



AMERICAN  
SOCIETY FOR  
MICROBIOLOGY

# Complete Genome Sequences of the Obligate Symbionts “*Candidatus Sulcia muelleri*” and “*Ca. Nasuia deltocephalinicola*” from the Pestiferous Leafhopper *Macrostelus quadripunctulatus* (Hemiptera: Cicadellidae)

Gordon M. Bennett,<sup>a</sup> Simona Abbà,<sup>b</sup> Michael Kube,<sup>c</sup> Cristina Marzachi<sup>b</sup>

University of Hawaii at Manoa, Department of Plant and Environmental Protection Sciences, Honolulu, Hawaii, USA<sup>a</sup>; Istituto per la Protezione Sostenibile delle Piante, CNR, Torino, Italy<sup>b</sup>; Albrecht Daniel Thaer-Institute, Humboldt-Universität zu Berlin, Berlin, Germany<sup>c</sup>

Two bacterial symbionts of the European pest leafhopper, *Macrostelus quadripunctulatus* (Hemiptera: Cicadellidae), were fully sequenced. “*Candidatus Sulcia muelleri*” and “*Ca. Nasuia deltocephalinicola*” represent two of the smallest known bacterial genomes at 190 kb and 112 kb, respectively. Genome sequences are nearly identical to strains reported from the closely related host species, *M. quadrilineatus*.

Received 20 November 2015 Accepted 27 November 2015 Published 21 January 2016

Citation Bennett GM, Abbà S, Kube M, Marzachi C. 2016. Complete genome sequences of the obligate symbionts “*Candidatus Sulcia muelleri*” and “*Ca. Nasuia deltocephalinicola*” from the pestiferous leafhopper *Macrostelus quadripunctulatus* (Hemiptera: Cicadellidae). *Genome Announc* 4(1):e01604-15. doi:10.1128/genomeA.01604-15.

Copyright © 2016 Bennett et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](#).

Address correspondence to Gordon M. Bennett, gordon.bennett@hawaii.edu.

Insects that feed on plant-sap diets (Hemiptera) rely on obligate microbial symbionts to provide essential nutrition (1). In the Auchenorrhyncha suborder, many hosts harbor two co-primary symbionts, “*Candidatus Sulcia muelleri*” (*Bacteroidetes*) and a *betaproteobacterium*. Together, these symbionts provide insect hosts with the ten essential amino acids (EAAs) (2–4). Recent genomic analysis revealed that these symbionts have the smallest known genomes, ranging between 112 and 245 kb (2–8). Both originated with their hosts over 260 million years ago (6, 9, 10).

We sequenced “*Ca. Sulcia*” and the *betaproteobacterium*, “*Ca. Nasuia deltocephalinicola*,” strains from the leafhopper *Macrostelus quadripunctulatus* (Cicadellidae: Deltocephalinae: PUNC) (9, 11). This species is a pest throughout Europe, vectoring economically important “*Candidatus Phytoplasma*” plant pathogens (12, 13). Genomes were sequenced in order to characterize bacterial symbiont populations infecting *M. quadripunctulatus*.

Ten insect specimens were pooled from lab-reared colonies collected in Torino, Italy. Genomic DNA was extracted with a CTAB protocol and RNase treatment (14). DNA was sequenced with PacBio (405,502 reads, average length 991 bp) and paired-end Illumina HiSeq (2 × 100 bp reads, 300 bp insert, ~13 million reads). Illumina-derived reads were adaptor trimmed and quality filtered in Trimmomatic v0.33 (15). PacBio reads were corrected with post-processed Illumina reads through PacBioToCA (16). *De novo* hybrid genome assemblies were done with SPAdes v3.6.1 (17). BLASTn was used to bin symbiont contigs (18). “*Ca. Sulcia*” and “*Ca. Nasuia*” were assembled into one and three contigs, respectively. Contigs were merged into single scaffolds in Geneious v9.0.2 (19). Reads were mapped iteratively to reordered symbiont scaffolds with SOAP2 v2.22 to verify consistent chromosomal

coverage (20). Average read depth for completed genomes was 94-fold for “*Ca. Sulcia*” and 130-fold “*Ca. Nasuia*.” Refinement revealed a 3-fold coverage spike in “*Ca. Nasuia*” starting at nucleotide position 106,992 and corresponding to an 85 bp AT-rich, noncoding repeat. Coding content was predicted via RAST with Glimmer, and verified manually with respect to previously sequenced strains (21, 22). Pair-wise divergence for all protein-coding genes was estimated following previously described pipelines (23).

The circular chromosomes of “*Ca. Nasuia*” and “*Ca. Sulcia*” PUNC comprise 112,031 bp and 190,671 bp, respectively. The “*Ca. Nasuia*” PUNC genome is characterized by a G+C content of 16.6%, and it encodes 137 protein-coding genes (CDS), 29 tRNAs, and a 16S-23S-5S rRNA operon. It retains the metabolic pathways for two EAAs, histidine and methionine. In contrast, the “*Ca. Sulcia*” PUNC genome has a G+C content of 24.4% and encodes 190 CDS, 30 tRNAs, and a 16S-23S-5S rRNA operon. It retains the complementary pathways for synthesizing the remaining eight EAAs.

Results confirm that the leafhopper genus *Macrostelus* hosts obligate nutritional symbionts with two of the smallest known genomes (6, 8). Both PUNC symbiont genomes exhibit perfectly conserved sequence synteny and coding content in line with strains previously sequenced from *Macrostelus quadrilineatus* (ALF) (6). However, as observed in other leafhoppers, the average pair-wise divergence for protein-coding genes is lower between “*Ca. Nasuia*” PUNC and ALF strains (90.47%) than it is for the highly conserved “*Ca. Sulcia*” symbiont (99.68%) (24).

**Nucleotide sequence accession numbers.** Complete genome sequences have been deposited in DDBJ/ENA/GenBank under accession numbers CP013211 to CP013212.

## ACKNOWLEDGMENTS

We thank Saskia A. Hogenhout (John Innes Centre), Assunta Bertaccini of the COST action FA0807 (*alma mater* Studiorum, University of Bologna), and Nancy Moran (University of Texas at Austin) for project support.

## FUNDING INFORMATION

University of Hawaii, Manoa provided funding to Gordon Morse Bennett.

This work was supported by a Short-Term Mobility Program (STM 2014) from the National Research Council of Italy (CNR) for C.M. and G.M.B. G.M.B. was partially supported by the National Science Foundation award IOS1347116 and the U.S. Department of Agriculture, AFRI fellowship TEXQ-2013-0342.

## REFERENCES

- Baumann P. 2005. Biology bacteriocyte-associated endosymbionts of plant sap-sucking insects. *Annu Rev Microbiol* 59:155–189. <http://dx.doi.org/10.1146/annurev.micro.59.030804.121041>.
- Wu D, Daugherty SC, Van Aken SE, Pai GH, Watkins KL, Khouri H, Tallon LJ, Zaborsky JM, Dunbar HE, Tran PL, Moran NA, Eisen JA. 2006. Metabolic complementarity and genomics of the dual bacterial symbiosis of sharpshooters. *PLoS Biol* 4:e188. <http://dx.doi.org/10.1371/journal.pbio.0040188>.
- McCutcheon JP, Moran NA. 2007. Parallel genomic evolution and metabolic interdependence in an ancient symbiosis. *Proc Natl Acad Sci USA* 104:19392–19397. <http://dx.doi.org/10.1073/pnas.0708855104>.
- McCutcheon JP, Moran NA. 2010. Functional convergence in reduced genomes of bacterial symbionts spanning 200 million years of evolution. *Genome Biol Evol* 2:708–718.
- Woyke T, Tighe D, Mavromatis K, Clum A, Copeland A, Schackwitz W, Lapidus A, Wu D, McCutcheon JP, McDonald BR, Moran NA, Bristow J, Cheng J. 2010. One bacterial cell, one Complete genome. *PLoS One* 5:e10314. <http://dx.doi.org/10.1371/journal.pone.0010314>.
- Bennett GM, Moran NA. 2013. Small, smaller, smallest: the origins and evolution of ancient dual symbioses in a phloem-feeding insect. *Genome Biol Evol* 5:1675–1688. <http://dx.doi.org/10.1093/gbe/evt118>.
- Moran NA, Bennett GM. 2014. The tiniest tiny genomes. *Annu Rev Microbiol* 68:195–215. <http://dx.doi.org/10.1146/annurev-micro-091213-112901>.
- Chang H, Cho S, Canale MC, Mugford ST, Lopes JRS, Hogenhout SA, Kuo C. 2015. Complete genome sequence of “*Candidatus Sulcia muelleri*” ML, an obligate nutritional symbiont of maize leafhopper (*Dalbulus maidis*). *Genome Announc* 3(1):e01483-14. <http://dx.doi.org/10.1128/genomeA.01483-14>.
- Moran NA, Tran P, Gerardo NM. 2005. Symbiosis and insect diversification: an ancient symbiont of sap-feeding insects from the bacterial phylum *Bacteroidetes*. *Appl Environ Microbiol* 71:8802–8810. <http://dx.doi.org/10.1128/AEM.71.12.8802-8810.2005>.
- Koga R, Bennett GM, Cryan JR, Moran NA. 2013. Evolutionary replacement of obligate symbionts in an ancient and diverse insect lineage. *Environ Microbiol* 15:2073–2081. <http://dx.doi.org/10.1111/1462-2920.12121>.
- Noda H, Watanabe K, Kawai S, Yukuhiro F, Miyoshi T, Tomizawa M, Koizumi Y, Nikoh N, Fukatsu T. 2012. Bacteriome-associated endosymbionts of the green rice leafhopper *Nephrotettix cincticeps* (Hemiptera: Cicadellidae). *Appl Entomol Zool* 47:217–225. <http://dx.doi.org/10.1007/s13355-012-0110-1>.
- Bosco D, Galetto L, Leoncini P, Saracco P, Raccah B, Marzachi C. 2007. Interrelationships between “*Candidatus Phytoplasma asteris*” and its leafhopper vectors (Homoptera: Cicadellidae). *J Econ Entomol* 100:1504–1511.
- Batlle A, Altabella N, Sabaté J, Laviña A. 2008. Study of the transmission of stolbur phytoplasma to different crop species, by *Macrosteles quadripunctulatus*. *Ann Appl Biol* 152:235–242. <http://dx.doi.org/10.1111/j.1744-7348.2007.00210.x>.
- Marzachì C, Veratti F, Bosco D. 1998. Direct PCR detection of phytoplasmas in experimentally infected insects. *Ann Appl Biol* 133:45–54. <http://dx.doi.org/10.1111/j.1744-7348.1998.tb05801.x>.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 30:2114–2120. <http://dx.doi.org/10.1093/bioinformatics/btu170>.
- Koren S, Schatz MC, Walenz BP, Martin J, Howard JT, Ganapathy G, Wang Z, Rasko DA, McCombie WR, Jarvis ED, Phillippy AM. 2012. Hybrid error correction and *de novo* assembly of single-molecule sequencing reads. *Nat Biotechnol* 30:693–700. <http://dx.doi.org/10.1038/nbt.2280>.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: A new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 19:455–477. <http://dx.doi.org/10.1089/cmb.2012.0021>.
- Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. *BMC Bioinformatics* 10:421. <http://dx.doi.org/10.1186/1471-2105-10-421>.
- Kearse M, Moir R, Wilson A, Stones-Havas S, Cheung M, Sturrock S, Buxton S, Cooper A, Markowitz S, Duran C, Thirer T, Ashton B, Meintjes P, Drummond A. 2012. Geneious basic: an integrated and extendable desktop software platform for the organization and analysis of sequence data. *Bioinformatics* 28:1647–1649. <http://dx.doi.org/10.1093/bioinformatics/bts199>.
- Li R, Li Y, Kristiansen K, Wang J. 2008. SOAP: short oligonucleotide alignment program. *Bioinformatics* 24:713–714. <http://dx.doi.org/10.1093/bioinformatics/btn025>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsmma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <http://dx.doi.org/10.1186/1471-2164-9-75>.
- Delcher AL, Bratke KA, Powers EC, Salzberg SL. 2007. Identifying bacterial genes and endosymbiont DNA with glimmer. *Bioinformatics* 23:673–679. <http://dx.doi.org/10.1093/bioinformatics/btm009>.
- Bennett GM, McCutcheon JP, McDonald BR, Moran NA. 10 August 2015. Lineage-specific patterns of genome deterioration in obligate symbionts of sharpshooter leafhoppers. *Genome Biol Evol*.
- Bennett GM, McCutcheon JP, MacDonald BR, Romanovicz D, Moran NA. 2014. Differential genome evolution between companion symbionts in an insect-bacterial symbiosis. *mBio* 5:e01697-14. <http://dx.doi.org/10.1128/mBio.01697-14>.