

Research Article



## Effects of Environmental Factors on Soluble Expression of a Humanized Anti-TNF- $\alpha$ scFv Antibody in *Escherichia coli*

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### Abstract

**Purpose:** The bacterial cultivation conditions for obtaining anti-TNF- $\alpha$  single chain variable fragment (scFv) antibody as the soluble product in *E. coli* was investigated.

**Methods:** To avoid the production of inclusion bodies, the effects of lactose, IPTG, incubation time, temperature, shaking protocol, medium additives (Mg<sup>2+</sup>, sucrose), pH, osmotic and heat shocks were examined. Samples from bacterial growth conditions with promising results of soluble expression of GST-hD2 scFv were affinity purified and quantified by SDS-PAGE and image processing for further evaluation.

**Results:** The results showed that cultivation in LB medium under induction by low concentrations of lactose and incubation at 10 °C led to partial solubilization of the expressed anti-TNF- $\alpha$  scFv (GST-hD2). Other variables which showed promising increase in soluble expression of GST-hD2 were osmotic shock and addition of magnesium chloride. Furthermore, addition of sucrose to medium suppressed the expression of scFv completely. The other finding was that the addition of sorbitol decreased the growth rate of bacteria.

**Conclusion:** It can be concluded that low cultivation temperature in the presence of low amount of inducer under a long incubation time or addition of magnesium chloride are the most effective environmental factors studied for obtaining the maximum solubilization of GST-hD2 recombinant protein.

### Introduction

Anti-TNF- $\alpha$  antibodies have reported as promising factors in the treatment of some diseases like rheumatoid arthritis and Crohn's disease.<sup>1,2</sup> But the possibility of long-term utilization of antibodies, depends on the rate of toxicity in body and the production cost.<sup>2</sup> The total cost of antibody injection per year for each patient is markedly high and it can be considered as an obstacle for its use.<sup>2</sup> To overcome this dilemma, recombinant DNA technology along with possibility of cloning and the expression of monoclonal antibody fragments in prokaryotes, have revealed a new promising platform for the development of more affordable drugs.<sup>3</sup> For example, the variable regions of heavy (VH) and light (VL) fragments of monoclonal antibody can be coupled by a flexible peptide linker to form a polypeptide known as single chain fragment variable (scFv).<sup>4,5</sup> Although the scFv antibodies can be produced in both prokaryotic and eukaryotic hosts, however, the prokaryotic systems can be economic and easy to handle, compared to eukaryotic biological systems.<sup>6</sup> However, the expression of scFv antibodies in bacterial hosts sometimes leads to the aggregation of recombinant proteins due to partial folding. These aggregates called inclusion bodies are

needed to be refolded into their proper structure. Refolding of inclusion body to a soluble protein is costly, laborious and time consuming.<sup>6-8</sup> There are several approaches to avoid the production of inclusion body in bacteria such as optimization of the cultivation temperature, type and the concentration of inducers, medium additives, and utilization of heat and osmotic shocks.<sup>9-13</sup> *Escherichia coli* is one of the common host prokaryotic organisms for the production of recombinant proteins because of facile genetic engineering and optimization of recombinant protein expression.<sup>14,15</sup> Total amount of recombinant protein has an important role in its degradation rate. It has revealed that when the protein content exceeds 50 percent of the total protein, it reduces the opportunity for proper folding of the product.<sup>6</sup> Expression of scFv in *E. coli* cytoplasm in numerous cases have shown leads to insoluble inclusion bodies, and therefore some strategies were adopted to achieve soluble expression of the target protein.<sup>16,17</sup> Using low concentration of the inducers like IPTG and lactose reduces the rate of recombinant protein production in the cells.<sup>6</sup> The soluble expression rate of scFv in bacteria can vary based on medium and cultivation conditions.<sup>18-20</sup>

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The presence of some additives like sorbitol and sodium chloride in the medium triggers osmotic shock, and the high incubation temperature for a short period of time produces a heat shock response.<sup>12</sup> Either osmotic or heat shocks cause increased manufacturing of chaperone molecules which play essential roles in proper folding of *de novo* synthesized proteins in cells at harsh conditions and eventually catalyze proper folding of proteins.<sup>12,18</sup> Metal ions have been described as solubilizing factors for some recombinant proteins.<sup>20</sup> Especially magnesium solubilization activity is proposed to be higher than any other metal ions.<sup>20</sup> Sucrose has been shown to increase the solubility of scFv molecules ranging from 15 to 150 folds.<sup>9</sup> Some studies have proved that the low temperatures (10 °C or lower) increase the solubility of recombinant proteins which are otherwise expressed in the form of inclusion body when the incubation temperature is set to 15 and 20 °C.<sup>11,21</sup> As mentioned above, the expression of scFv antibodies in *E. coli* cytoplasm generally leads to the production of inclusion bodies,<sup>17,22-24</sup> so optimized growth conditions are required to achieve soluble expression of the target scFv. Therefore, in this study, we have examined the effects of temperature, shaking condition, concentration of inducers (lactose or IPTG), incubation time, pH, sucrose and Mg<sup>2+</sup> concentrations, and osmotic or heat shocks on the soluble expression of a 52 kD GST-fusion recombinant protein called GST-hD2 scFv in *Escherichia coli*. GST-hD2 is a humanized anti-TNF- $\alpha$  single chain variable fragment (scFv) antibody fused to GST protein, which is designed<sup>25</sup> using CDRs replacement strategy based on a murine scFv anti-TNF- $\alpha$  antibody.<sup>26</sup>

## Materials and Methods

### Bacterial strain, plasmid, and target protein

*E. coli* Origami (DE3) harboring recombinant plasmid pGEX-6P-1/hD2 was obtained generously from Biotechnology Research Center Tabriz (BRC, Iran). hD2 is a GST fusion protein cloned in pGEX-6P-1 and encodes a single chain fragment variable antibody against TNF- $\alpha$ .

### Optimization parameters

A single colony of the recombinant bacteria was cultivated overnight in LB medium containing 100  $\mu$ g mL<sup>-1</sup> ampicillin at 37 °C, while shaking at 175 rpm. Then, it was diluted in 1:100 ratio by 50 mL fresh medium in a 500 mL flask. To explore the effects of inducers, the induction of GST-hD2 protein expression was performed at final concentrations 0.1 and 1 mM for IPTG, or 0.03, 0.15, 0.3 and 0.6 mM for lactose in subculture grown at 37 °C while shaking at 180 rpm until an OD<sub>600</sub> = 0.6-0.8.

To study the effect of temperature and incubation time on scFv solubilization, the above mentioned cultures were incubated at 10, 20 and 37 °C, while samples were harvested at 0, 3, 6 and 18 h after induction. Also, at 10°C, the incubation time was extended further for 48 h

after the induction.<sup>10</sup> To examine the shaking effect, two different approaches were carried out. Subculture was grown in absence of inducer at 37 °C, while shaking at 200 rpm for 4 h. Then, the shaking speed was decreased to 50 rpm for 18 h and the samples were harvested at 0, 4 and 22 h after subculturing. In another procedure, starter culture and subculture were grown at 30 °C, 120 rpm. The subculture was induced by 0.1 mM IPTG and the cultivation condition was changed to 30 rpm at 25 °C for 15 h.<sup>27</sup> To assess the effects of MgCl<sub>2</sub> and sucrose on the soluble expression of GST-hD2, 0.1 and 10 mM of MgCl<sub>2</sub> and 0.4 M sucrose in final concentration were added to each subculture, respectively.<sup>9,20</sup> In this procedure, induction was achieved using 0.6 mM lactose and the cells were harvested 6 and 18 h after induction. The medium pH was adjusted at 5, 6, 7 or 8 and its effects on soluble expression of GST-hD2 were investigated.<sup>28</sup> To examine heat and osmotic shock responses, the study were performed according to the protocol by Oganessian et al<sup>12</sup> with some minor modification. Harvesting of cells were performed 3 and 15 h after induction.

### GST-hD2 expression and SDS-PAGE analyses

To harvest equal number of cells from different cultures, first the optical densities of the cultures were measured at 600 nm and then the adjusted volume withdrawn from each culture was equated to be 1/OD<sub>600</sub> mL (equal to 0.3 mg cell pellet). Then the cells were harvested by centrifugation at 13000 g for 3 min at room temperature. The bacterial pellet was resuspended in 60  $\mu$ L of lysis buffer (50 mM Tris (pH=8.0), 100 mM NaCl, 1.4 mM PMSF, 0.1% B-Mercaptoethanol, 1% Triton X-100) and kept for 30 min on ice. After three freeze-thaw cycles, the soluble fraction was isolated from insoluble fraction by centrifuging at 12000 g for 10 min at 4 °C. Then, to the soluble and insoluble fractions were added 20 and 80  $\mu$ L of phosphate saline buffer (PBS), respectively. Subsequently, to the samples was added 10  $\mu$ L of 2X sample solvent buffer (SDS 4%, Glycerol 20%, 2-mercaptoethanol 10%, Bromophenol blue 0.2%, Tris-Base 50 mM) and heated at 95 °C for 10 min. The expression of GST-hD2 fusion protein was investigated using SDS-PAGE analysis on 12% (V/V) polyacrylamide gel using an electrophoresis apparatus (Bio-Rad, USA) according to Sambrook and Russell instruction after staining by Coomassie Brilliant Blue G250.<sup>29</sup> The purified GST-hD2 from the previous study<sup>25</sup> called Ab in the current study and was used as a protein size marker in electrophoresis analyses.

### Purification and quantification of soluble GST-hD2

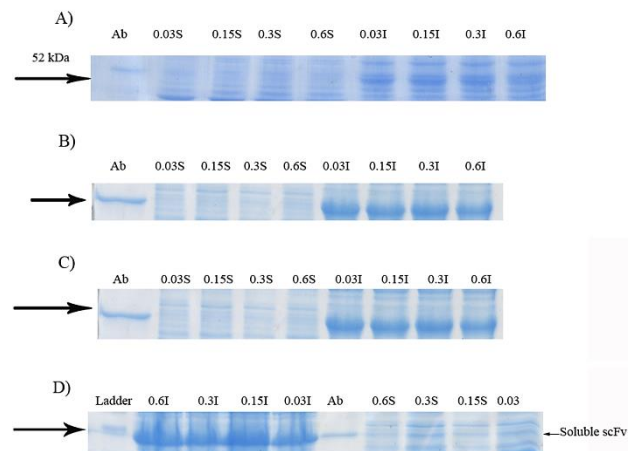
For purification of fusion GST-hD2, cells were collected from a 50 mL culture grown as outlined above by centrifugation at 7000 g for 15 min at 4 °C and then were resuspended in lysis buffer (7 mL lysis buffer for each gram of cell pellet) while incubating for 30 min on ice. For maximum cell disruption, 3 cycles of freeze-thaw steps were applied following by intermittently ultra-

sonication on ice for 30 s at 60 W with 30 s allowed for cooling for total of 5 min. Afterward lysate were centrifugation at 10,000 g for 20 min at 4 °C and supernatants was used for concentrating and affinity purification. Supernatant was loaded on 20 µL of Glutathione Sepharose 4B bead for 2 h at 4 °C. The bead was isolated by centrifugation at 2300 g for 3 min and washed 5 times with wash buffer (50 mM Tris, 100 mM NaCl, 0.1% 2-β mercaptoethanol). Presence of recombinant antibody in soluble fraction was analyzed using SDS-PAGE. The gels from SDS-PAGE analyses were scanned and their images were saved as TIF files. The relative concentrations of the soluble and purified recombinant antibody samples were quantified based on the band densities to that of known concentration of the standard Ab protein using image processing program called ImageJ (National Institute of Health, Bethesda, MD).

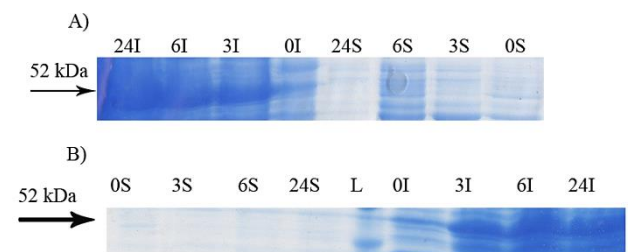
**Results**

SDS-PAGE analyses showed that the production of GST-hD2 as inclusion body was proportionally increased with the increase of IPTG or lactose concentrations as well as the incubation time as shown in Figure 1. Different concentrations of IPTG (0.1 and 1 mM) did not led to soluble expression of GST-hD2. The cultivation of bacteria at different concentrations of lactose ranging from 0.03 to 0.3 mM caused soluble expression of GST-hD2 after 48 h incubation at 10 °C. To optimize culture temperature, the experiments were performed at 20 and 37 °C, however, no sign of soluble expression of recombinant protein was detected (Figure 2). The best result for soluble expression of GST-hD2 was achieved when cells induced by 0.03 to 0.3 mM lactose and incubated at 10 °C for 48 h. Analyzing the soluble fraction obtained from growing the cells in the presence of 0.3 mM lactose as the inducer for 48 h at 10 °C showed that the amount of GST-hD2 expressed as soluble protein was 26.15 µg per one g of bacteria (Figures 3 and 4). Soluble expression of the recombinant protein was not observed after reaching to the OD<sub>600</sub> of 0.6-0.8 in bacterial culture with no added inducers (Figure 5). Different shaking speed protocols had no effect on solubilizing the recombinant protein (Figure 6). The various concentrations of magnesium chloride caused the production of slight amount of the recombinant protein. This finding was verified by affinity purification experiment using GSH-Sepharose bead. As shown in Figures 4 and 7, GST-hD2 was observed in the samples prepared from the soluble fraction of cultures grown in the presence of 10 mM magnesium chloride at the levels of 25.21 and 13.92 µg per g bacteria after 6 and 18 h induction times, respectively. By addition of 0.4 M sucrose into the culture media, the expression of recombinant scFv antibody (GST-hD2) was suppressed totally and even the aggregate form of the antibody was not observed as shown in Figure 8 compared to the control (i.e., in the absence of sucrose). The solubility of the recombinant

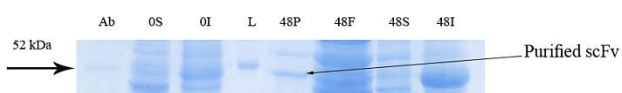
protein did not show any remarkable changes when the pH was varied from 5 to 8 (Figure 9). Sorbitol at final concentration of 0.5 M in the LB medium caused diminished growth rate of cells in subculturing step. A faint band parallel to our standard size marker protein of GST-hD2 was detected 3 h after induction in all shock treatments. According to the results shown in Figures 10 and 11, applying the affinity purification step on all types of shock experiments (osmotic shock, osmotic shock plus sorbitol, heat shock, heat shock plus sorbitol) revealed that only slight soluble expression of the recombinant antibody was achieved (5.78 µg per g bacteria) under osmotic shock treatment after 3 h incubation (Figures 4 and 11).



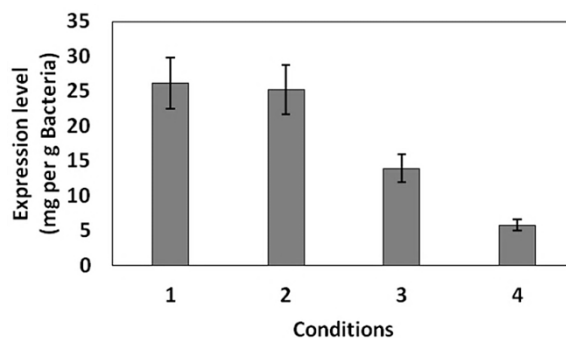
**Figure 1.** Effect of cultivations at the presence of different concentrations of lactose on soluble expression of GST-hD2. Proteins were resolved on SDS-PAGE prepared using 12% acrylamide and visualized by Coomassie Brilliant Blue G250<sup>29</sup>. Cells grown at 10 °C. The digital part of the alphanumerical codes above the lanes show the concentrations of lactose. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. Gels in panels A, B, C and D show the results for 3, 6, 24 and 48 h inductions, respectively.



**Figure 2.** Effect of cultivations at different incubation time on soluble expression of GST-hD2. The cells were induced by 0.1 mM IPTG at 20 °C (panel A) and 37 °C (Panel B). The digital part of the alphanumerical codes above the lanes show incubation times after induction. (L), (S) and (I) symbols of the code denote 50 kDa ladder, soluble and insoluble fractions, respectively.



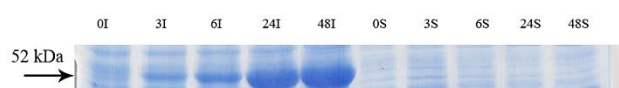
**Figure 3.** Further assessment of soluble expression of GST-hd2 by applying affinity purification step to the soluble fraction prepared from the culture grown at 10 °C and induced by 0.3 mM lactose (extension to experiment shown in panel D of Figure 1). The digital part of the alphanumerical codes above the lanes show incubation times after induction. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. L, P and F represent respectively a 50 kDa ladder protein, purified GST-hd2 and flowthrough.



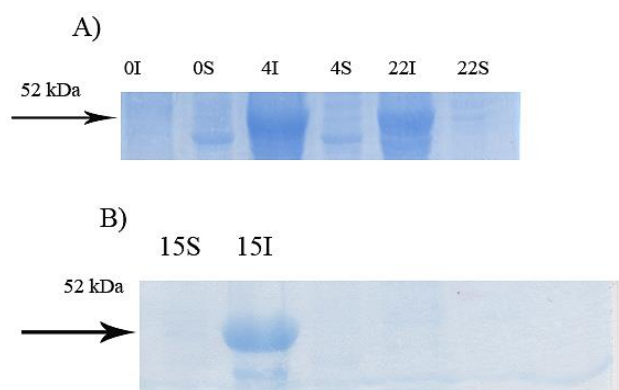
**Figure 4.** Levels of the soluble GST-hd2 under different conditions explained in Table 1.

**Table 1.** Growth conditions where the soluble expression of GST-hd2 was detected.

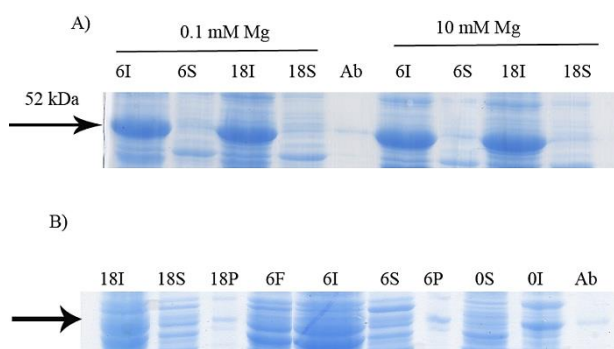
Condition number	Condition	Figure showing the results
1	Induction by 0.03 to 0.3 mM lactose and Incubation for 48 h at 10 °C	1.D
2	Addition 10 mM magnesium and Induction by 0.6 mM lactose and Incubation for 6 at 20 °C	7
3	Addition 10 mM magnesium and Induction by 0.6 mM lactose and Incubation for 15 h at 20 °C	7
4	Osmotic shocks (0.5 M NaCl and 1 mM glycinebetaine) and induction by 0.3 mM IPTG for 3 h at 20 °C	11



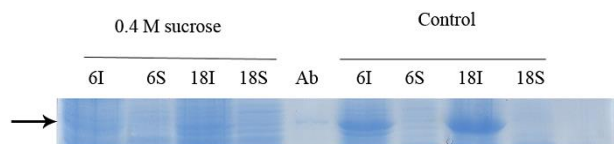
**Figure 5.** Samples from culture grown in the absence of inducer at 10 °C at 0, 3, 6, 24 and 48 h after reaching to OD<sub>600</sub>=0.6. The digital part of the alphanumerical codes above the lanes show incubation times. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively.



**Figure 6.** Effect of shaking on soluble expression of GST-hd2. The digital part of the alphanumerical codes above the lanes show incubation times after induction. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. **A** Subculture in absence of inducer and cultivated at 37 °C, 200 rpm for 4 h then, the shaking speed was decreased to 50 rpm. **B** starter culture and subculture were grown at 30 °C, 120 rpm. The subculture was induced by 0.1 mM IPTG and the cultivation condition was changed to 30 rpm at 25 °C.

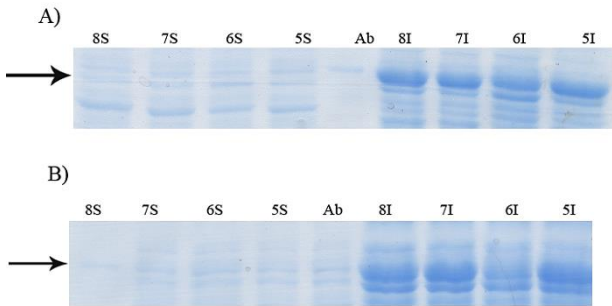


**Figure 7.** Effect of Mg<sup>2+</sup> on soluble expression of GST-hd2. The cells were induced by 0.6 mM lactose at 20 °C. The digital part of the alphanumerical codes above the lanes show incubation times after induction. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. **A** Magnesium chloride was used at 0.1 and 10 M concentrations. **B** Further assessment of soluble expression of GST-hd2 by applying affinity purification step to the soluble fraction prepared from the culture grown at 20 °C and induced by 0.6 mM lactose in the presence of magnesium chloride in 10 mM final concentration. P and F represent purified GST-hd2 and Flowthrough respectively.

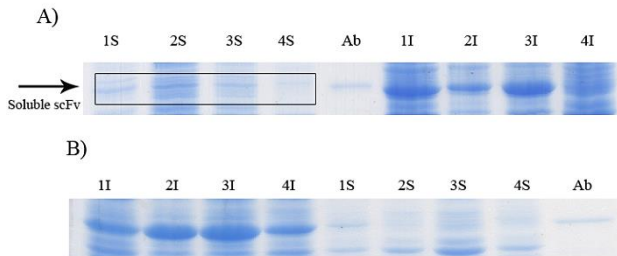


**Figure 8.** Effect of sucrose on soluble expression of GST-hd2. Sucrose was used at 0.4 M concentration. The culture was induced with 0.6 mM lactose at 20 °C. The digital part of the alphanumerical codes above the lanes show incubation times after induction. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. Control experiment was performed without addition of sucrose in subculture process.

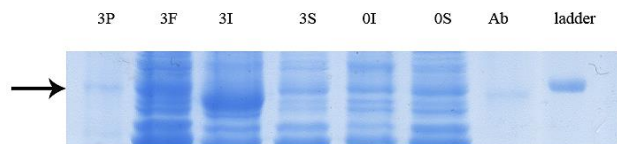




**Figure 9.** Effect of pH on soluble expression of GST-hD2 at 20°C using 0.6 mM lactose as the inducer. The incubation times were 6 (panel A) and 18 h (panel B) after induction. The digital part of the alphanumeric codes above the lanes show pH of the medium. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively.



**Figure 10.** Effect of osmotic and heat shocks on soluble expression of GST-hD2 at 20 °C. The incubation times were 3 (panel A) and 15 h (panel B) after induction. The digital part of the alphanumeric codes above the lanes are as follow, 1: Osmotic shock, 2: Heat shock, 3: Osmotic shock plus sorbitol, 4: Heat shock with sorbitol. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively.



**Figure 11.** Further assessment of soluble expression of GST-hD2 by applying affinity purification step to the soluble fraction prepared from osmotic shocks (0.5 M NaCl and 1mM glycinebetaine) (extension to experiment shown in panel A of Figure 10). The digital part of the alphanumeric codes above the lanes show incubation times after induction. (S) and (I) symbols of the code denote soluble and insoluble fractions, respectively. P and F represent respectively, purified GST-hD2 and flowthrough.

**Discussion**

The correct, active and soluble expression of scFv antibodies and prevention of inclusion body formation in cytoplasm of *E. coli* have been the focus of many investigations.<sup>17,21,24</sup> Inclusion bodies are misfolded proteins due to inefficient interaction between nascent protein and chaperone molecules which causes unsuitable folding of protein.<sup>30</sup> Some protocols have

been developed based on environmental modifications to produce the soluble form of the expressed proteins in *E. coli*.<sup>6,10,13</sup> In our previous study, no soluble expression of GST-hD2 was observed in cytoplasm when the standard expression methods were applied.<sup>29</sup> Very low concentration of inducers has shown to be a good candidate for promoting soluble expression of scFv antibody.<sup>31</sup> The results illustrated in Figure 1D show that the extended incubation time of 48 h in the presence of different concentrations of lactose ranging from 0.03 to 0.3 mM at 10 °C was resulted in soluble expression of GST-hD2. This may be the result of slow production rate of scFv antibody at such low temperature and low inducer concentrations providing enough time for the proper folding of the recombinant protein as reported by other investigators as well.<sup>9,18,20,32</sup> Soluble expression of GST-hD2 at 20 and 37 °C which were induced by IPTG was not observed (Figure 2). Raising the shaking speed led to increased aeration and growth rate of bacteria and, finally to the increased amount of recombinant protein.<sup>33</sup> Therefore, the effect of shaking speed on solubilization of GST-hD2 was examined. However, different shaking speed protocols studied in the current work had no effect on soluble expression of GST-hD2 (Figure 6). Magnesium ion has been used to solubilize recombinant proteins in the study of Yang et al.<sup>20</sup> This is probably due to cellular mechanism and magnesium function as a cofactor in folding of recombinant protein. In a similar manner, we also observed beneficial effect of addition of magnesium chloride (0.1 and 10 mM) on soluble expression of GST-hD2 as shown in Figure 7. In our investigation, sucrose suppressed totally the expression of scFv antibody and the recombinant protein was not detectable in either of soluble and insoluble fractions (Figure 8). This finding is consistent with that of the Heo et al. which described a significant decrease in production of insoluble scFv antibody when sucrose was added to the medium.<sup>23</sup> Further study are required to understand the mechanism of suppression of inclusion body by sucrose. Based on our results, pH did not affect the solubility of the recombinant protein at the studied pH range (Figure 9). But some previous studies have shown that pH can affect the solubility rate of the recombinant proteins, this controversy can be due to the differences of the target recombinant proteins.<sup>28,35</sup> Osmotic or heat shocks lead to production of heat shock protein (HSP) and also uptake of osmolytes molecules like glycine betaine (as chemical chaperone) which stabilize native protein and probably assist in correct, active and soluble folding of proteins.<sup>12</sup> We showed an slight amount of soluble recombinant antibody production 3 h after induction in osmotic shock condition (NaCl 0.5 M, glycine betaine 0.1 mM) which can be attributed to the combined effect of osmotic shock and the role of glycine betaine as a chemical chaperone.<sup>12,19</sup> One may deduce that both increased concentration of chaperones due to high concentration of NaCl and chaperone like function of glycine betaine may assist the correct folding of scFv antibody.

## Conclusion

In conclusion, our study described the role of temperature, type and concentration of inducer, incubation time, pH, Mg<sup>2+</sup>, sucrose, heat and osmotic shocks on soluble expression of GST-hD2 recombinant protein. The results showed that the induction by 0.03 to 0.3 mM lactose for 48 h at 10 °C or addition of magnesium chloride into medium can demonstrate solubilizing effect on expression of scFv antibody. In general, incubation for 48 h induction by 0.03 to 0.3 mM lactose at low temperature was more effective than incubation time and different concentration of inducers or cultivation without inducer.

Also slight soluble GST-hD2 expression was detected by applying the osmotic shock protocol. Based on experiments mentioned above it is concluded that, the proper folding of GST-hD2 was under the influence the traffic of nascent protein expression and other factor like chaperone molecule which assist correct folding of nascent protein. As the levels of soluble expression of the target protein under the studied conditions were not extremely appropriate for large scale protein production, other strategies such as co-expression of chaperone molecules need to be evaluated for further improvement.<sup>11,17,23</sup>

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## Ethical Issues

Not applicable.

## Conflict of Interest

The authors declare that they have no conflict of interest.

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