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REVIEW

Mechanisms of intrahepatic triglyceride accumulation

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Abstract

Hepatic steatosis defined as lipid accumulation in

hepatocytes is very frequently found in adults and obese adolescents in the Western World. Etiologically, obesity and associated insulin resistance or excess alcohol intake are the most frequent causes of hepatic steatosis. However, steatosis also often occurs with chronic hepatitis C virus (HCV) infection and is also found in rare but potentially life-threatening liver diseases of pregnancy. Clinical significance and outcome of hepatic triglyceride accumulation are highly dependent on etiology and histological pattern of steatosis. This review summarizes current concepts of pathophysiology of common causes of hepatic steatosis, including non-alcoholic fatty liver disease (NAFLD), alcoholic fatty liver disease, chronic HCV infections, drug-induced forms of hepatic steatosis, and acute fatty liver of pregnancy. Regarding the pathophysiology of NAFLD, this work focuses on the close correlation between insulin resistance and hepatic triglyceride accumulation, highlighting the potential harmful effects of systemic insulin resistance on hepatic metabolism of fatty acids on the one side and the role of lipid intermediates on insulin signalling on the other side. Current studies on lipid droplet morphogenesis have identified novel candidate proteins and enzymes in NAFLD.

Key words: Steatosis; Lipid accumulation; Fatty acids; Insulin resistance; Triglycerides

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Core tip: Fatty liver disease is a highly prevalent condition in the Western World. This article summarizes the most frequent causes and states of hepatic steatosis, including non-alcoholic fatty liver disease (NAFLD), alcoholic fatty liver, drug-induced forms, hepatitis C virus infections, and acute fatty liver of pregnancy. Important pathophysiological and cellular aspects of various forms of fatty liver disease are reviewed as well as the clinically relevant close interaction between hepatic triglyceride accumulation and insulin resistance in NAFLD.

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INTRODUCTION

Hepatic steatosis is defined by the accumulation of triglycerides resulting in more than 5% of hepatocytes containing visible lipid droplets in either a micro- or macrovesicular pattern. Most frequently, steatosis is histologically assessed by using a classification that was originally suggested for grading of nonalcoholic fatty liver disease (NAFLD) by Javor *et* $a^{[1]}$: grade 0: < 5% hepatocytes involved; grade 1: 5%-33% hepatocytes involved; grade 2: > 33%-66% hepatocytes involved, grade 3: > 66% hepatocytes involved. At the cellular level, steatosis is an imbalance of hepatic fatty acid uptake, lipogenesis, β-oxidation, and triglyceride export in very low density lipoprotein (VLDL) particles.

Fatty liver might be caused by several conditions, diseases, or drugs; however, obesity, insulin resistance, and dyslipidemia as well as excess alcohol intake are the most frequent causes of NAFLD or alcoholic fatty liver disease (AFLD). Beyond metabolic or alimentary causes, fatty liver can also be found in patients with hepatitis C virus (HCV) infection and rarely in pregnancy. Other rare causes of fatty liver disease are summarized in Table $1^{[2]}$. This review will focus on the most frequent and important causes of fatty liver disease.

The natural course of fatty liver is strongly dependent on etiology and comorbidities. For example, in NAFLD, 20%-30% of patients with steatosis will develop non-alcoholic steatohepatitis (NASH), which is associated with increased total and cardiovascular mortality^[3]. NASH is complicated by liver cirrhosis in 2% -5% of patients^[2]. Co-occurrence of steatosis and chronic HCV infection has been found to be associated with poorer outcome when compared to patients without steatosis $[4]$.

NAFLD

NAFLD has become the most common cause of elevated liver enzymes in the Western World, affecting 30%-40% of men and 15%-20% of women in the general population^[5] and up to 70% of type 2 diabetics $^{[6]}$. It is now considered the hepatic manifestation of the metabolic syndrome, which comprises a spectrum of metabolic disorders, including overweight or obesity, insulin resistance, dyslipidemia, and hypertension $^[7,8]$. According to estimates, NAFLD</sup> will become the most frequent indication for liver transplantation by 2030^{9} . Beyond excess mortality rate from advanced liver disease, NAFLD is associated

Table 1 Overview on potential causes of fatty liver (adapted from[2,121,122])

with significantly increased cardiovascular risk, mostly due to associated disturbances in glucose and lipid metabolism^[9].

Intracellular lipid accumulation in NAFLD results from an imbalance between hepatic fatty acid uptake, lipid synthesis, lipid oxidation, and export *via* VLDL particles. Several studies have stressed the importance of adipose tissue lipolysis in the development of hepatic steatosis. Increased total fat mass in obesity is associated with elevated whole body lipolysis $[10,11]$ and excess fatty acid uptake into the liver. Further highlighting the role of adipose tissue in triglyceride metabolism, Nye and colleagues $[12]$ found that glycerol 3-phosphate, which is essential for triglyceride synthesis, primarily originates from glyceroneogenesis and only to a lesser extent from glycolysis. Accordingly, mice lacking fatty acid transporter protein 5 (FATP 5), which is the primary fatty acid transporter in the liver, are protected from diet-induced steatosis $[13]$. Underlining the importance of the adipose tissue in development of NAFLD, lipodystrophic patients who lack visceral and peripheral fat accumulation due to leptin deficiency display severe hepatic steatosis and insulin resistance^[14,15]. Similarly, subjects with mutations of perilipin-1 - an inhibitor of adipose triglyceride lipase (ATGL) and stabilizer of lipid droplets in adipose tissue - have also profound hepatic steatosis and insulin resistance^[16].

In addition to increased influx of fatty acids and their esterification, *de novo* lipogenesis (DNL), which from a quantitative point of view only plays a minor role in hepatic lipid accumulation, is also increased in NAFLD. In affected patients, DNL is significantly increased when compared to healthy controls $[17-19]$. Increased levels of ligand-activated transcription factor α (LXR α) and sterol regulating element binding protein 1c (SREBP-1c) have been described as major contributors to increased DNL in NAFLD^[20,21]. Further highlighting the close connection between glucose and fatty acid metabolism, high glucose levels induce pyruvate production *via* stimulation of carbohydrate response element binding protein (ChREBP). Pyruvate

is the principle source of acetyl-CoA in fatty acid synthesis^[22]. Very recently, Perry and colleagues^[23] found that white adipose tissue-derived hepatic acetyl-CoA is the critical determinant of insulininduced suppression of hepatic glucose production. By decreasing oxidation of fatty acids on the one side and inducing DNL on the other side, fructose consumption is a well-known highly effective driver of hepatic steatosis^[24,25].

DNL is closely related to β-oxidation *via* SREBP-1c. In addition to induction of lipogenic enzymes, SREBP-1c activates acetyl-CoA carboxylase 2 (ACC2), resulting in increased levels of malonyl-CoA, which decreases β-oxidation *via* diminishing shuttling of fatty acids into the mitochondrion by inhibiting carnitine palmitoyl transferase 1 (CPT-1). AMP activated protein kinase (AMPK) is also a key regulator of intracellular lipid metabolism. By sensing cellular AMP levels, activation of AMPK stimulates fatty acid β-oxidation and inhibits ATP-consuming processes, such as DNL^[26]. Mitochondrial dysfunction and resulting impaired fatty acid oxidation have been suggested to significantly contribute to the pathogenesis of NAFL $D^{[27,28]}$.

On a cellular level, triglycerides are stored in lipid droplets (LD) that supply fatty acids for oxidation and VLDL secretion. As described in detail in a very recent review by Mashek and colleagues^[29], LDs are formed within the lipid bilayer of the endoplasmic reticulum (ER) and subsequently budded. The latter process requires action of certain fat storage-inducible transmembrane proteins as well as seipin $[30]$. Growth of LDs is dependent on triglyceride and phospholipid synthetic enzymes that are present on the LD surface. LDs are catabolized *via* three different mechanisms: (1) ATGL/ patatin-like phospholipase domain-containing protein 2 (PNPLA2) catalyzes the first step of triglyceride hydrolysis and partitions hydrolyzed fatty acids to β-oxidation[31]. Activity of ATGL is strongly influenced by LD-associated proteins, such as perilipins 2 and 5^{32} . Sirtuin 1 (SIRT1) has been shown to regulate ATGL activity, probably *via* peroxisome proliferator-activated receptor α (PPAR α)^[33]; (2) LDs are degraded by lysosomal lipases, a process that is determined by the small guanosine triphosphatase RAB7 $[34,35]$; and (3) LD catabolism is mediated by carboxylesterase 3 and cell death inducing DFF45-like effector B (CIDEB), which are involved in repacking of cytosolic LDs in the ER leading to synthesis of VLDL particles^[36,37]. Increased expression of LD associated proteins, such as 17 β hydroxysteroid dehydrogenase 13 and perilipins 1, 2, 3, and 5, have been reported in NAFLD^[38-42]. PNPLA3 polymorphisms have been identified as the strongest genetic factor associated with NAFLD^[43].

Apolipoprotein A5 is a protein with extra- and intracellular effects on lipid metabolism $[44]$. In obese subjects undergoing bariatric surgery, improvements of hepatic steatosis were associated with decreased hepatic apoA5 expression. Direct involvement of apolipoprotein A5 in hepatic triglyceride accumulation

has been shown in a hepatic cell culture model^[45]. In adipocytes, apolipoprotein A5 has been found to surround lipid droplets^[46], suggesting a potential role of apolipoprotein A5 in hepatic lipid droplet metabolism.

Stressing the metabolic consequences of steatosis, hepatic triglyceride accumulation is associated with increased gluconeogenesis, decreased glycogen synthesis, and inhibition of insulin signalling $[47,48]$ as well as alterations in hepatokine production $[49]$. Lipid intermediates of long-chain fatty acids (LCAF) have a crucial role in hepatic insulin resistance in NAFLD. In hepatocytes, LCAF are esterified with glycerol-3 phosphate to form monoacylglycerol, diacylglycerol (DAG), and triacylglycerol^[9]. Hepatic DAG content has been positively correlated with inhibition of insulin signalling in mice and humans with NAFLD^[48,50,51]. Mechanistically, DAG induces increased translocation of protein kinase C ε to the plasma membrane, where it binds and inhibits the activity of the intracellular kinase domain of the insulin receptor, resulting in diminished activation of the canonical insulin pathway^[52,53]. As a consequence, activation of glycogen synthesis and inhibition of gluconeogenesis is reduced^[48]. Unesterified lipid intermediates also exert proinflammatory properties by inducing ER stress, which leads to the activation of c-janus N-terminal kinases and nuclear factor kappa B, resulting in decreased phosphorylation of insulin receptor substrate-2 (IRS-2)^[9,54,55]. Accumulation of ceramides in the plasma membrane have also been found to alleviate insulin signalling^[56]. Pathophysiological aspects of NAFLD are summarized in Figure 1.

AFLD

Chronic alcohol exposure is another frequent cause of fatty liver disease. More than 90% of alcoholics have fatty liver disease that will potentially resolve with abstinence from alcohol^[57-59]. Chronic consumption of more than 30 g of pure alcohol was demonstrated to significantly increase the risk of chronic liver disease $[60]$. Susceptibility factors included female sex, obesity, and cigarette smoking as well as coexistence of other hepatic disorders, such as hepatitis B or C virus infection, NAFLD, or hemochromatosis $[61]$.

Alcohol is metabolized *via* two main pathways in the liver, the oxidative pathway, which is mediated by the alcohol dehydrogenase (ADH) and acetaldehyde dehydrogenase (ALDH), and the non-oxidative cytochrome P450 2E1 pathway $^{[62-64]}$.

Ethanol is converted by the cytosolic enzyme ADH to acetaldehyde and further metabolized to acetate by the mitochondrial enzyme ALDH. The reactions are both coupled to the reduction of nicotinamide adenine dinucleotide (NAD) to nicotinamide adenine dinucleotide-hydrogen (NADH)^[65,66].

Excess NADH has deleterious effects on gluconeogenesis and fatty acid synthesis^[67] Furthermore, acetaldehyde was found to induce lipogenesis by

Figure 1 Pathophysiological aspects of hepatic triglyceride accumulation in non-alcoholic fatty liver disease. Both increased uptake of fatty acids due to elevated whole body lipolysis in states of insulin resistance and enhanced fatty acid synthesis are key features of non-alcoholic fatty liver disease. Increased de novo lipogenesis results from enhanced activation of LXR, SREBP-1c, and ChREBP in insulin resistance. SREBP-induced activation of ACC2 leads to accumulation of malonyl-CoA, which in turn inhibits CPT-1 activity, resulting in reduced β-oxidation. In the liver, triglycerides are stored in LDs that are formed within the lipid bilayer of the ER. Stabilization and growth of LDs are dependent on transmembrane proteins, seipin, and triglyceride and phospholipid synthetic enzymes that are located on the LD surface. LDs are catabolized *via* ATGL-dependent hydrolysis and β-oxidation of fatty acids, lysosomal lipases, and carboxylesterase 3- and CIDEB- mediated repacking of cytosolic LDs in the ER, leading to synthesis of VLDL particle. Intermediates of long chain fatty acids (DAG) inhibit insulin signalling further, exacerbating hepatic insulin resistance by exerting proinflammatory effects and reducing activation of the insulin receptor. LXRα: Ligand-activated transcription factor α; SREBP1c: Sterol regulating element binding protein 1c; CPT-1: Carnitine palmitoyl transferase 1; LD: Lipid droplet; ATGL: Adipose triglyceride lipase; CIDEB: Cell death inducing DFFA like effector B; DAG: Diacylglycerol; IRS: Insulin receptor substrate; ER: Endoplasmic reticulum; ChREBP: Carbohydrate response element binding protein; LXR: Ligand-activated transcription factor; DNL: *De novo* lipogenesis; ACC2: Acetyl-CoA carboxylase 2; PNPLA2: Patatin-like phospholipase domain-containing protein 2; ROS: Reactive oxygen species.

increasing SREBP-1c expression^[68]. Increased fatty acid synthesis leads to the accumulation of fatty acid intermediates, such as malonyl-CoA, which suppresses fatty acid transport into the mitochondria and their oxidation by CPT-1[69]. Alcohol mainly *via* its metabolite acetaldehyde further inhibits PPAR α , which is the main transcription factor for genes involved in oxidation, transport, and export of fatty acids^[70-74]. In PPAR α deficient mice, chronic ethanol feeding was associated with progressive intrahepatic triglyceride accumulation due to reduced β-oxidation and alterations in tricarboxylic acid cycle and the electron transfer chain^[75]. In a mouse model, alcohol feeding resulted in dysfunction of ALDH2 and aldehyde accumulation. Pharmacological activation of ALDH2 reversed alcoholic steatosis in these mice, underlining the deleterious effects of accumulating metabolic products of alcohol^[76]. Very recently, Guo et al^[77] reported that the beneficial role of ALDH2 on hepatic steatosis and inflammation might be mediated through regulation of autophagy.

Ceramide-induced inhibition of AMPK and impaired hepatic adiponectin effects might further contribute to increased fatty acid synthesis and reduced fatty acid oxidation^[78-80]. Besides β-oxidation and lipogenesis, VLDL export is also affected by alcohol: triglyceride export is impaired due to suppression of phosphatidylcholine, acetaldehyde-induced inhibition of microtubular assembly, and reduced apolipoprotein synthesis^[81]. The role of perilipin 2, which is a major LD protein, was recently demonstrated in perilipindeficient mice that are protected from ethanolinduced hepatic steatosis^[82]. Several studies suggest that lipin-1, which is critically involved in triglyceride synthesis, might have a significant role in ALD. Liver-specific deficiency of lipin 1 was associated

with increased hepatic triglyceride and cholesterol accumulation and inflammation in ethanol fed mice $[83]$. SIRT1, which is downregulated in mice models of ALD, was found to affect hepatic steatosis, inflammation, and fibrosis *via* lipin-1^[84].

Chronic HCV infection

Steatosis is also found in up to 80% of patients with chronic HCV infection^[85]. Co-occurrence of steatosis and chronic HCV infection are associated with progression of liver disease and poor response in interferon- α based treatment^[4,86-89]. Prevalence of steatosis in chronic HCV infection is double than that found in patients with hepatitis B virus infection, suggesting that cooccurrence of NAFLD and HCV does not fully explain the high prevalence of steatosis in patients with chronic HCV infection^[85,90]. Additionally, probability of steatosis is significantly higher in genotype 3 than in non-3 genotype HCV infections, which argues in favor of viral induction of fatty liver in genotype 3 HCV infection $[4]$. Accordingly, fatty liver was found to be associated with HCV RNA levels in genotype 3a and body mass index in genotype 1 HCV infection^[91]. Interestingly, infection of the immortalized hepatoma cell line HepG2 with HCV genotype 1b was associated with suppressor of cytokine signalling 3 mediated impairment of insulin signalling when compared to HCV genotype 2 infected cells^[92]. All HCV genotypes, but genotype 3 to a higher extent, exert direct effects on hepatic lipid metabolism resulting in occurrence of large droplets in hepatocytes $[85,93]$. HCV makes use of VLDL assembly and secretion pathways in production of infectious virus. In virion morphogenesis, LDs are targeted by virus-encoded proteins. LDs have also been shown to influence the course of the infection. The role of LDs in HCV infection has recently been reviewed extensively by Filipe *et al*[93]. Increased intracellular triglyceride accumulation in HCV infection might be due to both direct viral effects and indirect effects resulting from accompanying insulin resistance^[93-105]. Decreased β-oxidation, increased gluconeogenesis, and DNL are key features of steatosis in HCV patients. Recently, it was shown that HCV blocks phosphorylation of forkhead box protein O1 (FOXO1), resulting in enhanced gluconeogenesis and promotion of lipogenesis by increasing the levels of SREBP-1c and fatty acid synthase $(FAS)^{[94,96,105]}$. In addition to reduced phosphorylation of FOXO1, increased expression of gluconeogenic enzymes phosphoenolpyruvate carboxykinase and glucose-6-phosphatase was found to be due to enhanced cyclic AMP responsive element-binding protein phosphorylation, linking HCVinduced ER stress with increased hepatic glucose output $[106]$. Viral proteins were also found to stimulate transcription of key enzymes of gluconeogenesis and diacylglycerol acyltransferase-1, which catalyses the final step in triglyceride synthesis and affects LD formation. Resulting insulin resistance might further

contribute to increased lipogenesis. However, the exact mechanisms have not been fully elucidated in this setting. Furthermore infection with genotype 3 HCV strains has been found to be associated with reduced levels of phosphatase and tensin homolog, which is a negative regulator of insulin/phosphoinositide 3-kinase signalling. Remarkably, interaction of HCV with DDX3, a DEAD box protein, led to phosphorylation of inhibitor k B kinase α (IKK α) and, as a consequence, the induction of SREBP-induced lipogenesis and LD formation $[102]$. Beyond quantitative effects, HCV infection was recently shown to alter hepatic metabolic zonation by redistributing FAS from perilobular to mid-zone of the lobule^[104]. Beta-oxidation is also reduced in virusinfected cells due to decreased expression of shortchain acyl-CoA dehydrogenase and medium-chain acyl-CoA dehydrogenase. *In vivo* data suggested that $PPAR\alpha$ expression is also decreased as a consequence of increased suppression by microRNA 27 (miR27) in chronic HCV infection^[99]. In contrast, Tanaka and colleagues^[100] reported from a mouse model that permanent PPARα activation plays an essential role in development of hepatic steatosis in chronic HCV infection. Remarkably, HCV infection may also directly affect VLDL secretion, resulting in intracellular lipid accumulation. Triglyceride content of VLDL particles have been shown to be lower in HCV infected patients $[101]$. In the same study, hepatic lipogenesis was significantly higher and cholesterol synthesis lower in patients with chronic HCV infection when compared to healthy controls^[101]. Low cholesterol levels have been associated with genotype 3 infection and, in contrast to non-responders, successful HCV treatment was accompanied by normalization of cholesterol levels^[107].

Drug-induced hepatic steatosis

Several drugs have been identified that may induce reversible intrahepatic triglyceride accumulation. These include antimicrobials, such as tetracycline, glucocorticoids, tamoxifen, chemotherapeutic agents, methotrexate, anti-arrhythmics such as amiodarone as well as antiepileptic valproic acid, several antiviral agents, lipid lowering mipomersen and lomitapide, and non-steroidal anti-inflammatory drugs such as acetylsalicylic acid $[108]$. Mechanistically, druginduced (*e.g.,* valproic acid) diminished mitochondrial β-oxidation of fatty acids leads to excessive reactive oxygen species generation and depletion of ATP, commonly resulting in development of microvesicular steatosis. Other pathophysiological mechanisms include enhanced cellular uptake of fatty acid due to increased fatty acid translocase expression or increased DNL resulting from elevated expression levels of SREBP1c or decreased AMPK levels. In contrast to the microvesicular steatotic pattern, macrovesicular steatosis is usually considered benign and is often reversible. Detailed mechanisms and outcome have

recently been reviewed by Amacher *et al*^[108] (Table 2).

Acute fatty liver of pregnancy

Antiretroviral agents (*e.g.*, didanosine, stavudine)

Methotrexate

Acute fatty liver of pregnancy is a rare life-threatening disease in late pregnancy. It is characterized by microvesicular pattern of hepatic steatosis^[109]. Incidence was estimated as 5/100000 pregnancies in the United Kingdom $[110]$. Mechanistically, fatty liver is due to a defect in mitochondrial β-oxidation. In late pregnancy, when the woman is increasingly dependent on fat as an energy source, the hitherto compensated defect in fatty acid metabolism results in mitochondrial dysfunction^[111,112]. Remarkably, risk of acute fatty liver of pregnancy (AFLP) is highly increased when the fetus is homozygous or compound heterozygous for a defect in fatty acid oxidation $(FAO)^{[113,114]}$. Urgent termination of pregnancy usually leads to significant improvements or normalization of $AFLP^{[115]}$, suggesting a pathophysiological role of the placenta in this disease. Mitochondrial dysfunction due to a defect in placental FAO leads to oxidative stress and, in parallel, accumulation of circulating toxic fatty acid intermediates, such as arachidonic acid. The latter has been shown to induce damage, apoptosis, and lipid accumulation in hepatocytes^[116].

FUTURE PERSPECTIVES

Although knowledge on pathophysiology of fatty liver disease has significantly improved in the past years, treatment options, especially of NAFLD, are still very limited. Several studies have suggested the beneficial effects of weight loss on the course of NAFLD. Pharmacologically, thiazolidinedione (glitazones) and antioxidative vitamin E are the most promising therapies today. Novel expectant concepts include activation of SIRT-1 (*e.g.*, reservatrol), which was found to be beneficial in murine models of NAFLD by exerting insulin-sensitizing, anti-inflammatory, and antioxidative effects^[117,118]. Very recently, a liver specific LXR inverse

agonist significantly reduced hepatic steatosis by reducing *de novo* lipogenesis^[119]. The effect of simtuzumab, which is a humanized antifibrotic monoclonal antibody against lysyl oxidase like molecule 2, is currently under investigation in patients with advanced $NAFLC^{[120]}$. FXR agonists, such as obeticholic acid, are probably the most promising therapeutic option under investigation in NAFLD.

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