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# **REGULATORY T CELLS IN NON-LYMPHOID TISSUES**

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# Abstract

Both Foxp3<sup>+</sup>CD4<sup>+</sup> regulatory T cells ( $T_{reg}$  cells) and local immune responses within nonlymphoid tissues have long been recognized as important elements of a well-orchestrated immune system, but only recently have these two fields of study begun to intersect. There is growing evidence that  $T_{reg}$  cells are present in various non-lymphoid tissues in health and disease, that they have a unique phenotype, and that their functions go beyond the classical modulation of immune responses. Thus, tissular  $T_{reg}$  cells might add yet another level to classification of the  $T_{reg}$  cell compartment into functional and/or phenotypic subtypes. This review summarizes recent findings in this new field, discussing knowns and unknowns about the origin, phenotype, function and memory of non-lymphoid tissue-resident  $T_{reg}$  cells.

## Introduction

Regulatory T cells that express the Foxp3 transcription factor, affectionately termed  $T_{reg}$  cells, are one of the immune system's main bastions against inappropriate or over-exuberant responses. They control autoimmunity, allergic and inflammatory reactions, and responses to infectious agents and tumors. Over the past decade, innumerable studies have addressed differentiation of the majority of  $T_{reg}$  cells in the thymus, generation of a minority in the periphery, homeostasis of the  $T_{reg}$  cell compartment, and cellular and molecular mechanisms of  $T_{reg}$ -mediated suppression<sup>1</sup>. For the most part, these explorations have taken the average  $T_{reg}$  cell residing in the spleen or lymph nodes to be paradigmatic.

However, it eventually became impossible to ignore the considerable heterogeneity of the Foxp3<sup>+</sup>CD4<sup>+</sup> compartment, especially once transcriptome analysis had become a routine tool<sup>2</sup>. Initially,  $T_{reg}$  cell subphenotypes were delineated based on expression of activation or memory markers; adhesion molecules, notably CD103; or homing receptors. But a major advance was the discovery of  $T_{reg}$  cell functional diversity matched to the type of response being reined in. A subtype of  $T_{reg}$  cells was discovered that depends on the transcription factor IRF4 to control T helper ( $T_H$ )2 cells, which also critically rely on IRF4 (ref.<sup>3</sup>). In parallel, a discrete CXCR3<sup>+</sup>  $T_{reg}$  cell subtype was found, dependent on the T-bet transcription factor, that is specialized in regulating the activities of  $T_H$ 1 cells, which also require T-bet for their differentiation and functions<sup>4</sup>.  $T_{reg}$  cells that optimally regulate

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interleukin 17 (IL-17)- or IL-27-dependent responses may be yet different subtypes<sup>5, 6</sup>. The relevance of these various subtypes was serendipitously confirmed in a recent study showing a mutation of Foxp3 to dampen arthritis in an IL-17, IL-4 dependent mouse model, while exacerbating type-1 diabetes in NOD mice, a  $T_H$ 1-type disorder<sup>7</sup>. Another striking match between the cells that regulate and those that are regulated is found in germinal centers: follicular regulatory ( $T_{FR}$ ) and helper ( $T_{FH}$ ) cells both depend on Blimp-1 and Bcl6 for their differentiation/homeostasis and CXCR5 for their localization<sup>8, 9</sup>. The advantage of such a matching is probably that it provokes co-localization to and/or co-survival within discrete locations. Arming regulatory and effector cells with the same capabilities could be dangerous, but safeguards are in place, for example  $T_H$ 1-type  $T_{reg}$  cells poorly up-regulate IL-12R $\beta$ 2 upon interferon- $\gamma$  induction of STAT1, meaning that their differentiation to  $T_H$ 1 effector cells is aborted<sup>10</sup>.

This review focuses on studies that go one step further, highlighting the phenotype and functions – sometimes exquisitely adapted – of  $T_{reg}$  cells residing in non-lymphoid tissues. We will survey the populations of tissue-resident  $T_{reg}$  cells, focusing on a few particularly interesting examples; consider their origin; discuss potential cellular targets; and weigh the concept of  $T_{reg}$  cell memory. Lastly, we will highlight some general principles and knowledge gaps to fill.

### Tissue-resident T<sub>reg</sub> cells: the landscape

The presence of a distinct population of  $T_{reg}$  cells has been documented in several nonlymphoid tissues of both mice and humans: skin, intestinal mucosa, lung, liver, adipose tissue, autoimmune target tissues, infected tissues, grafts, placenta, tumors, atherosclerotic plaques and injured muscle are just some examples (<sup>11–21</sup>, D.B., C.B. and D.M., manuscript submitted). It is clear from this extensive list that  $T_{reg}$  cells can localize in healthy tissues, in tissues with various types and grades of inflammation, and in immunoprivileged sites. In every case where the comparison has been made, tissue-resident  $T_{reg}$  cells are distinguishable from classical lymphoid-organ  $T_{reg}$  cells in phenotype and function. While they display some features of activated/effector cells<sup>22</sup>, certain properties make each tissueresident  $T_{reg}$  cell population unique, such as by the expression of specific transcription factors, chemokine receptors or effector molecules; or by a distinct T cell receptor (TCR) repertoire, migration pattern, mechanism of action or targets.

Currently, one of the best-characterized examples of tissue-resident  $T_{reg}$  cells is the population found in visceral adipose tissue (VAT)<sup>12</sup>.  $T_{reg}$  cells are enriched in VAT of lean, aged, male mice, constituting more than 50% of the CD4<sup>+</sup> T cell compartment at that site, a significantly higher fraction than the 5–15% routinely observed in lymphoid organs, including in aged individuals. Microarray-based gene-expression profiling, confirmed by flow cytometry analyses, revealed VAT  $T_{reg}$  cells to have a unique phenotype. While they are *bona fide*  $T_{reg}$  cells, as shown by their recapitulation of most of the classical  $T_{reg}$ signature and by their functionality in standard *in vitro* suppression assays, they differentially express a panoply of genes in comparison with lymphoid-organ  $T_{reg}$  cells. The set of loci overexpressed in VAT  $T_{reg}$  cells includes those encoding several chemokine receptors (for example, CC chemokine receptor (CCR)1, CCR2, CCR9),

immunomodulatory cytokines (such as IL-10), and the transcription factor peroxisome proliferator-activated receptor (PPAR)- $\gamma$ , all of which confer VAT T<sub>reg</sub> cells with unique functional properties<sup>12, 23</sup>. As flow cytometry studies showed that some of those markers are not expressed by 100% of the fat T<sub>reg</sub> cells<sup>12, 23</sup>, it is not clear how heterogeneous the population is and whether this has functional significance or more prosaically marks cells at different cell-cycle or differentiation stages. Their TCR repertoire is another feature contributing to a distinct phenotype: the TCR sequences they employ have little overlap with those used by lymphoid-organ T<sub>reg</sub> cells (<sup>12</sup> and Kolodin *et al.*, personal communication). In addition, there is a striking expansion of certain T<sub>reg</sub> clones in the adipose tissue, suggesting that recognition of particular antigens may have an important role in the accumulation of T<sub>reg</sub> cells in abdominal fat (further discussed below).

The major driver of VAT  $T_{reg}$  cell accumulation, phenotype and function is PPAR- $\gamma^{23}$ . A member of the nuclear receptor superfamily characterized as the "master regulator" of adipocyte differentiation and function<sup>24</sup>, PPAR- $\gamma$  interacts with Foxp3 to promote up-regulation of the VAT  $T_{reg}$  signature in *in vitro* transduction experiments. Mice lacking PPAR- $\gamma$  only in  $T_{reg}$  cells show a substantial and specific reduction in the frequency and number of  $T_{reg}$  cells in VAT vis-à-vis their wild-type littermates, without any changes in lymphoid-organ  $T_{reg}$  populations. Moreover, the few VAT  $T_{reg}$  cells remaining in PPAR- $\gamma$  mutant mice show an under-representation of the VAT  $T_{reg}$  signature. The fact that PPAR- $\gamma$  expression in VAT  $T_{reg}$  cells is much higher than in anyother  $T_{reg}$  population, together with the observation that they can take up lipids by expressing CD36, the scavenger receptor<sup>23</sup>, suggests that tissue-resident  $T_{reg}$  cells are attuned to local cues, which they can exploit for phenotypic and functional specialization, as well as for preferential survival within the tissue microenvironment (Fig.1).

Another tissue-resident population of great interest are  $T_{reg}$  cells that infiltrate injured and regenerating skeletal muscle (D.B., C.B. and D.M., unpublished data). Within days after skeletal muscle injury (of various types),  $T_{reg}$  cells start to accumulate locally, their frequency among CD4<sup>+</sup> T cells increasing steadily to up to 50–60% and remaining at that frequency for weeks, long after local inflammation has resolved. Microarray-based gene-expression profiling revealed that muscle  $T_{reg}$  cells have a unique transcriptome, more closely related to that of VAT  $T_{reg}$  cells than to that of lymphoid-organ  $T_{reg}$  cells. For example, genes encoding IL-10, amphiregulin (Areg) and T-cell immunoglobulin domain and mucin domain 3 (TIM-3) are highly expressed by  $T_{reg}$  cells accumulated in muscle. Muscle  $T_{reg}$  cells have a skewed TCR repertoire, which appears not to overlap with that of muscle conventional CD4<sup>+</sup> T cells or of splenic  $T_{reg}$  cells, and shows clear signs of clonal expansion. Interestingly,  $T_{reg}$  cells also accumulate in skeletal muscle of dystrophic mice (such as dystrophin-deficient mdx mice and dysferlin-deficient mice), whose chronic muscle injury has a genetic origin.

Given recent successes with immunomodulatory anti-tumor strategies (see Gajewski et al. reviewed in this issue), there is a growing interest in the more heterogeneous group of tumor-infiltrating  $T_{reg}$  cells. In solid tumors of non-lymphoid origin,  $T_{reg}$  cells can account for 30–50% of CD4<sup>+</sup> T cells, depending on the tumor type<sup>19</sup>. Like other tissue-resident  $T_{reg}$  cells, those found in tumors have a distinctive phenotype that differentiates them from  $T_{reg}$ 

cells found in the circulation or in lymphoid organs. Microarray-based gene-expression profiling of tumor-associated  $T_{reg}$  cells is still lacking<sup>25</sup>, but Foxp3<sup>+</sup>CD4<sup>+</sup> T cells from several mouse and human tumors show increased expression of cell-surface markers such as Cytotoxic T-Lymphocyte-Antigen 4 (CTLA-4), Inducible T-cell Costimulator, (ICOS), TIM-3, glucocorticoid-induced TNFR family related gene (GITR); as well as of suppressive cytokines such as IL-10 and transforming growth factor (TGF)- $\beta$ ; and a variety of chemokine receptors<sup>19</sup>, <sup>26–29</sup>.

#### Who gets there, when and how?

 $T_{reg}$  cells are found in healthy tissues, although usually in very small numbers<sup>11</sup>. Upon challenges of different types (autoimmunity, infection, injury),  $T_{reg}$  cell numbers, and often their frequencies, increase considerably. Diverse mechanisms, not mutually exclusive, have been proposed to explain the accumulation of  $T_{reg}$  cells at different tissular sites: chemokine-based recruitment of circulating  $T_{reg}$  cells, local TCR- or cytokine-driven expansion, conversion of local or circulating conventional CD4<sup>+</sup> T cells, or prolonged survival (Fig. 1). In some cases, for example tumor  $T_{reg}$  cells, all of these mechanisms have been championed by one investigator or another<sup>19, 25, 26, 30–34</sup>.

As for other types of immune-system cells, recruitment of Treg cells to non-lymphoid tissues is governed by specific combinations of chemotactic molecules and their receptors (reviewed in<sup>35</sup>).  $T_{reg}$  cells residing in various tissues express distinct patterns of chemokine receptors and adhesion molecules in comparison with their lymphoid-organ counterparts, both at single cell level and in terms of population frequencies<sup>11, 23</sup>. This finding is true even for T<sub>reg</sub> cells residing in non-inflamed tissues like skin, lung and liver, wherein an enrichment for CCR4<sup>+</sup>CD103 (integrin  $\alpha_E$ )<sup>+</sup> T<sub>reg</sub> cells is observed<sup>11</sup>. Ccr4<sup>-/-</sup> T<sub>reg</sub> cells show impaired accumulation in healthy non-lymphoid tissues, and this dearth has functional consequences as mice defective in CCR4<sup>+</sup> T<sub>reg</sub> cells develop severe skin inflammation and a less severe lung infiltrate. It seems that antigen-specific activation of T<sub>reg</sub> cells in subcutaneous lymph nodes under inflammatory conditions induces up-regulation of CCR4, priming Treg cells to migrate to the affected tissue and to suppress immune responses locally. However, the origin of the small fraction of CCR4+CD103+ T<sub>reg</sub> cells found in lymphoid tissues in the absence of antigen challenge remains unclear. In this regard, a study showed that, in humans, only few Treg cells express CCR4 at birth but the majority expresses the gut-homing receptor  $\alpha_4\beta_7$ , while the reverse pattern is true for adults<sup>36</sup>. The switch commences between 1.5 and 3 years of age and correlates with a change from a naïve to memory Treg phenotype, which has been suggested to depend on the recognition of microbial antigens in the gut, although this point has yet to be definitively demonstrated.

In addition to CCR4, an array of chemokine receptors can be involved, usually in a redundant fashion, in the recruitment of  $T_{reg}$  cells to non-lymphoid tissues under various inflammatory conditions<sup>37</sup>. In some cases, the expression of a particular receptor by tissue-resident  $T_{reg}$  cells is directly associated with the type of tissue, while in others it is related to the type of T cell response (T<sub>H</sub>1, T<sub>H</sub>2, T<sub>H</sub>17) occurring in the tissue and the accompanying  $T_{reg}$  cell polarization, for example, the preferential recruitment of CCR6<sup>+</sup> and CXCR3<sup>+</sup>  $T_{reg}$  cells to sites with an ongoing  $T_{H}17$ - or  $T_{H}1$ -driven inflammation, respectively<sup>4, 5</sup>.

The pattern and timing of trafficking between lymphoid and non-lymphoid tissues is not well-defined in most cases. Interestingly, in a model of pancreatic islet transplantation,  $T_{reg}$  cells first migrate from blood to the inflamed allograft, in a process dependent on CCR2, CCR4, CCR5 and P- and E-selectin ligands<sup>38</sup>. Upon activation in the allograft, they subsequently move to the draining lymph nodes in a CCR2-, CCR5-, and CCR7-dependent fashion. This sequential migration is required for an efficient suppression of alloimmunity, since impairment of either of the steps by abolishing expression of the relevant receptor(s) in  $T_{reg}$  cells results in decreased graft survival.

Although not universally accepted, it has been suggested that the T<sub>reg</sub>-cell TCR repertoire is biased towards the recognition of self-antigens<sup>1</sup>, which could facilitate the localization of Treg cells in non-lymphoid tissues to prevent harmful autoimmune attacks, and collateral damage to the tissue resulting from inflammation or injury. Sequencing of the TCR repertoire of VAT T<sub>reg</sub> cells in either wild-type mice or a mouse line engineered to have only limited TCR diversity yielded several interesting observations (<sup>12</sup>, Kolodin *et al*, personal communication). First, the sequences expressed by VAT Treg cells have little overlap with those expressed by conventional CD4<sup>+</sup> T cells, either in adipose tissue or in lymphoid organs, arguing against Treg cell conversion in this context. Second, there are striking clonal expansions of Treg cells in VAT even in standard mice. Lastly, in the "limited" line, it is possible to find examples, in the same or different individuals, of a repeated complementary-determining region (CDR)-3 protein sequence specified by different nucleotide sequences. These findings argue that VAT Treg cells are responding to one or more local antigens, whether they be derived from lipids or proteins. By combining an ovalbumin (OVA)-specific TCR transgenic model and the expression of OVA in the skin, a recent study showed that engineered expression of a self-antigen in a peripheral tissue can drive the activation and proliferation of TCR-transgenic T<sub>reg</sub> cells, which mediate resolution of organ-specific autoimmunity<sup>39</sup>. These antigen-specific  $T_{reg}$  cells remain in the target tissue and are primed to attenuate subsequent autoimmune reactions when the antigen is reexpressed (discussed in the next section), suggesting that antigen recognition is important for tissue T<sub>reg</sub> cell function and memory.

A recent report provided the first clear identification of the antigen specificity of a natural population of tissue-resident  $T_{reg}$  cells<sup>40</sup>. By sequencing the TCR repertoire of prostate-tumor-infiltrating  $T_{reg}$  cells, the authors identified a TCR  $\alpha\beta$  pair recurrently enriched in independent mice, whose sequences they used to generate a TCR transgenic line. In tumor-free TCR-transgenic mice,  $T_{reg}$  cells are found selectively in the prostate and its draining lymph nodes, suggesting that the cloned TCR is specific for an antigen expressed in the normal prostate in addition to the malignant tissue. Interestingly, selection of the TCR-transgenic  $T_{reg}$  cells is dependent on expression of Aire in the thymus, thereby linking two major mechanisms of immunological tolerance - one central, one peripheral. Although similar demonstrations for other tissue-resident  $T_{reg}$  specificities will be necessary before one can definitively generalize, this work supports the hypothesis that tissue-resident  $T_{reg}$  cells are generated in the thymus in response to self-antigens, and that this TCR specificity facilitates their localization and amplification in non-lymphoid tissues, especially in

pathological situations such as inflammation or cancer in which an enhanced self-antigen presentation can occur in the periphery.

While the majority of the Treg cells differentiates in the thymus, a small fraction of them can be generated in the periphery, by conversion of Foxp3<sup>-</sup>CD4<sup>+</sup> T cells into Foxp3<sup>+</sup>CD4<sup>+</sup> Tregs, termed "peripheral Treg cells" (pTreg cells) to distinguish them from their thymicderived counterparts. There has been a lot of interest in conversion as a mechanism of T<sub>reg</sub> cell accumulation in non-lymphoid tissues, although few studies have clearly demonstrated that, in certain sites and/or conditions, pTreg cells constitute a substantial proportion of tissular  $T_{reg}$  cells. The intestinal lamina propria  $T_{reg}$  pool is one of the main populations of non-lymphoid-tissue Treg cells impacted by conversion, and pTreg cells have been shown to contribute substantially to oral tolerance (reviewed in<sup>41</sup>). Exposure to agonist administered orally and specific microbial products strongly induce conversion in the lamina propria, which is reflected in the increased frequency of Treg cells, as well as in the increased proportion of Helios<sup>-</sup> T<sub>reg</sub> cells in that tissue<sup>42–45</sup>. The analysis of a mutant mouse bearing a deletion in the CNS1 enhancer region of the Foxp3 locus necessary for peripheral induction confirmed previous observations on lamina propria T<sub>reg</sub> cells and suggested that the accumulation and function of pTreg cells might be important at mucosal and feto-maternal interfaces, and perhaps little else<sup>17, 46, 47</sup>. Different populations of tolerogenic antigenpresenting cells promote Foxp3 induction at mucosal surfaces, such as CD103<sup>+</sup> dendritic cells that produce retinoic acid in the intestinal lamina propria<sup>42, 48</sup>, and tissue-resident macrophages in the lung mucosa<sup>49</sup>. Interestingly, a report showed that neurons can elicit conversion of encephalitogenic Foxp3<sup>-</sup>CD4<sup>+</sup> T cells into T<sub>reg</sub> cells in vitro, although evidence for in vivo conversion was weak<sup>50</sup>. The generation of pT<sub>reg</sub> cells has also been proposed to play a role in the accumulation of  $T_{reg}$  cells in tumors<sup>34, 51</sup>, although the exact contribution of this mechanism versus recruitment/expansion of thymic-derived T<sub>reg</sub> cells remains controversial<sup>25, 41</sup>. In fact, a study using a mouse tumor model showed that both mechanisms can coexist and that the induction of tumoral pT<sub>reg</sub> cells is intrinsically influenced by the tumor microenvironment and the presence of a tumor-specific antigen<sup>51</sup>.

Lastly, the question remains as to how and when the few  $T_{reg}$  cells found in healthy, unchallenged tissues are recruited to these sites. It is possible that a combination of specific expression of chemokine receptors and antigen specificity constantly brings a small number of  $T_{reg}$  cells to the tissues throughout life. Alternatively, early seeding with  $T_{reg}$  cells precursors during fetal life might occur, paralleling the phenomenon observed in tissue macrophages<sup>52</sup>, followed by self-renewal of the tissue-resident  $T_{reg}$  cells. In theory, seeding during embryonic life could occur after or before thymic development, although extrathymic differentation of TCR $\alpha\beta$  T cells remains controversial.

#### Immunological and non-immunological targets

One of the major functions of  $T_{reg}$  cells residing in non-lymphoid tissues is to control local inflammation. Since their initial discovery, Foxp3<sup>+</sup>CD4<sup>+</sup> cells were recognized to be potent suppressors of T cell responses, and this activity has been extended to tissular Foxp3<sup>+</sup> cells. In addition, tissue-resident  $T_{reg}$  cells can strongly impact myeloid populations in the

vicinity, inhibiting neutrophils and pro-inflammatory macrophages, and promoting the activities of anti-inflammatory macrophages and monocyte subsets (Fig. 2).

As a particular example, VAT  $T_{reg}$  cells may influence a broad spectrum of targets. Their elevated expression of factors like IL-10 and CTLA-4, and their effectiveness in the typical *in vitro* suppression assay<sup>12</sup> suggest that VAT  $T_{reg}$  cells are capable of controlling conventional CD4<sup>+</sup> and CD8<sup>+</sup> T cell populations in the adipose tissue, although *in vivo* experiments to substantiate this point are still lacking. They may also control co-resident myeloid cells, as suggested by an inverse correlation between the frequency of  $T_{reg}$  cells and that of pro-inflammatory myeloid populations (CD11b<sup>+</sup>CD11c<sup>+</sup>F4/80<sup>+</sup> macrophages and pro-inflammatory CD11b<sup>+</sup>Ly6c<sup>hi</sup> monocytes) observed in VAT depots<sup>23, 53, 54</sup>. Similarly, muscle  $T_{reg}$  cells are important in controlling the pro-inflammatory to anti-inflammatory switch that occurs in the myeloid infiltrate of injured muscle; punctual ablation of  $T_{reg}$  cells at the time of muscle injury results in prolonged accumulation of pro-inflammatory Ly6c<sup>hi</sup> monocytes in the injured tissue (D.B., C.B. and D.M., unpublished data).

Because of the ability of  $T_{reg}$  cells to dampen immune responses that can eradicate malignancies, their accumulation in tumors is considered to be a tumor 'escape' mechanism. Tumor-infiltrating  $T_{reg}$  cells can be as effective as peripheral-blood  $T_{reg}$  cells in T cell-suppression assays<sup>30</sup> or even significantly better<sup>29</sup>; and, in many cases, a high frequency of intra-tumoral  $T_{reg}$  cells is correlated with poor prognosis<sup>30, 55, 56</sup>. On the other hand,  $T_{reg}$  cells can also have anti-tumoral effects by inhibiting inflammation in the tumor environment. A strong link between inflammation and cancer exists in many tissues: inflammation can promote proliferation and survival of malignant cells, angiogenesis and metastasis<sup>57, 58</sup>. In line with these observations, in certain tumors, like colorectal cancers, a high density of  $T_{reg}$  cells correlates with a favorable prognosis<sup>59, 60</sup>. It has recently been proposed that  $T_{reg}$ -mediated suppression of pro-inflammatory  $T_H 17$  responses to the dense microbiome of the large intestine is key to preventing tumor growth in the intestinal epithelium<sup>60</sup>.

There is growing evidence that  $T_{reg}$  cells (in particular those residing in tissues) not only control T lymphocytes and other immune-system cells, but also regulate certain nonimmunological processes, including systemic metabolic indices<sup>12, 23, 54, 61</sup>, ischemic stroke<sup>62, 63</sup>, formation of atherosclerotic plaques<sup>64, 65</sup>, cardiac remodeling after myocardial infarction<sup>66</sup>, liver cholestasis and fibrosis<sup>67</sup> and skeletal muscle regeneration (D.B., C.B. and D.M., manuscript submitted). It is still unclear to what extent such influences reflect direct interaction between  $T_{reg}$  cells and their non-immunological target cells or an indirect effect of  $T_{reg}$  suppression of local inflammation (Fig. 2). In any case, these non-traditional roles can have profound impact on both homeostatic and pathophysiological processes and should be considered an important facet of  $T_{reg}$  cell function.

An unusual property of VAT  $T_{reg}$  cells is their control of metabolic indices, inhibiting local and systemic insulin resistance and glucose intolerance. VAT of obese individuals exhibits low grade, chronic inflammation which has been directly linked to the appearance of metabolic abnormalities, ultimately type 2 diabetes and the metabolic syndrome<sup>68, 69</sup>. That the frequency of VAT, but not lymphoid-organ  $T_{reg}$  cells drops severely upon feeding mice

with a high-fat diet (HFD) or in genetic models of obesity first suggested that this T<sub>reg</sub> subset might influence local and/or systemic metabolism<sup>12, 53</sup>. Loss-of-function experiments, relying on punctual and specific Treg ablation in a Foxp3-DTR mouse model, demonstrated increases in inflammatory mediators in the visceral fat depot, a decrease in insulin-stimulated insulin receptor tyrosine phosphorylation in VAT and the liver, and insulin resistance upon loss of T<sub>reg</sub> cells<sup>12</sup>. Conversely, gain-of-function experiments, which entailed treatment of HFD-fed mice with IL-2-anti-IL-2 complexes, resulted in enhanced IL-10 expression in VAT and improved insulin sensitivity and glucose tolerance. These results were mirrored in a more recent report on experiments manipulating PPAR- $\gamma$ expression or signaling: mice devoid of PPAR-y only in Treg cells show a strong reduction in VAT Treg cells and degradation of metabolic indices, while HFD-fed wild-type mice injected with a PPAR- $\gamma$  agonist, pioglitazone, have a substantially more robust T<sub>reg</sub> cell population and metabolic improvements. Strikingly, and unexpectedly, the restoration of metabolic indices typically induced by treatment of HFD-fed mice with pioglitazone is only very partial in mice lacking PPAR- $\gamma$  specifically in T<sub>reg</sub> cells<sup>23</sup>, demonstrating not only that the beneficial effects of this drug occur in part by targeting VAT Treg cells, but also that VAT (and not lymphoid) Tregs are important players in the control of the metabolic indices downstream of obesity and inflammation of the adipose tissue. At least part of this  $T_{reg}$  cell influence may be directly on adipocytes, as one of their major mediators, IL-10, can engage receptors on adipocytes to down-regulate pro-inflammatory cytokines and increase glucose uptake.

 $T_{reg}$  cells that accumulate in injured skeletal muscle also seem to impact non-immunological processes, in this case tissue regeneration (D.B., C.B. and D.M., unpublished data). Mice punctually depleted of  $T_{reg}$  cells mice show impaired muscle repair according to multiple criteria, notably the functionality of satellite cells, key players in the regeneration of skeletal muscle. Muscle  $T_{reg}$  cells overexpress Areg, a growth factor that induces *in vitro* differentiation of satellite cells, which express Areg receptors (D.B., C.B. and D.M., manuscript in preparation), suggesting that local  $T_{reg}$  cells could have a direct effect on skeletal muscle precursor cells.

The influence of  $T_{reg}$  cells on the pathophysiology of ischemic stroke provides yet another example of non-immune processes regulated by  $T_{regs}$ . However, in this case, opposite effects have been reported by different investigators, so whether  $T_{reg}$  cells are beneficial or detrimental to the outcome of ischemic stroke remains controversial. One study showed that  $T_{reg}$  cells can be found in the post-ischemic brain; and that their depletion profoundly enhances brain damage and exacerbates the functional outcome<sup>62</sup>. IL-10 production by  $T_{reg}$ cells may be their major mechanism of immunomodulation in the brain and of  $T_{reg}$  mediated cebroprotection. Unfortunately, these results conflict with another study published subsequently that indicated that indicating that  $T_{reg}$  cells promote acute ischemic stroke in mice by inducing dysfunction of the cerebral microvasculature<sup>63</sup>. Notably, this negative impact of  $T_{reg}$  cells on recovery from stroke is unrelated to their classical immunoregulatory functions; in fact, the CD4<sup>+</sup> T cell and macrophage populations infiltrating the ischemic tissue are not affected by the depletion of  $T_{reg}$  cells. Rather, the negative influence seems to result from an interaction between  $T_{reg}$  cells and activated cerebral endothelial cells and

platelets. These contradictory findings on the role of  $T_{reg}$  cells in stroke issue from different models of  $T_{reg}$  depletion (anti-CD25-mediated depletion vs a Foxp3-DTR line), by the different stroke models used (permanent vs temporal; different ischemia times), or the timing of read-out (late vs acute stages).

A liaison between  $T_{reg}$  cells and endothelial cells has also been observed within the tumor microenvironment, in this case reflecting a pro-angiogenic effect by tumor-infiltrating Foxp3<sup>+</sup>CD4<sup>+</sup> cells<sup>70</sup>. Tumor hypoxia promotes the recruitment of  $T_{reg}$  cells through induction of expression of the chemokine, CC-chemokine ligand 28 (CCL28), in tumor cells. Elevated numbers of Treg cells expressing CCR10 accumulate in CCL28-positive tumors, which exhibit increased growth and angiogenesis in comparison with CCL28negative tumors. Angiogenesis is dependent on the presence of tumor-infiltrating T<sub>reg</sub> cells, which not only enhance expression of vascular endothelial growth factor A (VEGFA) in tumor cells, but also directly contribute to the tumor VEGFA pool by producing the proangiogenic factor themselves<sup>70</sup>. Tumor-infiltrating  $T_{reg}$  cells also promote mammary carcinoma metastasis in the lung by producing receptor activator of nuclear factor kappa-B ligand (RANKL)<sup>71</sup>. Thus, the impact of tumor-resident  $T_{reg}$  cells on tumor growth and patient prognosis reflects both traditional (on immune targets) and non-traditional (on nonimmune targets) Treg functions. Dissection of the individual pathways regulated by tissue Treg cells will be important for the design of effective and specific therapies aimed at modulating T<sub>reg</sub> cells and their products in cancer and other diseases.

### T<sub>req</sub> cell memory in tissues

The generation of memory is one of the hallmarks of adaptive immunity, and it is becoming clear that Treg cells are no exception to the rule. During viral infections, the expansion of antigen-specific  $T_{reg}$  cells is followed by a contraction phase and the formation of a memory  $pool^{72, 73}$ . Upon re-infection, these memory  $T_{reg}$  cells rapidly expand and effectively control specific anti-viral responses by CD4<sup>+</sup> and CD8<sup>+</sup> T cells. An accelerated accumulation of memory T<sub>reg</sub> cells is observed in the non-lymphoid tissues targeted by the infection; furthermore, memory Treg cells are capable of suppressing the collateral tissue damage and inflammation elicited by recall expansion of non-Treg memory CD4<sup>+</sup> T cells. A link between Treg memory and tissue-resident Treg cells has also been demonstrated in the context of autoimmunity<sup>39</sup>. Using a TCR-neo-self-antigen (OVA) double-transgenic model, a recent study showed that upon local induction of ovalbumin expression, ovalbumin-specific  $T_{reg}$ cells accumulate in the skin. After resolution of the inflammatory response, activated  $T_{reg}$ cells remain in the target tissue in high frequencies and are primed to attenuate subsequent autoimmune responses when the antigen is re-expressed. These memory  $T_{reg}$  cells can survive in the non-lymphoid tissues in the absence of overt expression of antigen, and show enhanced functional activity.

In human skin, resident  $T_{reg}$  cells comprise 5–10% of total skin T cells<sup>74</sup>. These  $T_{regs}$  express the memory T cell marker CD45RO, and are thus typically referred to as 'effector-memory  $T_{reg}$  cells', although their true memory nature has not been formally demonstrated. They reside preferentially in the epidermis, where Langerhans cells (LCs) induce their proliferation in an MHC-class-II- and IL-15-dependent manner<sup>75</sup>. It has been proposed that

self-peptides or the normal skin microbiome might be the source of antigens presented by LCs. Human skin-resident  $T_{reg}$  cells also proliferate in response to pathogens<sup>75</sup> and during the recall response induced by injection of tuberculin purified-protein derivative (PPD) in individuals immunized with bacille Calmette-Guérin (BCG)<sup>76</sup>.

Memory  $T_{reg}$  cells seem also to have an important function in preventing fetal rejection in successive pregnancies<sup>77</sup>. Fetal-antigen-specific  $T_{reg}$  cells, generated by conversion of Foxp3<sup>-</sup>CD4<sup>+</sup> T cells, expand >100 fold through parturition, remain at enriched numbers after delivery, and re-accumulate with accelerated kinetics during subsequent pregnancies. The precise localization of memory  $T_{reg}$  cells in feto-maternal non–lymphoid tissues remains unknown. Although in the case of pregnancy the target tissue is present only transiently, and memory  $T_{reg}$  cells seem to be maintained in lymphoid organs, their fetal-antigen specificity and existing evidence of the presence of  $T_{reg}$  cells in placenta and utero<sup>78–80</sup> suggest that memory  $T_{reg}$  cells might migrate to these tissues in recurrent pregnancies to exert their protective role.

Thus, evidence suggests that there is a close association between tissue-resident  $T_{reg}$  cells and memory  $T_{reg}$  cells. The generation of regulatory memory is emerging as an important factor to control the accelerated and enhanced recall responses to secondary exposure to antigens. The localization of memory  $T_{reg}$  cells in the target non-lymphoid tissues seems to be key for their function and, in some cases, for the maintenance of the memory  $T_{reg}$  cell pool during the remission period.

#### Perspectives

Tissue-resident  $T_{reg}$  cells is a relatively new area of  $T_{reg}$  cell biology. It is not surprising, then, that some key questions remain unanswered, or even unaddressed. We would like to highlight three questions that particularly intrigue us:

First, how widespread is the phenomenon? Given the distinct  $T_{reg}$  cell compartments already found to be associated with adipose tissue, muscle, the colon, etc, it is theoretically possible that there is a specialized type of  $T_{reg}$  cell dedicated to maintaining homeostasis within each tissue. Or it might be that evolution has provided dedicated  $T_{reg}$  cell types only for those tissues particularly prone to insult (for example, the skin and muscle) or in especially close communication with the environment (such as the skin and the gut). An alternative possibility is that there are a limited number of  $T_{reg}$  cell types and each tissue  $T_{reg}$ compartment represents a different optimum mix of the various subsets – that is what we are seeing are really only unique "averages." A broader survey of tissue  $T_{reg}$  cells, especially in single-cell-analysis mode, should yield critical information bearing on this question. It may also prove informative to compare  $T_{reg}$  compartments from tumors and their matched normal tissues.

Second, where do tissue-resident  $T_{reg}$  cells come from? So far, it seems clear that, except for the case of the gut and placental compartment, tissue  $T_{reg}$  cells do not emanate from conversion of conventional Foxp3<sup>-</sup>CD4<sup>+</sup> T cells. It is perhaps simplest to envisage that a precursor cell, already Foxp3<sup>+</sup> but not committed to a particular  $T_{reg}$  cell sub-phenotype, gets retained within a particular tissue, likely because its TCR reacts to an antigen therein

and, in response to tissue-specific cues, takes on a tissue-specific sub-phenotype. But it is difficult to rule out the possibility that small numbers of precursors precommitted to a particular  $T_{reg}$  cell sub-phenotype are generated in the thymus and monitor their corresponding peripheral tissues, either as residents or as members of the circulating T cell pool. Such a scenario is reminiscent of the recent finding that populations of tissue-resident macrophages are generated before birth and are important elements in enforcing tissue homeostasis<sup>52</sup>. Perhaps these macrophages need  $T_{reg}$  cell monitors to keep them in line. Identifying the origin of tissue-resident  $T_{reg}$  cells would be aided by generating appropriate inducible lineage-tracer mice, for example in PPAR- $\gamma$  reporters for VAT  $T_{reg}$  cells.

Third, what is the advantage of having unique populations of tissue-resident  $T_{reg}$  cells? It makes perfect evolutionary sense to optimize  $T_{reg}$  compartments for both effective survival and appropriate effector activities. This is especially true for  $T_{reg}$  cells that will reside in a given tissue long-term, either as tissue-resident sentries or in response to chronic insult. For example, adipose tissue is an environment not generally hospitable for lymphocytes – VAT  $T_{reg}$  cells co-opting of PPAR- $\gamma$  provides these cells with properties (such as expression of the lipid transport, CD36) that promote survival in adipose tissue. Similarly, high-level expression of Areg by  $T_{reg}$  cells in regenerating tissues (for example muscle or intestine) arms them with the capacity to favorably impact their local environment. We wonder how common it is for tissue-resident  $T_{reg}$  cells to co-opt transcriptional programs characteristic of tissular cells (like VAT  $T_{reg}$  use of PPAR- $\gamma$ , the "master-regulator" of adipocytes), and whether there might be something special about the chromatin organization or epigenetic make-up of  $T_{reg}$  cells that favors such adaptability.

Interesting realms to explore!

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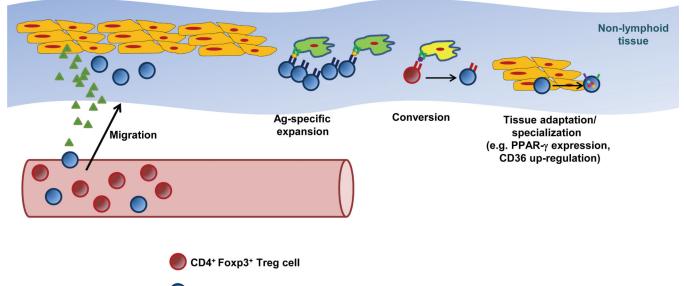
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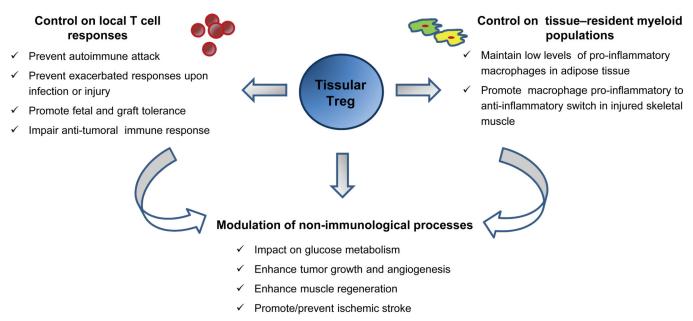
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CD4<sup>+</sup>Foxp3<sup>+</sup> T conventional cell

# Figure 1. Mechanisms involved in the accumulation of $T_{\mbox{reg}}$ cells in non-lymphoid tissues

Several mechanisms (alone or in combination) have been shown to increase the frequency and numbers of tissue-resident  $T_{reg}$  cells: 1) Migration and retention of circulating  $T_{regs}$ , promoted by the expression of specific chemotactic molecules by parenchymal or stromal cells, and the expression of the specific receptors on  $T_{reg}$  cells; 2) Expression of tissuespecific antigens that induce expansion of particular  $T_{reg}$  clones; 3) Antigen-induced conversion of CD4<sup>+</sup>Foxp3<sup>-</sup> T conventional cells into p $T_{regs}$ ; and 4) Acquisition of a tissuespecific phenotype that allows adaptation or specialization in the new environment, promoting survival and enhancing  $T_{reg}$  functions.



✓ Modulate liver cholestasis and fibrosis

#### Figure 2. Functions of tissue-resident T<sub>reg</sub> cells

The functions of tissue resident  $T_{reg}$  cells can be divided in three main groups (representative examples of each group are listed in the figure): control on local T-cell responses, control on tissue-resident myeloid populations and modulation of nonimmunological processes. The latter may occur by direct interaction between  $T_{reg}$  cells and their non-immune targets, or indirectly through the regulation of other tissue-resident leukocytes which in turn could affect those targets.