

Complete Genome Sequence of Porcine Parvovirus 2 Recovered from Swine Sera

F. S. Campos,^a M. Kluge,^a A. C. Franco,^a A. Giongo,^b F. P. Valdez,^c T. M. Saddi,^d W. M. E. D. Brito,^d P. M. Roehé^a

Laboratório de Virologia, Instituto de Ciências Básicas da Saúde, Universidade Federal do Rio Grande do Sul, Porto Alegre, Rio Grande do Sul, Brazil^a; Instituto do Petróleo e dos Recursos Naturais, Pontifícia Universidade Católica do Rio Grande do Sul, Porto Alegre, Rio Grande do Sul, Brazil^b; Pontifícia Universidade Católica do Rio Grande do Sul, Faculdade de Biociências, Porto Alegre, Rio Grande do Sul, Brazil^c; Instituto de Patologia Tropical e Saúde Pública, Universidade Federal de Goiás, Goiânia, Goiás, Brazil^d

A complete genomic sequence of porcine parvovirus 2 (PPV-2) was detected by viral metagenome analysis on swine sera. A phylogenetic analysis of this genome reveals that it is highly similar to previously reported North American PPV-2 genomes. The complete PPV-2 sequence is 5,426 nucleotides long.

Received 25 November 2015 Accepted 7 December 2015 Published 28 January 2016

Citation Campos FS, Kluge M, Franco AC, Giongo A, Valdez FP, Saddi TM, Brito WMED, Roehé PM. 2016. Complete genome sequence of porcine parvovirus 2 recovered from swine sera. *Genome Announc* 4(1):e01627-15. doi:10.1128/genomeA.01627-15.

Copyright © 2016 Campos et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 3.0 Unported license](https://creativecommons.org/licenses/by/3.0/).

Address correspondence to F. S. Campos, camposvet@gmail.com.

Porcine parvovirus (PPV) is a small nonenveloped virus associated with reproductive problems and is endemic in virtually all swine-producing regions worldwide (1). The viral genome is a single-stranded DNA molecule of approximately 5 kb (2). Porcine parvovirus 2 (PPV-2) was first detected in 2001 and is probably distributed worldwide; however, links between PPV-2 and disease remain unclear (3).

Here, the complete sequence of a PPV-2 genome, identified in sera of Brazilian sows, is reported. Sera from ten clinically healthy, 5-months-old gilts (*Sus scrofa*) of the Large White breed were collected in Senador Canedo, Goiás, Brazil. Samples were pooled and viral particles were pelleted by centrifugation. The pellet was resuspended in Tris-EDTA and treated with DNase I (Roche) and RNase (Invitrogen). Viral genomes were extracted with a commercial kit (PureLink Viral RNA/DNA minikit) and amplified by random PCR (4). The products were purified and sequenced on an Ion Torrent platform with a 316 Ion chip. A total of 261,836 raw reads were generated and reduced to 126,661 after trimming with the Geneious version 8.0.2. From these, 18,862 reads were filtered by closest matching, where 95% of those fit within the family *Parvoviridae*. The reads were *de novo* assembled, with a mean coverage of at least 347×. A contig of 5,426 nucleotides (nt) comprised the full Brazilian PPV-2 (BrPPV-2) genome. Phylogenetic analyses performed with MEGA version 6.0 revealed that BrPPV-2 is closely related to genomes of North American PPV-2, with an overall 98.3 to 98.7% nt identity. The complete sequence of BrPPV-2 reveals two putative open reading frames (ORF): ORF1 (1,985 nt) and ORF2 (3,098 nt). In addition, two 14 nt-long

palindromic sequences were identified, one at each extremity of the genome.

Nucleotide sequence accession number. The GenBank accession number is [KM926355](https://www.ncbi.nlm.nih.gov/nuclseq/KM926355).

ACKNOWLEDGMENTS

This work was supported by the FAPERGS, CAPES, CNPq, and FINEP. F.S.C. is a post-doc grantee from FAPERGS.

FUNDING INFORMATION

MCTI | Conselho Nacional de Desenvolvimento Científico e Tecnológico (CNPq) provided funding to Ana Claudia Franco. Fundação de Amparo à Pesquisa do Estado do Rio Grande do Sul (FAPERGS) provided funding to Fabrício Souza Campos. MCTI | Financiadora de Estudos e Projetos (FINEP) provided funding to Paulo Michel Roehé.

REFERENCES

- Mengeling WL, Lager KM, Vorwald AC. 2000. The effect of porcine parvovirus and porcine reproductive and respiratory syndrome virus on porcine reproductive performance. *Anim Reprod Sci* 60–61:199–210. [http://dx.doi.org/10.1016/S0378-4320\(00\)00135-4](http://dx.doi.org/10.1016/S0378-4320(00)00135-4).
- Bergeron J, Menezes J, Tijssen P. 1993. Genomic organization and mapping of transcription and translation products of the NADL-2 strain of porcine parvovirus. *Virology* 197:86–98. <http://dx.doi.org/10.1006/viro.1993.1569>.
- Saekhow P, Mawatari T, Ikeda H. 2014. Coexistence of multiple strains of porcine parvovirus 2 in pig farms. *Microbiol Immunol* 58:382–387. <http://dx.doi.org/10.1111/1348-0421.12159>.
- Allander T, Tammi MT, Eriksson M, Bjerkner A, Tiveljung-Lindell A, Andersson B. 2005. Cloning of a human parvovirus by molecular screening of respiratory tract samples. *Proc Natl Acad Sci USA* 102:12891–12896. <http://dx.doi.org/10.1073/pnas.0504666102>.