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Novel terpenes generated by heterologous expression of bacterial terpene synthase genes in an engineered *Streptomyces* host

Yuuki Yamada[1], **Shiho Arima**[2], **Tohru Nagamitsu**[2], **Kohei Johmoto**[3], **Hidehiro Uekusa**[3], **Tadashi Eguchi**[3], **Kazuo Shin'ya**[4], **David E. Cane**[5], and **Haruo Ikeda**[1,§]

[1]Kitasato Institute for Life Sciences, Kitasato University, Kitasato, Sagamihara, Kanagawa 252-0373, Japan

[2]School of Pharmacy, Kitasato University, Shirokane, Minato-ku, Tokyo 108-8641 Japan

[3]Department of Chemistry and Materials Science, Tokyo Institute of Technology, O-okayama, Meguro-ku, Tokyo 152-8551, Japan

[4]National Institute of Advanced Industrial Science and Technology, Aomi, Koto-ku, Tokyo 135-0064, Japan

[5]Department of Chemistry, Box H, Brown University, Providence, RI 02912-9108, USA

Abstract

Mining of bacterial genome data has revealed numerous presumptive terpene synthases. Heterologous expression of several putative terpene synthase genes in an engineered *Streptomyces* host has revealed 13 newly discovered terpenes whose GC-MS and NMR data did not match any known compounds in the spectroscopic databases. Each of the genes encoding the corresponding terpene synthases were silent in their parent microorganisms. Heterologous expression and detailed NMR spectroscopic analysis allowed assignment of the structures of 13 new cyclic terpenes. Among these newly identified compounds, two were found to be linear triquinane sesquiterpenes that have never previously been isolated from bacteria or any other source. The remaining 11 new compounds were shown to be diterpene hydrocarbons and alcohol, including hydropyrene (**1**), hydropyrenol (**2**), tsukubadiene (**11**), and odyverdienes A (**12**) and B (**13**) each displaying a novel diterpene skeleton that had not previously been reported.

Keywords

heterologous expression; novel terpenes; *Streptomyces*; bacterial terpene synthase

INTRODUCTION

Among the tens of thousands of known terpenoid metabolites, many with medically or agriculturally useful activity, the vast majority have been isolated from plants or fungi, with

[§]To whom correspondence should be addressed: ikeda@ls.kitasato-u.ac.jp, tel: +81-42-778-9345, fax: +81-42-778-9930.

only a relative handful having been obtained from bacterial sources. Among the bacteria, *Actinomycetales* microorganisms have been known for many years as producers of volatile odoriferous metabolites, foremost among these the degraded sesquiterpene alcohol geosmin that is largely responsible for the characteristic odor of moist soil[1,2]. Also contributing to this odor is the well-recognizable musty scent of the methylated monoterpene alcohol 2-methylisobornenol also produced by many *Actinomycetales* microorganisms[3]. These two terpene natural products have also been detected from *Cyanobacteria*[4,5] and *Myxobacteria*[6].

The numerous parent cyclic monoterpene, sesquiterpene, and diterpene hydrocarbons and alcohols are all formed by variations of a universal cyclization mechanism that is initiated by enzymatic ionization of the relevant acyclic allylic diphosphate precursors, geranyl diphosphate, farnesyl diphosphate, and geranylgeranyl diphosphate, respectively. Intramolecular attack by the resultant allylic cations on the remaining double bonds, followed by a cascade of electrophilic reactions, frequently in combination with common carbocation rearrangements, terminated by quenching of the positive charge by deprotonation or by capture of water generates the enormous variety of cyclic terpenes, with nearly 400 parent skeletons being known to date. The experimental or bioinformatics search for the responsible bacterial terpene synthases has been hindered since these microbial terpene synthases exhibit at most low and most often insignificant levels of overall amino acid sequence similarity to those from fungi and plants as well as relatively low levels of mutual sequence similarity even among the bacterial terpene synthases. To address this challenge, we have developed a powerful genome mining method using hidden Markov models (HMMs) to carry out Pfam searches of bacterial sequence databases. As originally reported, the initial formulation of this model resulted in the recognition of 41 bacterial terpene synthases, including for the first time those for 2-methylisoborneol and 2-methylenebornane[7]. While the original PF3936 profile (Terpene synthase C terminal domain) had been based exclusively on an alignment of plant terpene synthase sequences, we subsequently generated a new HMM profile for searching for bacterial terpene synthases using the alignment data derived from the 41 bacterial terpene synthase sequences that had been discovered by the first round of HMM searching[8]. Most recently, we have used the 140 bacterial terpene synthase sequences uncovered in the second round of Pfam searching to create a third generation HMM, thereby uncovering 262 prospective bacterial terpene synthases by screening of more than 8.6 million predicted protein sequences found within the most recent public and in-house bacterial genome databases[9]. These results establish that terpene synthases are widely distributed in bacteria. Although many of the responsible genes appear to be silent in their host microorganisms, at least under common laboratory culture conditions, we have been able to express a large number of bacterial genes encoding terpene synthases in an engineered *Streptomyces* host, thereby enabling the identification of many terpenoid metabolites from this heterologous expression system. Although the majority of the newly detected bacterial terpenes corresponded to known compounds previously isolated from fungi or plants, several novel terpenes were also isolated[9].

We now report the detailed structure elucidation of a more than a dozen novel terpenes generated by heterologous expression of newly identified *Streptomyces* genes that encode terpene synthases.

MATERIALS AND METHODS

Bacterial strains and growth conditions

The previously described large-deletion derivative of *S. avermitilis*, SUKA22,[10] was used as the heterologous host for the expression of genes encoding presumptive terpene synthases derived from *Streptomyces* microorganisms. Cloning of genes encoding terpene synthase and the cultivation conditions for the heterologous expression of terpene synthase genes were described previously[10,11]. Two integrating vectors, pKU1021*fps* (AB982125), also harboring a gene for farnesyl diphosphate synthase, and pKU1021*ggs* (AB982126), harboring the gene for geranylgeranyl diphosphate synthase, were used for expression in *S. avermitilis* SUKA22 of genes encoding sesquiterpene synthases and diterpene synthases, respectively, *S. avermitilis* SUKA22 carrying pKU460[11] served as the negative control.

Isolation of terpenoid metabolites

Terpene hydrocarbons and alcohols were extracted with methanol from the mycelium of *S. avermitilis* SUKA22 transformants carrying individual genes encoding heterologous terpene synthases. The methanol extract was re-extracted with hexane and a portion of the hexane extract was directly analyzed by GC-MS using previously described analytical conditions[12]. Terpenes for which there was no match in the current GC-MS databases, NIST/EPA/NIH MS Library (2014 version), were purified by silica gel chromatography, as described previously[9].

Physico-chemical analysis

Nuclear magnetic resonance (NMR) spectra were obtained on a JEOL JNM-ECP 500 FT NMR System (^{1}H: 500 MHz, ^{13}C: 125 MHz) and Agilent Technologies UNITY-400 (^{1}H: 400 MHz). Chemical shifts are referenced to $CDCl_3$ at room temperature. HR-MS spectra by electron ionization (EI) mode were obtained on a JEOL JMS-700 Mstation. Infrared (IR) spectra were recorded on a Horiba FT-720 infrared spectrometer and optical rotations were recorded on a Horiba SEPA-300 polarimeter.

Preparation of hydropyrene 4,12-epoxide

Purified hydropyrene (5.2 mg, 0.019 mmol) was dissolved in 1 ml of CH_2Cl_2 under N_2 atmosphere at 0°C. *m*-Chloroperbenzoic acid (4.9 mg, 0.023 mmol) was added and the mixture was stirred for 1 hr at 0°C. After the reaction was stopped by addition of 1 ml $Na_2S_2O_3$-saturated water, the product was extracted twice with 5 ml of CH_2Cl_2. The organic layers were combined, dried over anhydrous Na_2SO_4, and evaporated to dryness. The residue was dissolved in small volume of hexane and the oxidized product was separated by silica gel column chromatography. The purified product eluted with hexane/ethyl acetate (50:1) to give 4.1 mg of hydropyrene-4,12-epoxide, which was obtained as white powder (yield 79%; HR-MS (EI) $[M]^+$ calcd. for $C_{20}H_{32}O$ 288.2453, found 288.2455).

X-ray crystallographic analysis

Hydropyrene-4,12-epoxide (3.5 mg) was crystallized from chloroform. The colorless crystal having approximate dimension of 0.18 × 0.14 × 0.10 mm was mounted. The single crystal

X-ray diffraction data were collected on a Rigaku R-AXIS Varimax RAPID-II with IP area detectors system using Cu-Kα radiation (λ = 1.54178Å). The crystal structure was solved by the direct method by *SHELXS*[13]. Refinement was performed by full-matrix least-squares using *SHELXL-2014*. Crystallographic data for the structure of hydropyrene-4,12-epoxide have been deposited with the Cambridge Crystallographic Data Center, deposition No. CCDC-1022903.

RESULTS

Diterpenes from transformants carrying *sclav_p0765*

Cultures of the large-deletion mutant, *S. avermitilis* SUKA22, carrying *sclav_p0765* from *S. clavuligerus* ATCC 26074, known as a producer of the clinically important antibiotics, clavulanic acid and cephamycin C, produced ten diterpenes (Fig. S1). Although six of these were obtained in only very low quantities, the four major compounds were purified. After cultivation of the transformants in 35 liters (5 × 7 liters), the terpene products were extracted from the mycelium and the four major components were purified by silica gel column chromatography. The HR-MS (electron ionization; EI) of peak 5 (hydropyrene; **1,** 51.7 mg, $[\alpha]_D^{24}$ −60.7 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) showed a molecular ion peak at *m/z* 272.2505 $[M]^+$, consistent with a molecular formula of $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon with five degrees of unsaturation. The ^1H NMR spectrum (Fig. S8) of **1** showed two olefinic protons (δ_H 4.42, d, *J* = 2.0 Hz, 1H, H-12; 4.65, t, *J* = 2.0 Hz, 1H, H-12; Fig. 1A) assigned to an exomethylene. Analysis of the ^{13}C NMR (Fig. S8) and DEPT spectra of **1** showed 20 resolved signals and confirmed the presence of three methyls, eight sp^3 methylenes, one sp^2 methylene, five sp^3 methines, and one sp^2 and two sp^3 quaternary carbons. One double bond accounted for one degree of unsaturation, while the remaining degrees of unsaturation were thus attributed to four rings in **1**. The proton and carbon connectivity were assigned by analysis of the ^1H-^{13}C hetero-nuclear multiple quantum coherence (HMQC) spectrum (Fig. S9), as shown in Fig. 1A. Analysis of the ^1H-^1H COSY (Fig. S10) revealed one partial structure C-1 to C-11 (Fig. 1A) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C hetero-nuclear multiple-bond connectivity (HMBC) experiments (Fig. S11) gave the following information: The cross peaks from H_3-11 (δ_H 0.83) to C-1 (δ_C 28.2), C-2 (δ_C 35.9) and C-10a (δ_C 44.0), from H_2-12 (δ_H 4.42, 4.65) to C-3a (δ_C 47.6), C-4 (δ_C 151.8) and C-5 (δ_C 49.8), from H_3-13 (δ_H 0.77) to C-5, C-5a (δ_C 36.2), C-6 (δ_C 42.4) and C-10c (δ_C 45.8), and from H_3-14 (δ_H 0.90) to C-8 (δ_C 42.3), C-8a (δ_C 34.6), C-9 (δ_C 39.8) and C-10c supported the structure as shown in Fig. 1A.

The molecular formula of peak 10 (hydropyrenol; **2,** 22.3 mg, $[\alpha]_D^{24}$ −124.7 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 3434, 2921, 1440, 1382, 890 cm^{-1}) was deduced as $C_{20}H_{32}O$ (calcd. 290.2610) by HR-MS (EI) *m/z* 290.2612 $[M^+]$ for a diterpene alcohol with four degrees of unsaturation. The ^1H and ^{13}C NMR spectra (Fig. S13) of **2** were similar to those of **1**. Analysis of ^1H-^1H COSY (Fig. S14) revealed one partial structure C-1 to C-11 (Fig. 1B), ^1H-^{13}C HMQC (Fig. S15) and ^1H-^{13}C HMBC (Fig. S16) revealed that **2** is the C-4 alcohol derivative of **1** (Fig. 1B). The ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC experiments gave the following information: The cross

peaks from H_3-11 (δ_H 0.82) to C-1 (δ_C 28.3), C-2 (δ_C 36.1) and C-10a (δ_C 44.1), from H_3-12 (δ_H 1.08) to C-3a (δ_C 49.9), C-4 (δ_C 73.8) and C-5 (δ_C 52.3) from H_3-13 (δ_H 1.13) to C-5, C-5a (δ_C 34.9), C-6 (δ_C 43.2) and C-10c (δ_C 45.2), and from H_3-14 (δ_H 0.99) to C-8 (δ_C 42.6), C-8a (δ_C 34.6), C-9 (δ_C 39.8) and C-10c supported the structure as shown in Fig. 1B. The stereochemical assignments of the methyl protons (H_3-12) were elucidated from the ^{1}H-^{1}H NOESY data (Fig. S17). Two pairs of correlations between two protons H_3-12/H_1-3a and H_3-12/H_3-13, respectively, were observed by ^{1}H-^{1}H NOESY, suggesting that the hydroxyl residue at C-4 is equatorial (Fig. 1B). Since NMR analysis was not sufficient to elucidate the full structures of **1** and **2**, X-ray crystallographic analysis of **1** was undertaken. Although neither **1** nor **2** formed fine crystals in variety of solvent system, the corresponding 4,12-epoxide of **1** (**1-epoxide**; ^{1}H NMR ($CDCl_3$, 400 MHz) δ_H 2.70 (1H, d, J = 5.0 Hz), 2.69 (1H, d, J = 5.0 Hz), 2.04 (1H, dt, J = 12.8, 5.0 Hz), 1.93 (1H, d, J = 12.8 Hz), 1.79–0.83 (18H, m), 0.96 (3H, s), 0.95 (3H, s), 0.85 (3H, d, J = 6.2 Hz), 0.64 (1H, dd, J = 12.8, 1.4 Hz)) prepared by oxidation of the C-4,12 olefinic bond formed tiny monoclinic crystals (Fig. S19). Crystal data for **1-epoxide**: Empirical formula; $C_{20}H_{32}O$; Molecular weight: 288.46; Crystal system: monoclinic; Space group: $P2_1$; Unit cell dimensions: a = 6.2239(1)Å, b = 8.9584(2)Å, c = 14.6575(3)Å, β = 93.696(1)°, V = 815.55(3)Å^3; Z value = 2; D_{calc} = 1.175 g/cm^3; T = 173(2) K, no. of unique reflections = 2924, R_{int} = 0.0357, no. of parameters = 193, no. of restraints = 1, R_1 = 0.0398, wR_2 = 0.1091, S = 1.091 for 2402 reflections, max/min. residual density 0.16/−0.15 eÅ^{-3}. The absolute structure was unable to be determined from Flack parameter 0.1(2). Diterpenes, **1** and **2** each, have novel 6-6-6-6 ring skeletons.

The compound corresponding to peak 2 purified (8.0 mg) was identical to the known diterpene isoelisabethatriene[14] (isolated from sea plumes *Pseudopterogorgia elisabethae*). The ^{1}H and ^{13}C NMR data of peak 3 (isoelisabethatriene B, **3**, 4.8 mg, $[\alpha]_D^{24}$ +9.9 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) were very similar to those of isoelisabethatriene. The HR-MS (EI) of **3** showed a molecular ion peak at *m/z* 272.2505 $[M]^+$ consistent with $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon with five degrees of unsaturation. The ^{1}H NMR spectrum (Fig. S20) of **3** showed two olefinic protons (δ_H 5.11, t, J = 7.0 Hz, 1H, H-14; 5.40, brs, 1H, H-5; Fig. 1C). Analysis of ^{13}C NMR (Fig. S20) and DEPT spectra of **3** showed 20 resolved signals and confirmed the presence of five methyls, six sp^3 methylenes, three sp^3 methines, two sp^2 methines, and four sp^2 quaternary carbons. Three double bonds accounted for three degrees of unsaturation, indicating a bicyclic skeleton for **3**. The connectivity of proton and carbon were assigned by analysis of the ^{1}H-^{13}C HMQC spectrum (Fig. S21) which combined with analysis of the ^{1}H-^{1}H COSY (Fig. S22) and ^{1}H-^{13}C HMBC (Fig. S23) spectra. The ^{1}H-^{1}H COSY revealed two partial structures C-2 to C-14 and C-7 to C-8 (Fig. 1C). The ^{1}H-^{13}C long range couplings of *2J* and *3J* observed in the ^{1}H-^{13}C HMBC experiments gave the following information: The cross peaks from H_1-4 (δ_H 1.16) to C-12 (δ_C 35.8), from H_1-5 (δ_H 5.40) to C-7 (δ_C 31.9) and C-19 (δ_C 23.5), from H_1-11 (δ_H 1.24) to C-12, from H_2-12 (δ_H 1.30) to C-4 (δ_C 43.2), C-11 (δ_C 31.5), C-13 (δ_C 26.2), C-14 (δ_C 124.9) and C-18 (δ_C 13.9), from H_2-13 (δ_H 2.01, 1.96) to C-12, from H_1-14 (δ_H 5.11) to C-12, C-16 (δ_C 18.0) and C-17 (δ_C 25.7), from H_3-16 (δ_H 1.60) to C-14, C-15 (δ_C 131.2) and C-17, from H_3-17 (δ_H 1.69) to C-14, C-15 and C-16, from H_3-18 (δ_H 0.77) to C-4, C-11 and C-12, from H_3-19 (δ_H 1.66) to

C-5, C-6 (δ_C 134.2) and C-7, and from H_3-20 (δ_H 1.64) to C-1 (δ_C 134.2), C-2 (δ_C 32.4) and C-9 (δ_C 124.5) supported the structure, a isomer of isoelisabethatriene, as shown in Fig. 1C.

Diterpenes from transformants carrying *sclav_p1169*

S. avermitilis SUKA22 transformants carrying *sclav_p0765* of *S. clavuligerus* ATCC 26074 produced seven diterpene compounds (Fig. S2). These metabolites, which accumulated in the mycelium, were purified from a 7-liter culture by silica gel column chromatography. Six components, peak 4 (clavulatriene A; **4**, 22.3 mg, $[\alpha]_D^{24}$ +42.0 (*c* 0.15, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}), peak 7 (clavulatriene B; **5**, 4.2 mg, $[\alpha]_D^{24}$ −88.3 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}, λ_{max}^{MeOH} nm (ε) 246 (1,003)), peak 1 (prenyl-β-elemene, **6**; 0.9 mg, $[\alpha]_D^{24}$ +15.1 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}), peak 2 (prenylgermacrene B; **7**, 1.3 mg, $[\alpha]_D^{24}$ −110.2 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}), peak 3 (prenylgermacrene[15]; 1.2 mg), and peak 5 (lobophytumin C[16]; 4.8 mg), were isolated. Each of the latter two metabolites has previously been isolated from plant and soft coral sources, respectively.

The HR-MS (EI) of **4** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, corresponding to $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon with five degrees of unsaturation. The ^1H NMR spectrum (Fig. S25) of **4** showed two olefinic protons (δ_H 4.74, d, *J* = 1.6 Hz, 1H, H-18; 4.80, brs, 1H, H-18; Fig. 2A) assignable to an exomethylene. An additional olefinic proton (δ_H 5.14, m, 1H, H-14) was also observed. Analysis of the ^{13}C NMR (Fig. S25) and DEPT spectra of **4** showed 20 resolved signals and confirmed the presence of four methyls, eight sp^3 methylenes, one sp^2 methylene, one sp^3 methine, one sp^2 methine, and four sp^2 and one sp^3 quaternary carbons. Three double bonds accounted for three degrees of unsaturation, indicating that **4** possessed a bicyclic skeleton. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S26), as shown in Fig. 2A. Analysis of the ^1H-^1H COSY (Fig. S27) revealed two partial structures C-6 to C-8 and C-13 to C-14 (Fig. 2A) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S28) experiments gave the following information: The cross peaks from H_2-6 (δ_H 1.60) to C-4 (δ_C 124.5), C-5 (δ_C 135.1), C-8 (δ_C 28.2) and C-10 (δ_C 34.5), from H_1-7 (δ_H 1.80) to C-9 (δ_C 40.3), from H_2-12 (δ_H 2.07) to C-7 (δ_C 45.6), C-11 (δ_C 154.8), C-13 (δ_C 26.9), C-14 (δ_C 124.3) and C-18 (δ_C 106.8), from H_2-13 (δ_H 2.11) to C-12 (δ_C 35.0), from H_1-14 (δ_H 5.14) to C-16 (δ_C 25.7) and C-17 (δ_C 17.7), from H_3-16 (δ_H 1.69) to C-14, C-15 (δ_C 131.5) and C-17, from H_3-17 (δ_H 1.62) to C-14, C-15 and C-16, from H_3-19 (δ_H 1.04) to C-1 (δ_C 42.4), C-5 (δ_C 135.1), C-9 (δ_C 40.3) and C-10 (δ_C 34.5), and from H_3-20 (δ_H 1.60) to C-3 (δ_C 33.2) and C-4 and C-5 supported the structure as shown in Fig. 2A. The stereochemical assignment of the methyl protons (H_3-19; two pairs of correlations between two protons H_3-19/H_2-2 and H_3-19/H_2-8, respectively) was elucidated from ^1H-^1H NOESY data (Fig. 2A and Fig. S29). Since a correlation between two protons H_1-7 and H_1-9 was observed by ^1H-^1H NOESY, the prenyl residue at C-7 was assigned an equatorial configuration (Fig. 2A).

The HR-MS (EI) of **5** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, consistent with a molecular formula of $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon with five

degrees of unsaturation. The ^{1}H NMR spectrum (Fig. S31) showed two olefinic protons (δ_H 5.14, m, 1H, H-14; 6.12, s, 1H, H-6; Fig. 2B). The ^{13}C NMR (Fig. S31) and DEPT spectra of **5** showed 20 resolved signals and indicated the presence of five methyls, seven sp^3 methylenes, one sp^3 methine, two sp^2 methines, and four sp^2 and one sp^3 quaternary carbons. Three double bonds accounted for three degrees of unsaturation, with the remaining two degrees of unsaturation was attributed to a bicyclic skeleton in **5**. The connectivity of proton and carbon were assigned by analysis of the ^{1}H-^{13}C HMQC spectrum (Fig. S32), as shown in Fig. 2B. Analysis of the ^{1}H-^{1}H COSY (Fig. S33) revealed three partial structures C-1 to C-20, C-8 to C-9 and C-12 to C-14 (Fig. 2B) and the ^{1}H-^{13}C long range couplings of *2J* and *3J* observed in the ^{1}H-^{13}C HMBC (Fig. S34) experiments gave the following information: The cross peaks from H_1-6 (δ_H 6.12) to C-4 (δ_C 38.2), C-8 (δ_C 22.9), C-10 (δ_C 34.6) and C-11 (δ_C 128.4), from H_3-16 (δ_H 1.67) to C-14 (δ_C 124.5), C-15 (δ_C 131.7) and C-17 (δ_C 17.6), from H_3-17 (δ_H 1.60) to C-14, C-15 and C-16 (δ_C 25.7), from H_3-18 (δ_H 1.78) to C-7 (δ_C 128.5), C-11 (δ_C 128.4) and C-12 (δ_C 34.9), from H_3-19 (δ_H 1.13) to C-1 (δ_C 41.7), C-5 (δ_C 148.5), C-9 (δ_C 41.5) and C-10 (δ_C 34.6), and from H_3-20 (δ_H 1.15) to C-3 (δ_C 33.2), C-4 and C-5 supported the structure as shown in Fig. 2B, while and the stereochemistry of two methyl protons (H_3-19 and H_3-20) was deduced from the ^{1}H-^{1}H NOESY data (Fig. 2B and Fig. S35).

The HR-MS (EI) of **6** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, consistent with a molecular formula of $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon with five degrees of unsaturation. The ^{1}H NMR spectrum (Fig. S37) showed six olefinic protons (δ_H 4.59, d, J = 1.2 Hz, 1H, H-3; 4.72, brs, 1H, H-18; 4.79, brs, 1H, H-18; 4.81, brs, 1H, H-3; 4.89, s, 1H, H-2; 4.91, d, J = 17.5 Hz, 1H, H-2; Fig. 2C) assignable to three pairs of exomethylene protons. Two additional olefinic protons (δ_H 5.13, m, 1H, H-14; 5.81, dd, J = 10.7, 17.7 Hz, 1H, H-1) were also observed. The ^{13}C NMR (Fig. S37) and DEPT spectra of **6** showed 20 resolved signals and indicated the presence of four methyls, five sp^3 methylenes, three sp^2 methylenes, two sp^3 methines, two sp^2 methines, and three sp^2 and one sp^3 quaternary carbons. The four double bonds accounted for four degrees of unsaturation, indicating the presence of one ring in **6**. The connectivity of proton and carbon were assigned by analysis of the ^{1}H-^{13}C HMQC spectrum (Fig. S38), as shown in Fig. 2C. Analysis of the ^{1}H-^{1}H COSY (Fig. S39) revealed three partial structures C-1 to C-2, C-5 to C-9 and C-12 to C-14 (Fig. 2C) and the ^{1}H-^{13}C long range couplings of *2J* and *3J* observed in the ^{1}H-^{13}C HMBC (Fig. S40) experiments gave the following information: The cross peaks from H_2-2 (δ_H 4.91, 4.89) to C-10 (δ_C 40.0), from H_2-3 (δ_H 4.81, 4.59) to C-5 (δ_C 52.8), from H_3-16 (δ_H 1.68) to C-14 (δ_C 124.2), C-15 (δ_C 131.6) and C-17 (δ_C 17.7), from H_3-17 (δ_H 1.61) to C-14, C-15 and C-16 (δ_C 25.7), from H_2-18 (δ_H 4.79, 4.72) to C-7 (δ_C 44.4), C-11 (δ_C 154.5) and C-12 (δ_C 34.9), from H_3-19 (δ_H 0.99) to C-1 (δ_C 150.1), C-5 (δ_C 52.8), C-9 (δ_C 40.1) and C-10 (δ_C 40.0), and from H_3-20 (δ_H 1.69) to C-3 (δ_C 112.0), C-4 (δ_C 147.8) and C-5 (δ_C 52.8) supported the structure as shown in Fig. 2C. The stereochemical assignments of two methyl protons (H_3-19 and H_3-20) were deduced from the ^{1}H-^{1}H NOESY data (Fig. S41), while the C-7 prenyl residue was assigned an equatorial configuration, based on the observed correlation of H-7 with H-5 and H-9.

The HR-MS (EI) of **7** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, consistent with a molecular formula of $C_{20}H_{32}$ (calcd. 272.2504) for a diterpene hydrocarbon having five degrees of unsaturation. The ^1H NMR spectrum (Fig. S43) showed three olefinic protons (δ_H 4.51, m, 1H, H-5; 4.71, m, 1H, H-1; 5.04, m, 1H, H-14; 4.81, brs, 1H, H-3; 4.89, s, 1H, H-2; 4.91, d, J = 17.5 Hz, 1H, H-2; Fig. 2D). The ^{13}C NMR (Fig. S43) and DEPT spectra of **7** showed 20 resolved signals and confirmed the presence of five methyls, seven sp^3 methylenes, three sp^2 methines, and five sp^2 quaternary carbons. With four double bonds the remaining degree of unsaturation was attributed to a ring in **7**. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S44), as shown in Fig. 2D. Analysis of the ^1H-^1H COSY (Fig. S45) revealed four partial structures C-1 to C-3, C-5 to C-6, C-8 to C-9, and C-12 to C-14 (Fig. 2D) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S46) experiments gave the following information: The cross peaks from H_3-16 (δ_H 1.69) to C-14 (δ_C 124.6), C-15 (δ_C 131.6) and C-17 (δ_C 17.7), from H_3-17 (δ_H 1.62) to C-14, C-15 and C-16 (δ_C 25.8), from H_3-18 (δ_H 1.68) to C-7 (δ_C 124.7), C-11 (δ_C 131.7) and C-12 (δ_C 34.7), from H_3-19 (δ_H 1.58) to C-1 (δ_C 128.4), C-9 (δ_C 40.0) and C-10 (δ_C 131.4), and from H_3-20 (δ_H 1.53) to C-3 (δ_C 39.0), C-4 (δ_C 135.8) and C-5 (δ_C 124.6) supported the structure as shown in Fig. 2D.

Sesquiterpenes from transformants carrying *sclav_p1407*

Although *S. avermitilis* SUKA22 transformants carrying *sclav_p1407* produced eight sesquiterpenes, one component was produced as a major product (Fig. S3; peak 1). From the mycelial extracts obtained from 4 liters culture, the compound corresponding to peak 1 (isohirsut-1-ene; **8**, 1.7 mg, $[\alpha]_D^{24}$ −162.0 (*c* 0.05, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) was purified by silica gel column chromatography. The HR-MS (EI) of **8** showed a molecular ion peak at *m/z* 204.1879 $[M]^+$ consistent with a sesquiterpene hydrocarbon of molecular formula $C_{15}H_{24}$ (calcd. 204.1878) with four degrees of unsaturation. The ^1H NMR spectrum (Fig. S49) showed a single olefinic proton (δ_H 5.01, s, 1H, H-1; Fig. 3A). ^{13}C NMR (Fig. S49) and DEPT spectra of **8** showed 15 resolved signals and confirmed the presence of four methyls, four sp^3 methylenes, three sp^3 methines, one sp^2 methine, and one sp^2 and two sp^3 quaternary carbons. In addition to the one double bond, the remaining degrees of unsaturation in **8** were attributed to three rings. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S50), as shown in Fig. 3A. Analysis of the ^1H-^1H COSY (Fig. S51) revealed one partial structure C-8 to C-10 (Fig. 3A) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S52) experiments gave the following information: The cross peaks from H_1-1 (δ_H 5.01) to C-2 (δ_C 154.1) and C-9 (δ_C 47.9), from H_1-3 (δ_H 1.69) to C-1 (δ_C 127.4) and C-2, from H_2-6 (δ_H 1.50) to C-4 (δ_C 43.7), from H_2-8 (δ_H 1.82) to C-2 and C-3 (δ_C 57.7), from H_1-9 (δ_H 3.16) to C-1 and C-2, from H_2-10 (δ_H 1.76) to C-1 and C-2, from H_3-12 (δ_H 1.08) to C-1, C-10 (δ_C 47.7), C-11 (δ_C 50.8) and C-13 (δ_C 28.1), from H_3-13 (δ_H 1.02) to C-1, C-10, C-11 and C-12 (δ_C 30.1), from H_3-14 (δ_H 1.06) to C-3 (δ_C 57.7), C-5 (δ_C 35.1), C-7 (δ_C 55.3) and C-8 (δ_C 47.8), and from H_3-15 (δ_H 1.03) to C-3, C-4 and C-5 supported the structure with a novel linear triquinane skeleton as shown in Fig. 3A.

Sesquiterpenes from transformants carrying *slt18_1880*

S. avermitilis SUKA22 transformants carrying *slt18_1880* of *S. lactacystinaeus* OM-6159 produced four sesquiterpenes, with one component being the predominant metabolite (Fig. S4; peak 2). The compound corresponding to peak 2 (isohirsut-4-ene; **9**, 1.8 mg, $[\alpha]_D^{24}$ −188.0 (*c* 0.05, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) was purified from 2 liters of culture by silica gel column chromatography. All spectroscopic data were very similar to those of **8**. The HR-MS (EI) of **9** showed a molecular ion peak at *m/z* 204.1878 $[M]^+$, consistent with a sesquiterpene hydrocarbon of molecular formula $C_{15}H_{24}$ (calcd. 204.1878) with four degrees of unsaturation. The ^1H NMR spectrum (Fig. S54) showed one olefinic proton (δ_H 5.06, brs, 1H, H-5; Fig. 3B). Analysis of the ^{13}C NMR (Fig. S54) and DEPT spectra of **9** showed 15 resolved signals and confirmed the presence of four methyls, four sp^3 methylenes, three sp^3 methines, one sp^2 methine, and one sp^2 and two sp^3 quaternary carbons. Besides the one double bond, the remaining three degrees of unsaturation in **9** were attributed to three rings. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC (Fig. S55) spectrum, as shown in Fig. 3B. Analysis of the ^1H-^1H COSY (Fig. S56) revealed two partial structures C-1 to C-10 and C-5 to C-6 (Fig. 3B) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S57) experiments gave the following information: The cross peaks from H_3-12 (δ_H 0.92) to C-1 (δ_C 48.8), C-10 (δ_C 47.4), C-11 (δ_C 41.6) and C-13 (δ_C 30.5), from H_3-13 (δ_H 1.06) to C-1, C-10, C-11 and C-12 (δ_C 29.2), from H_3-14 (δ_H 1.17) to C-6 (δ_C 47.5) C-7 (δ_C 52.9) and C-8 (δ_C 48.9), and H_3-15 (δ_H 1.68) to C3 (δ_C 66.7), C-4 (δ_C 142.8) and C-5 (δ_C 121.8) supported the structure, an isomer of **8**, as shown in Fig. 3B.

Cyclooctat-7(8),10(14)-diene (10), a diterpene isolated from transformants carrying *slt18_1078*

S. avermitilis SUKA22 transformants carrying *slt18_1078* of *S. lactacystinaeus* OM-6159 produced a diterpene hydrocarbon (Fig. S5), assigned as cyclooctat-7(8),10(14)-diene, **10** (24.0 mg, $[\alpha]_D^{24}$ +70.7 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}), which was isolated from 7 liters of culture and purified by silica gel column chromatography. The HR-MS (EI) of **10** showed a molecular ion peak at *m/z* 272.2505 $[M]^+$ consistent with a diterpene hydrocarbon of molecular formula $C_{20}H_{32}$ (calcd. 272.2504) with five degrees of unsaturation. The ^1H NMR spectrum (Fig. S60) showed an olefinic proton (δ_H 5.44, dd, J = 7.7, 12.8 Hz, 1H, H-8; Fig. 4). ^{13}C NMR (Fig. S60) and DEPT spectra of **10** showed 20 resolved signals and confirmed the presence of five methyls, six sp^3 methylenes, four sp^3 methines, one sp^2 methine, and three sp^2 and one sp^3 quaternary carbons. Besides the two double bonds, the remaining degrees of unsaturation in **10** were attributed to three rings. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S61), as shown in Fig. 4. Analysis of the ^1H-^1H COSY (Fig. S62) revealed four partial structures C-1 to C-18, C-8 to C-9, C-12 to C-13 and C-15 to C-17 (Fig. 4) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S63) experiments gave the following information: The cross peaks from H_2-1 (δ_H 1.60) to C-3 (δ_C 43.0), C-6 (δ_C 49.1) and C-12 (δ_C 37.3), H_2-1 (δ_H 1.73) to C-2 (δ_C 53.5), C-6, C-12 and C-20 (δ_C 26.0), from H_1-8 (δ_H 5.44) to C-7 (δ_C 139.6), C-9 (δ_C 23.4) and C-19 (δ_C 20.6), from H_2-9 (δ_H 2.45) to C-7, C-8 (δ_C 123.8), C-10 (δ_C

141.9) and C-11 (δ_C 51.3), from H_2-12 (δ_H 1.40, 1.51) to C-10 and C-13 (δ_C 26.3), from H_2-13 (δ_H 1.99, 2.11) to C-10, from H_1-15 (δ_H 2.63) to C-10 and C-13, from H_3-16 (δ_H 0.90) to C-14 (δ_C 140.0), C-15 (δ_C 27.8) and C-17 (δ_C 21.5), from H_3-17 (δ_H 0.97) to C-14, C-15 and C-16 (δ_C 20.9), from H_3-18 (δ_H 0.93) to C-2, C-3 and C-4 (δ_C 35.9), from H_3-19 (δ_H 1.65) to C-6, C-7 and C-8, and from H_3-20 (δ_H 0.92) to C-1 (δ_C 48.2), C-10, C-11 and C-12 supported the structure as shown in Fig. 4.

Diterpenes from transformants carrying *stsu_20912*

S. avermitilis SUKA22 transformants carrying *stsu_20912* of *S. tsukubaensis* NRRL 18488, the producer of the immunosuppressant polyketide tacrolimus, generated one major diterpene product along with another seven would be minor diterpene components (Fig. S6). The major component, tsukubadiene (**11**, 9.4 mg, $[\alpha]_D^{24}$ −120.3 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) was isolated from 2 liters of culture by silica gel column chromatography. The HR-MS (EI) of **11** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, consistent with a diterpene hydrocarbon of molecular formula $C_{20}H_{32}$ (calcd. 272.2504) with five degrees of unsaturation. The ^1H NMR spectrum (Fig. S65) showed two olefinic protons (δ_H 5.11, t, *J* = 2.0, 2.0 Hz, 1H, H-14; 5.49, t, *J* = 7.3, 1H, H-7; Fig. 5). ^{13}C NMR (Fig. S65) and DEPT spectra of **11** showed 20 resolved signals and confirmed the presence of five methyls, six sp^3 methylenes, three sp^3 methines, two sp^2 methines, and two sp^2 and two sp^3 quaternary carbons. Besides the two double bonds, the remaining three degrees of unsaturation were attributed to three rings in **11**. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S66), as shown in Fig. 5. Analysis of the ^1H-^1H COSY (Fig. S67) revealed three partial structures C-1 to C-16, C-7 to C-9, C-12 and C-11 to C-12 (**********Fig. 5) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S68) experiments gave the following information: The cross peaks from H_2-5 (δ_H 2.25, 2.01) to C-3 (δ_C 37.0), C-6 (δ_C 137.5), C-7 (δ_C 124.7), C-15 (δ_C 143.7) and C-17 (δ_C 25.3), from H_1-7 (δ_H 5.49) to C-5 (δ_C 41.3) and C-17, from H_1-9 (δ_H 2.34) to C-18 (δ_C 32.1) and C-19 (δ_C 25.7), from H_2-11 (δ_H 1.48, 1.37) to C-19, from H_2-12 (δ_H 1.47) to C-20 (δ_C 25.4), from H_1-14 (δ_H 5.11) to C-9 (δ_C 53.3), C-12 (δ_C 40.7) and C-20, from H_3-16 (δ_H 1.05) to C-1 (δ_C 41.7), C-2 (δ_C 31.9) and C-15 (δ_C 143.7), from H_3-18 (δ_H 1.00) to C-9, C-10 (δ_C 40.8), C-11 (δ_C 39.9) and C-19, from H_3-19 (δ_H 0.93) to C-9, C-10 and C-18, and from H_3-20 (δ_H 0.96) to C-9 (δ_C 53.3), C-12, C-13 (δ_C 48.1) and C-14 (δ_C 132.4) supported the structure with a novel 5-9-5 ring skeleton as shown in Fig. 5.

Diterpenes from transformants carrying *nd90_0354*

S. avermitilis SUKA22 transformants carrying *nd90_0354* of *Streptomyces* sp. ND90 produced two major and one minor diterpene components (Fig. S7). The two major components were isolated from 2 liters culture, with odyverdienes A (**12**, 1.2 mg, $[\alpha]_D^{24}$ −18.7 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) and B (**13**, 1.2 mg, $[\alpha]_D^{24}$ −44.2 (*c* 0.1, $CHCl_3$), IR ν_{max} (attenuated total reflection) 2921, 1440, 1382, 890 cm^{-1}) each being obtained as a colorless oil. The HR-MS (EI) of **12** showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, corresponding to a diterpene hydrocarbon of molecular formula $C_{20}H_{32}$ (calcd. 272.2504) with five degrees of unsaturation. The ^1H

NMR spectrum (Fig. S71) showed four olefinic protons (δ_H 4.63, s, 1H, H-17; 4.66, s, 1H, H-17; 4.85, s, 1H, H-18; 4.96, s, 1H, H-18; Fig. 6A) assignable to two exomethylenes. ^{13}C NMR (Fig. S71) and DEPT spectra of **12** showed 20 resolved signals and confirmed the presence of three methyls, seven sp^3 methylenes, two sp^2 methylenes, five sp^3 methines, and two sp^2 and one sp^3 quaternary carbons. Besides the two double bonds the remaining degrees of unsaturation in **12** were attributed to three rings. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC spectrum (Fig. S72), as shown in Fig. 6A. Analysis of the ^1H-^1H COSY (Fig. S73) revealed two partial structures C-1 to C-3 and C-5 to C-19 (Fig. 6A) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S74) experiments gave the following information: The cross peaks from H_2-1 (δ_H 1.57) to C-13 (δ_C 44.9) and C-20 (δ_C 26.8), from H_2-2 (δ_H 2.07) to C-1 (δ_C 33.8) and C-4 (δ_C 150.5), from H_1-3 (δ_H 2.36) to C-20, from H_2-9 (δ_H 1.62) to C-10 (δ_C 32.4), from H_2-13 (δ_H 1.36) to C-1, C-3 (δ_C 47.8) and C-7 (δ_C 49.9), from H_3-16 (δ_H 1.58) to C-11 (δ_C 52.9), C-15 (δ_C 150.3) and C-17 (δ_C 110.8), from H_2-17 (δ_H 4.66, 4.63) to C-11 and C-16 (δ_C 19.1), from H_2-18 (δ_H 4.96, 4.85) to C-3 and C-5 (δ_C 40.6), from H_3-19 (δ_H 0.90) to C-7, C-8 (δ_C 39.2) and C-9 (δ_C 35.1), and from H_3-20 (δ_H 1.29) to C-1, C-3, C-13 (δ_C 44.9) and C-14 (δ_C 41.2) supported the structure with a novel 6-8-4 ring skeleton as shown in Fig. 6A.

The HR-MS (EI) of **13** also showed a molecular ion peak at *m/z* 272.2506 $[M]^+$, corresponding to a diterpene hydrocarbon of molecular formula $C_{20}H_{32}$ (calcd. 272.2504) with five degrees of unsaturation. The ^1H NMR spectrum (Fig. S76) showed four olefinic protons (δ_H 4.58, s, 1H, H-17; 4.70, s, 1H, H-17; 4.75, s, 1H, H-19; 4.89, s, 1H, H-19; Fig. 6B) assignable to two exomethylenes. Analysis of ^{13}C NMR (Fig. S76) and DEPT spectra of **13** showed 20 resolved signals and confirmed the presence of three methyls, six sp^3 methylenes, two sp^2 methylenes, seven sp^3 methines, and two sp^2 quaternary carbons. Besides the two double bonds the remaining degrees of unsaturation in **12** were attributed to three rings in **13**. The connectivity of proton and carbon were assigned by analysis of the ^1H-^{13}C HMQC (Fig. S77) spectrum, as shown in Fig. 6B. Analysis of the ^1H-^1H COSY (Fig. S78) revealed one partial structure C-1 to C-20 (Fig. 6B) and the ^1H-^{13}C long range couplings of *2J* and *3J* observed in the ^1H-^{13}C HMBC (Fig. S79) experiments gave the following information: The cross peaks from H_3-16 (δ_H 1.64) to C-12 (δ_C 56.0), C-15 (δ_C 150.7) and C-17 (δ_C 110.8), from H_2-17 (δ_H 4.70, 4.58) to C-12 and C-16 (δ_C 20.1), from H_3-18 (δ_H 0.83) to C-1 (δ_C 35.1), C-2 (δ_C 29.5) and C-14 (δ_C 59.6), H_2-19 (δ_H 4.89, 4.75) to C-4 (δ_C 42.4) and C-6 (δ_C 38.1), and from H_3-20 (δ_H 0.90) to C-8 (δ_C 46.0), C-9 (δ_C 37.5) and C-10 (δ_C 35.4) supported the structure with a novel 6-7-5 ring skeleton as shown in Fig. 6B.

DISCUSSION

Actinomycetales microorganisms are prolific producers of an enormous variety of natural products, including aminoglycosides, polyketides, peptides, shikimate-derived aromatic compounds, among many others. By contrast, reports describing the production of terpenoid metabolites by these microorganisms have been very limited to date. Genome sequencing has established that *Actinomycetales* microorganisms harbor many genes, many within extensive gene clusters, dedicated to secondary metabolite biosynthesis. Our recent studies

have revealed that genes encoding presumptive terpene synthase are in fact widely distributed in bacteria[9]. *Actinomycetales* microorganisms are especially rich in genes encoding terpene synthases, although the majority of such genes appear to be cryptic in the parent microorganisms when studied under standard laboratory culture conditions. To assign the biochemical function of these apparently silent terpene synthases, we have therefore used heterologous expression to evaluate the intrinsic ability of these bacterial genes to control the biosynthesis of terpenes[9].

The closely related diterpene hydrocarbon and alcohol, hydropyrene (**1**) and hydropyrenol (**2**), were efficiently produced in a heterologous *Streptomyces* host carrying *sclav_p0756*, although they were not produced by the parent microorganism *S. clavuligerus* ATCC 26074 under any culture conditions. The unique structures of **1** and **2** have not previously been reported, with characterized by a skeleton consisting of four fused cyclohexane rings. The same heterologous transformants also produced the known diterpene hydrocarbon, isoelisabethatriene (which has previously been isolated from sea plumes), and its isomer, the novel diterpene hydrocarbon, isoelisabethatriene B (**3**) as minor components. Each of these minor components may be generated by alternative cyclization of the allylic cation formed upon initial ionization of geranylgeranyl diphosphate. The actual cyclization mechanism leading to these unusual diterpenes should be amenable to evaluation by NMR analysis of the corresponding ^{13}C-labeled compounds obtained by feeding ^{13}C-labeled precursors to the *S. avermitilis* host carrying *sclav_p0756*. It is predicted that the six diterpenes, **4**, **5**, **6**, and **7** as well as prenylgermacrene and lobophytumin C, which were produced by transformants carrying *sclav-p1169*, are likely generated by a common cyclization mechanism involving the prenylgermacrene cation.

The production of sesquiterpenes possessing a triquinane skeleton by bacteria has not previously been reported. Interestingly, two triquinane-type sesquiterpene hydrocarbons, isohirsut-1-ene (**8**) and isohirsut-4-ene (**9**), were generated by different terpene synthases, SCLAV_p1407 and SLT18_1880, respectively which exhibited limited amino acid sequence similarity, with SLT18_1880 (325 aa) showing 32% identity and 47% similarity to SCLAV_p1407 (367 aa). SLT18_1880 was also similar to a monoterpene synthase SCLAV_p0982 (31% identity and 48% similarity), which catalyzes the generation of 1,8-cineole, camphene and β-pinene[9].

Metabolites with a fused three ring 5-9-5, 6-8-4 and 6-7-5 skeletons have not previously been reported. In these studies, we have described the production and structure elucidation of three such unusual diterpene hydrocarbons, tsukubadiene (**11**) and odyverdines A (**12**) and B (**13**), The 6-8-4 ring system of odyverdiene A **(12**) differs from the 6-7-5 fused rings of odyverdiene B (**13**), representing alternative modes of cyclization of their common geranylgeranyl diphosphate precursor.

Given the wealth of new terpenes uncovered in this initial survey, it is evident that bacterial terpene synthases have the potential to generate unique structures and that the genes encoding these terpene synthase are a very attractive source for the discovery of new natural products.

Supplementary Material

Refer to Web version on PubMed Central for supplementary material.

Acknowledgments

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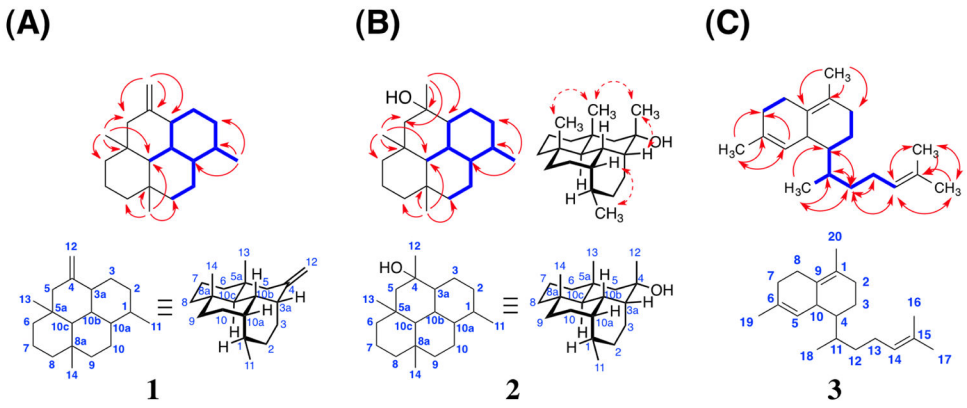
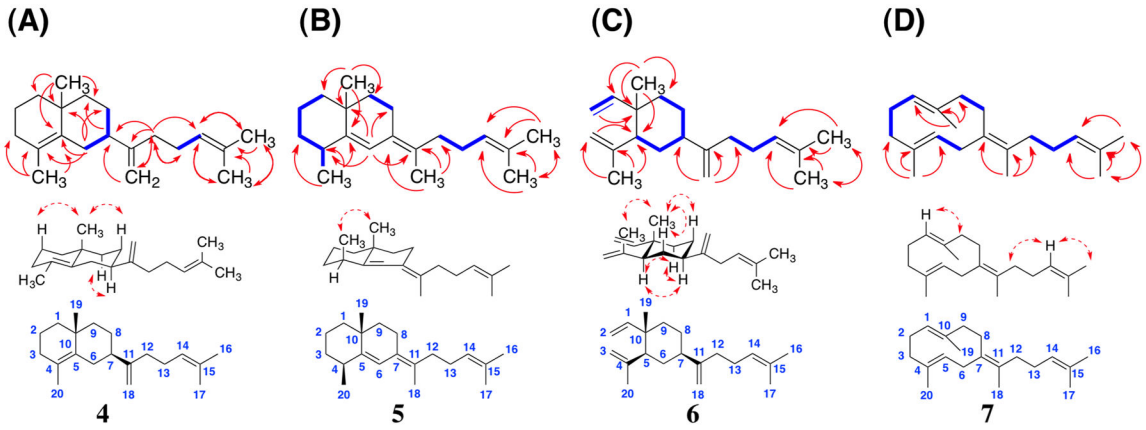


NMR chemical shifts in $CDCl_3$

	1		2		3		
No.	δ_C	δ_H (multi, *J* in Hz)	δ_C	δ_H (multi, *J* in Hz)	No.	δ_C	δ_H (multi, *J* in Hz)
1	28.2	1.66 (1H, m)	28.3	1.67 (1H, m)	1	130	-
2	35.9	1.02 (1H, m), 1.66 (1H, m)	36.1	0.97 (1H, m), 1.68 (1H, m)	2	32.4	1.92 (1H, m), 1.89 (1H, m)
3	29.6	1.26 (1H, m), 1.74 (1H, m)	25.1	1.26 (1H, m), 1.74 (1H, m)	3	21.5	1.53 (1H, m), 1.17 (1H, m)
3a	47.6	2.27 (1H, m)	49.9	1.47 (1H, m)	4	43.2	1.16 (1H, m)
4	151.8	-	73.8	-	5	124.4	5.40 (1H, brs)
5	49.8	1.50 (1H, m), 2.26 (1H, m)	52.3	1.14 (1H, m), 1.33 (1H, m)	6	134.2	-
5a	36.2	-	34.9	-	7	31.9	1.98 (1H, m), 1.83 (1H, m)
6	42.4	1.17 (1H, m), 1.35 (1H, m)	43.2	1.02 (2H, m)	8	26.7	2.70 (2H, ddd, *J* = 12.9, 4.5, 2.5)
7	18.9	1.44 (2H, m)	18.2	1.41 (2H, m)	9	124.5	-
8	42.3	1.15 (1H, m), 1.33 (1H, m)	42.6	1.11 (1H, m), 1.36 (1H, m)	10	39.2	2.53 (1H, brs)
8a	34.6	-	34.6	-	11	31.5	1.24 (1H, m)
9	39.8	0.91 (1H, m), 1.24 (1H, m)	39.8	0.92 (1H, m), 1.29 (1H, m)	12	35.8	1.30 (2H, m)
10	22.7	1.51 (1H, m), 1.63 (1H, m)	22.8	1.51 (1H, m), 1.63 (1H, m)	13	26.2	2.01 (1H, m), 1.96 (1H, m)
10a	44.0	1.23 (1H, m)	44.1	1.25 (1H, m)	14	124.9	5.11 (1H, brt, *J* = 7.0)
10b	36.1	1.89 (1H, m)	31.6	2.17 (1H, m)	15	131.2	-
10c	45.8	1.37 (1H, m)	45.2	1.25 (1H, m)	16	18.0	1.60 (3H, s)
11	20.7	0.83 (3H, d, *J* = 7.6)	20.6	0.82 (3H, d, *J* = 7.6)	17	25.7	1.69 (3H, s)
12	106.7	4.42 (1H, d, *J* = 2.0), 4.65 (1H, t, *J* = 2.0)	31.1	1.08 (3H, s)	18	13.9	0.77 (3H, d, *J* = 6.8)
					19	23.5	1.66 (3H, s)
13	20.4	0.77 (3H, s)	24.1	1.13 (3H, s)	20	18.5	1.64 (3H, s)
14	21.0	0.90 (3H, s)	22.0	0.99 (3H, s)			

Figure 1.

Structures of hydropyrene (A; **1**), hydropyrenol (B; **2**) and isoelisabethatriene B (C; **3**) produced by *S. avermitilis* SUKA22 carrying *sclav_p0765*. The upper panel shows ^{1}H-^{1}H COSY (bold line), ^{1}H-^{13}C HMBC (arrow) and ^{1}H-^{1}H NOESY (dashed double arrow) data for each compound. Double arrows indicate a cross peak in HMBC.

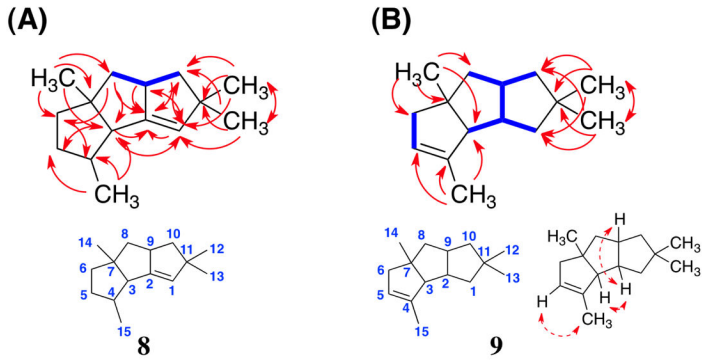


NMR chemical shifts in $CDCl_3$

	4		5		6		7	
No.	δ_C	δ_H (multi, *J* in Hz)	δ_C	δ_H (multi, *J* in Hz)	δ_C	δ_H (multi, *J* in Hz)	δ_C	δ_H (multi, *J* in Hz)
1	42.4	1.26 (1H, m), 1.55 (1H, m)	41.7	1.15 (1H, m), 1.53 (1H, m)	150.1	5.81 (1H, dd, *J* = 10.7, 17.7)	128.4	4.71 (1H, m)
2	19.1	1.55 (2H, m)	17.5	1.47 (1H, m), 1.54 (1H, m)	109.7	4.91 (1H, d, *J* = 17.5), 4.89 (1H, s)	26.9	2.30 (1H, m), 2.08 (1H, m)
3	33.2	1.83 (2H, m)	33.2	1.55 (2H, m)	112.0	4.81 (1H, brs,), 4.59 (1H, d, *J* = 1.2)	39.0	2.18 (1H, m), 2.13 (1H, m)
4	124.5	-	38.2	2.50 (1H, m)	147.8	-	135.8	-
5	135.1	-	148.5	-	52.8	1.99 (1H, m)	124.6	4.51 (1H, m)
6	31.3	2.55 (1H, dt, *J* = 14.0, 2.4), 1.74 (1H, m)	120.8	6.12 (1H, s)	33.3	1.57 (1H, m), 1.53 (1H, m)	28.4	2.00 (2H, m)
7	45.6	1.80 (1H, m)	128.5	-	44.4	1.95 (1H, m)	124.7	-
8	28.2	1.54 (1H, m), 1.63 (1H, m)	22.9	2.20 (1H, m), 2.48 (1H, m)	26.8	1.62 (1H, m), 1.41 (1H, m)	39.8	1.90 - 2.00 (2H, m)
9	40.3	1.28 (1H, m), 1.50 (1H, m)	41.5	1.35 (1H, m), 1.45 (1H, m)	40.1	1.44 (1H, m), 1.41 (1H, m)	40.0	1.90 - 2.00 (2H, m)
10	34.5	-	34.6	-	40.0	-	131.4	-
11	154.8	-	128.4	-	154.5	-	131.7	-
12	35.0	2.07 (2H, m)	34.9	2.01 (1H, m), 2.15 (1H, m)	34.9	2.06 (1H, m), 2.03 (1H, m)	34.7	2.08 (1H, m), 2.02 (1H, m)
13	26.9	2.11 (2H, m)	26.7	2.02 (1H, m), 2.13 (1H, m)	27.3	2.11 (2H, m)	26.0	2.02 (2H, m)
14	124.3	5.14 (1H, m)	124.5	5.14 (1H, m)	124.2	5.13 (1H, m)	124.6	5.04 (1H, m)
15	131.5	-	131.7	-	131.6	-	131.6	-
16	25.7	1.69 (3H, s)	25.7	1.67 (3H, s)	25.7	1.68 (3H, s)	25.8	1.69 (3H, s)
17	17.7	1.62 (3H, s)	17.6	1.60 (3H, s)	17.7	1.61 (3H, s)	17.7	1.62 (3H, s)
18	106.8	4.74 (1H, d, *J* = 1.6), 4.80 (1H, brs)	17.7	1.78 (3H, d, *J* = 2.1)	107.0	4.79 (1H, brs), 4.72 (1H, brs)	18.0	1.68 (3H, s)
19	24.7	1.04 (3H, s)	26.2	1.13 (3H, s)	16.6	0.99 (3H, s)	16.4	1.58 (3H, s)
20	19.3	1.60 (3H, s)	23.3	1.15 (3H, d, *J* = 12.2)	24.7	1.69 (3H, s)	16.1	1.53 (3H, s)

Figure 2.

Structures of clavulatriene A (A; **4**), clavulatriene B (B; **5**), prenyl-β-elemene (C; **6**) and prenylgermacrene B (D; **7**) produced by *S. avermitilis* SUKA22 carrying *sclav_p1169*. The upper panel shows ^{1}H-^{1}H COSY (bold line), ^{1}H-^{13}C HMBC (arrow) and ^{1}H-^{1}H NOESY (dashed arrow) data for each compound. Double arrows indicate a cross peak in ^{1}H-^{13}C HMBC.

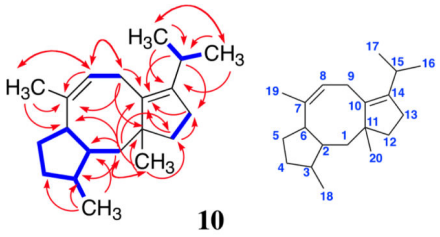


NMR chemical shifts in $CDCl_3$

	8		9	
No.	δ_C	δ_H (multi, J in Hz)	δ_C	δ_H (multi, J in Hz)
1	127.4	5.01 (1H, s)	48.8	1.58 (1H, dd, J = 13.0, 8.6), 1.30 (1H, m)
2	154.1	-	50.1	2.34 (1H, q, J = 10.0)
3	57.7	1.69 (1H, m)	66.7	2.11 (1H, brs)
4	43.7	*not assigned*	142.8	-
5	35.1	1.67 (1H, m), 1.20 (1H, m)	121.8	5.06 (1H, brs)
6	41.2	1.50 (2H, m)	47.5	2.19 (1H, dq, J = 15.7, 2.2), 2.01 (1H, dt, J = 15.7, 2.1)
7	55.3	-	52.9	-
8	47.8	1.22 (1H, m), 1.82 (1H, dd, J = 7.0, 11.5)	48.9	1.81(1H, dd, J = 13.0, 7.8), 1.32 (1H, dd, J = 13.0, 7.7)
9	47.9	3.16 (1H, quin, J = 8.3)	44.3	2.52 (1H, m)
10	47.7	0.84 (1H, m), 1.76 (1H, dd, J = 7.1, 11.5)	47.4	1.64 (1H, ddd, J = 11.6, 8.7, 1.1), 1.27 (1H, m)
11	50.8	-	41.6	-
12	30.1	1.08 (3H, s)	29.2	0.92 (3H, s)
13	28.1	1.02 (3H, s)	30.5	1.06 (3H, s)
14	30.1	1.06 (3H, s)	29.4	1.17 (3H, s)
15	20.1	1.03 (3H, d, J = 6.8)	15.9	1.68 (3H, s)

Figure 3.

Structure of isohirsut-1-ene (A; **8**) produced by *S. avermitilis* SUKA22 carrying *sclav_p1407* and isohirsut-4-ene (B; **9**) produced by *S. avermitilis* SUKA22 carrying *slt18_1880*. The upper panel shows ^{1}H-^{1}H COSY (bold line), ^{1}H-^{13}C HMBC (arrow) and ^{1}H-^{1}H ROESY (dashed arrow) data for each compound. Double arrows indicate a cross peak in ^{1}H-^{13}C HMBC.



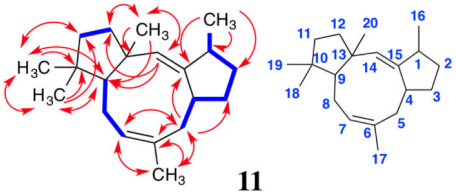
10

NMR chemical shifts in $CDCl_3$

No.	δ_C	δ_H (multi, *J* in Hz)
1	48.2	1.60 (1H, m), 1.73 (1H, m)
2	53.5	0.91 (1H, m)
3	43.0	1.36 (1H, m)
4	35.9	1.05 (1H, m), 1.69 (1H, m)
5	29.7	1.24 (1H, m), 1.62 (1H, m)
6	49.1	2.73 (1H, dd, *J* = 5.5, 8.3)
7	139.6	-
8	123.8	5.44 (1H, dd, *J* = 12.8, 7.7)
9	23.4	2.45 (2H, m)
10	141.9	-
11	51.3	-
12	37.3	1.40 (1H, m), 1.51 (1H, m)
13	26.3	1.99 (1H, m), 2.11 (1H, m)
14	140.0	-
15	27.8	2.63 (1H, m)
16	20.9	0.90 (3H, d, *J* = 7.6)
17	21.5	0.97 (3H, d, *J* = 7.6)
18	18.0	0.93 (3H, d, *J* = 7.6)
19	20.6	1.65 (3H, s)
20	26.0	0.92 (3H, s)

Figure 4.

Structure of cyclooctat-7(8),10(14)-diene (**10**) produced by *S. avermitilis* SUKA22 carrying *slt18_1078*. The upper panel shows ^{1}H-^{1}H COSY (bold line) and ^{1}H-^{13}C HMBC (arrow) data for **10**. Double arrows indicate a cross peak in ^{1}H-^{13}C HMBC.

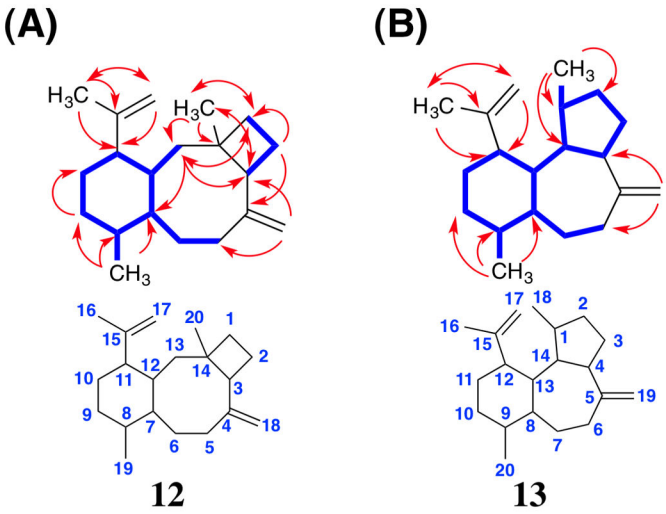


NMR chemical shifts in $CDCl_3$

No.	δ_C	δ_H (multi, *J* in Hz)
1	41.7	2.32 (1H, m)
2	31.9	1.70 (1H, m), 1.13 (1H, m)
3	37.0	1.78 (1H, m), 1.58 (1H, m)
4	40.9	2.96 (1H, m)
5	41.3	2.25 (1H, dd, *J* = 13.5, 7.7), 2.01 (1H, dd, *J* = 13.5. 2.8)
6	137.5	-
7	124.7	5.49 (1H, t, *J* = 7.3)
8	24.0	1.90 (2H, m)
9	53.3	2.34 (1H, t, *J* = 7.0)
10	40.8	-
11	39.9	1.48 (1H, m), 1.37 (1H, m)
12	40.7	1.47 (2H, m)
13	48.1	-
14	132.4	5.11 (1H, t, *J* = 2.0)
15	143.7	-
16	21.7	1.05 (3H, d, *J* = 7.0)
17	25.3	1.64 (3H, s)
18	32.1	1.00 (3H, s)
19	25.7	0.93 (3H, s)
20	25.4	0.96 (3H, s)

Figure 5.

Structure of tsukubadiene (**11**) produced by *S. avermitilis* SUKA22 carrying *stsu_20912*. The upper panel shows ^{1}H-^{1}H COSY (bold line) and ^{1}H-^{13}C HMBC (arrow) data for **11**. Double arrows indicate a cross peak in ^{1}H-^{13}C HMBC.



NMR chemical shifts in $CDCl_3$

	12		13	
No.	δ_C	δ_H (multi, *J* in Hz)	δ_C	δ_H (multi, *J* in Hz)
1	33.8	1.57 (2H, m)	35.1	2.29 (1H, m)
2	16.9	2.07 (1H, m), 1.79 (1H, m)	29.5	1.80(1H, m), 1.24 (1H, m)
3	47.8	2.36 (1H, dd, *J* = 9.8, 3.5)	27.0	1.58 (2H, m)
4	150.5	-	42.4	2.52 (1H, m)
5	40.6	2.17 (1H, m), 1.96 (1H, ddd, *J* = 14.0, 14.0, 3.0) 1.53 (1H, m),	151.6	- 2.36 (1H, m),
6	31.0	1.22 (1H, m)	38.1	2.11 (1H, ddd, *J* = 13.4, 13.4, 5.8)
7	49.9	0.42 (1H, m)	28.3	1.70 (1H, m), 1.44 (1H, m)
8	39.2	1.06 (1H, m)	46.0	0.75 (1H, m)
9	35.1	1.62 (1H, m), 0.96 (1H, m)	37.5	1.14 (1H, m)
10	32.4	1.49 (1H, m), 1.29 (1H, m)	35.4	1.67 (1H, m), 0.98 (1H, m)
11	52.9	1.55 (1H, m)	32.9	1.43 (1H, m), 1.35 (1H, m)
12	41.1	1.10 (1H, m)	56.0	1.64 (1H, m)
13	44.9	1.36 (1H, m), 1.14 (1H, m)	45.7	1.28 (1H, m)
14	41.2	-	59.6	1.11 (1H, m)
15	150.3	-	150.7	-
16	19.1	1.58 (3H, s)	20.1	1.64 (3H, s)
17	110.8	4.66 (1H, s), 4.63 (1H, s)	107.0	4.70 (1H, s), 4.58 (1H, s)
18	108.1	4.96 (1H, s), 4.85 (1H, s)	19.8	0.83 (3H, d, *J* = 12.0)
19	20.4	0.90 (3H, d, *J* = 12.0)	110.0	4.89 (1H, s), 4.75 (1H, s)
20	26.8	1.29 (3H, s)	20.6	0.90 (3H, d, *J* = 12.0)

Figure 6.

Structures of odyverdienes A (A; **12**) and B (B; **13**) produced by *S. avermitilis* SUKA22 carrying *nd90_0354*. The upper panel shows ^{1}H-^{1}H COSY (bold line) and ^{1}H-^{13}C HMBC (arrow) data for each compound. Double arrows indicate a cross peak in ^{1}H-^{13}C HMBC.