

Preparation, characterization, and in ovo vaccination of dextran-spermine nanoparticle DNA vaccine coexpressing the fusion and hemagglutinin genes against Newcastle disease

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Abstract: Plasmid DNA (pDNA)-based vaccines have emerged as effective subunit vaccines against viral and bacterial pathogens. In this study, a DNA vaccine, namely plasmid internal ribosome entry site-HN/F, was applied in ovo against Newcastle disease (ND). Vaccination was carried out using the DNA vaccine alone or as a mixture of the pDNA and dextran-spermine (D-SPM), a nanoparticle used for pDNA delivery. The results showed that in ovo vaccination with 40 µg pDNA/egg alone induced high levels of antibody titer ($P < 0.05$) in specific pathogen-free (SPF) chickens at 3 and 4 weeks postvaccination compared to 2 weeks postvaccination. Hemagglutination inhibition (HI) titer was not significantly different between groups injected with 40 µg pDNA + 64 µg D-SPM and 40 µg pDNA at 4 weeks postvaccination ($P > 0.05$). Higher antibody titer was observed in the group immunized with 40 µg pDNA/egg at 4 weeks postvaccination. The findings also showed that vaccination with 40 µg pDNA/egg alone was able to confer protection against Newcastle disease virus strain NDIBS002 in two out of seven SPF chickens. Although the chickens produced antibody titers 3 weeks after in ovo vaccination, it was not sufficient to provide complete protection to the chickens from lethal viral challenge. In addition, vaccination with pDNA/D-SPM complex did not induce high antibody titer when compared with naked pDNA. Therefore, it was concluded that DNA vaccination with plasmid internal ribosome entry site-HN/F can be suitable for in ovo application against ND, whereas D-SPM is not recommended for in ovo gene delivery.

Keywords: Newcastle disease, DNA vaccine, in ovo vaccination, Newcastle disease virus, dextran-spermine nanoparticle, hemagglutinin and fusion

Introduction

The administration of DNA vaccines is an advanced approach to achieve specific immune system activation.^{1,2} The advantages of DNA vaccines such as the capacity to be delivered in ovo, the potential to overcome maternal immunity, non-requirement for cold chain transport, and the ability to mix with immunological factors to reach high efficiency have made them more attractive for use in the poultry industry. It has been demonstrated that recombinant DNA transfection into avian embryos is a problematic approach.² Hence, in adult animals, DNA vaccines are most commonly delivered via intradermal or intramuscular injections; a few studies have focused on embryonic administration.³⁻⁵ Administration of plasmid DNA (pDNA) containing the reporter beta galactosidase gene into the avian embryo body produced the protein, although it can increase the mortality rate.³ Avian embryo is competent immunologically after the 17th day of embryonic stages.⁶⁻⁸

Therefore, it was hypothesized that a DNA vaccine could be administered into the amniotic sac of avian embryos to induce immunity at this stage.

Few factors such as cell membrane, in vivo stability, endosome escape, nuclear entry, and cytosolic transport act as barriers that inhibit the delivery of DNA into the host cells. DNA vaccines can be efficiently delivered using cationic lipids and/or cationic polymers. A study showed that DNA when covered by these particles resulted in a complex with positive charge that enhanced the endocytosis process through which the DNA entered the cell. Cationic polymers created more stable complexes than cationic lipids; thus, they provide more protection to the vaccines through cellular trafficking.⁷⁻¹⁰ Cationic polysaccharides are biodegradable and nontoxic and can be easily modified to exhibit different physicochemical properties.¹¹ Usage of biodegradable polysaccharide carriers are especially appropriate for biological applications such as transfection, because they are soluble in water and freely transported to the cells.¹² Polysaccharide dextran is a complex, diverged glucan with a chain length from 3 to 2,000 kDa and is prepared by using many glucose molecules. Recently, dextran-spermine (D-SPM) has been developed as a novel cationic polymer for gene delivery. A recent study has suggested a new biodegradable polycation made up of natural components, which is efficient in transfecting tissues and cells in vitro and in vivo.¹³ This polycation has shown an effective transfection for a range of cell lines and genes in serum-poor or serum-free medium.¹⁴ D-SPM polycations have been found to be active in transfecting a wide range of cell lines in vitro.¹⁵ Therefore, the objectives of the present study were: 1) to determine that a DNA plasmid coexpressing the fusion (F) and hemagglutinin (HN) proteins of Newcastle disease virus (NDV) can be a potential vehicle for successful vaccination in ovo against ND; and 2) to test whether the cationic polysaccharide D-SPM can be used to enhance the efficacy of in ovo delivery of the DNA vaccine against ND.

Materials and methods

Construction of DNA vaccine

The plasmid internal ribosome entry site (pIRES) coexpression vector containing multiple cloning sites (MCS) such as MCS A and MCS B with a size of 6.1 kb (Clontech, Palo Alto, CA, USA) was used to construct a DNA plasmid encoding the *F* and *HN* genes of NDV AF2240 strain. The full lengths of the *F* and *HN* genes were amplified using reverse transcription polymerase chain reaction (RT-PCR) using a pair of *F*-specific primers and *HN*-specific primers with sequence for the restriction enzymes *Nhe*I and *Mlu*I, and *Sal*I and *Not*I,

respectively, introduced into the forward and reversed primers regard to conserving frameshift of sequences:

F-For: 5'AATTCGGCTAGCACCATGGGCTCCAAGTCTT3'

F-Rev: 5'GGCACGCGTCTAGCTGCCAGAATTGACGCGCA3'

HN-For: 5'CAGTCGACGTCATGGGGAACCAGGCCTCACAA3'

HN-Rev: 5'GAGCGGCCGCCCTATTGACAAGAATTCAGGCCAT3'.

The RT-PCR products of *F* and *HN* genes with a length of 1,722 and 1,950 base pairs (bp), respectively, were amplified and cloned into the *Nhe*I and *Mlu*I (inserted into MCS A) and *Sal*I and *Not*I (inserted into MCS B) of the pIRES vector to the construct pIRES-HN/F DNA plasmid. The construct was purified using an endotoxin-free plasmid purification kit (Qiagen NV, Venlo, the Netherlands) following verification of the orientation and nucleotide sequence of the inserts by double-stranded sequencing. Expression of both *HN* and *F* genes together in a pIRES-HN/F plasmid was confirmed using both indirect immunofluorescence and Western blotting techniques.

Preparation of pDNA/D-SPM complex

D-SPM was prepared as previously described by Abedini et al.¹⁶ pDNA/D-SPM nanoparticles were prepared by mixing pDNA and D-SPM at various concentrations in aqueous solution. A volume of 100 μ L of DNase-free water was taken in five separate tubes containing 8, 12, 16, 18, and 20 μ g of D-SPM and placed in a sonicator (Branson, Danbury, CT, USA) for 30 minutes. To each test tube, 10 μ g of pDNA was added and the solutions were pipetted up and down three to five times and then placed in an orbital mixer for 10 minutes at room temperature. The solutions of pDNA and D-SPM were mixed and gently agitated for 30 minutes to form self-assembled pDNA/D-SPM complexes.

Characterization of the self-assembled pDNA

The reliability of covering pDNA by D-SPM was tested on 1% agarose gel. The formation of DNA complexes was also confirmed by transmission electron microscopy (TEM). Fifty microliters of the sample was kept on a copper grid for 5 minutes; excess solution was blotted off using filter paper and air-dried for 5 minutes before viewing by TEM.

Particle size assayed by NANOPHOX

Fresh pDNA/D-SPM complex was prepared with a fixed concentration of pDNA and a varying concentration of

D-SPM and then the mean particle size was analyzed by a particle size analyzer (NANOPHOX, Sympatec, Germany). The size of all the dispersed samples in nuclease-free water was determined at 25°C in triplicate. Photon cross correlation sensor present in this analyzer allowed for the simultaneous determination of particle size and stability in a range, approximately, of 1 nm to several micrometers in opaque suspensions and emulsions.¹⁷

Zeta potential and size measurement

Zeta potential is commonly used to characterize the surface charge property of nanoparticles.¹⁸ Size and zeta potential of nanoparticles were determined using a laser particle size analyzer (Malvern, Zeta, Worcestershire, UK). A tenfold dilution of the sample in pure water in a total volume of 1 mL was subjected to a particle size analyzer at 25°C. The measurement was based on the electrophoretic mobility ($\mu\text{m/s}$) of the particles which was converted to zeta potential by inbuilt software based on the Helmholtz–Smoluchowski equation.

In ovo vaccination of SPF embryo

Eighteen-day-old embryonated specific pathogen-free (SPF) eggs were randomly divided into four groups (15 eggs per group). The eggs were inoculated with 40 μg pIRES-HN/F, 20 μg pIRES-HN/F + 32 μg D-SPM, and 40 μg pIRES-HN/F + D-SPM complex or the empty plasmid. The egg shells were disinfected and the vaccines were injected via the amnio-allantoic cavity through a small hole made at the air sacs with 21-gauge needle followed by sealing the holes and continued the incubation of the eggs. After hatching, the chicks had free access to feed and water. Bleeding was carried out at 2, 3, and 4 weeks post-immunization; total serum antibody titers were measured by an indirect enzyme-linked immunosorbent assay (ELISA) Kit (IDEXX, Westbrook, ME, USA) and hemagglutination inhibition (HI) test as previously described.¹⁹ The chickens were treated and handled according to the protocols approved by the Institutional Animal Care and Use Committee (IACUC) of the Faculty of Veterinary Medicine (AUP no. 12R1541), Universiti Putra Malaysia.

Virus challenge

After 4 weeks of in ovo vaccination, the vaccinated chickens in all groups were challenged with intranasal administration of 10^5 mean egg infection dose (EID_{50}) of either NDV strain AF2240 or NDV strain IBS002 in a volume of 0.1 mL viral suspensions per chick. The chickens were monitored daily after challenge for 10 days and the numbers of dead chickens were recorded.

Statistical analysis

Data were analyzed by Student's *t*-test and statistical significance was set at $P < 0.05$. The results were expressed as mean \pm standard error of the mean. All the analyses were carried out using Minitab 15 Statistical Software (Minitab Inc., University Park, PA, USA) and Microsoft Excel 2010 (Microsoft Corporation, Redmond, WA, USA).

Results

Agarose gel electrophoresis of pDNA/D-SPM complex at different ratios

The DNA vaccine pIRES-HN/F was constructed to express HN and F genes of NDV strain AF2240. The pDNA/D-SPM nanoparticles were prepared by mixing pIRES-HN/F and D-SPM at various concentrations. The formation of nanoparticles was tested by performing electrophoresis of the samples on 1% agarose gel, where the absence of DNA band on the gel indicates the initial formation of a stable complex (Figure 1).

Particle size analysis

The formation of nanoparticles and particle sizes of the pDNA/D-SPM complexes were also checked by TEM. Good agglomeration and a particle size of ~70–130 nm were obtained when a mixture of 10 μg of pDNA and 16 μg of D-SPM was used to prepare the nanoparticles (Figure 2). In addition, the particle size of the pDNA/dextran complexes was determined by nanophox equipment (NANOPHOX). As shown in Figure 3, a particle size of 10–100 nm was obtained when a mixture of 10 μg of pDNA and 16 μg of D-SPM was used.

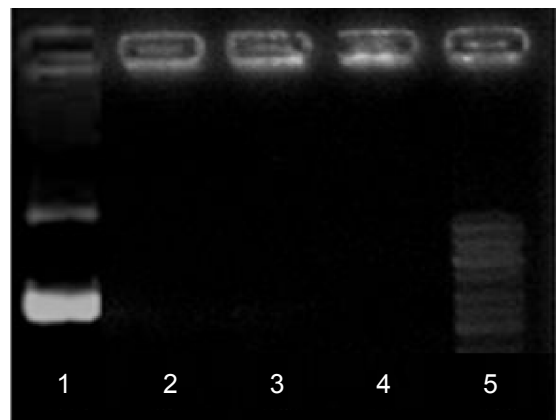


Figure 1 Agarose gel electrophoresis of the pDNA/D-SPM complex at different ratios.

Notes: Line 1, free pDNA plasmid; lines 2–4, pDNA/D-SPM complexes at ratios 1:2, 1:6, and 1:18 μg , respectively; line 5, GeneRuler GeneRuler™ DNA ladder 1 kb (Fermentas Canada, Burlington, ON, Canada).

Abbreviations: pDNA, plasmid DNA; D-SPM, dextran-spermine.

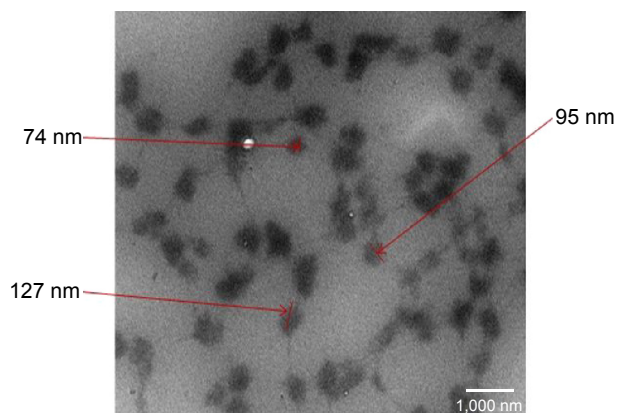


Figure 2 Particle size analysis of pDNA/D-SPM complexes by transmission electron microscopy (TEM).

Notes: Nanoparticle sizes between 74 and 127 nm were obtained from the pDNA/dextran complexes. Scale bar =1,000 nm. These arrows point to the size of pDNA/D-SPM complexes.

Abbreviations: pDNA, plasmid DNA; D-SPM, dextran-spermine.

Size distribution of pDNA with D-SPM nanoparticle complexes

In addition, the size distribution of the pDNA/D-SPM complexes were determined with a Zetasizer 3000 HSA (Malvern); when a mixture of 10 µg of pDNA and 16 µg of D-SPM nanoparticles was used, DNA nanoparticles of 75 nm size were obtained, as shown in Figure 4A.

Zeta potential report of pDNA with D-SPM nanoparticle complexes

A zeta potential in the range of 25–30 mV was used for the preparation of pDNA/D-SPM complexes in a ratio of 10 µg pDNA to 16 µg of D-SPM (Figure 4B).

As expected, the zeta potential of pDNA alone was found to be in a range of –55 to –60 mV (Figure 5A). The backbone of DNA is highly charged and each bp carries two unit negative charges and each phosphate group carries one unit negative charge. The surface charge of nanoparticles with a zeta potential around ±30 mV has been revealed to be stable in suspension, and this surface charge stops aggregation of the particles. For D-SPM alone, the surface charge was determined to be in a range of 30–35 mV (Figure 5B).

In ovo immunization of SPF chickens

In ovo immunization was carried out using naked DNA plasmid and pDNA + D-SPM complexes. After hatching, an immune response was assessed at 2, 3, and 4 weeks postvaccination.

NDV antibody response by ELISA

The antibody titer was evaluated at a serum dilution of 1:500 using commercial ELISA kit (IDEXX). The positive control mean was 0.165 and the negative control mean was 0.046.

ELISA results were presented as an S/P ratio. An S/P ratio is determined by dividing the absorbance units (ELISA score) of the sample in question by the ELISA score of a positive control analyzed alongside the test sample(s). An S/P ratio is directly correlated with the amount of target antigen present in the sample.

As shown in Table 1 and Figure 6, no anti-NDV immune response was detected in the controls vaccinated with the empty plasmid. Antibody levels remained low 2 weeks after

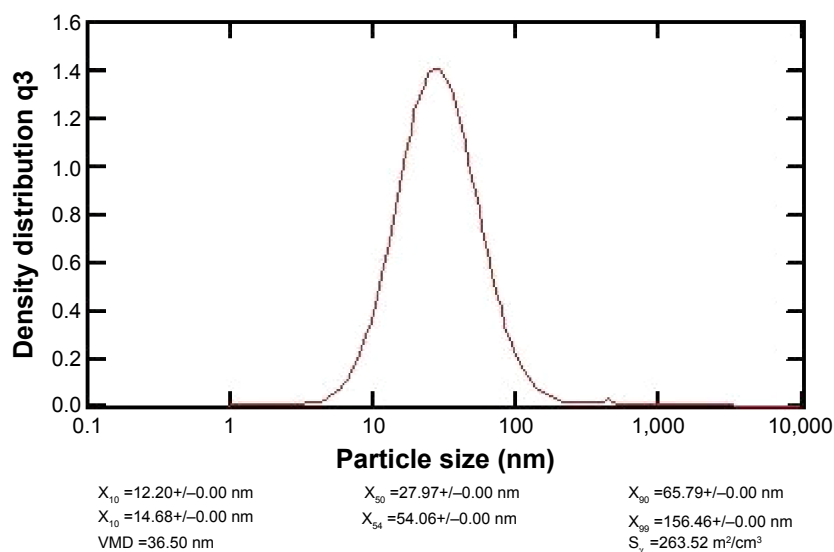


Figure 3 NANOPHOX particle size analysis of the pDNA/D-SPM complexes.

Abbreviations: pDNA, plasmid DNA; D-SPM, dextran-spermine; VMD, volume mean diameter.

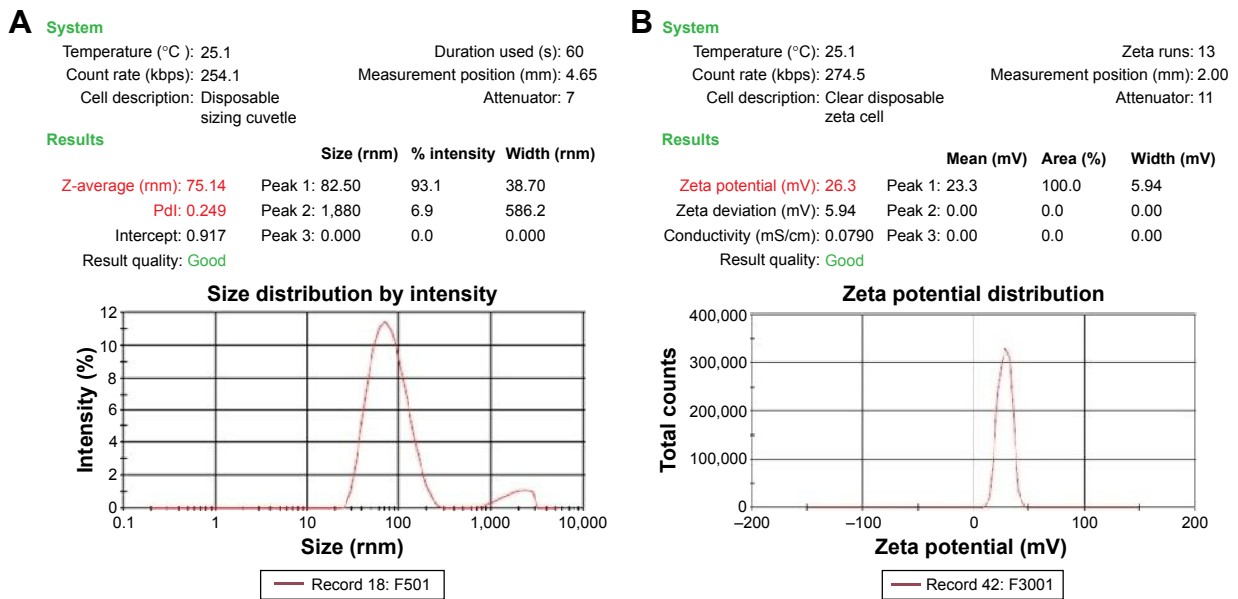


Figure 4 Zeta potential of pDNA and D-SPM.

Notes: (A) Size distribution of pDNA with D-SPM nanoparticle complexes by Zetasizer 3000 HSA. Plasmid DNA and D-SPM nanoparticles when used in a ratio of 10:16 produced a complex of size 75 nm. (B) Zeta potential report of pDNA/D-SPM complexes. The range of zeta potential of the pDNA/D-SPM complexes at a ratio of 10 of µg plasmid DNA to 14 µg or 16 µg of D-SPM was 25–30 mV.

Abbreviations: pDNA, plasmid DNA; D-SPM, dextran-spermine.

in ovo immunization in all the vaccinated groups. A higher antibody titer was observed in the group immunized with 40 µg pDNA at 4 weeks postvaccination; however, it was not significant compared to the chickens immunized with

40 µg pDNA +64 µg dextran complex ($P>0.05$). The lowest antibody titer was obtained in groups vaccinated with 20 µg pDNA +32 µg dextran complex compared to other vaccinated groups ($P<0.05$).

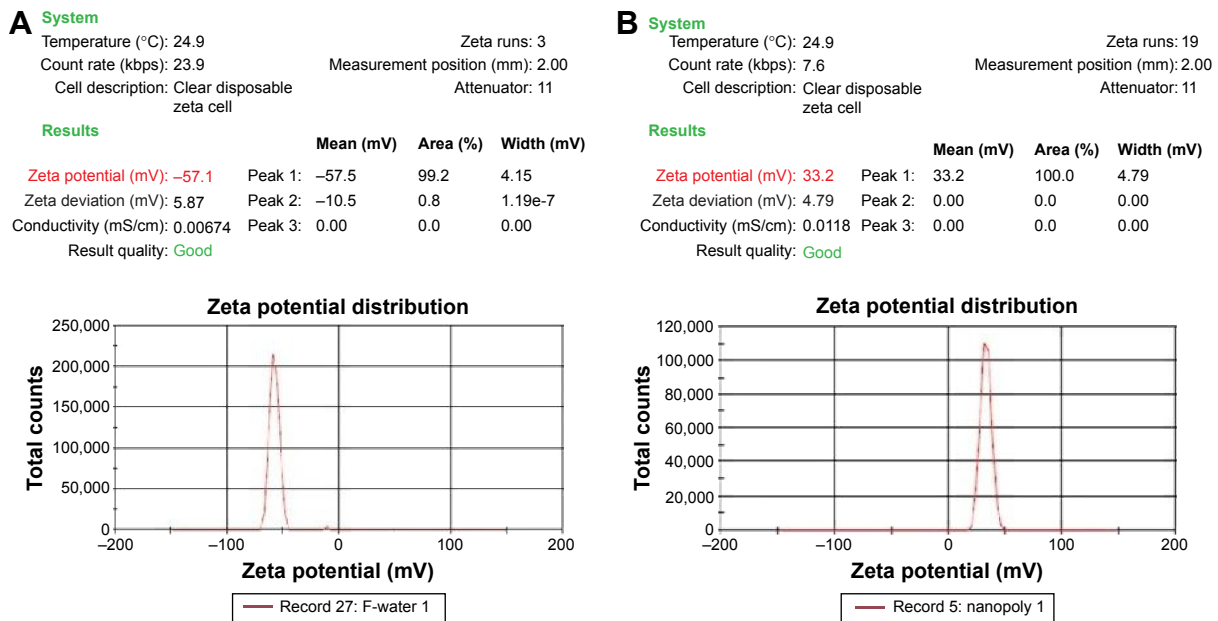


Figure 5 Zeta potential of pDNA and D-SPM.

Notes: (A) Zeta potential report of pDNA alone. The zeta potential of backbone plasmid DNA ranged from -55 to -60 mV. (B) Zeta potential report of D-SPM alone. The surface charge of D-SPM alone was determined to be in a range of 30–35 mV.

Abbreviations: pDNA, plasmid DNA; D-SPM, dextran-spermine.

Table 1 Analysis of S/P ratios of ELISA results

Group	S/P ratio of ELISA results		
	2 weeks	3 weeks	4 weeks
pIRES-HN/F (40 µg pDNA)	0.038±0.00	0.248±0.02	0.272±0.02
pIRES-HN/F (40 µg pDNA + D-SPM)	0.035±0.00	0.209±0.01	0.251±0.02
pIRES-HN/F (20 µg pDNA + D-SPM)	0.022±0.04	0.063±0.00	0.085±0.02
pIRES	0.00	0.00	0.00

Note: S/P ratio = Sample mean (mean of optical absorbance) – negative control mean/positive control mean – negative control mean.

Abbreviations: ELISA, enzyme-linked immunosorbent assay; pIRES, plasmid internal ribosome entry site; HN, hemagglutinin; F, fusion; pDNA, plasmid DNA; D-SPM, dextran-spermine.

NDV-HN antibody titers by HI test

The HI test was carried out on serum samples with four hemagglutination (HA) units of the mucosal LaSota vaccine. A twofold dilution of each serum was made and titers were expressed as log 2 values of the highest dilution of serum inhibiting hemagglutination. The mean HI titers are shown in Table 2. HI titer was not significantly different between the groups injected with 40 µg pDNA +64 µg D-SPM (3.5±0.51) and 40 µg pDNA (3.43±0.22). However, the mean HI titer was significantly lower in the groups vaccinated with 20 µg pDNA +32 µg D-SPM (2.73±0.04) compared to the other vaccinated groups.

Virus challenge

Four weeks post-immunization, chickens were challenged with NDV AF2240 and IBS002 strains. In the AF2240-challenged chickens, regular death was first detected 3 days post-challenge and the last death was recorded on the fifth day for all the control chickens. Clinical signs such as the wing or leg

paralysis and weakness were observed on the third day post-challenge and the chickens began to die. After 1 week post-challenge, all the chickens in all the vaccinated groups died (Table 3). However, chickens immunized with pIRES-HN/F (40 µg pDNA/egg) and challenged with IBS002 were more active and showed no signs of disease even after 3 days post-challenge and finally two chickens survived. Furthermore, one chicken in each pIRES-HN/F group vaccinated with 40 µg pDNA + D-SPM/egg or 20 µg pDNA + D-SPM/egg died 8 days post-challenge (Table 4).

In all the challenged chickens, depression and loss of appetite were observed. Severe manifestations such as edema, hemorrhages, and necrosis of respiratory tissue were found in dead chickens of the control group, whereas no significant gross lesions were found in the vaccinated group.

However, pIRES-HN/F-immunized chickens showed no signs of the disease even after 3 days of the challenge and finally two chickens survived (Table 4).

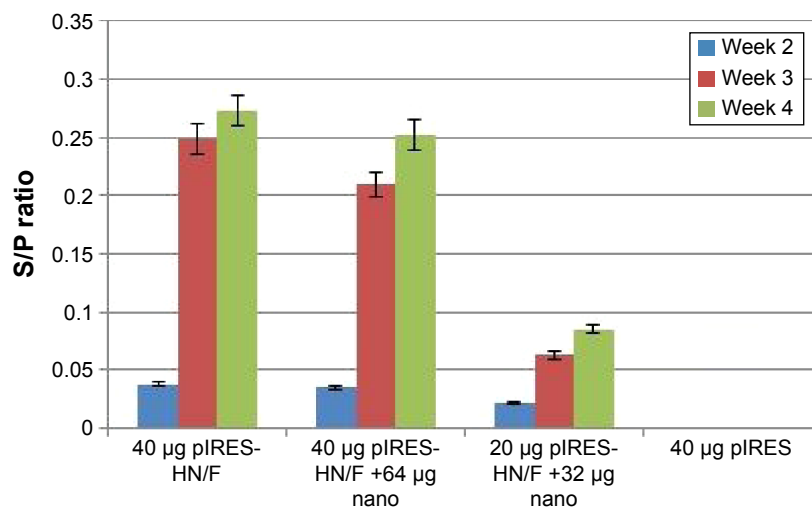


Figure 6 Mean S/P ratio of ELISA result after in ovo immunization with pIRES-HN/F or pIRES-HN/F-D-SPM complexes.

Notes: The error bars show standard deviation of means. S/P ratio = Sample mean (mean of optical absorbance) – negative control mean/positive control mean – negative control mean.

Abbreviations: ELISA, enzyme-linked immunosorbent assay; pIRES, plasmid internal ribosome entry site; HN, hemagglutinin; F, fusion; D-SPM, dextran-spermine; Nano, nanoparticle.

Table 2 Mean antibody titers determined by HI test (\log_2) of SPF embryonated eggs after immunization with different vaccination programs (geometric mean \pm SD)

Group	Mean HI titer in different weeks of postvaccination (\log_2)		
	2 weeks	3 weeks	4 weeks
pIRES-HN/F (40 μ g pDNA)	1.93 \pm 0.47	2.14 \pm 0.53	3.43 \pm 0.22
pIRES-HN/F (40 μ g pDNA + D-SPM)	1.36 \pm 0.49	1.71 \pm 0.61	3.5 \pm 0.51
pIRES-HN/F (20 μ g pDNA + D-SPM)	1.64 \pm 0.50	1.64 \pm 0.5	2.73 \pm 0.04
pIRES	1.1 \pm 0.31	1.1 \pm 0.31	1.1 \pm 0.31

Abbreviations: HI, hemagglutination inhibition; SPF, specific pathogen-free; pIRES, plasmid internal ribosome entry site; HN, hemagglutinin; F, fusion; pDNA, plasmid DNA; D-SPM, dextran-spermine.

Discussion

It is thought that F and HN glycoproteins interact with one another to promote F activity and, therefore, immunization with a DNA plasmid encoding both F and HN genes would be more effective than either component alone.^{20–22} Therefore, in this research, the DNA plasmid pIRES-HN/F coexpressing F and HN proteins was constructed and used for in ovo immunization against NDV. Immunization with 40 μ g of pIRES-HN/F per egg induced a significant level of antibody titer; however, it was not effective in protecting against the virus challenge. This deficiency may be correlated to the relatively low transfection efficiency of the pDNA into the cells. Enzymes and serum proteins such as endonucleases may degrade the plasmid, and thereby reduce the available plasmids in the host cells that express the antigen.

In the present study, 1×10^5 EID₅₀ of the virulent NDV was used by the nasal route for challenge trials.

This dose was considered to be high to test the efficacy of the DNA vaccines. Therefore, the trial infection can be applied on statistically significant number (≥ 10) of adult and young chickens (contact challenge) with a viral standard dose as 10^5 EID₅₀.²³ Thus, virus challenges via contact route rather than nasal route may provide more protection against virulent viruses.

For most DNA vaccines, direct inoculation has been shown not to be effective enough due to electrostatic repulsion between the negatively charged DNA and positively charged cell membrane. Enzymatic degradation is another negative point that subsequently inhibits effective cellular

uptake. DNA hydrolyzing activity by DNase I is also observed in blood plasma. Hence, the amount of foreign DNA that crosses the cytoplasm varies and depends on the DNase I activity.²⁴

Therefore, to deliver DNA molecules into cells, a studies have used D-SPM, which is a polycationic polysaccharide.^{25,26} D-SPM can bind to DNA through a combination of electrostatic attraction and intercalation. In this study, D-SPM was used as a nanoparticle for in ovo delivery of a DNA vaccine. The results showed that pDNA/D-SPM complex can induce antibody titer in a low level compared to pDNA alone. The results of in ovo immunization are summarized in Table 4; the table shows ($P > 0.05$) low S/P ratio obtained by ELISA at 4 weeks post-immunization of groups injected with pDNA + D-SPM compared to groups injected with pDNA alone, suggesting that D-SPM may not be effective in delivering pDNA to the cell by an in ovo method, thus resulting in low pDNA transfection efficiency.

This deficiency in vaccine efficacy may be due to the number of positive charges and the charge spacing of polyamine molecules in pDNA/D-SPM complex within the amniotic liquid that surrounds the chicken embryo. Studies on λ -DNA concentration by spermine homologues showed that the mid-point concentration of polyamine homologues for contracting DNA and the hydrodynamic ranges of the condensates are dependent on the polyamine structure.²⁶ On the other hand, the most abundant protein in white egg is ovalbumin, representing 54% of the total protein content.²⁷ It has been shown that at pH levels between 9.6 and 6.6, there is transference

Table 3 Results of virus challenge with the NDV AF2240 strain after in ovo vaccination

Vaccinated groups	Sample numbers	2 days	3 days	4 days	5 days	6 days	Survived
pIRES-HN/F (40 μ g pDNA)	7	0	2	3	2	0	0
pIRES-HN/F (40 μ g pDNA + D-SPM)	7	0	2	3	1	1	0
pIRES-HN/F (20 μ g pDNA + D-SPM)	6	0	2	3	1	0	0
pIRES	10	0	3	5	2	0	0

Note: Sample numbers are the numbers of hatched chicks after in ovo vaccination.

Abbreviations: NDV, Newcastle disease virus; pIRES, plasmid internal ribosome entry site; HN, hemagglutinin; F, fusion; pDNA, plasmid DNA; D-SPM, dextran-spermine.

Table 4 Number of dead chickens at days post-challenge with IBS002 strain

Vaccinated groups	Sample numbers	2 days	3 days	4 days	5 days	6 days	8 days	Survived
pIRES-HN/F (40 µg pDNA)	7	0	0	2	2	1	0	2
pIRES-HN/F (40 µg pDNA + D-SPM)	7	0	0	2	3	1	1	0
pIRES-HN/F (20 µg pDNA + D-SPM)	6	0	0	2	1	1	1	0
pIRES	10	0	0	4	4	2	–	0

Note: Sample numbers are the numbers of hatched chicks after in ovo vaccination.

Abbreviations: pIRES, plasmid internal ribosome entry site; HN, hemagglutinin; F, fusion; pDNA, plasmid DNA; D-SPM, dextran-spermine.

of albumin to dextran when the two components are mixed, the amount of albumin lost being nearly equal to the gain in the new component, albumin plus dextran.²⁸ Polysaccharide such as dextran shows high viscosity, which helps in the stabilization of emulsion with ovalbumin.²⁹

Therefore, in the present study, low antibody titer induced in the chickens immunized with the pDNA nanoparticle can be related to the low efficacy of D-SPM in delivering pDNA to the embryo cell. According to the manufacturer's instructions, S/P values lower than 0.5 are considered negative. Thus, in ovo vaccination resulted in a slight increase in antibody titers, which are too low to be considered as seroconversion.

The basis of immunity against NDV is circulating antibodies and cell-mediated immunity. In this study, the stimulation of B cells (antibodies) by the constructed plasmid was not strong enough to elicit T cell-independent induction of IgM antibodies. In summary, protection against lethal challenge could not be expected when DNA vaccines were delivered in ovo.

Conclusion

This study revealed that although in ovo vaccination did not confer significant protection from the subsequent NDV challenge, a significant increase in the production of antibodies in the chicks vaccinated with pIRES-HN/F was achieved. This is the first study to report on the testing of D-SPM for the delivery of pDNA in ovo. This study described a set of experiments investigating a new DNA vaccine containing NDV F and HN genes and D-SPM to support DNA delivery for in ovo vaccination to protect chickens against the lethal Newcastle disease. Further investigation using a different virus titer and contact route for challenge to test the efficacy of different immunizations will help improve the nanoparticle DNA vaccine. Biotechnological studies today have mainly focused on safe and efficient nano-delivery of pDNA into chicken embryo cells.

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Author contributions

AI, SDH, MHB, and ARO conceived and designed the study. MF, HM, PM, and MEZ carried out the experiments. MF, and HM performed the molecular genetic studies, immunoassays, sequence alignment, statistical analysis. MF, MEZ and PM performed the vaccination experiments. MF, HM, MEZ drafted the manuscript, all authors have contributed to writing and revising the manuscript. MEZ, MF, TJW, and AI revised the manuscript during all stages of submission. All authors read and approved the final paper.

Disclosure

The authors report no conflicts of interest in this work.

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