Determination of the phylogenetic relationships among Pacific salmonids by using short interspersed elements (SINEs) as temporal landmarks of evolution

(retroposon/PCR/phylogeny/orthologous locus/salmon)

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ABSTRACT Several subfamilies of the salmonid Hpa I short interspersed element (SINE) family were isolated from salmonid genomes and were sequenced. For each genomic locus that represented the subfamily, amplification by PCR of the orthologous loci in the 12 fish allowed us to determine the order of branching of the Pacific salmonid species. The deduced phylogeny suggests three evolutionary lines, namely, a line of chum salmon, pink salmon, and kokanee; a line of coho salmon and chinook salmon; and a line of steelhead trout. Our data also support a change in the phylogenetic assignment of steelhead trout from Salmo to Oncorhynchus. We present here an extensive phylogenetic tree constructed from an analysis of differential insertion of SINEs, and we propose that SINE insertion analysis is one of the best available methods for clarifying the order of divergence of closely related species.

A retroposon is defined as ^a nucleotide sequence, present initially as ^a cellular RNA transcript, that has been reincorporated into the genome, presumably via ^a cDNA intermediate. Retroposons constitute roughly 10% of the human genome and are similarly abundant in other mammalian genomes (1, 2). As a result, the remarkable fluidity of eukaryotic genomes reflects the contributions of retroposition (2) as well as mechanisms operating at the DNA level such as mutation and recombination (1-4). Retroposons can be unique to one species, a few species, a genus, or in some cases a family. Retroposition is therefore a specialized form of gene duplication, which is believed to be of major importance in the creation of genetic diversity during evolution (5).

Nonviral retroposons are classified into three main groups: processed retropseudogenes, LINEs (long interspersed elements), and SINEs (short interspersed elements) (6). Except for the rodent type ¹ and human Alu families (7, 8), all of the SINE families examined to date have been shown to be derived from tRNAs (9-14). In contrast to DNA transposable elements, which are often capable of being excised precisely, SINEs appear to be inserted irreversibly and should therefore provide an ideal evolutionary and phylogenetic marker (4).

The Pacific salmon and trout (Oncorhynchus) are a group of closely related species with complex life histories and an interesting global distribution (reviewed in ref. 15). Previously, in an attempt to elucidate a possible role of SINEs in the genomic organization and speciation of salmonids, we characterized three families of tRNA-derived SINEs in salmonid genomes $(16, 17)$. The salmon Sma I family is restricted to the genomes of chum salmon and pink salmon. The charr Fok I family is present only in species that belong to the genus Salvelinus. The third family, the salmonid Hpa ^I family, is present in all species in the family Salmonidae but

not in other species (16). These results suggest that these SINEs were amplified specifically within certain salmonid lineages during evolution.

Our data prompted us to attempt to construct a phylogenetic tree for the salmonid species by using SINE insertions as irreversible events that would serve as informative markers of evolution. In this report, we present a characterization of the four subfamilies of the Hpa I family.[†] These subfamilies were amplified in the four different ancestral species within the genus Oncorhynchus. Such characterization provides a highly reliable order of branching of the various species of Oncorhynchus.

MATERIALS AND METHODS

Experiments were performed by using standard techniques $(18-21)$.

The fish species examined in this study and their geographic sources are listed in Table 1. The family Salmonidae consists mainly of four genera: Oncorhynchus, Salmo, Salvelinus, and Hucho. The genus Oncorhynchus includes eight species, of which six species were analyzed in this study. Genomic DNAs from chum salmon (O, keta), kokanee $(0.$ nerka adonis), and coho salmon $(0.$ kisutch) were used to construct three genomic libraries. Each genomic library was screened for phage clones that contained the salmonid Hpa ^I family and their sequences were determined by the chaintermination method (19). When a unit of the family appeared to be integrated into a unique region of the genome, we synthesized ⁵' and ³' 20-meric primers that flanked the unit. Then PCR was performed (20), using the DNAs from the ¹² listed species as templates. Each locus was named after the number of the clone and the name of the species from which it was isolated. For example, when aphage clone was isolated from the genomic library of kokanee and the number of the clone was 345, the locus was named Hpa(ON)-345 (where ON stands for 0. nerka). The orthologous loci of chum salmon $(0. \text{ keta})$, pink salmon $(0. \text{ gorbuscha})$, coho salmon $(0. \text{ keta})$ kisutch), and steelhead trout $(O.$ mykiss), which could be detected by PCR, were named Hpa(OK)-345, Hpa(OG)-345, Hpa(OKi)-345, and Hpa(OMy)-345, respectively. To confirm the presence or absence of a SINE unit, Southern hybridization experiments were performed (21) and several sequences of products of PCR at the orthologous loci were determined. To distinguish different loci from one another, different numbering systems were adopted for the different genomic libraries.

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Abbreviation: SINE, short interspersed element.

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tThe sequences reported in this paper have been deposited in the GenBank data base (accession nos. D16238-D16246).

Table 1. Fish species analyzed

RESULTS

The Genomes of the Pacific Salmon and Trout Are Distinct
from Those of the Atlantic Salmon and Trout. The Hpa-345 locus was isolated from a genomic library of kokanee. The locus was isolated from a genomic horary of Kokanee. The sequence of this locus, which contains a unit of the Hpu subfamily, and that of flanking regions were determined $[Hpa(ON)-345]$ in Fig. 1A], and two primers flanking the unit $[HPa(011)-375$ in Fig. 1A], and two primers haining the unit (Table 2) were synthesized. Using the 12 fish DNAs as templates, we amplified by PCR. As shown in Fig. 2A, the unit of the Hpa I subfamily is apparently integrated in the genomes of the Pacific salmon and trout, whereas it is not integrated in those of the other fish. To confirm this result, two Southern hybridization experiments were performed. two Southern hybridization experiments were performed. First, an agarose gel of the products of the PCR experiment
decembed in Fig. 2.4 year blotted ante a mambrone filter. The described in Fig. 2A was blotted onto a membrane filter. The filter was then hybridized with a T7 transcript that contained filter was then hybridized with a T_7 transcript that contained ϵ only the unit sequence of the Hpu I family (from nucleotides)
 $\frac{1}{2}$ and $\frac{1}{2}$ f the separatus sequence in ref. 16) as a probability 8-101 of the consensus sequence in ref. 16) as a probe. Fig.

2B shows that the probe hybridized specifically to the upper bands (395 bp) in the gel, confirming that the unit is integrated bands (335 bp) in the gel, community that the unit is integrated α in the genomes of the Pacific salmon and trout. Second, after being washed, the same filter was hybridized with labeled cloned DNA of the Hpa-345 locus that contained the unit sequence as well as the unique flanking regions as a probe. This experiment was performed to ascertain that the loci into This experiment was performed to ascertain that the loci into which the unit was not integrated were intact. Fig. 2C shows that the probe hybridized to the lower bands as well. Fur-
thermore, to confirm that the genomes of the Atlantic salmon thermore, to confirm that the genomes of the Atlantic salmon and trout do not contain the unit of the Hpa I subfamily at this locus, the sequence of ^a 255-bp fragment of DNA from brown trout was determined [Hpa(ST)-345 in Fig. 1A].
The above results clearly indicated that all the present-day

Pacific salmon and trout, including the steelhead trout, which Γ acific salmon and trout, including the steelhead trout, which was recently moved from the genus Salmo to the genus Oncorhynchus (see Discussion) (15), form a monophyletic group. The Hp a I subfamily, including Hpa-345 as a member was amplified in one individual of the ancestral species of all

FIG. 1. Sequences of the orthologous loci of Hpa-345 (A), Hpa-19 (B), Hpa-51 (C), and Hpa-341 (D). Primer sequences are underlined. A unit sequence of the Hpa I family is given in boldface. Identical nucleotides are indic nucleotides in DNAs amplified by PCR, which are different from those in phage DNA clones, are shown by lowercase letters.

Table 2. Sequences of primers used for detection of the four loci analyzed

Locus	5' flanking	3' flanking
Hpa-345	5'-GCACTGTTACCACATAGTTAG-3'	3'-TCACGACGAACTCACAAATA-5'
Hpa-19	5'-AAACACATGGTCACGGTGTG-3'	3'-CACCCATGTCCTTTGTATGT-5'
$Hpa-51$	5'-GAGTTGAACATTACCAGTGACTACCATT-3'	3'-ATACCTTAGTACATCATTGGTTTTTCC-5'
Hpa-341	5'-CGAGTGTATCTGAAGTGTCC-3'	3'-GTCACCTATGAAAGACGGTG-5'

the present-day Pacific salmon and trout, and the unit in this locus was fixed among the population, most likely by random genetic drift. We have isolated several other loci that indicate a clear distinction between the genomes of the Pacific and Atlantic groups. A compilation of these loci, together with Atlantic groups. A compilation of these loci, together with
other loci that specify other subfamilies, will be published other loci that specify other subfamilies, will be published

elsewhere.
Chum Salmon, Pink Salmon, and Kokanee Form a Monophyletic Group. The Hpa-19 locus was isolated from a genomic library of chum salmon and its nucleotide sequence was determined. Primer DNAs were synthesized (Table 2) and PCR was performed. Judging from additional DNA and PCR was performed. Judging from additional DN.
moducts amplified by PCP (Fig. 34), sequence polymoi products amplified by PCR (Fig. 3A), sequence polymor-
mbigme are present in this locus among several selmoni phisms are present in this locus among several salmonid species. However, it is evident that a unit of this *Hpa* I subfamily is integrated in the genomes of chum salmon, pink salmon, and kokanee (Fig. 3). These results suggest the existence of a common ancestral species for the three present-day species. The nucleotide sequences of the Hpa(ON)-19 locus in kokanee and the Hpa(\overline{OT})-19 locus in chinook salmon were determined and confirmed the validity of this inference (Fig. $1B$).

Coho Salmon and Chinook Salmon Are Sister Species. The Hpa-51 locus was isolated from a genomic library of coho salmon and its nucleotide sequence was determined (Fig. 1 C). As shown in Fig. 4, it is evident that a unit of the Hpa I subfamily at this locus was integrated only in the genomes of subtaining at this recupe was integrated only in the generate of

FIG. 2. The Pacific salmonids form a monophyletic group. (A) Analysis of the products of PCR of the orthologous loci of Hpa-345 by electrophoresis in an agarose gel. (B) Southern hybridization of a blot of the gel in A , using the unit sequence of the Hpa I family as a probe. (C) Southern hybridization of the same blot, using the Hpa(ON)-345 DNA as a probe. Hybridization was performed in buffer [50% (vol/vol) formamide/1% SDS/1 M NaCl] at 42°C for 16 hr. In the experiment in C , the blot was treated with an alkaline solution (0.4 \bar{M} NaOH) at 42°C for 30 min before hybridization. Black and white arrowheads indicate positions of DNA with and without an and white arrowheads indicate positions of DNA with and without and integrated unit of the *Hpa* I subfamily, respectively. \ddotsc

orthologous loci in species other than $Oncorhynchus$ were lost during evolution (Fig. 4C). The sequence of the orthologous locus Hpa(ON)-51 indicates that the unit is present in the genome of coho salmon but not in that of kokanee (Fig. $1C$).

The Five Pacific Salmon Form a Monophyletic Group. The Hpa-341 locus was isolated from a genomic library of ko-Hpa-341 locus was isolated from a genomic library of ko-kanee. A unit ofthe Hpa ^I subfamily was demonstrated to be integrated into the orthologous loci of the five Pacific salmon (Fig. 5). Thus, chum salmon, pink salmon, kokanee, chinook salmon, and coho salmon form a monophyletic group. The band in the lane for chum salmon in Fig. 5A has higher mobility than the other four bands, but a hybridization signal was detected in this lane (Fig. $5B$). The nucleotide sequence of the Hpa (OK) -341 locus in chum salmon indicated that a deletion of 160 bp of DNA has occurred specifically in this lineage, with only 42 nucleotides of the unit at the 5' end being present at this locus. Fig. $1D$ shows the sequences of the present at this locus. Fig. 1D shows the sequences of the three orthologous loci of Kokanee, chum salmon, and steel-
head-trout head trout.
This locus does not provide information about the diver-

gence of the former Salmo (steelhead trout) from the genus Oncorhynchus (see Discussion), because the unit was not integrated at the orthologous locus of cherry salmon $(O,$ masou), which belongs to Oncorhynchus (data not shown). Therefore, the data indicate the existence of an ancestral species that was common to the above five salmon after the divergence of cherry salmon and steelhead trout. The relationship between cherry salmon and steelhead trout is curtionship between cherry salmon and steelhead trout is cur-
rently unknown rently unknown.

DISCUSSION
SINE Insertion Analysis Is an Excellent Method for Deter-SINE INSTRUMENT INSERT IN AN EXCELUNT METHOD FOR DUCTION

FIG. 3. Chum salmon, pink salmon, and kokanee form a mono-
phyletic group. (A) Analysis of the products of PCR of the orthologous loci of Hpa-19 by electrophoresis in an agarose gel. (B) Southern hybridization of a blot of the gel in A , using the unit sequence of the Hpa I family as a probe. (C) Southern hybridization of the same blot, using the Hpa (OK)-19 DNA as a probe. The hybridization conditions using the $H_{\text{Pd}}(X, Y, Y, D)$ DIM as a probe. The hybridization conditions were the same as those described in the legend to Fig. 2. were the same as those described in the legend to \mathcal{L}

FIG. 4. Coho salmon and chinook salmon are sister species. (A) Analysis of the products of PCR of the orthologous loci of Hpa-51 by electrophoresis in an agarose gel. (B) Southern hybridization of a blot of the gel in A , using the unit sequence of the Hpa I family as a probe. (C) Southern hybridization of the same blot, using the Hpa (OKi) -51 DNA as a probe. The hybridization conditions were the same as those described in the legend to Fig. 2.

Several types of variation in DNA and protein have been used to infer phylogenetic relationships among species (22). The methods based on comparisons of sequence data among species must, however, always involve statistical errors and often require a large quantity of sequence data to ensure the validity of the branching orders. Moreover, the fact that the

FIG. 5. The five Pacific salmon form a monophylic Analysis of the products of PCR of the orthologous l by electrophoresis in an agarose gel. (B) Southern hybridization of a blot of the gel in A , using the unit sequence of the Hpa I family as a probe. (C) Southern hybridization of the same Hpa(ON)-341 DNA as a probe. The hybridization conditions in the experiment of C were the same as those described in the legend to Fig. 2. In the experiment of B , hybridization was performed in buffer (1% SDS/0.9 M NaCI/0.09 M sodium citrate, pH 7.0, serum albumin/0.1% Ficoll/0.1% polyvinylpyrrolidone/Escherichia coli DNA at 5 mg/ml) at 37° C for 16 hr. FIG. 5. The five Pacific salmon form a monophyletic group. (A) variations in the DNA to be fixed among the population.

rate of molecular evolution is sometimes not constant makes it very difficult to generate an accurate phylogeny.

We propose that SINE insertion analysis, as presented here, is one of the best methods for determining the branching orders of closely related species. Previously, several laboratories demonstrated that the human Alu family can be divided into several subfamilies, with each being inserted into the host genome at a different time during evolution (23–26).
Britten et al. (24) and Jurka and Smith (25) clearly showed 450 bp
that the human Alu repeats that arose at earlier times shared
 $\sqrt{250 \text{ bp}}$ correlated blocks of nucleotides that were different from the -tJ 250 bp correlated blocks of nucleotides that were different from the current consensus sequence at diagnostic positions. On the basis of these findings, Ryan and Dugaiczyk (27) suggested the possibility that the distribution of Alu elements might be used to resolve the branching order in the evolution of primates. In this case, however, only the loci specific to the human genome were analyzed, and no extensive classification using this strategy has been attempted to date. We believe that our phylogeny is the first example of the full application of this methodology. We have isolated and characterized several loci per branching point, greatly reducing the probability of phylogenetic incongruence due to poly-
morphisms in the ancestral species (28).

> With regard to the four loci described in the present study, it is not precisely determined whether each inserted SINE is fixed or polymorphic in the lineages of salmonid species.
Recently we isolated one locus which contains a member of the Hpa I subfamily amplified specifically in the chum salmon, and we showed that the inserted SINE in this locus salmon, and we showed that the inserted SINE in this locus is fixed in all the populations of chum salmon from various places in the Pacific Ocean (unpublished results). This result strongly suggests that the SINE members in the four loci described above may be fixed in all the populations of salmonid species, because they are much older than the member of the species-specific SINE in the chum salmon.

Phylogeny of Pacific Salmonids. Western American trout e fact that the Phylogeny of Pacific Salmonian troute a method is consist of various species, which include steelhead troute (rainbow trout), cutthroat trout, and allied subspecies. The phylogenetic position of these species, represented by the steelhead trout in this study, has long been disputed because
of their close relatedness both to Pacific salmon (Oncorhynof their close relatedness both to Pacific salmon (Oncorhynchus) and to Atlantic salmon (Salmo) (reviewed in ref. 15). Recently, these species, which were previously classified as Salmo, were officially renamed Oncorhynchus (29). The decision was based on studies of the osteology (30) and decision was based on studies of the osteology (30) and biochemistry (31) of trout and salmon, which showed that 1435 bp steelhead trout and their close relatives are more closely related to Pacific salmon than to brown trout and Atlantic salmon.

 $\sum_{z \to z}$ bp salmon.
The SINE insertion analyses described herein support this change in classification. The data presented here and other data to be published elsewhere demonstrate that there are several genomic loci that show clear distinctions between the Pacific salmonids and the Atlantic ones. Our results indicate that all the Pacific salmonids, including steelhead trout, have that all the Pacific salmonids, including steelhead trout, have a common ancestral species and form a monophyletic group. Since all the present-day Pacific salmonids contain the inserted SINE in the Hpa-345 locus, it is likely that the ancestral species lived as a unique species in the Pacific Ocean during a long enough period of time for several

Chum salmon, pink salmon, and kokanee were shown to have a common ancestral species and to form a monophyletic group (Fig. 6). With respect to the relationship among these three species, we suggest that the kokanee lineage diverged before the divergence of the remaining two species, because the salmon Sma I family is present only in the genomes of Formed in buffer the salmon Δma I family is present only in the genomes of contract the salmon Δma I family $1/0.1\%$ bovine chum salmon and pink salmon (16). The salmon Sma I family $\text{me}/\text{Escherichia}$ may have been amplified in a common ancestral species of the theory than the concession and the common ancestral species of the theory than the concession and the common species of the common species of th these two species, suggesting that kokanee is distinct from

FIG. 6. Phylogenetic tree of the salmonid species as deduced from SINE insertion analyses. Three different kinds of tRNAderived SINEs, namely, the Sma I family, the Fok I family, and the Hpa ^I family, were amplified at specific stages of evolution of salmonid species (16). Multiple filled arrows show the Hpa I subfamilies described in the text and ref. 16.

that pair of species and might occupy an intermediate evolutionary position (16) . The phylogenetic relationship of these three species in Fig. 6 is well correlated with that of Thomas et al. (32), obtained from analysis of mitochondrial DNA, but not with that obtained from allozyme variations by Utter et al. (33) or with that derived from morphological comparisons (29).

On the basis of ecological, morphological, behavioral, and biochemical data, there is general agreement that coho salmon and chinook salmon are the most closely related species (29, 30, 32-34). Our SINE insertion data support this conclusion.

With respect to the phylogenetic position of steelhead trout, our phylogeny (Fig. 6) indicates that steelhead trout diverged before the divergence of the other five Pacific salmon. This conclusion is in accordance with that from morphological comparisons (29) and with that from allozyme variations (33, 34) but not with that of Thomas et al. (32), who showed, by analysis of mitochondrial DNA, that rainbow trout (steelhead trout), coho salmon, and chinook salmon form a monophyletic group. When a root of the phylogenic tree of Thomas et al. (32) is moved to the evolutionary line of rainbow trout, their phylogeny becomes the same as ours. Therefore, we suggest that the rate of mitochondrial molecular evolution altered in the lineages ofrainbow and cutthroat trout. With respect to the order of branching, we believe that our phylogeny (Fig. 6) is the most reliable of those presented to date.

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