigenesis. *Curr Opin Genet Dev* 2002; **12**: 22-29 [PMID: 11790550 DOI: 10.1016/S0959-437X(01)00259-3]
- 210 **Engel ME, McDonnell MA, Law BK, Moses HL.** Interdependent SMAD and JNK signaling in transforming growth factor-beta-mediated transcription. *J Biol Chem* 1999; **274**: 37413-37420 [PMID: 10601313 DOI: 10.1074/jbc.274.52.37413]
- 211 **Furukawa F, Matsuzaki K, Mori S, Tahashi Y, Yoshida K, Sugano Y, Yamagata H, Matsushita M, Seki T, Inagaki Y, Nishizawa M, Fujisawa J, Inoue K.** p38 MAPK mediates fibrogenic signal through Smad3 phosphorylation in rat myofibroblasts. *Hepatology* 2003; **38**: 879-889 [PMID: 14512875 DOI: 10.1002/hep.1840380414]
- 212 **Aroor AR, Shukla SD.** MAP kinase signaling in diverse effects of ethanol. *Life Sci* 2004; **74**: 2339-2364 [PMID: 15027449]
- 213 **Mandrekar P, Szabo G.** Signaling pathways in alcohol-induced liver inflammation. *J Hepatol* 2009; **50**: 1258-1266 [PMID: 19398236 DOI: 10.1016/j.jhep.2009.03.007]
- 214 **Yao J, Mackman N, Edgington TS, Fan ST.** Lipopolysaccharide induction of the tumor necrosis factor-alpha promoter in human monocytic cells. Regulation by Egr-1, c-Jun, and NF-kappaB transcription factors. *J Biol Chem* 1997; **272**: 17795-17801 [PMID: 9211933]
- 215 **Chen J, Ishac EJ, Dent P, Kunos G, Gao B.** Effects of ethanol on mitogen-activated protein kinase and stress-activated protein kinase cascades in normal and regenerating liver. *Biochem J* 1998; **334** (Pt 3): 669-676 [PMID: 9729476]
- 216 **Spector MS, Auer KL, Jarvis WD, Ishac EJ, Gao B, Kunos G, Dent P.** Differential regulation of the mitogen-activated protein and stress-activated protein kinase cascades by adrenergic agonists in quiescent and regenerating adult rat hepatocytes. *Mol Cell Biol* 1997; **17**: 3556-3565 [PMID: 9199291]
- 217 **Ding W, Shi W, Bellusci S, Groffen J, Heisterkamp N, Minoo P, Warburton D.** Sprout2 downregulation plays a pivotal role in mediating crosstalk between TGF-beta1 signaling and EGF as well as FGF receptor tyrosine kinase-ERK pathways in mesenchymal cells. *J Cell Physiol* 2007; **112**: 796-806 [PMID: 17516543 DOI: 10.1002/jcp.21078]
- 218 **Masoumi-Moghaddam S, Amini A, Morris DL.** The developing story of Sprouty and cancer. *Cancer Metastasis Rev* 2014; **33**: 695-720 [PMID: 24744103 DOI: 10.1007/s10555-014-9497-1]
- 219 **Arellanes-Robledo J, Reyes-Gordillo K, Shah R, Domínguez-Rosales JA, Hernández-Nazara ZH, Ramirez F, Rojkind M, Lakshman MR.** Fibrogenic actions of acetaldehyde are beta-catenin dependent but Wingless independent: a critical role of nucleoredoxin and reactive oxygen species in human hepatic stellate cells. *Free Radic Biol Med* 2013; **65**: 1487-1496 [PMID: 23880292 DOI: 10.1016/j.freeradbiomed.2013.07.017]
- 220 **Patel R, Gao M, Ahmad I, Fleming J, Singh LB, Rai TS, McKie AB, Seywright M, Barnetson RJ, Edwards J, Sansom OJ, Leung HY.** Sprout2, PTEN, and PP2A interact to regulate prostate cancer progression. *J Clin Invest* 2013; **123**: 1157-1175 [PMID: 23434594 DOI: 10.1172/jci63672]
- 221 **Chen N, Geng Q, Zheng J, He S, Huo X, Sun X.** Suppression of the TGF-beta/Smad signaling pathway and inhibition of hepatic stellate cell proliferation play a role in the hepatoprotective effects of curcumin against alcohol-induced hepatic fibrosis. *Int J Mol Med* 2014; **34**: 1110-1116 [PMID: 25069637 DOI: 10.3892/ijmm.2014.1867]
- 222 **Reyes-Gordillo K, Shah R, Arellanes-Robledo J, Hernández-Nazara Z, Rincón-Sánchez AR, Inagaki Y, Rojkind M, Lakshman MR.** Mechanisms of action of acetaldehyde in the up-regulation of the human alpha2(I) collagen gene in hepatic stellate cells: key roles of Ski, SMAD3, SMAD4, and SMAD7. *Am J Pathol* 2014; **184**: 1458-1467 [PMID: 24641900 DOI: 10.1016/j.ajpath.2014.01.020]
- 223 **Svegliati-Baroni G, Inagaki Y, Rincon-Sanchez AR, Else C, Saccomanno S, Benedetti A, Ramirez F, Rojkind M.** Early response of alpha2(I) collagen to acetaldehyde in human hepatic stellate cells is TGF-beta independent. *Hepatology* 2005; **42**: 343-352 [PMID: 16025520 DOI: 10.1002/hep.20798]
- 224 **Abhilash PA, Harikrishnan R, Indira M.** Ascorbic acid suppresses endotoxemia and NF-kB signaling cascade in alcoholic liver fibrosis in guinea pigs: a mechanistic approach. *Toxicol Appl Pharmacol* 2014; **274**: 215-224 [PMID: 24239723 DOI: 10.1016/j.taap.2013.11.005]
- 225 **Gu H, Fortunato F, Bergmann F, Büchler MW, Whitcomb DC, Werner J.** Alcohol exacerbates LPS-induced fibrosis in subclinical acute pancreatitis. *Am J Pathol* 2013; **183**: 1508-1517 [PMID: 24091223 DOI: 10.1016/j.ajpath.2013.07.023]
- 226 **Xu XB, He ZP, Leng XS, Liang ZQ, Peng JR, Zhang HY, Zhang HY, Xiao M, Zhang H, Liu CL, Zhang XD.** Effects of Smad4 on liver fibrosis and hepatocarcinogenesis in mice treated with CCl4/ethanol. *Zhonghua Gan Zang Bing Za Zhi* 2010; **18**: 119-123 [PMID: 20196951 DOI: 10.3760/cma.j.issn.1007-3418.2010.02.010]
- 227 **Tang Y, Li Y, Yu H, Gao C, Liu L, Chen S, Xing M, Liu L, Yao P.** Quercetin prevents ethanol-induced iron overload by regulating hepcidin through the BMP6/SMAD4 signaling pathway. *J Nutr Biochem* 2014; **25**: 675-682 [PMID: 24746831 DOI: 10.1016/j.jnutbio.2014.02.009]
- 228 **Gerjevic LN, Liu N, Lu S, Harrison-Findik DD.** Alcohol Activates TGF-Beta but Inhibits BMP Receptor-Mediated Smad Signaling and Smad4 Binding to Hepcidin Promoter in the Liver. *Int J Hepatol* 2012; **2012**: 459278 [PMID: 22121494 DOI: 10.1155/2012/459278]
- 229 **Zhu L, Wang L, Wang X, Luo X, Yang L, Zhang R, Yin H, Xie D, Pan Y, Chen Y.** Hepatic deletion of Smad7 in mouse leads to spontaneous liver dysfunction and aggravates alcoholic liver injury. *PLoS One* 2011; **6**: e17415 [PMID: 21386907 DOI: 10.1371/journal.pone.0017415]
- 230 **Ciuclan L, Ehnert S, Ilkavets I, Weng HL, Gaitantzi H, Tsukamoto H, Ueberham E, Meindl-Beinker NM, Singer MV, Breitkopf K, Dooley S.** TGF-beta enhances alcohol dependent hepatocyte damage via down-regulation of alcohol dehydrogenase I. *J Hepatol* 2010; **52**: 407-416 [PMID: 20129692 DOI: 10.1016/j.jhep.2009.12.003]
- 231 **Szuster-Ciesielska A, Mizerska-Dudka M, Daniluk J, Kandefers-Szerszeń M.** Butein inhibits ethanol-induced activation of liver stellate cells through TGF-beta, NFkB, p38, and JNK signaling pathways and inhibition of oxidative stress. *J Gastroenterol* 2013; **48**: 222-237 [PMID: 22722906 DOI: 10.1007/s00535-012-0619-7]
- 232 **Szuster-Ciesielska A, Plewka K, Daniluk J, Kandefers-Szerszeń M.** Betulin and betulinic acid attenuate ethanol-induced liver stellate cell activation by inhibiting reactive oxygen species (ROS), cytokine (TNF-alpha, TGF-beta) production and by influencing intracellular signaling. *Toxicology* 2011; **280**: 152-163 [PMID: 21172400 DOI: 10.1016/j.tox.2010.12.006]
- 233 **Cederbaum AI, Lu Y, Wang X, Wu D.** Synergistic toxic interactions between CYP2E1, LPS/TNFalpha, and JNK/p38 MAP kinase and their implications in alcohol-induced liver injury. *Adv Exp Med Biol* 2015; **815**: 145-172 [PMID: 25427906 DOI: 10.1007/978-3-319-09614-8_9]
- 234 **Luo Z, Dong X, Ke Q, Duan Q, Shen L.** Chitoooligosaccharides inhibit ethanol-induced oxidative stress via activation of Nrf2 and reduction of MAPK phosphorylation. *Oncol Rep* 2014; **32**: 2215-2222 [PMID: 25189124 DOI: 10.3892/or.2014.3463]
- 235 **Yan SL, Yang HT, Lee HL, Yin MC.** Protective effects of maslinic acid against alcohol-induced acute liver injury in mice. *Food Chem Toxicol* 2014; **74**: 149-155 [PMID: 25301236 DOI: 10.1016/

- j.fct.2014.09.018]
- 236 **Li Y**, Gao C, Shi Y, Tang Y, Liu L, Xiong T, Du M, Xing M, Liu L, Yao P. Carbon monoxide alleviates ethanol-induced oxidative damage and inflammatory stress through activating p38 MAPK pathway. *Toxicol Appl Pharmacol* 2013; **273**: 53-58 [PMID: 23994557 DOI: 10.1016/j.taap.2013.08.019]
- 237 **Chang YY**, Lin YL, Yang DJ, Liu CW, Hsu CL, Tzang BS, Chen YC. Hepatoprotection of noni juice against chronic alcohol consumption: lipid homeostasis, antioxidation, alcohol clearance, and anti-inflammation. *J Agric Food Chem* 2013; **61**: 11016-11024 [PMID: 24152092 DOI: 10.1021/jf4038419]
- 238 **Morio Y**, Tsuji M, Inagaki M, Nakagawa M, Asaka Y, Oyamada H, Furuya K, Oguchi K. Ethanol-induced apoptosis in human liver adenocarcinoma cells (SK-Hep1): Fas- and mitochondria-mediated pathways and interaction with MAPK signaling system. *Toxicol In Vitro* 2013; **27**: 1820-1829 [PMID: 23726865 DOI: 10.1016/j.tiv.2013.05.009]
- 239 **Park EJ**, Kim YM, Park SW, Kim HJ, Lee JH, Lee DU, Chang KC. Induction of HO-1 through p38 MAPK/Nrf2 signaling pathway by ethanol extract of *Inula helenium* L. reduces inflammation in LPS-activated RAW 264.7 cells and CLP-induced septic mice. *Food Chem Toxicol* 2013; **55**: 386-395 [PMID: 23298677 DOI: 10.1016/j.fct.2012.12.027]
- 240 **Aroor AR**, Jackson DE, Shukla SD. Elevated activation of ERK1 and ERK2 accompany enhanced liver injury following alcohol binge in chronically ethanol-fed rats. *Alcohol Clin Exp Res* 2011; **35**: 2128-2138 [PMID: 21790671 DOI: 10.1111/j.1530-0277.2011.01577.x]
- 241 **Hung HF**, Hou CW, Chen YL, Lin CC, Fu HW, Wang JS, Jeng KC. Elephantopus scaber inhibits lipopolysaccharide-induced liver injury by suppression of signaling pathways in rats. *Am J Chin Med* 2011; **39**: 705-717 [PMID: 21721151 DOI: 10.1142/s0192415x11009147]
- 242 **Aroor AR**, James TT, Jackson DE, Shukla SD. Differential changes in MAP kinases, histone modifications, and liver injury in rats acutely treated with ethanol. *Alcohol Clin Exp Res* 2010; **34**: 1543-1551 [PMID: 20586759 DOI: 10.1111/j.1530-0277.2010.01239.x]
- 243 **Wang X**, Lu Y, Xie B, Cederbaum AI. Chronic ethanol feeding potentiates Fas Jo2-induced hepatotoxicity: role of CYP2E1 and TNF-alpha and activation of JNK and P38 MAP kinase. *Free Radic Biol Med* 2009; **47**: 518-528 [PMID: 19477265 DOI: 10.1016/j.freeradbiomed.2009.05.021]
- 244 **Lu Y**, Cederbaum AI. CYP2E1 potentiation of LPS and TNF α -induced hepatotoxicity by mechanisms involving enhanced oxidative and nitrosative stress, activation of MAP kinases, and mitochondrial dysfunction. *Genes Nutr* 2010; **5**: 149-167 [PMID: 19798529 DOI: 10.1007/s12263-009-0150-5]
- 245 **Hsu MK**, Qiao L, Ho V, Zhang BH, Zhang H, Teoh N, Dent P, Farrell GC. Ethanol reduces p38 kinase activation and cyclin D1 protein expression after partial hepatectomy in rats. *J Hepatol* 2006; **44**: 375-382 [PMID: 16226824 DOI: 10.1016/j.jhep.2005.07.031]
- 246 **Yao P**, Nussler A, Liu L, Hao L, Song F, Schirmeier A, Nussler N. Quercetin protects human hepatocytes from ethanol-derived oxidative stress by inducing heme oxygenase-1 via the MAPK/Nrf2 pathways. *J Hepatol* 2007; **47**: 253-261 [PMID: 17433488 DOI: 10.1016/j.jhep.2007.02.008]
- 247 **Kenny S**, Steele I, Lyons S, Moore AR, Murugesan SV, Tiszlavicz L, Dimaline R, Pritchard DM, Varro A, Dockray GJ. The role of plasminogen activator inhibitor-1 in gastric mucosal protection. *Am J Physiol Gastrointest Liver Physiol* 2013; **304**: G814-G822 [PMID: 23494120 DOI: 10.1152/ajpgi.00017.2013]

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