Detection of blood AFPmRNA in nude mice bearing human HCC using nested RT-PCR and its significance

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Prognosis of patients with HCC is estimated by several factors, such as histological differentiation of tumor cells, tumor size, and extent of lymhatic or hemotogenous spread. In this study, we detected the AFPmRNA in the blood of nude mice bearing human hepatocellular cacinoma (HCC) using nested reverse transcriptase polymerase chain reaction (nested RT-PCR) and study its significance and role in blood spread and distant metastasis.

MATERIALS AND METHODS

Animal SMMC-LINM cell lines which can secrete AFP (provided by the pathology department of our university^[1]) were inoculated into the neck and back of nude mice (BALB/C/NU, 4 weeks after birth), (107 HCC cells to each mouse). After 6 weeks, 20 nude mice (15g-20g in weight) with tumors growing to 2cm-4cm were used in the experiment.

Collection of samples Both eyes of the nude mice bearing HCC were scooped out and blood (about 1ml) was collected and placed into a 12ml centrifuge tube. The subcutaneous tumors were resected, and their integrity and relationship with the surrounding tissues were observed. The liver, lung, kidneys and other oragans of the mice were cut and examined pathologically.

Methods One ml whole blood was collected from the peripheralvein of each subject into a centrifuge tube. AFPmRNA was detected with nested RT-PCR. The detailed procedures were as follows.

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Detection of AFPmRNA Heparinized whole blood was centrifuged and the plasma fraction was removed. The cellular fraction was enriched for mononuclear cells or possible tumor cells according to the method by Komeda^[2]. Total cellular RNA was extracted by a single-step method of RNA isolation^[3]. The reverse transcription reaction was carried out in 20 µl reaction mixture using a first-strand cDNA synthesis kit (Promega USA) according to the manufacturer's instructions. Nested PCR was conducted by addition of 5 μ l solution of cDNA to 100 µl reaction mixture containing 10mM Tris HCl (pH 9.0), 50mM potassium chloride, 4.5mM magnesium chloride, 250nM dNTP 15pmol of each outer primer (EX-sense and EX-antisense) and 2.5 units of Taq DNA polymerase (Promega, USA). The reaction mixtures were subjected to 35 cycles of amplification in a programmable thermal cycler (Perking-Elmer Cetus, USA) using the following sequence: 94 °C for 1.5min, 57 °C for 1.5min and 72°C for 2.5min, and a final extension step at 72 °C for 10min. A sample of 10 μ l of the first amplification product was further amplified using an inner pair of primers (IN-sense and INantisense). To verify the amplified AFP DNA fragment, the samples were digested with the restriction enzyme Pst I and analysed by electrophoresis on a 2% agrose gel and stained with ethidium bromide for the specific bands of 174 base pairs (first amplification product) and 101 base pairs (second amplification product). Nested PCR was performed two or three times for samples with conflicting results. The external and inner pair of primers were designed as follows:

EX-sense 5'-ACTGAATCCACAACACTGCATAG-3' EX-antisense 5'-TGCAGTCAATGCATCTTCACCA-3' IN-sense 5'-TGGAATAGCTTCCATATTGGATTC-3' IN-antisense 5'-AAGTGGCTTCTTGAACAAACTGG-3'

According to this design, the PCR products of 176 and 101 base pairs were amplified from AFPcDNA by external (EX-sense and EX-antisense) and internal (INsense and IN-antisense) primer pairs, respectively. EXsense was located in exon 1 (AFPmRNA nucleotides 90-112), EX-antisense in exon 2 (AFPmRNA nucleotides 241-263), IN-sense over exon 1 and exon 2 (AFPmRNA nucleotides 122-145) and IN-antisense in exon 3

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(AFPmRNA nucleotides 200-222). cDNA sequences followed the method reported previously^[4].

Statistical analysis The relationship between the AFPmRNA in peripheral blood and various clinical parameters was examined by Chi-square test.

RESULTS

AFP mRNA was detected in the blood of 6 (30. 0%) mice bearing HCC, 4 (66.67%) of 6 nude mice had distant metastasis in lungs, liver or kidneys (Figures 1-3).

Table 1 The relationship between detectable rate of AFPmRNA and distant metastasis

Types	Cases	Cases of metastasis	Metastasis rate
AFPmRNA (+)	6	4	66.67 ^b
AFPmRNA (-)	14	0	0.00

^b*P*<0.01, *vs* AFPmRNA (-).

The diameters of HCC in 6 mice with positive AFP mRNA in blood were more than 3cm, no distant metastasis occurred in tumors below 3cm in diameter (Table 2).

Table 2The relationship between tumor diameters and distantmetastasis

Diameter	Cases	Cases of AFPmRNA (+)	Positive rate
>3cm	9	6	66.67 ^b
<3cm	11	0	0.00

^b*P*<0.01, *vs* tumors below 3cm in diameter.

AFPmRNA was detected among 12 (50%) nude mice bearing HCC with serum AFP levels beyond 4 000 μ g/L while no AFPmRNA was found in 8 nude mice with serum AFP levels below 4 000 μ g/L (Table 3).

Table 3 The relationship between AFP levels in serum anddetectable AFPmRNA

AFP levels(µg/L)	Cases	AFPmRNA (+)	Positive rate
>4000	12	6	50.0 ^b
<4000	8	0	0.0

^b*P*<0.01, vs nude mice with serum AFP levels below 4 000 μ g/L.

The first-cycle PCR products, 174 base pairs, can be cut into two pieces of 102 and 72 base pairs by restriction enzyme Pst I. The second-cycle PCR product, 101 base pairs, can be cut into two pieces of 60 and 41 base pairs (Figure 4).



Figure 1 Metastasis in liver. (×30)

Figure 2 Metastasis in lungs. (×30)

Figure 3 Metastasis in kidney. (×30)

Figure 4 The first-cycle PCR products was cut into two pieces of 102 bp and 72 bp by restriction enzyme Pst I; and the final PCR products was cut into two pieces of 60 bp and 41 bp.

DISCUSSION

Recently, there have been some reports about the employment of the reverse transcriptasepolymerse chain reaction (RT-PCR) technique to detect tumor cells spreading into the peripheral blood, bone marrow, and lymph nodes ^[5-7]. There trials have aimed to amplify tumorspecific gene transcripts which can not be detected in these tissues under normal conditions. Free mRNA is so fragile under conditions of abundant RNAse activity that the specific mRNA in blood can indicate the presence of intact cells producing such proteins just before the extraction of RNA.

This study showed that AFPmRNA was detected in 6 (30.0%) of 20 nude mice bearing human HCC 4 (66.67%) of them had distant metastasis. None of 14 nude mice with negative AFPmRNA in blood had distant metastasis (P<0.01), suggesting that the distant metastasis occurred via blood circulation. The detectable rates of AFPmRNA was significantly related with AFP levels, tumor size and distant metastasis (P<0.01). In other words, AFPmRNA in blood may be a prerequisite for distant metastasis of HCC.

In conclution, AFPmRNA may be an effective and sensitive marker for HCC matestasis in blood and distant metastasis as well.

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