RESEARCH PAPER

The novel, orally available and peripherally restricted selective cannabinoid CB₂ receptor agonist LEI-101 prevents cisplatin-induced nephrotoxicity

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BACKGROUND AND PURPOSE

Here, we have characterized 3-cyclopropyl-1-(4-(6-((1,1-dioxidothiomorpholino)methyl)-5-fluoropyridin-2-yl)benzyl) imidazolidine-2,4-dione hydrochloride (LEI-101) as a novel, peripherally restricted cannabinoid CB₂ receptor agonist, using both *in vitro* and *in vivo* models.

EXPERIMENTAL APPROACH

We investigated the effects of LEI-101 on binding and functional activity. We assessed its *in vitro* and *in vivo* selectivity. Efficacy of LEI-101 was determined in a mouse model of cisplatin-induced nephrotoxicity.

KEY RESULTS

LEI-101 behaved as a partial agonist at CB₂ receptors using β-arrestin and GTPγS assays and was ~100-fold selective in CB₂/CB₁ receptor-binding assays. It did not display any activity on endocannabinoid hydrolases and nor did it react with serine hydrolases in an activity-based protein profiling assay. In mice, LEI-101 had excellent oral bioavailability reaching high concentrations in the kidney and liver with minimal penetration into the brain. LEI-101 up to a dose of 60 mg·kg⁻¹ (p.o.) did not exert any CNS-mediated effects in the tetrad assay, in mice. LEI-101 (p.o. or i.p.) at 3 or 10 mg·kg⁻¹ dose-dependently prevented kidney dysfunction and/or morphological damage induced by cisplatin in mice. These protective effects were associated with improved renal histopathology, attenuated oxidative stress and inflammation in the kidney. These effects were absent in CB₂ receptor knockout mice.

CONCLUSION AND IMPLICATIONS

These results indicate that LEI-101 is a selective, largely peripherally restricted, orally available CB₂ receptor agonist with therapeutic potential in diseases that are associated with inflammation and/or oxidative stress, including kidney disease.

Abbreviations

4-HNE, 4-hydroxy-2-nonenal; Δ^9-THC, Δ^9-tetrahydrocannabinol; ABPP, activity-based protein profiling; BUN, blood urea nitrogen; DAGL, diacylglycerol lipase; hFAAH, human fatty acid amide hydrolase; LEI-101, 3-cyclopropyl-1-(4-(6-((1,1-dioxidothiomorpholino)methyl)-5-fluoropyridin-2-yl)benzyl)imidazolidine-2,4-dione hydrochloride; MAGL, monoacylglycerol lipase; NAPE-PLD, N-acylphosphatidylethanolamine-specific phospholipase D; Vss, volume of distribution in steady state

Tables of Links

TARGETS	
GPCRs[a]	DAGL diacylglycerol lipase
CB$_1$ receptors	FAAH, fatty acid amide hydrolase
CB$_2$ receptors	MAGL monoacylglycerol lipase
Enzymes[b]	NAPE-PLD, N-acylphosphatidylethanolamine-specific phospholipase D
Caspase 3	
Caspase 7	

LIGANDS
Cisplatin
CP55940
IL-1β
JZL184
TNF-α
URB597
Δ^9-THC, Δ^9-tetrahydrocannabinol

These Tables list key protein targets and ligands in this article which are hyperlinked to corresponding entries in http://www.guidetopharmacology.org, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Pawson et al., 2014) and are permanently archived in the Concise Guide to PHARMACOLOGY 2013/14 ([a][b]Alexander et al., 2013a,b).

Introduction

In recent years, drug discovery efforts have focused on the design and synthesis of selective cannabinoid CB$_2$ receptor agonists (Han et al., 2013; Dhopeshwarkar and Mackie, 2014). Selective activation of this receptor has been associated with anti-inflammatory and tissue protective effects without the CNS-mediated side effects, which are associated with stimulation of psychoactive CB$_1$ receptors in the CNS (Hanus et al., 1999; Malan et al., 2001).

Although numerous putative CB$_2$ receptor agonists from different chemical series have been characterized in vitro, only a few have been used in inflammatory (Hanus et al., 1999) and neuropathic pain models (van der Stelt et al., 2011) as well as in experimental paradigms of ischemia-reperfusion injury (Batkai et al., 2007; Pacher et al., 2007), neurodegeneration (Zhang et al., 2009) and gastrointestinal disorders (Storr et al., 2009) and other diseases (Teixeira-Clerc et al., 2010; Louvet et al., 2011).

Furthermore, very limited information is available on in vivo selectivity and oral availability of these compounds (Pacher and Mechoulam, 2011; Pacher and Kunos, 2013), and limited selectivity for CB$_2$ over CB$_1$ receptors and/or bioavailability may have contributed to the failure of clinical trials with peripherally restricted mixed CB$_1$ /CB$_2$ receptor agonists for pain and to recent controversies related to CB$_2$ receptor function in the brain (Pacher and Kunos, 2013).

Recently, we reported the identification of 3-cyclopropyl-1-(4-(6-((1,1-dioxidothiomorpholino)methyl)-5-fluoropyridin-2-yl)benzyl)imidazolidine-2,4-dione hydrochloride (LEI-101) as a novel, peripherally restricted CB$_2$ receptor agonist (van der Stelt et al., 2011). In the present study, we have further characterized the activity and selectivity of LEI-101 in both in vitro and in vivo models, using various pharmacological tools and CB$_2$ receptor knockout mice. LEI-101 is ~100-fold more potent in binding to CB$_2$ receptors than to CB$_1$ receptors. It was a partial agonist in the β-arrestin recruitment and GTPγS assays and did not display any activity on the other proteins of the endocannabinoid system. LEI-101 up to a dose of $60\ mg\cdot kg^{-1}$ (p.o.) did not exert any CNS-mediated side effects in the mouse tetrad assay of cannabimimetic activity.

In addition, LEI-101 dose-dependently ameliorated kidney dysfunction and morphological damage in the cisplatin-induced nephrotoxicity model. At 3 or $10\ mg\cdot kg^{-1}$ (p.o.), LEI-101 largely prevented cisplatin-induced increases in serum creatinine and blood urea nitrogen levels, improved renal histopathological injury and attenuated oxidative/nitrative stress and inflammation in the kidney. These protective effects were absent in CB$_2$ receptor knockout mice, which is indicative of a CB$_2$ receptor-mediated effect. Taken together, our findings give strength to the potential of LEI-101 for the prevention or treatment of oxidative stress and inflammation-induced tissue damage and establish LEI-101 as an excellent tool to study CB$_2$ receptor biology.

Methods

Cell culture

We used the CNR1 PathHunter® CHO-K1 CNR1 β-Arrestin Cell Line (catalogue number 93-0959C2) and the CNR2 PathHunter® CHO-K1 CNR2 β-Arrestin Cell Line (catalogue number 93-0706C2) stably expressing human CB$_1$ receptors (CHOK1hCB$_1$R_bgal) or human CB$_2$ receptors

(CHOK1hCB$_2$R_bgal) respectively, obtained from DiscoveRx Corporation (Fremont, CA, USA). CHOK1hCB$_1$R_bgal and CHOK1hCB$_2$R_bgal cells were cultured in Ham's F12 Nutrient Mixture supplemented with 10% fetal calf serum, 1 mM glutamine, 50 μg·mL^{-1} penicillin, 50 μg·mL^{-1} streptomycin, 300 mg·mL^{-1} hygromycin and 800 μg·mL^{-1} geneticin in a humidified atmosphere at 37°C and 5% CO$_2$. Cells were subcultured twice a week at a ratio of 1:20 on 10cm diameter plates by trypsinization. For membrane preparation, the cells were subcultured 1:10 and transferred to large 15cm diameter plates. Membrane fractions were prepared exactly as described before (Zweemer et al., 2013).

[³H]CP55940 displacement assays

[³H]CP55940 displacement assays to determine CB$_2$ receptor and CB$_1$ receptor binding affinity were performed as reported previously (Bonger et al., 2007), with the following changes:

Membrane aliquots containing 5 μg (CHOK1hCB$_1$R_bgal) or 1 μg (CHOK1hCB$_2$R_bgal) of membrane protein in 100 μL assay buffer (50 mM Tris–HCl, 5 mM Mgl$_2$, 0.1% BSA, pH 7.4) were incubated at 30°C for 1 h. Non-specific binding was determined in the presence of 10 μM AM630. Filtration was performed on GF/C filters, presoaked for 30 min with 0.25% PolyEthyleneImine (PEI), using a Brandel harvester. Filter-bound radioactivity was determined in a β-counter.

PathHunter® β-arrestin recruitment assay

The assay was performed using the PathHunter hCB$_1$R_bgal CHOK1 or hCB$_2$R_bgal CHOK1 β-arrestin recruitment assay kit (DiscoveRx Corporation, Fremont, CA, USA), according to the manufacturer's protocol.

[³⁵S]GTPγS binding assay

The assay was performed as reported previously (Zweemer et al., 2013), with the following changes: 5 μg of homogenized CHOK1hCB$_2$R_bgal membranes, pretreated with 5 μg saponin and 1 μM GDP, were used in 100 μL assay buffer per assay point (50 mM Tris–HCl buffer (pH 7.4), 5 mM MgCl$_2$, 150 mM NaCl, 1 mM EDTA, 0.05% BSA and 1 mM DTT, freshly prepared every day). The reference agonist CP55940 (10 μM) was added to each filter for 100% stimulation.

Off-target profiling

The enzymatic activity of anandamide and 2-arachidonoylglycerol metabolic enzymes was assayed in human THP-1 macrophages and was expressed as pmol of product released per min per mg of protein. Fatty acid amide hydrolase (FAAH) activity was assayed in cell extracts (100 μg of protein/test) at 37°C for 30 min by measuring the release of ethanolamine-1-³H from 10 μM [ethanolamine-1-³H]-anandamide (Gattinoni et al., 2010). The synthesis of anandamide through the activity of N-acylphosphatidylethanolamine-specific phospholipase D (NAPE-PLD) was assayed in cell homogenates (100 μg protein/test) at 37°C for 30 min by measuring the release of [³H]-anandamide from 100 μM [³H]N-arachidonoyl-phosphatidylethanolamine (Tsuboi et al., 2011). The activity of diacylglycerol lipase (DAGL) was assayed in cell extracts (200 μg protein/test) with 100 μM of sn-1-stearoyl-2-[¹⁴C]arachidonoyl-glycerol at 37°C for 30 min (Bisogno

et al., 2003). Monoacylglycerol lipase (MAGL) activity was assayed (100 μg protein/test) with 10 μM [³H]2-oleoylglycerol at 37°C for 30 min (Dinh et al., 2002).

Activity based protein profiling (ABPP) selectivity assay was performed as described previously (Baggelaar et al., 2013). Briefly, mouse brain proteome (2.0 mg·mL^{-1}, 20 μL) was preincubated for 30 min with vehicle (0.5 μL DMSO) or ligand in 0.5 μL DMSO at room temperature and subsequently treated with 250 nM (final concentration) MB064 1 or 500 nM (final concentration) TAMRA-FP (from Thermo Fischer Scientific) for 15 min at room temperature. The reactions were quenched with 10 μL standard 3 × SDS page sample buffer. The samples were directly loaded and resolved on SDS page gel (10% acrylamide).

Animals

All animal care and experimental procedures conformed to National Institutes of Health (NIH) guidelines and were approved by the Institutional Animal Care and Use Committee of the National Institute on Alcohol Abuse and Alcoholism (Bethesda, MD, USA). Studies involving animals are reported in accordance with the ARRIVE guidelines for reporting experiments involving animals (Kilkenny et al., 2010; McGrath et al., 2010). A total of 206 animals were used in the experiments described here.

Six- to 8-week-old male C57BL/6J mice were obtained from The Jackson Laboratory (Bar Harbor, ME, USA). CB$_2$ receptor knockout mice (CB$_2$R$^{-/-}$) and their wild-type littermates (CB$_2$R$^{+/+}$) were developed as described previously and had been backcrossed to a C57BL/6J background. (Batkai et al., 2007). All animals were kept in a temperature-controlled environment with a 12 h light–dark cycle and were allowed free access to food and water at all times.

Pharmacokinetics of LEI-101 in mice

LEI-101 was given i.p. or by oral gavage (p.o.). The drug was dissolved in a vehicle of DMSO: Tween 80: saline, 1:1:18. After administration of LE-101, mice were killed by cervical dislocation under deep anaesthesia with 5% isoflurane, for collection of blood and tissue samples at the time described in the figure. Both tissues and plasma from the blood were stored at −70°C for further processing for mass spectrometry analyses.

Plasma and tissues were extracted and levels of LEI-101 were quantified by LC-MS/MS, using multiple reactions monitoring under conditions described previously (Godlewski et al., 2010) Briefly, the mass spectrometer was set for electrospray ionization operated in the positive ion mode. The molecular ion and fragments for each compound measured were as follows:

m/z 473.2 ➔ 200.1 and 473.2 ➔148 for LEI-101 (CID energy 28 and 44 V, respectively.

The amounts of LEI-101 in the samples were determined from standard curves using linear regression. Values were expressed as ng·ml^{-1} for serum or ng·g^{-1} for tissues.

Tetrad assay. ICR mice (counterbalanced Latin square within subject design), from The Jackson Laboratory, Bar Harbor, ME) were housed individually overnight. The behavioural testing was conducted in the following order: bar test (catalepsy), tail withdrawal test, rectal temperature

and locomotor activity. Testing was performed according to previously described procedures (Long *et al.*, 2009; Schlosburg *et al.*, 2010; Ignatowska-Jankowska *et al.*, 2014). Mice received LEI-101 or vehicle (DMSO,Tween 80 and saline, in 1:1:18 ratio) p.o. (via gavage) at dose of $60 \, mg \cdot kg^{-1}$ of each drug, 2 h before testing in the tetrad assay.

Catalepsy was assessed on a bar 0.7 cm in diameter placed 4.5 cm off of the ground. The mouse was placed with its front paws on the bar and a timer (Timer no. 1) was started. A second timer (Timer no. 2) was turned on only when the mouse was immobile on the bar, with the exception of respiratory movements. If the mouse moved off the bar, it was placed back on in the original position. The assay was stopped when either Timer no. 1 reached 60 s, or after the fourth time the mouse moved off the bar, and the cataleptic time was scored as the amount of time on Timer no. 2. Nociception was then assessed in the tail immersion assay. The mouse was placed head first into a small bag fabricated from absorbent under pads (VWR Scientific Products; 4 cm diameter, 11 cm length) with the tail out of the bag. Each mouse was hand-held and 1 cm of the tail was submerged into a 52°C water bath. The latency for the mouse to withdraw its tail within a 10 s cut off time was scored. Rectal temperature was assessed by inserting a thermocouple probe 2 cm into the rectum, and temperature was determined by telethermometer (BAT-10 Multipurpose Thermometer, Clifton, NJ, USA). Locomotor activity was assessed 120 min after treatment, for a 5 min period in a Plexiglas cage (42.7 × 21.0 × 20.4 cm) and Anymaze (Stoelting, Wood Dale, Illinois, USA) software was used to determine the percentage of time spent immobile and distance travelled.

Nephropathy model

Mice (CB$_2$R$^{-/-}$ and CB$_2$R$^{+/+}$) were killed (as described above) 72 h after a single injection of cisplatin (cis-diammineplatinum(II) dichloride $20 \, mg \cdot kg^{-1}$ i.p.; Sigma). LEI-101 was dissolved as described previously and was used at 0.3, 3.0 and $10 \, mg \cdot kg^{-1}$, i.p. and p.o. every day, starting 1.5 h before the cisplatin exposure.

Renal function monitoring

Blood was collected under deep anaesthesia with 5% isoflurane, by retro-orbital bleeding and serum levels of creatinine and blood urea nitrogen (BUN) were measured using a Prochem-V chemical analyser (TX, USA) or Idexx VetTest 8008 (Idexx Laboratories, Westbrook, ME, USA).

Histological examination for tubular damage

After fixation of the kidneys with 10% formalin, renal tissues were sectioned (5μm) and stained with periodic acid–Schiff (PAS) reagents for histological examination. Tubular damage in PAS-stained sections was examined under the microscope (200× magnification) as described (Pan *et al.*, 2010).

Nitrotyrosine staining of tissue samples

Paraffin-embedded sections (5μm) of kidney were cut, deparaffinized and hydrated in descending gradations of ethanol, followed by microwave treatment. Next, sections were incubated in 0.3% H$_2$O$_2$ in PBS to block endogenous peroxidase activity. The sections were then incubated with anti-nitrotyrosine (1:200 dilution; Cayman Chemical, Ann Arbor, MI, USA) overnight at 4°C in a moist chamber. Biotinylated secondary antibodies and ABC reagent were added according to the kit instructions (Vector Laboratories, Burlingame, CA, USA). Colour development was induced by incubation with a DAB kit (Vector Laboratories) for 3–5 min, and specific staining was visualized by light microscopy as described previously (Mukhopadhyay *et al.*, 2010b; Mukhopadhyay *et al.*, 2010a).

Lipid peroxidation/ROS production

4-Hydroxynonenal (4-HNE) in the renal tissues was determined using a kit from Cell Biolabs (San Diego, CA, USA). Renal tissue (50-80 mg) was homogenized in 1 mL of T-PER Reagent (Fisher Scientific) in a Precellys Ceramic Bead (1.4 mm) homogenizer and centrifuged at 10,000 g for 20 min. Supernatants were used as tissue lysate and renal tissue lysates (protein concentration, $10 \, \mu g \, mL^{-1}$) were adsorbed onto a 96-well plate for 12 h at 4°C. 4-HNE adducts present in the sample or standard were probed with anti-4-HNE antibody, followed by a horseradish peroxidase-conjugated secondary antibody. 4-HNE-protein adduct content in an unknown sample was determined by comparison with a standard curve as described in the manufacturer's protocol (Mukhopadhyay *et al.*, 2012).

Detection of apoptosis by renal DNA fragmentation and caspase 3/7 activity assays

Renal tissue (50-100 mg) was homogenized in 1 ml of TRIZOL® Reagent(Life technologies) in a Precellys Ceramic Bead (1.4 mm) homogenizer and centrifuged at 10,000 g for 20 min. Supernatants were used as tissue lysate. Caspase 3/7 activity in tissue lysates was measured using the Apo-One Homogenous Caspase-3/7 assay kit (Promega Corp., Madison, WI, USA). An aliquot of caspase reagent was added to each well and mixed on a plate shaker for 1 h at room temperature shielded from light, and the fluorescence was measured.

The DNA fragmentation assay was based on measuring the amount of mono- and oligonucleosomes in the cytoplasmic fraction of tissue extracts using a commercially available kit (Roche Diagnostics, Basel, Switzerland) according to the manufacturer's instructions as described by Mukhopadhyay *et al.* (2012).

Real-time PCR analyses

Total RNA was isolated from kidney homogenate (50-100 mg of renal tissue were homogenized in 1 ml of TRIZOL® Reagent (Life Technologies) in Precellys Ceramic Bead(1.4 mm) homogenizer), using Trizol reagents (Invitrogen; ThermoFisher Scientific, Waltham, MA, USA) according to the manufacturer's instructions. The isolated RNA was treated with RNase-free DNase (Ambion, Austin, TX, USA) to remove traces of genomic DNA contamination. One microgram of total RNA was reverse-transcribed to cDNA using SuperScript II (Invitrogen). The target gene expression was quantified with Power Sybr green PCR master mix using an ABI 7500 real-time PCR instrument. Each amplified sample was analysed for homogeneity using dissociation curve analysis. After denaturation at 95°C for 2 min, 40 cycles were performed at 95°C for 10 s and 60°C for 30 s. Relative quantification was

calculated using the comparative C_t method ($2^{-\Delta\Delta C^t}$: $\Delta\Delta C_t = \Delta C_{t\ sample} - \Delta C_{t\ reference}$). Lower ΔC_t values and lower $\Delta\Delta C_t$ reflect a relatively higher amount of gene transcript. Statistical analyses were carried out for at least 6 to 15 replicate experimental samples in each set.

The following primers were used: TNF-α, 5′-AAGCCTGTAGCCCACGTCGTA-3′ and 5′-AGGTACAACCC-ATCGGCTGG-3′; IL-1β, 5′-AAAAAAGCCTCGTGCTGTCG-3′ and 5′-GTCGTTGCTTGGTTCTCCTTG-3′; gp91phox, 5′-GACCATTGCAAGTGAACACCC-3′ and 5′-AAATGAAGTG-GACTCCACGCG-3′; NOX4, 5′-TCATTTGGCTGTCCCTAAA-CG-3′ and 5′-AAGGATGAGGCTGCAGTTGAG-3′; and actin, 5′-TGCACCACCAACTGCTTAG-3′ and 5′-GGATGCAGGGAT-GATGTTC-3′.

Data analysis

Results are reported as means ± SEM. Statistical significance between two measurements was determined by the two-tailed unpaired Student t-test (and among groups it was determined by ANOVA followed by *post hoc* Student–Newman–Keuls) using GraphPad Prism 4.3 or 5.0 software (San Diego, CA, USA). Probability values of $P < 0.05$ were considered significant.

Materials

LEI-101, 3-cyclopropyl-1-(4-(6-((1,1-dioxidothiomorpholino) methyl)-5-fluoropyridin-2-yl)benzyl)imidazolidine-2,4-dione hydrochloride was synthesized as described previously (van der Stelt *et al.* 2011). For *in vivo* studies, LEI-101 was dissolved in DMSO/Tween80/saline (1/1/18 v/v/v). [³H]CP55940 (specific activity 141.2 Ci mmol⁻¹) and GTPγS (specific activity 1250 Ci mmol⁻¹) were purchased from Perkin Elmer (Waltham, MA, USA). The other labelled compounds - [ethanolamine-1-³H]-anandamide (60 Ci mmol⁻¹); [³H]N-arachidonoyl-phos-phatidylethanolamine (200 Ci mmol⁻¹); *sn*-1-stearoyl-2-[¹⁴C] arachidonoyl-glycerol (55 mCi mmol⁻¹); [³H]2-oleoylglycerol (20 Ci mmol⁻¹) - were from ARC (St. Louis, MO). Bicinchoninic acid (BCA) and BCA protein assay reagent were obtained from Pierce Chemical Company (Rochford, IL, USA). All other materials were obtained from standard commercial sources and were of analytical grade.

Results

LEI-101 is a selective CB$_2$ receptor partial agonist

As shown in Table 1, LEI-101 behaves as a CB$_2$ receptor partial agonist in the β-arrestin recruitment and GTPγS assays, compared with the reference mixed CB$_1$/ CB$_2$ receptor agonist CP-55940. LEI-101 had more than 100-fold selectivity for CB$_2$ receptors over CB$_1$ receptors in the binding assay. The compound does not interact with other proteins of the endocannabinoid system: human FAAH (hFAAH), MAGL, NAPE-PLD and DAGL. LEI-101 did not inhibit serine hydrolases as determined in an ABPP with mouse membrane proteome and MB064 and TAMRA-FP as fluorescent probes (data not shown).

LEI-101 has excellent oral bioavailability in mice

Previously, we have determined the pharmacokinetic profile of LEI-101 in rats (van der Stelt *et al.*, 2011). Because the efficacy models presented in this study are in mice, we determined the plasma concentrations of LEI-101 after $10\,mg\cdot kg^{-1}$ doses, given p.o. or i.p. to mice using DMSO/

Table 1

Molecular pharmacology profile of LEI-101

Assay	Parameter	LEI-101	CP-55.940
Binding	hCB$_2$R pK$_i$ ± SEM	7.5 ± 0.1	8.4 ± 0.1
	hCB$_1$R pK$_i$ ± SEM	<5	9.6 ± 0.1
cAMP inhibition[a]	hCB$_2$R pEC$_{50}$ ± SEM	8.0 ± 0.1	N.D.[b]
	hCB$_1$R pEC$_{50}$ ± SEM	<5	N.D.
β-Arrestin recruitment	hCB$_2$R pEC$_{50}$ ± SEM (Emax: % ± SEM)	7.0 ± 0.3 (41 ± 6)	8.5 ± 0.1 (100 ± 1)
	hCB$_1$R pEC$_{50}$ ± SEM (Emax: % ± SEM)	<5 (−1% @ 10 μM)	8.3 ± 0.2 (99 ± 1)
GTPγS binding	hCB$_2$R pEC$_{50}$ ± SEM (Emax: % ± SEM)	6.6 ± 0.2 (65 ± 8)	8.3 ± 0.2 (96 ± 3)
Substrate hydrolysis[c]	hFAAH1/2 pK$_i$ ± SEM	<5	N.D.
	MAGL pK$_i$ ± SEM	<5	N.D.
	DAGL pK$_i$ ± SEM	<5	N.D.
	NAPE-PLD pK$_i$ ± SEM	<5	N.D.
ABPP	Serine hydrolases	No off targets detected	No off targets detected

[a]Data from van der Stelt *et al.* 2011; [b]N.D., not determined; [c]Residual activity values with reference inhibitors were as follows (vs. controls, set to 100%): 24% for FAAH (with 0.1 nM URB597), 27% for MAGL (with 0.1 nM JZL184) and 38% for DAGL (with 1 μM tetrahydrolipstatin). At present, inhibitors for NAPE-PLD are not commercially available.

Tween80/Saline (1/1/18 v/v/v) as a vehicle. As depicted in Figure 1A–C, plasma levels were found to peak after 2 h of either administration route. In view of its low plasma protein binding (fraction unbound = 0.44), these plasma concentrations should be more than sufficient to activate CB$_2$ receptors. Of note, the compound seemed to accumulate in the kidney (Figure 1). Lowest levels were found in the brain, thereby confirming the peripherally restricted nature of LEI-101.

LEI-101 does not induce cannabimimetic effects in vivo

To determine whether the compound maintains its selectivity for CB$_2$ receptors over CB$_1$ receptors in vivo, we tested LEI-101 in the mouse tetrad assay for CB$_1$ receptor activity. This is a very sensitive assay in which four consecutive behavioural tests, related to anti-nociception, hypothermia, catalepsy and spontaneous activity, are performed 120 min after administration of the compound. A positive result in all four tests indicates that the compound or one of its metabolites activates CB$_1$ receptors in the CNS. LEI-101 (60 mg·kg^{-1}, p.o.) did not produce any effects in the tetrad assay as compared with vehicle. There were no effects on nociceptive behaviour assessed in tail withdrawal test (Figure 2A) ($P = 0.2$ in both cases) and no changes in body temperature (Figure 2B) ($P = 0.4$ and $P = 0.8$ for each respective drug). No effects on locomotor behaviour (Figure 2C) were found in case of distance travelled ($P = 0.6$ and $P = 1.0$, respectively), time spent mobile ($P = 0.8$ and $P = 0.5$, respectively), or running speed of mice ($P = 0.3$ and $P = 0.9$, respectively). No catalepsy was observed following administration of LEI-101. The results indicate that LEI-101 at high doses administered orally did not produce CB$_1$ receptor-mediated CNS-side effects up to 60 mg·kg^{-1} (p.o.). Overall, we can conclude that LEI-101 maintains its selectivity in vivo.

LEI-101 attenuates biomarkers and histological damage of cisplatin-induced nephropathy

The known CB$_2$ receptor agonists HU-308 (Hanus et al., 1999; Mukhopadhyay et al., 2010a,b) and the natural product β-carophyllene (Horvath et al., 2012) ameliorate cisplatin-induced nephrotoxicity after i.p. administration. Therefore we sought to test whether our novel synthetic CB$_2$ receptor partial agonist, which endowed with a different chemical scaffold, could also protect animals against cisplatin-induced nephropathy after oral gavage. For assessment of cisplatin-induced nephrotoxicity, the serum creatinine and total blood urea levels, both biomarkers of renal dysfunction, were measured. 72 h following the cisplatin injection (20 mg·kg^{-1}, i.p.), a marked increase of these biomarkers was observed in C57Bl6/J mice, compared with vehicle-treated control animals. LEI-101 dose-dependently attenuated these increases in creatinine and blood urea nitrogen levels in cisplatin-treated animals (Figure 3). At 3.0 or 10 mg·kg^{-1} (p.o. and i.p.), LEI-101 largely prevented cisplatin-induced increases in serum creatinine and BUN levels. This attenuation was not observed in CB$_2$R$^{-/-}$ mice (Figure 4). Of note, the CB$_2$R$^{-/-}$ mice had an increased susceptibility to cisplatin nephrotoxicity (Figure 4).

Renal dysfunction was accompanied by morphological damage to the kidney (Figure 5). Histological examination
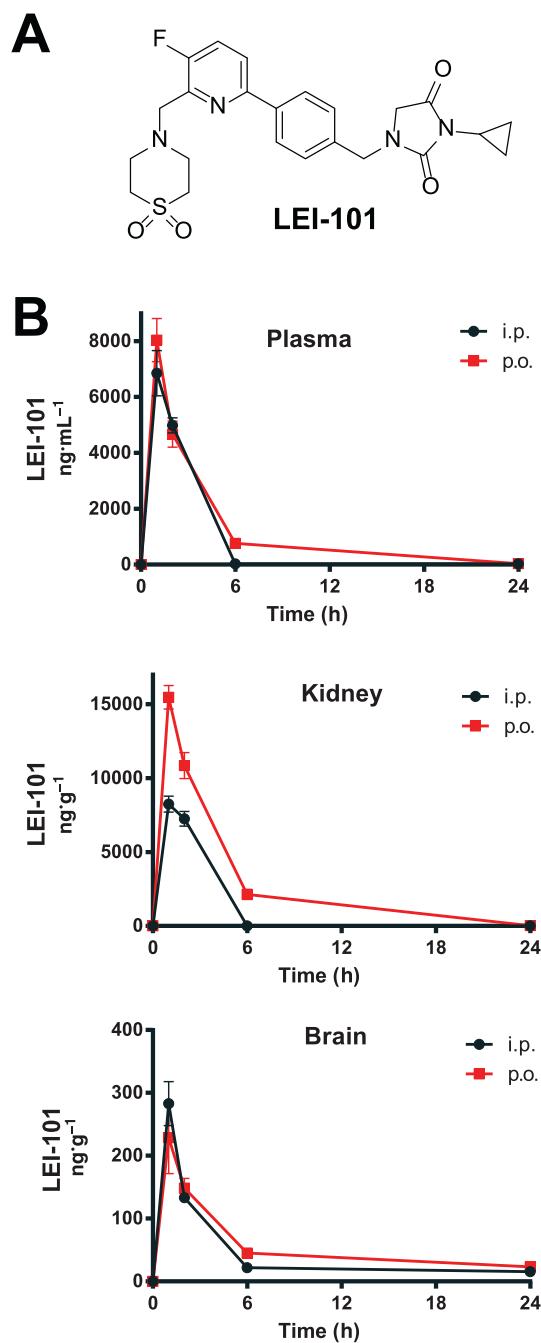

Figure 1

The CB$_2$ receptor agonist LEI-101 is an orally bioavailable and a peripherally restricted CB$_2$ receptor agonist with high kidney exposure. Chemical Structure of LEI-101 (A) and time-course of tissue distribution (B) in plasma, kidney and brain after 10 mg·kg^{-1} p.o. and i.p. administration of LEI-101 in mice. Results are means ± SEM; $n = 4$ per group.

revealed necrosis, protein casts, vacuolization and desquamation of epithelial cells in the renal tubules. LEI-101 at 10 mg·kg^{-1} decreased tubular damage as determined by PAS staining of the kidney (Figure 5). This effect was not observed in the CB$_2$R$^{-/-}$ mice, thereby indicating that the protective effect of LEI-101 was mediated through CB$_2$ receptors.
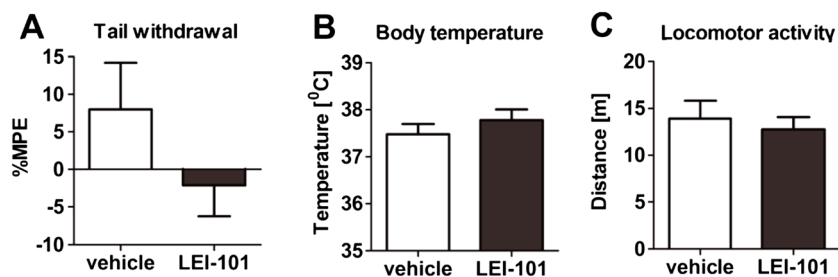
Figure 2

The CB$_2$ receptor agonist LEI-101 does not induce cannabimimetic effects *in vivo*. LEI-101 (60 mg·kg^{-1}, p.o.) did not affect nociceptive behaviour assessed in tail withdrawal test (A) and did not change body temperature (B), as compared with mice receiving vehicle. No effects on locomotor behaviour (C) were found. Results are means ± SEM; *n* = 8 per group.
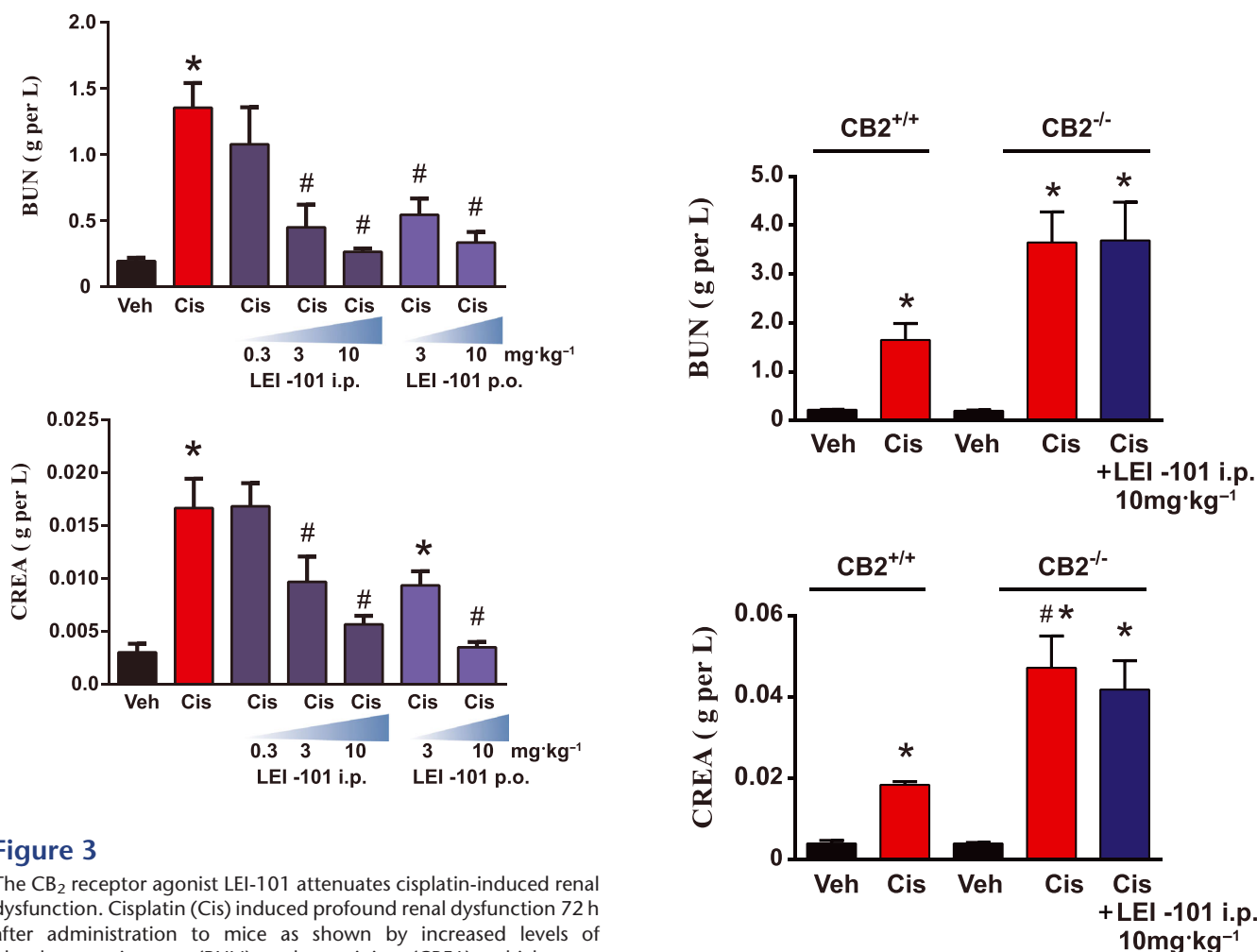

Figure 3

The CB$_2$ receptor agonist LEI-101 attenuates cisplatin-induced renal dysfunction. Cisplatin (Cis) induced profound renal dysfunction 72 h after administration to mice as shown by increased levels of blood urea nitrogen (BUN) and creatinine (CREA), which were dose-dependently attenuated by the CB$_2$ receptor agonist LEI-101 administered i.p. or by gavage (p.o.). Results are means ± SEM; *n* = 6 per group. * $P < 0.05$, significantly different from vehicle; # $P < 0.05$, significantly different from cisplatin.

LEI-101 reduces renal oxidative and nitrative stress

Reactive oxygen species play an important role in the pathology of cisplatin-induced tubular damage (Mukhopadhyay *et al.*, 2010 and 2012). Consistent with previous reports

Figure 4

The protective effects of The CB$_2$ receptor agonist LEI-101 against cisplatin (Cis)-induced nephrotoxicity are absent in CB$_2$R$^{-/-}$ mice. The renal dysfunction was aggravated in CB$_2$R$^{-/-}$ mice compared with CB$_2$R$^{+/+}$ littermates. LEI-101 alone had no effect on blood urea nitrogen (BUN) and creatinine (CREA) levels compared with the vehicle (Veh)-treated group and had no protective effect in CB$_2$R$^{-/-}$ mice against cisplatin-induced nephrotoxicity. Results are means ± SEM; *n* = 6 per group. * $P < 0.05$, significantly different from vehicle; # $P < 0.05$, significantly different from cisplatin.

CB$_2$R$^{+/+}$ CB$_2$R$^{-/-}$
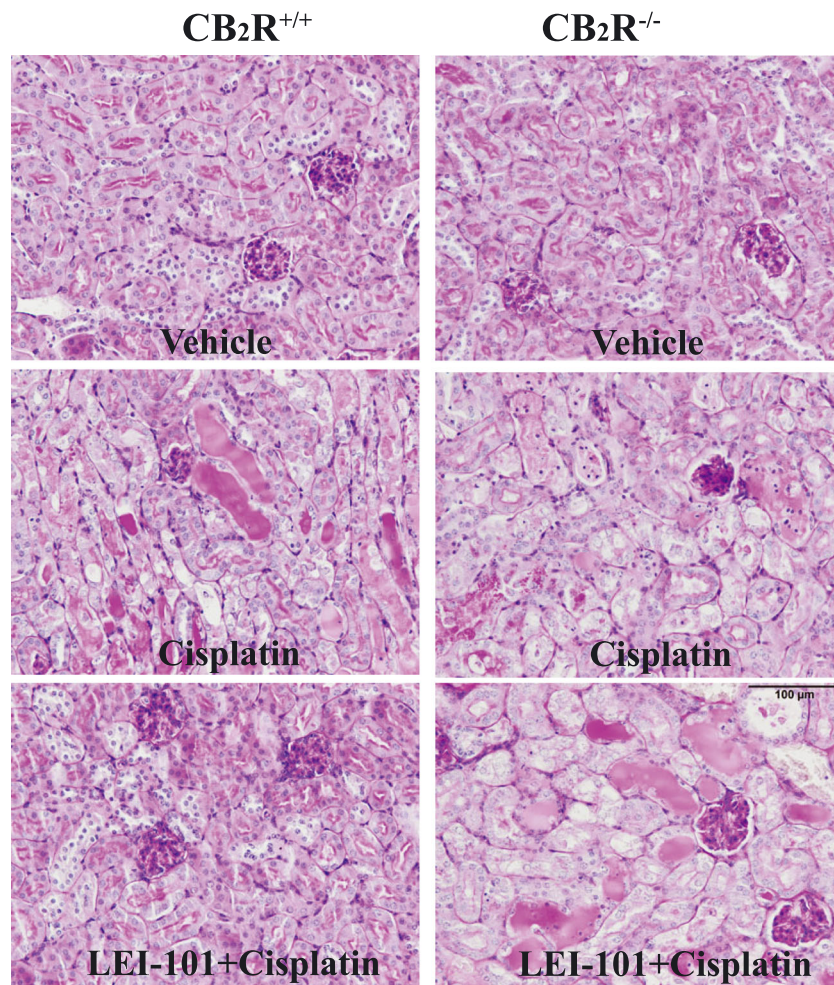
Figure 5

The CB$_2$ receptor agonist LEI-101 attenuates cisplatin-induced renal histopathological injury in wild type, but not in CB$_2$R$^{-/-}$ mice. Representative images of PAS staining show that cisplatin induced profound tubular injury 72 h after administration to mice, as evidenced by protein cast, vacuolation and desquamation of epithelial cells in the renal tubules of CB$_2$R$^{+/+}$ and CB$_2$R$^{-/-}$ mice. Note the decreased histopathological injury by LEI-101 (10 mg·kg^{-1} i.p.) in cisplatin-treated CB$_2$R$^{+/+}$, but not in CB$_2$R$^{-/-}$ mice, and enhanced injury in CB$_2$R$^{-/-}$ mice compared with their wild-types.

(Mukhopadhyay *et al.*, 2010, 2012), we also observed a marked increase in staining of 4-HNE, a marker of lipid peroxidation, and 3-nitrotyrosine, a marker of protein nitration, in the tubular cells of kidneys of cisplatin-treated animals (Figures 6 and 7). The mRNA levels of NADPH oxidase isoform 2 (NOX2) and 4 (NOX4), enzymes known to generate ROS, were also significantly upregulated 3 days after cisplatin injection (Figure 7). These cisplatin-induced pathological alterations were significantly reduced by LEI-101 (10 mg·kg^{-1}, i.p. or p.o.) pretreatment (Figures 5 and 8). Conversely, LEI-101 did not affect the oxidative and nitrative stress markers in CB$_2$R$^{-/-}$ mice treated with cisplatin.

LEI-101 attenuates renal inflammatory response and apoptotic cell death

Next, we tested whether LEI-101 was able to limit cisplatin-induced inflammation in the kidney. As observed in Figure 9, cisplatin increased the expression of mRNA for the

proinflammatory cytokines, TNF-α and IL-1β. Such an increase in proinflammatory markers was reversed by LEI-101 (10 mg·kg^{-1}, i.p. or p.o.).

Apoptotic cell death in the kidney following cisplatin-induced injury was evaluated by monitoring caspase 3/7 activity and DNA fragmentation (Figure 9). All these markers were significantly increased 3 days after cisplatin injection and largely attenuated by treatment with LEI-101 (10 mg·kg^{-1}, i.p. or p.o.).

Discussion

In this study, we have comprehensively characterized a novel CB$_2$ receptor ligand that may be used as a tool to interrogate CB$_2$ receptor biology in more detail. We have shown that LEI-101 is an orally bioavailable, largely peripherally restricted, selective CB$_2$ receptor partial agonist that does not
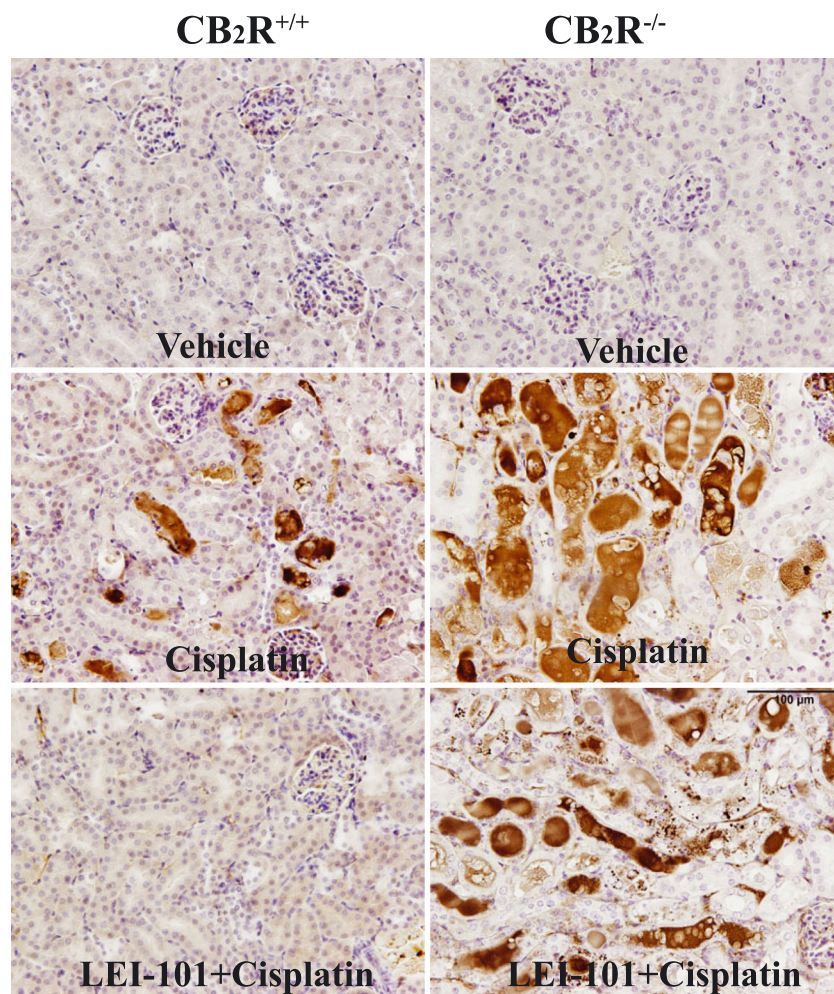
Figure 6

The CB_2 receptor agonist LEI-101 attenuates cisplatin-induced renal 3-nitrotyrosine (3-NT) formation in wild type, but not in $CB_2R^{-/-}$ mice. Representative images show marked accumulation of 3-NT (marker of protein nitration) predominantly in damaged renal proximal tubules (dark brown staining) 72 h after cisplatin administration to mice. Note the decreased renal 3-NT staining by LEI-101 (10 mg·kg^{-1} i.p.) in cisplatin-treated $CB_2R^{+/+}$, but not in $CB_2R^{-/-}$ mice, and enhanced staining in $CB_2R^{-/-}$ mice compared with their wild-types.

induce cannabimimetic effects in the mouse tetrad at high doses (up to 60 mg·kg^{-1} p.o.). This indicates that LEI-101 and its metabolites did not activate CB_1 receptors *in vivo* either directly, neither indirectly by raising endocannabinoid levels. LEI-101 significantly reversed cisplatin-induced nephropathy and attenuated inflammation and oxidative stress already at a dose of 3 mg·kg^{-1} p.o. and i.p. Importantly, this protective effect was mediated via CB_2 receptors, because it was not observed in $CB_2R^{-/-}$ animals. Currently, at least three CB_2 receptor agonists (GW81466, cannabinor and S-777469) have been tested in phase 2 clinical trials for pain and atopic dermatitis, albeit with a poor outcome (Dhopeshwarkar and Mackie, 2014). Peripherally restricted, mixed CB_1/ CB_2 receptor agonists induced severe CB_1 receptor-mediated cardiovascular and metabolic side effects leading to termination of the clinical trials (Pacher and Kunos, 2013). At this moment, the reasons for the failure to demonstrate clinical efficacy, for these CB_2 receptor agonists, are not exactly clear. One of them could be that the CB_2 receptor has not been properly validated for the disease under investigation, or that the clinical candidates could have been tested at the wrong stage of the disease. For example, once inflammation-induced tissue injury has progressed too far, CB_2 receptor agonists could be ineffective (Pacher and Mechoulam, 2011). In addition, the clinical candidates may not have reached the site of action. In fact, in chronic pain, the site of action of CB_2 receptor agonists as well as its mechanism of analgesic action is still under debate. At the moment, there is a lack of suitable biomarkers to correlate target engagement and modulation of CB_2 receptors with efficacy in chronic pain. It has been suggested that very low receptor occupancy of CB_1 receptors could be responsible, at least in part, for mediating the effect of commonly used 'selective' CB_2 receptor agonists in preclinical inflammatory pain models. Most previous *in vivo* studies of inflammation and tissue injury relied on the use of only very few CB_2 receptor agonists predominantly after i.p. administration, without characterization of their pharmacokinetics (Pacher and Mechoulam, 2011). In addition,
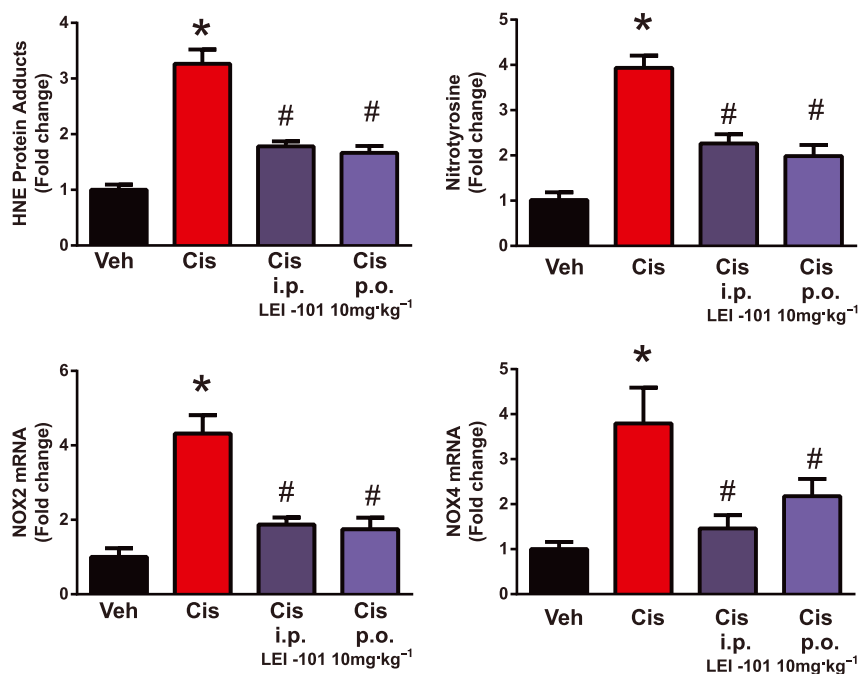
Figure 7

The CB$_2$ receptor agonist LEI-101 attenuates cisplatin-induced renal lipid peroxidation (as HNE adducts) and nitrotyrosine generation (upper panel). The lower panel shows overexpression of superoxide-generating enzymes NOX2 (gp91phox) and NOX4 (RENOX) after cisplatin. Cisplatin (Cis) induced marked increases in lipid peroxidation and ROS generation and in mRNA expression of NOX2 and NOX4 in the kidneys 72 h after administration to mice. These effects were attenuated by LE101 at 10 mg·kg^{-1} both upon i.p. and p.o. treatment (starting from 1.5 h before cisplatin injection). Results are means ± SEM; $n = 6$ per group; * $P < 0.05$, significantly different from vehicle; # $P < 0.05$, significantly different from cisplatin alone.

most of these studies either used a single dose of CB$_2$ receptor agonist or could not establish a clear dose-dependence, because of the biphasic response. Given the emerging evidence that supports the pro-inflammatory role of peripheral CB$_1$ receptors, such a biphasic response could be attributed (at least in part) to activation of CB$_1$ receptors at higher doses (Mukhopadhyay *et al.*, 2010; Jourdan *et al.*, 2014). Surprising, *in vivo* evidence for the latter has also been presented at a recent meeting on (endo)cannabinoids (Wiley *et al.*, 2014).

In this study, we demonstrate that LEI-101 dose-dependently attenuated cisplatin-induced kidney injury from 3 mg·kg^{-1} both p.o. or i.p., without inducing cannabimimetic effects in the mouse tetrad or drug discrimination paradigm up to 60 mg·kg^{-1} p.o. Thus, the compound and its metabolites did not activate CB$_1$ receptors *in vivo*, even at high concentrations. There is accumulating evidence on the opposing role of CB$_1$ and CB$_2$ receptors in kidney injury. It appears that while activation of CB$_1$ receptors promotes kidney injury, CB$_2$ receptor activation attenuates kidney inflammation and injury in rodent models of diabetic and tubular nephropathy (Barutta *et al.*, 2010, 2011 and 2014; Mukhopadhyay *et al.*, 2010; Jourdan *et al.*, 2014).

Cisplatin is a cornerstone in the treatment for many different types of cancer, but its use is dose-limited by the occurrence of nephropathy as a major adverse side effect. The exact mechanisms of cisplatin's renal toxicity are still not completely understood, and currently no effective medication is available to prevent this problem (Pabla and Dong, 2008).

Cisplatin accumulates in renal tubular cells because of its metabolism, reaching toxic levels (Pabla and Dong, 2008). Consistent with our recent results, enhanced superoxide production by mitochondria and various cellular NADPH oxidases, suhc as NOX2 and NOX4 (Mukhopadhyay *et al.*, 2010a,b, 2012), overproduction of NO via activation of inducible nitric oxide synthase, and increased peroxynitrite generation formed from diffusion limited reaction of nitric oxide with superoxide (Pacher *et al.*, 2007), and consequent lipid peroxidation and protein nitration have all been implicated in renal tubular cell death induced by cisplatin. Recent studies have also suggested a central role for the nuclear enzyme poly(ADP-ribose)polymerase 1 (PARP-1), which is activated by oxidative DNA injury and leads to depletion of cellular NAD+ and ATP stores, not only in cisplatin-induced tubular cell death but also in the pathological kidney inflammatory response (Mukhopadhyay *et al.*, 2011). In addition to damaging renal proximal tubular cells, cisplatin also injures renal endothelial cells leading to endothelial dysfunction and secondary functional hypoxia, amplifying the tubular cell death. These pathological events also trigger secondary inflammatory response in the kidneys, largely orchestrated by infiltrating immune cells. The pro-inflammatory cytokine TNF-α is produced by various cell types, including infiltrating immune cells, activated endothelium and renal parenchymal cells and appears to play a central role in further fuelling the inflammatory response and consequent tissue injury (Pabla and Dong, 2008). CB$_2$ receptor agonists appear to protect against cisplatin-induced kidney injury by attenuating
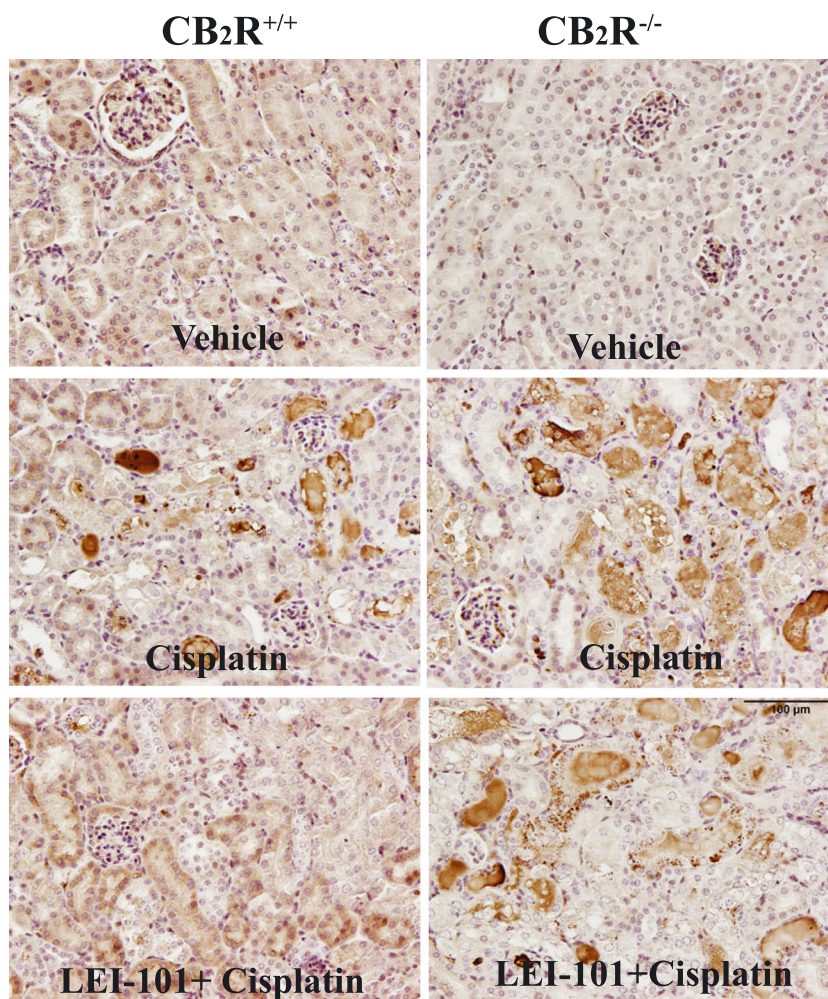
Figure 8

The CB$_2$ receptor agonist LEI-101 attenuates cisplatin (Cis)-induced renal HNE adducts formation in wild type, but not in CB$_2$R$^{-/-}$ mice. Representative images show marked accumulation of HNE adducts predominantly in damaged renal proximal tubules (dark brown staining) 72 h after cisplatin administration to mice. Note the decreased renal HNE staining by LEI-101 (10 mg·kg^{-1} i.p.) in cisplatin-treated CB$_2$R$^{+/+}$, but not in CB$_2$R$^{-/-}$ mice, and enhanced staining in CB$_2$R$^{-/-}$ mice compared with their wild-types.

endothelial cell activation and pro-inflammatory response, as well as by limiting inflammatory cell chemotaxis, adhesion to activated endothelium, transmigration to parenchyma and activation at the site of injury (Mukhopadhyay *et al.*, 2010a; Pacher and Mechoulam, 2011; Horvath *et al.*, 2012). In experimental models of diabetic nephropathy, in which initially the glomerular injury dominates the pathology, activation of CB$_2$ receptors protected glomerular podocytes, which wrap around the capillaries of the glomerulus and play critical role in maintaining the normal kidney function and filtration (Barutta *et al.*, 2011, 2014). In addition, CB$_2$ receptor agonists may attenuate monocyte chemokine signalling, which is also similar in the cisplatin-induced nephropathy model (Barutta *et al.*, 2011, 2014). Consistent with the importance of CB$_2$ receptors in kidney disease in this study, we demonstrate marked dose-dependent attenuation of kidney inflammation, oxidative stress and tubular damage induced by cisplatin in mice by LEI-101.

In summary, we have demonstrated, in this study, that LEI-101 is a selective, largely peripherally restricted, orally available agonist at CB$_2$ receptors and could be an excellent

tool to study the role of these receptors in inflammatory and other immunity-related diseases (Chiurchiù *et al.*, 2015). Of note, LEI-101 behaved as a partial CB$_2$ receptor agonist in the GTPγS-assay and β-arrestin recruitment assay, but as a full agonist in the cAMP-assay, a more downstream signalling pathway. It is not uncommon that some ligands behave as partial agonists at the G-protein activation while fully activating downstream signalling pathways. Whether a partial or full agonism was required in case of LEI-101 to observe *in vivo* efficacy in the cisplatin-induced nephrotoxicity model cannot be concluded at this moment, but would be very interesting to explore in future studies.

Our results also indicate that selective CB$_2$ receptor (partial) agonists could be excellent targets for treating kidney disease associated with oxidative stress and inflammation. This observation, coupled with the recent demonstration of the protective effects of a novel series of CB$_2$ receptor agonists in various kidney injury/fibrosis models (Adams *et al.*, 2014), further substantiates the strong translational potential of selective CB$_2$ receptor agonists in inflammatory kidney disease.
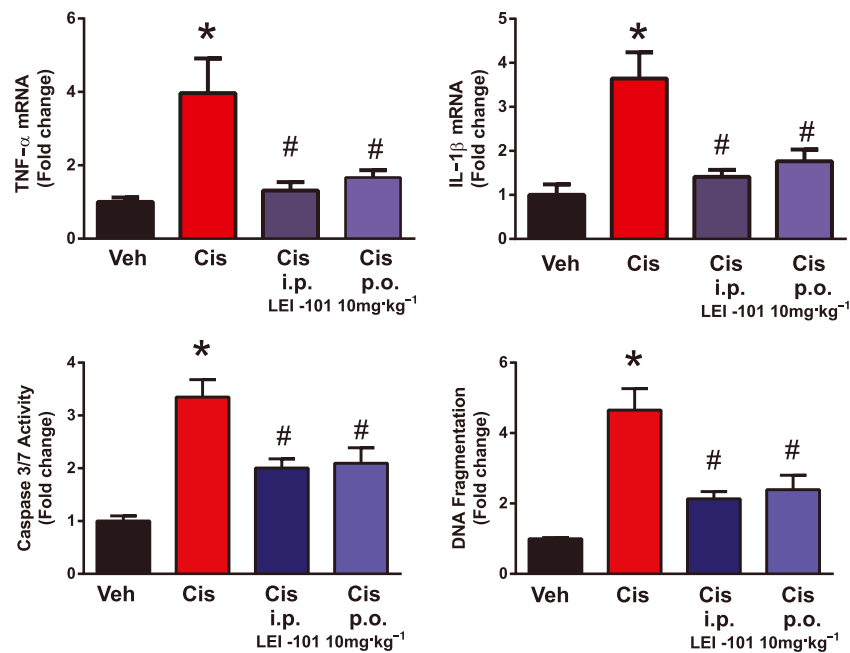
Figure 9

The CB$_2$ receptor agonist LEI-101 attenuates cisplatin-induced renal inflammation and cell death. Top panel: cisplatin (Cis) induced increases in renal TNFα and IL-1β mRNA expression, which were attenuated by LE-101 at 10 mg·kg^{-1}, both i.p. and p.o. treatment. Bottom panel: cell death in the kidneys was evaluated by caspase 3/7 activity and DNA fragmentation assay. Both markers of cisplatin-induced cell death in the kidneys were attenuated by LEI-101 treatment starting at 1.5 h before cisplatin injection. Results are means ± SEM; n = 5–8 per group; *$P < 0.05$, significantly different from vehicle; # $P < 0.05$, significantly different from cisplatin alone.

Conflict of interest

None

Author contributions

P.M., M.B., K.E., Z.C., F.F., B.I-J., J.W., N.G., T.H., M.R., M.S. designed, performed and analysed experiments: P.M., L.H., G.K., M.M., A.L., P.P., M.vd.S. designed and analysed the experiments and wrote the paper:

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