

Emergence of Mutations in the K13 Propeller Gene of *Plasmodium falciparum* Isolates from Dakar, Senegal, in 2013-2014

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The kelch 13 (K13) propeller gene is associated with artemisinin resistance. In a previous work, there were no mutations found in 138 *Plasmodium falciparum* isolates collected in 2012 and 2013 from patients residing in Dakar, Senegal (M. Torrentino-Madamet et al., *Malar J* 13:472, 2014, <http://dx.doi.org/10.1186/1475-2875-13-472>). However, the N554H, Q613H, and V637I mutations were identified in the propeller region of K13 in 92 (5.5%) isolates in 2013 and 2014. There were five polymorphisms identified in the *Plasmodium/Plasmodium*-specific domain (K123R, N137S, N142NN/NNN, T149S, and K189T/N).

Malaria resistance to most antimalarial drugs has developed in Southeast Asia and has spread to Africa. The World Health Organization (WHO) has recommended artemisinin-based combination therapy (ACT) as the first-line treatment for malaria since 2005. ACT has been recommended by the Senegalese National Malaria Control Program as the first-line treatment for uncomplicated malaria since 2006. The emergence of *Plasmodium falciparum* resistance to artemisinin and its derivatives manifests as delayed parasite clearance following treatment with artesunate monotherapy or ACT and has recently developed in Southeast Asia (1, 2). This clinical resistance was correlated with *in vitro* resistance, which manifests as an increase in the ring-stage survival rate after contact with artemisinin (3). Therefore, the spread of artemisinin resistance from Asia to Africa may be a serious threat for malaria control and elimination.

Mutations in the propeller domain of the kelch 13 (K13) gene (PF3D71343700) were recently associated with *in vivo* and *in vitro* resistance to artemisinin in Southeast Asia (4). The K13 propeller gene is located on chromosome 13 and encodes a kelch protein. There are 6 kelch motifs called blades at the C terminus (Fig. 1). Each propeller blade is composed of approximately 50 amino acids that form four antiparallel beta-sheet secondary structures arranged around a canal. The propeller domain harbors multiple protein-protein interaction sites (5). The Y493H, R539T, I543T, and C580Y mutations were also correlated with *in vivo* and *in vitro* artemisinin resistance in Southeast Asia (2, 4, 6–8). The C580Y mutation is located in the first beta-sheet of the fourth blade and is associated with delayed parasite clearance leading to *in vivo* resistance (4). A previous study used site-directed mutagenesis with zinc finger nucleases to modify the K13 propeller gene locus. The results confirmed the importance of the Y493H, R539T, I543T, and C580Y mutations in mediating *in vitro* artemisinin resistance (9). Although the four mutations are widely present in Southeast Asia, they are not observed in Africa (10–14). However, the P553L mutation, which is associated with delayed parasite clearance in Southeast Asia, was detected in Kenya and Malawi (12).

We collected 103 samples from *falciparum* malaria patients

attending the Hôpital Principal de Dakar, Senegal, from November 2013 to January 2014 and August 2014 to December 2014 (59 from 2013 and 44 from 2014). Sixty-four percent of the patients were recruited from the emergency department. The other patients were recruited from the intensive care unit (12%), pediatric department (7%), infectious diseases department (5%), maternity department (3%), and other units (9%). There was no information available on antimalarial treatment prior to admission. Despite the recommendations of the WHO, the patients were treated with quinine until November 2014 and with artesunate or artemether-lumefantrine at the Hôpital Principal de Dakar. Informed verbal consent was obtained from the patients or their parents/guardians before blood collection. The study was approved by the ethics committee of the Hôpital Principal de Dakar.

Venous blood samples were collected in Vacutainer acid citrate dextrose (ACD) tubes prior to patient treatment. The total genomic DNA of each isolate was extracted using the QIAamp DNA minikit, according to the manufacturer's recommendations (Qiagen, Germany). A malaria diagnosis was confirmed using a thin blood smear, rapid diagnosis test, and real-time quantitative PCR.

The K13 propeller gene was amplified using a PCR and nested-PCR method described previously (7). The following primers were used for PCR: 3'-GGG AAT CTG GTG GTA ACA GC-5' and 3'-CGG AGT GAC CAA ATC TGG GA-5', and 3'-GCC TTG TTG

Received 9 June 2015 Returned for modification 14 September 2015

Accepted 16 October 2015

Accepted manuscript posted online 26 October 2015

Citation Boussaroque A, Fall B, Madamet M, Camara C, Benoit N, Fall M, Nakoulima A, Dionne P, Fall KB, Diatta B, Diémé Y, Wade B, Pradines B. 2016. Emergence of mutations in the K13 propeller gene of *Plasmodium falciparum* isolates from Dakar, Senegal, in 2013-2014. *Antimicrob Agents Chemother* 60:624–627. doi:10.1128/AAC.01346-15.

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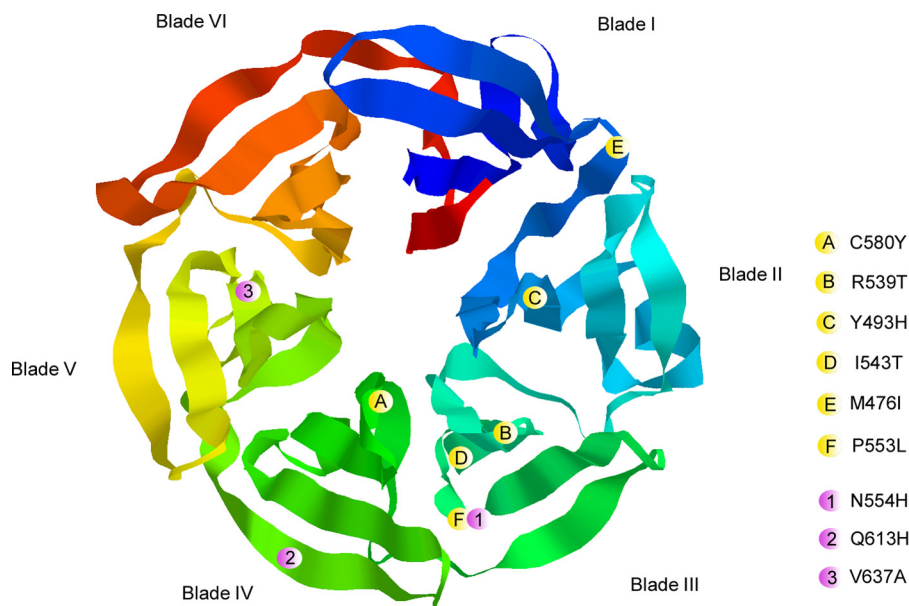


FIG 1 Locations of the mutations in the predicted three-dimensional (3D) model of the K13 propeller domain (RasTop software, Phyre 2 server). The locations of the various mutations are indicated by spheres, in which yellow represents mutations correlated with artemisinin resistance reported by Arley et al. (4) and pink represents mutations observed in Dakar, Senegal.

AAA GAA GCA GA-5' and 3'-GCC AAG CTG CCA TTC ATT TG-5'. The K13 propeller gene was successfully sequenced in 58 PCR and 92 nested-PCR amplicons, and the results were compared to the reference *P. falciparum* 3D7 strain. The K13 mutations were confirmed three times by sequencing the products of three different PCRs.

The polymorphisms identified in this study are reported in Table 1. The mutations associated with *in vitro* resistance in Southeast Asia, such as Y493H, R539T, I543T, and C580Y (2, 4–6), were not observed. The M476I mutation obtained *in vitro* on the F32 Tanzanian strain after artemisinin pressure was not identified (4).

The two novel mutations K123R and N137S in the N-terminal domain were each reported in 1.7% (1/58) of the isolates. One or

two asparagine (N or NN) insertions at codon 142 were found in 13.8% (8/58) of the isolates. These insertions at codon 142 were observed in 10.9% of the samples collected in the Hôpital Principal de Dakar in 2012 and 2013 (10). The T149S mutation was reported in 4.7% (4/64) of cases in 2012 and 2013 in Dakar (10); in this study, the mutation was found in 1.7% (1/58) of the isolates. The isolates with the T149S mutation (unknown location of collection) previously found were associated with a parasite clearance half-life of <5 h (2). There were two mutations at codon 189. We found that 31.0% (18/58) of isolates presented the K189T mutation, and 1.7% (1/58) presented the K189N mutation. In recent studies, the K189T mutation was found in 42.2% of the samples in Dakar in 2012 and 2013 (10) and in 34.5% (10/29) of the samples in Uganda (15). The mutation was also identified in one isolate in Bangladesh (8). Furthermore, the K189N mutation was reported in 3.4% (1/29) of the cases in Uganda (15). Among the 20 isolates previously found (unknown collection location), only one (in Bangladesh) was associated with a parasite clearance half-life of <5 h (2).

There were no previously reported mutations in the six K13 propeller blades in Dakar in 2012 and 2013 (10). In this study, we identified 5 isolates with mutations in blade 3, 4, or 5 (Fig. 1). The N554H mutation was found in 1.1% (1/92) of the isolates. The corresponding residue is located near the third beta-sheet structure of the 3rd blade and was found to already be mutated in African isolates. In Comoros (Grande Comore and Anjouan) in 2013, two isolates (6.9%) were found to carry the N554H and N554K mutations (16). A mutation in the corresponding codon (resulting in N554S) was found in one isolate from Ungoye, Kenya, in 2012 (11) and in one isolate from Mali in 2011 (17). A mutation in this codon has never been found in an isolate from Southeast Asia (18). This mutation is adjacent to the P553L mutation described in southern Asia and is associated with delayed parasite clearance following treatment with artesunate mono-

TABLE 1 Nonsynonymous mutations observed in the K13 gene in *P. falciparum* isolates from Dakar, Senegal, collected from November 2013 to January 2014 and August 2014 to December 2014^a

Amino acid change or insertion and location	Blade ^d	Referent sequence	Mutant sequence ^b	No. of isolates/total isolates (%)
K123R ^c	—	AAA	AGA	1/58 (1.7)
N137S ^c	—	AAT	AGT	1/58 (1.7)
N142NN	—	AAT	AAT	7/58 (12.1)
N142NNN	—	AAT	AAT AAT	1/58 (1.7)
T149S	—	ACT	TCT	1/58 (1.7)
K189T	—	AAA	ACA	18/58 (31.0)
K189N ^c	—	AAA	AAT	1/58 (1.7)
N554H	3	AAT	CAT	1/92 (1.1)
Q613H ^c	4	CAA	CAT	1/92 (1.1)
V637I	5	GTT	ATT	3/92 (3.3)

^a Mutations described were not found in the same isolates.

^b Mutated bases are in bold type.

^c Novel mutation.

^d —, absence of blade numbered only in the propeller domain.

therapy in Asia (2). The P553L mutation was found in 0.53% of the isolates in Kisumu, Kenya, and in 0.59% in Machinga, Malawi (8, 12). The Q613H mutation was observed in 1.1% (1/92) of the isolates. It is located in the 4th beta-sheet of the 4th blade. The Q613E and Q613L mutations were already found in Africa (18). The Q613E mutation was previously identified in one isolate with a parasite clearance half-life of <5 h (2). There are previous reports of different single nucleotide polymorphisms (SNP) at codon 637, whose corresponding residue is located in the second beta-sheet of the 5th blade. The SNP was found to be V637A in 3.22% of the isolates from Bas-Congo, Democratic Republic of Congo (12). The V637D SNP was found in 0.8% of the isolates in Uganda (15). In this study, the V637I mutation was present in 3.3% (3/92) of the isolates. The mutation at codon 637 has not been yet observed in Southeast Asia (18).

The propeller is composed of six blades and harbors protein-protein interaction sites. Thus, mutations in the beta-sheet structures may affect protein interactions. The *P. falciparum* phosphatidylinositol-3-kinase (PI3K) is a putative binding partner of K13 and a target for artemisinin derivatives (19). Artemisinin binding leads to decreased PI3K activity, decreased phosphatidylinositol-3-phosphate (PI3P), and inhibition of parasite growth. Wild-type K13 binds PI3K and delivers it to ubiquitin ligase, which polyubiquitinates K13 and marks it for proteasomal degradation. Mutant K13 fails to bind PI3K. Further studies are needed to characterize the roles of these three mutations in artemisinin resistance. While no mutation was found in the propeller region of K13 in parasites from Dakar in 2012 and 2013, three mutations (5.5%) were identified in this domain in 2013 and 2014. As a result, surveillance of K13 polymorphisms must be implemented.

ACKNOWLEDGMENTS

We thank the patients and the staff of the Hôpital Principal de Dakar and Ndeye Fatou Diop and Maurice Gomis from the Hôpital Principal de Dakar for technical support.

We declare no conflicts of interest.

FUNDING INFORMATION

This research was supported by the Schéma directeur Paludisme, Etat Major des Armées Françaises (grant LR 607a), and by the Ministère des Affaires Etrangères.

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