OPINION PAPER



# Alternatives to the 'water oxidation pathway' of biological ozone formation

Arnold N. Onyango<sup>1</sup>

Received: 18 April 2015 /Accepted: 5 June 2015 /Published online: 16 June 2015  $\oslash$  Springer-Verlag Berlin Heidelberg 2015

Abstract Recent studies have shown that ozone  $(O_3)$  is endogenously generated in living tissues, where it makes both positive and negative physiological contributions. A pathway for the formation of both  $O_3$  and hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) was previously proposed, beginning with the antibody or amino acid-catalyzed oxidation of water by singlet oxygen  $(^1O_2)$  to form hydrogen trioxide  $(H<sub>2</sub>O<sub>3</sub>)$  as a key intermediate. A key pillar of this hypothesis is that some of the  $H_2O_2$  molecules incorporate waterderived oxygen atoms. However,  $H_2O_3$  decomposes extremely readily in water to form  ${}^{1}O_{2}$  and water, rather than  $O_{3}$  and  $H_{2}O_{2}$ . This article highlights key literature indicating that the oxidation of organic molecules such as the amino acids methionine, tryptophan, histidine, and cysteine by  ${}^{1}O_{2}$  is involved in ozone formation. Based on this, an alternative hypothesis for ozone formation is developed involving a further reaction of singlet oxygen with various oxidized organic intermediates.  $H_2O_2$  having water-derived oxygen atoms is subsequently formed during ozone decomposition in water by known reactions.

Keywords Ozone . Singlet oxygen . Hydrogen peroxide . Amino acid oxidation . Baeyer-Villiger oxidation

#### Introduction

Ozone gas  $(O_3, {}^{6+}O = O-O^{6-})$  is an important component of the stratosphere where it protects organisms on earth from the

 $\boxtimes$  Arnold N. Onyango arnold.onyango@jkuat.ac.ke most damaging wavelengths of solar radiation [[1\]](#page-6-0). On the other hand, tropospheric ozone may be harmful to the respiratory system [\[1\]](#page-6-0). Nevertheless, ozone gas finds some application in alternative medicine [\[2](#page-6-0)] as well as in water and wastewater treatment [\[3](#page-6-0)].

Over a decade ago, Wentworth et al. [\[4\]](#page-6-0) reported that antibodies catalyze ozone formation in the presence of water and singlet oxygen  $({}^{1}O_{2})$ , partly based on the observed occurrence, under such conditions, of reactions that were thought to be unique to oxidation by ozone. These reactions included the conversion of indigo carmine to isatin sulfonic acid and the conversion of cholesterol to 3β-hydroxy-5-oxo-5,6 sechocholestan-6-al (secosterol aldehdye A) and 3β-hydroxy, 5β-hydroxy- B-norcholestane-6β-carboxaldehyde (secosterol aldehyde B). On the positive side, ozone generation in this manner contributes to bacterial killing by neutrophils [\[4](#page-6-0)]. Moreover, one of the best photodynamic therapy (PDT) strategies for cancer treatment involves the use of antibodies conjugated to photosensitizers [[5](#page-6-0)], and the anticancer effect of this type of PDT may partly be due to antibody-catalyzed ozone generation upon irradiation of the photosensitizers and formation of  ${}^{1}O_{2}$ . On the other hand, the secosterol aldehydes produced by cholesterol ozonolysis potentially contribute to the pathogenesis of disorders such as atherosclerosis and Alzheimer's disease [\[6,7\]](#page-6-0).

The concept of ozone generation in biological systems generated reasonable skepticism, and experimental results were obtained proving that conversion of indigo carmine to isatin sulfonic acid, or the formation of cholesterol secosterol aldehydes could also be mediated by oxidants other than ozone [\[8](#page-6-0),[9\]](#page-6-0). For example, it was demonstrated that cholesterol 5 hydroperoxide, obtained by the reaction of cholesterol with singlet oxygen, undergoes Hock cleavage to form the secosterol aldehydes [[9\]](#page-6-0). However, the singlet oxygen-Hock cleavage pathway predominantly generates secosterol aldehyde B [\[7](#page-6-0),[9\]](#page-6-0), while the ozonolysis pathway predominantly

<sup>1</sup> Department of Food Science and Technology, Jomo Kenyatta University of Agriculture and Technology, P. O. Box 62000, 00200 Nairobi, Kenya

generates secosterol aldehyde A [\[7](#page-6-0)]. The fact that the latter is the major secosterol aldehyde in atherosclerotic tissues [\[7\]](#page-6-0) and is also produced in vitro by neutrophils [[10\]](#page-6-0) supports the occurrence of ozone-mediated cholesterol oxidation in vivo [\[11\]](#page-6-0). More recently, the formation of endogenous ozone by plant leaves was proved directly by GC-MS-SIM [[1\]](#page-6-0). It was also reported that the antibiotic activity of a number of compounds including trans-resveratrol, salicylic acid, and cinnamic acid involves the endogenous formation of formaldehyde and ozone [\[2,12,13](#page-6-0)].

A pathway involving the oxidation of water by  ${}^{1}O_{2}$  was proposed for the antibody-catalyzed ozone formation [\[4](#page-6-0)], mainly based on the fact that some hydrogen peroxide molecules  $(H<sub>2</sub>O<sub>2</sub>)$  formed under the same conditions incorporate oxygen atoms from water molecules [\[14](#page-6-0)]. However, this proposal, which assumes that the  $O_3$  and  $H_2O_2$  are products of the same pathway, has not been unequivocally proved.

Here, I reinterpret available literature and develop an alternative hypothesis for  $O_3$  formation involving the oxidation and deoxidation of specific types of organic compounds. Pathways of methionine-, tryptophan-, and histidine-catalyzed ozone formation as well as a pathway of ozone formation during formaldehyde oxidation are proposed as examples of  $O<sub>3</sub>$  generation in this manner. A common feature of all the ozone-forming steps is that  ${}^{1}O_{2}$  reacts as an electrophile with a nucleophilic oxygen atom in the immediate precursor molecule, whose structure is such that loss of an  $O_3$  molecule results in co-formation of a non-charged product. I also propose that the decomposition of  $O_3$  in water may be largely responsible for the production of  $H_2O_2$  containing waterderived oxygen atoms.

## The antibody/amino acid-catalyzed water oxidation pathway and the need for alternative pathways of  $O_3$ formation

The antibody-catalyzed ozone formation occurs under conditions where  $H_2O_2$  is also generated in a high yield of  $>500$ molecules per antibody molecule [\[4,14](#page-6-0)]. Thus,  $O_3$  and  $H_2O_2$ were postulated to be formed by a common pathway, the antibody-catalyzed water oxidation pathway which begins with the reaction of  ${}^{1}O_{2}$  with water to generate hydrogen trioxide  $(H<sub>2</sub>O<sub>3</sub>)$  as a key intermediate (Fig. 1) [[4](#page-6-0)]. Two possible routes for the conversion of  $H_2O_3$  to  $H_2O_2$  and  $O_3$  have been proposed and shown to be feasible based on theoretical calculations [\[15,16\]](#page-6-0). The first of these routes involves a reaction of  $H_2O_3$ with  ${}^{1}O_{2}$  while the other involves a reaction of the former with another molecule of  $H_2O_3$  (Fig. 1). Notably, however, a third route for  $H_2O_3$  decomposition involving its water catalyzed conversion to  ${}^{1}O_{2}$  and water (Fig. 1) is well established both theoretically and experimentally [\[16\]](#page-6-0). In fact,  $H_2O_3$  is extremely unstable (with a half-life of less than a second) under aqueous conditions, where it basically decomposes to  $H_2O$  and  ${}^1O_2$  [\[16\]](#page-6-0). The antibody-catalyzed formation of  $O_3$  and  $H_2O_2$  was suggested to occur in a hydrophobic site where the  $H_2O_3$  is shielded from water [\[14](#page-6-0)]. However, even in organic solvents, all attempts to unambiguously detect  $H_2O_2$  and  $O_3$  as products of the decomposition of  $H_2O_3$  have failed [\[16\]](#page-6-0).

The possible role of amino acid oxidation in the antibodycatalyzed formation of  $H_2O_2$  as an alternative to the water oxidation pathway was discounted because of the consideration that, even if every photooxidizable amino acid residue (cysteine, methionine, histidine, tryptophan, and tyrosine) in



Fig. 1 The water oxidation pathway involving the initial formation of  $H<sub>2</sub>O<sub>3</sub>$  followed by decomposition of the latter by (i) reaction with singlet oxygen to form  $H_2O_2$  and  $O_3$ , (ii) reaction with another  $H_2O_3$  molecule to

form  $2H_2O_2$  and  ${}^{1}O_2$  with the intermediacy of ozone, or (iii) the water catalyzed decomposition to  ${}^{1}O_{2}$  and H<sub>2</sub>O [[4,15,16](#page-6-0)]. The latter reaction is extremely facile under aqueous conditions [\[16](#page-6-0)]

<span id="page-2-0"></span>an antibody molecule were consumed, this could not account for the  $>500$  mole equivalents of  $H_2O_2$  generated [[14](#page-6-0)]. Curiously, however, Yamashita et al. [\[17](#page-6-0)] found that out of 19 amino acids tested (excluding tyrosine), the photooxidizable amino acids methionine, cysteine, histidine, and tryptophan catalyzed the formation of ozone in the presence of singlet oxygen, comparably to antibodies, and that peptides lacking these amino acids did not generate ozone. They postulated that the amino acids catalyzed ozone formation through the water oxidation pathway but did not propose how the amino acids performed the catalysis. However, a major role for  $H_2O_3$  in ozone formation under these conditions is doubtful, even from the consideration that hydrophobic sites for its stabilization in amino acid solutions are unlikely to be present.

Hence, there is a need to explore alternative pathways for  $O_3$  and  $H_2O_2$  formation that do not involve  $H_2O_3$  as the key intermediate.

#### Evidence that the oxidation of amino acids and other organic molecules may be important for ozone generation

The common feature of the four amino acids that catalyze ozone formation [\[17\]](#page-6-0) is that they are photooxidizable [[14](#page-6-0)], and numerous studies have documented their oxidation by singlet oxygen under physiologically relevant conditions, both in the free form and as components of proteins [\[18](#page-6-0)–[32\]](#page-7-0).

As components of proteins, the reactivity of these amino acids with  ${}^{1}O_{2}$  greatly depends on their positions in the proteins, with residues exposed to solvent being oxidizable, while the less accessible residues remain unoxidized [\[20](#page-7-0),[21](#page-7-0)]. While only few amino acid residues in antibodies seem to get oxidized in the presence of singlet oxygen [\[22\]](#page-7-0), such few residues might be sufficient for  $O_3$  and  $H_2O_2$  formation. For example, just one exposed tryptophan residue was found to contribute 50 % of  $H_2O_2$  production by a monoclonal antibody [[23\]](#page-7-0). To date, the oxidative modification of specific tryptophan, methionine, and histidine residues in antibodies and monoclonal antibodies by singlet oxygen has been reported [\[23](#page-7-0)–[26\]](#page-7-0). Of particular interest is the recent discovery that  ${}^{1}O_{2}$  mediates the formation of cross-linked histidine-histidine dimers [[27\]](#page-7-0) and that two identical conserved histidine residues in the highly flexible and solvent-accessible hinge region of an

immunoglobulin G1 (IgG1) antibody undergo photooxidation to form such dimers [\[26](#page-7-0)].

Exposure of plant leaves to ozone causes damage to the leaves' photosystem 2 (PS II) [\[33](#page-7-0)]. PS II damage was also found to occur during light-induced oxidative stress in a process that involves  ${}^{1}O_{2}$ -mediated oxidation of specific tryptophan residues [[29](#page-7-0)]. PS II damage under the latter conditions is con-sistent with endogenous ozone production by the leaves [\[1](#page-6-0)] through a process involving tryptophan oxidation. In a related study, exogenous histidine was found to promote oxygen uptake and damage to PS II of the cyanobacterium Synechocystis PCC 6803, and the increased oxygen uptake was attributed to 'chemical trapping' of singlet oxygen by histidine [\[30\]](#page-7-0).

Tyihak et al. [\[12,13\]](#page-6-0) more recently reported that compounds such as formaldehyde, cinnamic acid, and resveratrol were precursors of ozone. They postulated a formaldehyde/ $O_3$  pathway in which formaldehyde reacts with  $H_2O_2$  to generate activated formaldehyde and  ${}^{1}O_{2}$ , followed by participation of the latter in the water oxidation pathway to generate ozone [\[12,13\]](#page-6-0). However, in atmospheric chemistry, formaldehyde is known as an important ozone precursor by a mechanism involving its oxi-dation [\[34](#page-7-0)] and not merely the production of  ${}^{1}O_{2}$ .

Thus, it is worthwhile to consider pathways of  ${}^{1}O_{2}$ -mediated amino acid and formaldehyde oxidation, with the aim of identifying potential steps that could be involved in  $O_3$  formation.

#### Suggested pathways of ozone formation via reactions of singlet oxygen with amino acids or formaldehyde

It was recently reported that various aliphatic and aromatic aldehydes react with  ${}^{1}O_{2}$  to form the corresponding organic acids [[35](#page-7-0)]. The initial reaction of the aldehyde with  ${}^{1}O_{2}$  involves a hydride transfer and generates a peroxyacid, which undergoes a Baeyer-Villiger oxidation with a second aldehyde molecule to form two molecules of acid [[35](#page-7-0)]. In the Baeyer-Villiger oxidation, the peroxyacid acts as a nucleophile while the aldehyde acts as an electrophile. Since  ${}^{1}O_{2}$  is a good electrophile, it may plausibly compete with the aldehyde in the Baeyer-Villiger oxidation step. Hence, formaldehyde 1 may be converted via peroxyformic acid 2 to formic acid 3 and ozone (Fig. 2). Thus, the formaldehyde/ $O_3$  pathway may involve formaldehyde reacting with  $H_2O_2$  to produce  ${}^{1}O_2$ [\[12,13](#page-6-0)], followed by reactions depicted in Fig. 2.



<span id="page-3-0"></span>

Fig. 3 Proposed pathway of  $O_3$  formation via the successive reactions of methionine (4) and methionine sulfoxide (7) with singlet oxygen ( ${}^{1}O_2$ ). The competing conversion of 7 to sulfone 10 normally occurs to a limited extent



Fig. 4 Proposed pathways of  $O_3$  formation via reactions of tryptophan 11 and its oxidation products with singlet oxygen  $(^1O_2)$ 

<span id="page-4-0"></span>Tomono et al. [36] found that decomposition of the pure <sup>1</sup>  ${}^{1}O_{2}$  generator, 1-methylnaphthalene-4-endoperoxide (MNPE) at 37 °C and pH 7.4 in the presence of cholesterol resulted in the formation of secosterol aldehyde B and a small amount of secosterol aldehyde A, and that addition of IgG resulted in a decrease in the former aldehyde and an increase in the latter. Increased formation of secosterol A in the presence of IgG is easily explained by the antibodycatalyzed ozone formation from  ${}^{1}O_{2}$ . On the other hand, formation of secosterol A even in the absence of IgG might be partly explained by secosterol aldehyde B reacting with singlet oxygen analogously to the ozone-producing reactions of formaldehyde with singlet oxygen.

Between pH 6 and 10, the reaction of methionine 4 with  ${}^{1}O_{2}$ in water proceeds via persulfoxide 5 and the cyclic dehydromethionine 6 to form methionine sulfoxide 7 [[36\]](#page-7-0) (Fig. [3](#page-3-0)). Interestingly, under similar conditions, methionine 4 also reacts efficiently with  $O_3$  to generate sulfoxide 7 and  ${}^{1}O_2$ [\[37](#page-7-0),[38\]](#page-7-0). Since the reaction between  $H_2S$  and  $O_3$  proceeds through an intermediate adduct  $H_2S-O_3$  [\[39\]](#page-7-0), the reaction of 4 with  $O_3$  to form 7 may also proceed through a trioxysulfoxide adduct, 8 (Fig. [3](#page-3-0)). Although methionine sulfoxide 7 may be further oxidized irreversibly to form methionine sulfone, this reaction is usually unfavorable [\[40\]](#page-7-0). A possible mechanism for

the latter reaction might begin with a nucleophilic attack of  $O_3$ on the S atom of sulfoxide 7 to generate intermediate 9 whose decomposition affords the sulfone 10 (Fig. [3\)](#page-3-0). Such ability of  $O<sub>3</sub>$  to react as a nucleophile has been previously reported [\[3\]](#page-6-0). On the other hand, it is conceivable that the reaction of  ${}^{1}O_{2}$  at the nucleophilic oxygen of sulfoxide 7 can produce trisulfoxide adduct 8 which decomposes to  $O_3$  and methionine 4. This proposed reversibility of the reaction of methionine  $4$  and  $O<sub>3</sub>$ to form sulfoxide  $7$  and  ${}^{1}O_{2}$  is indirectly supported by the fact that methionine sulfoxide reductases readily convert the sulf-oxide 7 back to methionine [[40](#page-7-0),[41](#page-7-0)]. The yield of  $O_3$  during the proposed reaction of 7 and  ${}^{1}O_{2}$  is expected to be good because ozone's co-product, 4, will react with  ${}^{1}O_{2}$  to regenerate reactant 7. Likewise, the reported high yield of  ${}^{1}O_{2}$  and methionine sulfoxide  $7$  from  $O_3$  and methionine 4 should be due to the very short lifetime of  ${}^{1}O_{2}$  (less than a second as compared to 4.8 min half-life of ozone in water) [\[37,38\]](#page-7-0).

Figure [4](#page-3-0) illustrates a suggested pathway for tryptophancatalyzed ozone formation. First, tryptophan 11 reacts with singlet oxygen to form hydroperoxide 12 [\[18](#page-6-0)[,28](#page-7-0),[32](#page-7-0)], which can exist in equilibrium with zwitterionic peroxide 13 and tricylic hydroperoxide 14. The latter is a well-known tryptophan photooxidation product [[31,32\]](#page-7-0). Zwitterionic peroxide 13 may cyclize to form dioxetane 15 whose decomposition



Fig. 5 Proposed pathways of  $O_3$  formation via reactions of histidine 23 and its oxidation products with singlet oxygen  $(^1O_2)$ 

affords N-formylkynurenine 16 [[28](#page-7-0),[32](#page-7-0)] or dioxindolylalanine 17 [\[32](#page-7-0)]. Alternatively, I propose that peroxide 13 may react with  ${}^{1}O_{2}$  to afford  $O_{3}$  and epoxide 18 in a concerted reaction like the above proposed reaction of peroxyacid 2 with  ${}^{1}O_{2}$ (Fig. [2](#page-2-0)). However, a stepwise reaction of  ${}^{1}O_{2}$  and 13 via a zwitterionic tetroxide is not ruled out. Epoxide 18 can exist in equilibrium with zwitterionic oxide 19, which may also react with  ${}^{1}O_{2}$  to form ozone and regenerate tryptophan 11 via trioxyanion <sup>20</sup>. A concerted reaction between <sup>19</sup> and <sup>1</sup>  ${}^{1}O_{2}$ , bypassing 20 is also a possibility. Compounds 18 and 19 can exist in equilibrium with isomeric alcohols 21 and 22, which are known products [\[28,31,32](#page-7-0)].

Figure [5](#page-4-0) illustrates some of the potential pathways involved in the histidine-catalyzed ozone formation. First, histidine 23 is converted to hydrated imidazolone 24 [\[18](#page-6-0),[27](#page-7-0),[28\]](#page-7-0), which is a precursor of the histidine-histidine cross-linked dimer 25 [\[26](#page-7-0)] that has been detected in the hinge region of an IgG 1 [\[26\]](#page-7-0). A cyclic oxidation-deoxidation pathway of this dimer will lead to  $O_3$  production via intermediates 26–28. Ozone formation may also occur during the conversion of compound 24 via zwitterionic oxide 29 to 2-oxohistidine 30, which is a known product [\[26\]](#page-7-0). The latter may be converted back to histidine 23 via zwitterionic oxide 31 and vinylic

anion 32. Alternatively, 30 may be converted back to 29, with O3 formation, via zwitterionic peroxide 33.

The pathways suggested in Figs. [4](#page-3-0) and [5](#page-4-0) for tryptophan and histidine-catalyzed ozone formation are not exhaustive, and other oxidized intermediates such as dioxindolylalanine 17 are also potential  $O<sub>3</sub>$  precursors.

### Ozone decomposition in water may play a key role in the formation of  $H_2O_2$  containing water-derived oxygen

The incorporation of oxygen atoms from water into  $H_2O_2$ molecules in the presence of  ${}^{1}O_{2}$  was considered as key evidence that water reacts with  ${}^{1}O_{2}$  to form  $H_{2}O_{3}$  according the water-oxidation pathway [[14](#page-6-0)]. However,  $H_2O_2$  containing water-derived oxygen may potentially be formed during the decomposition of  $O_3$  by known reactions as depicted in Fig. 6. For example, when  $O_3$  formation occurs during irradiation of antibodies, equation 1 is likely to be a major contributor of such  $H_2O_2$  [\[42](#page-7-0)].  $H_2O_2$  that does not contain water-derived oxygen can be generated during the oxidation of methionine (Fig. [3\)](#page-3-0) or by ozonolysis of tryptophan or histidine via  $\alpha$ -

$$
O_3 + H_2O \xrightarrow{hv} \text{HOOH} + O_2 \tag{1}
$$

$$
O_3 + HO_2^ \longrightarrow
$$
  $HO_5^ \longrightarrow$   $HO_2' + \text{C}_3$  (2)

$$
-O_3 \quad \longrightarrow \quad -O + O_2 \tag{3}
$$

$$
-O + H_2O \implies HO \qquad + HO^-(4)
$$

$$
HO^{\cdot} + H_2O \quad \Longleftrightarrow \quad H_2O \quad + \quad \cdot OH \tag{5}
$$

$$
2 \text{ HO} \cdot \longrightarrow H_2O_2 \tag{6}
$$

$$
HO \cdot + O^3 \longrightarrow HOOOO.
$$
 (1)

$$
HOOOO: \longrightarrow \qquad HOO \cdot + \qquad O_2 \tag{8}
$$

$$
2 \text{ HOO} \longrightarrow H_2O_2 + O_2 \tag{9}
$$

 $H_2O_2$  $2O<sub>3</sub>$  $(10)$ 2 HOOOO

$$
HOO \cdot + HOOOO \cdot \longrightarrow H_2O_2 + 2O_2 \tag{11}
$$

Fig. 6 Proposed formation of  $H_2O_2$  containing water-derived oxygen atoms as a consequence of the decomposition of  $O<sub>3</sub>$ 

<span id="page-6-0"></span>hydroxyhydroperoxide intermediates  $[43]$  $[43]$ . HO<sub>2</sub> anions derived from such  $H_2O_2$  molecules can initiate  $O_3$  decomposition and formation of hydroxyl radicals ( OH) according to equations 2–4 [[44](#page-7-0)]. Hydroxyl radicals undergo rapid hydrogen exchange with water molecules [\[45](#page-7-0)]. Thus, even if some hydroxyl radicals formed according to equation 4 have ozonederived oxygen atoms, their hydrogen exchange with water molecules will generate hydroxyl radicals having waterderived oxygen according to equation 5. There is reasonable chance that such hydroxyl radicals will meet and react to form  $H<sub>2</sub>O<sub>2</sub>$  (equation 6) [[46](#page-7-0)]. Other reactions that would lead to formation of  $H_2O_2$  with water-derived oxygen include equations 7–11 [\[47\]](#page-7-0). By such pathways therefore, the oxidation of water to form  $H_2O_2$  may occur as a consequence of  $O_3$  formation and decomposition, contrary to the previously proposed ozone formation as a consequence of water oxidation.

#### Concluding remarks

In the present article, potential pathways have been proposed for (i) the oxidation of aldehydes by  ${}^{1}O_{2}$  to form acids and  $O_{3}$ and (ii) the amino acid-catalyzed  $O_3$  formation via various oxidation and deoxidation reactions. Theoretical and experimental efforts to confirm these pathways will be possible because potential key intermediates have been identified.

Antibodies may play an important role in ozone production by neutrophils [4]. While efforts to understand the finer details of the antibody-catalyzed  $O_3$  formation have in the past focused on potential hydrophobic active sites for formation of  $H<sub>2</sub>O<sub>3</sub>$  [14,[22\]](#page-7-0), more attention should now shift to mapping solvent accessible sites where methionine, histidine, tryptophan, cysteine, methionine, and disulfide bridges undergo oxidation, and how chemical modifications to these sites affect  $O<sub>3</sub>$  generation.

Based on the types of  $O_3$ -generating reactions suggested here, it will be possible to predict the potential of various other organic compounds to participate in or catalyze ozone formation. It will also be possible to synthesize new ozone-generating molecules, which may help to advance the use of ozone in various fields. For example, such molecules may serve as alternatives to the antibiotics currently used in crop protection.

Compounds such as 1-butylnaphthalene-4-propionate endoperoxide or 1-methylnaphthalene-4-propionate endoperoxide generate  ${}^{1}O_{2}$  under physiological conditions in the dark [[48,49\]](#page-7-0). The potential application of such compounds to generate ozone for treatment of various diseases remains to be explored. For example, the malaria parasite, Plasmodium falciparum, which easily develops drug resistance, has been shown to be inhibited by ozone [[50](#page-7-0)] and, interestingly, it abundantly produces a histidine-rich protein [[51\]](#page-7-0) that might be a good catalyst of ozone formation in the presence of singlet oxygen.

#### References

- 1. Balla J, Tyihak E (2010) Direct measurement of emission of endogenous ozone from plants by GC-MS-SIM. Chromatographia Supp 71:S87–S91
- 2. Tyihak E, Moricz AM, Ott PG (2012) BioArena studies: unique function of endogenous formaldehyde and ozone in the antibiotic effect—a review. Med Chem 8:75–84
- Beltran FJ (2004) Ozone reaction kinetics for water and wastewater systems. CRC Press, pp 13–14
- 4. Wentworth P Jr, McDunn JE, Wentworth AD, Takeuchi C, Nieva J, Jones T, Bautista C, Ruedi JM, Gutierrez A, Janda KD, Babior BM, Eschenmoser A, Lerner RA (2002) Evidence for antibodycatalyzed ozone formation in bacterial killing and inflammation. Science 298:2195–2199
- 5. Pereira PMR (2015) Antibodies armed with photosensitizers: from chemical synthesis to photobiological applications. Org Biomol Chem. doi[:10.1039/C4OB02334](http://dx.doi.org/10.1039/C4OB02334)
- 6. Wentworth P Jr, Nieva J, Takeuchi C, Galve R, Wentworth AD, Dilley RB, DeLaria GA, Saven A, Babior BM, Janda KD, Eschenmoser A, Lerner RA (2003) Evidence for ozone formation in human atherosclerotic arteries. Science 302:1053–1056
- 7. Wentworth AD, Song B-D, Nieva J, Shafton A, Sangeetha T, Wentworth P Jr (2009) The ratio of cholesterol 5,6 secosterols formed from ozone and singlet oxygen offers insight into the oxidation of cholesterol in vivo. Chem Comm (Camb) 3098-3100
- 8. Kettle AJ, Clark BM, Winterbourn CC (2004) Superoxide converts indigo carmine to isatin sulfonic acid: implications for the hypothesis that neutrophils produce ozone. J Biol Chem 279:18521– 18525
- 9. Brinkhorst J, Nara SJ, Pratt DA (2008) Hock cleavage of cholesterol-5α-hydroperoxide: an ozone-free pathway to the cholesterol ozonolysis products identified in arterial plague and brain tissue. J Am Chem Soc 130:12224–12225
- 10. Tomono S, Miyoshi N, Shiokawa H, Iwabuchi T, Aratani Y, Higashi T, Nukaya H, Ohshima H (2011) Formation of cholesterol ozonolysis products in vitro and in vivo through a myeloperoxidase-dependent pathway. J Lipid Res 52:87–97
- 11. Miyoshi N, Iuliano L, Tomono S, Ohshima H (2014) Implications of cholesterol autoxidation products in the pathogenesis of inflammatory diseases. Biochem Biophys Res Comm 446:702–708
- 12. Tyihak E, Moricz AM, Ott PG, Katay G, Mincsovics E (2012) Biological characterization of ingredients in OPLC-BioArenagreenhouse-system: unique reactions of endogenous HCHO and  $O<sub>3</sub>$  in in vitro and in vivo conditions. Chromatographia 75:983–990
- 13. Tyihak E, Moricz AM, Ott PG, Kiraly-Veghely Z, Katay G (2013) BioArena system for knowing and understanding the biological world: a review with new experimental results. J AOAC Int 96: 1189–1199
- 14. Wentworth P Jr, Jones LH, Wentworth AD, Zhu X, Larsen NA, Wilson IA, Xu X, Goddard WA III, Janda KD, Eschenmoser A, Lerner RA (2001) Antibody catalysis of the oxidation of water. Science 293:1806–1811
- 15. Xu X, Muller RP, Goddard WA III (2002) The gas phase reaction of singlet dioxygen with water: a water-catalyzed mechanism. Proc Natl Acad Sci U S A 99:3376–3381
- 16. Cerkovnic J, Plesnicar B (2013) Recent advances in the chemistry of hydrogen trioxide (HOOOH). Chem Rev 113:7930–7951
- 17. Yamashita K, Miyoshi T, Arai T, Endo N, Itoh H, Makino K, Mizugishi K, Uchiyama T, Masataka S (2008) Ozone production by amino acids contributes to killing of bacteria. Proc Natl Acad Sci U S A 105:16912–16917
- 18. Wei C, Song B, Yuan J, Feng Z, Jia G, Li C (2007) Luminescence and Raman spectroscopic studies on the damage of tryptophan,

<span id="page-7-0"></span>histidine and carnosine by singlet oxygen. J Photochem Photobiol A Chem 189:39–45

- 19. Pattison DI, Rahmanto AS, Davies MJ (2012) Photo-oxidation of proteins. Photochem Photobiol Sci 11:38–53
- 20. Jensen RL, Arnbjerg J, Ogilby R (2012) Reaction of singlet oxygen with tryptophan in proteins: a pronounced effect of the local environment on the reaction rate. J Am Chem Soc 134:9820–9826
- 21. Lundeen RA, McNeill K (2013) Reactivity differences of combined and free amino acids: quantifying the relationship between threedimensional protein structure and singlet oxygen reaction rates. Environ Sci Technol 47:14215–14223
- 22. Zhu X, Wentworth P Jr, Wentworth AD, Eschenmoser A, Lerner RA, Wilson IA (2004) Probing the antibody-catalyzed water-oxidation pathway at atomic resolution. Proc Natl Acad Sci U S A 101: 2247–2252
- 23. Sreethara A, Lau K, Hosken B, Macchi F, Zhan D, Shen A, Steinmann D, Schoneich C, Lentz Y (2013) Role of surface exposed tryptophan as substrate generators for the antibody catalyzed water oxidation pathway. Mol Pharm 10:278–288
- 24. Khor HK, Jacoby ME, Squler TC, Chu GC, Chelius D (2010) Identification of methionine sulfoxide diastereomers in immunoglobin gamma antibodies using methionine sulfoxide reductase enzymes. mAbs 2:299–308
- 25. Amano M, Kobayashi N, Yabuta M, Uchiyama S, Fukui K (2014) Detection of histidine oxidation in a monoclonal immunoglobulin gamma (IgG) 1 antibody. Anal Chem 86:7536–7543
- 26. Liu M, Zhang Z, Cheetham J, Ren D, Zhou ZS (2014) Discovery and characterization of a photo-oxidative histidine-histidine crosslink in IgG1 antibody utilizing 18O-labeling and mass spectrometry. Anal Chem 86:4940–4948
- 27. Liu F, Lu W, Fang Y, Liu J (2014) Evolution of oxidation dynamics of histidine: nonreactivity in the gas phase, peroxides in hydrated clusters, and pH dependence in solution. Phys Chem Chem Phys 16:22179–22191
- 28. Kim J, Rodriquez ME, Guo M, Kenney ME, Oleinic NL, Anderson VE (2008) Oxidative modification of cytochrome c by singlet oxygen. Free Radic Biol Med 44:1700–1711
- 29. Kasson TMD, Rexroth S, Barry BA (2012) Light-induced oxidative stress, N-formylkynurenine, and oxygenic photosynthesis. PLoS ONE 7:e42220
- 30. Rehman AU, Cser K, Sass L, Vass I (2013) Characterization of singlet oxygen production and its involvement in photochemical damage of photosystem II in the cyanobacterium Cynechocystis PCC 6803 by histidine-mediated chemical trapping. Biochim Biophys Acta 1827:689–698
- 31. Ronsein GE, de Oliveira MCB, de Medeiros MHG, Mascio PD (2009) Characterization of <sup>1</sup>O2-derived oxidation products of tryptophan: a combination of tandem mass spectrometry analyses and isotopic labeling studies. J Am Soc Mass Spectrom 20:188–197
- 32. Ronsein GE, de Oliveira MCB, de Medeiros MHG, Mascio PD (2011) Mechanism of dioxindolylalanine formation by singlet molecular oxygen-mediated oxidation of tryptophan residues. Photochem Photobiol Sci 10:1727–1730
- 33. Kobayakawa H, Imai K (2011) Effects of the interaction between ozone and carbon dioxide on gas exchange, photosystem II and antioxidants in rice leaves. Photosynthetica 49:227–238
- 34. Liu L, Flatoy F, Ordonez C, Braathen GO, Hak C, Junkermann W, Andreani-Aksoyoglu S, Mellqvist J, Galle B, Prevot ASH, Isaksen ISA (2007) Photochemical modelling in the Po basin with focus on formaldehyde and ozone. Atmos Chem Phys 7:121–137
- 35. Hajimohammadi M, Safari N, Mofakham H, Shaabani A (2010) A new and efficient aerobic oxidation of aldehydes to carboxylic acids with singlet oxygen in the presence of porphyrin sensitizers and visible light. Tetrahedron Lett 51:4061–4065
- 36. Fang Y, Liu F, Bennet A, Ara S, Liu J (2011) Experimental and trajectory study on the reaction of protonated methionine with electronically excited singlet molecular oxygen  $(a1\Delta g)$ : reaction dynamics and collision energy effects. Phys Chem B 115:2671–2682
- 37. Kanofsky JR, Sima P (1991) Singlet oxygen production from the reactions of ozone with biological molecules. J Biol Chem 266: 9039–9042
- 38. Munoz F, Mvula E, Braslavsky SE, von Sonntag C (2001) Singlet dioxygen formation in ozone reactions in aqueous solution. J Chem Soc Perkin Trans 2:1109–1116
- 39. Vahedpour M, Baghary R, Khalili F (2013) Prediction of mechanism and thermochemical properties of  $O_3 + H_2S$  atmospheric reaction. J. Chem. Article ID 65968 http://dx.doi.org/[10.1155/2013/](http://dx.doi.org/10.1155/2013/659682) [659682](http://dx.doi.org/10.1155/2013/659682)
- 40. Ghesquere B, Jonckheere V, Colaert N, van Durme J, Timmerman E, Goethals M, Schymkowitz J, Rousseau F, Vanderkerckhove J, Gevaert K (2011) Redox proteomics of protein-bound methionine oxidation. Mol Cell Proteomics 10:M110.006866, http:// mcponline.org/content/10/5.toc
- 41. Lim JC, You Z, Kim G, Levine RL (2011) Methionine sulfoxide reductase A is a stereospecific methionine oxidase. Proc Natl Acad Sci U S A 108:10472–10477
- 42. Glaze WH, Kang J-W, Chapin DH (1987) The chemistry of water treatment processes involving ozone, hydrogen peroxide and ultraviolet radiation. Ozone Sci Eng 9:335–352
- 43. Sharma VK, Graham NJD (2010) Oxidation of amino acids, peptides and proteins by ozone: a review. Ozone Sci Eng 32:81–90
- 44. Merenyi G, Lind J, Sonntag C (2010) Reaction of ozone with hydrogen peroxide (peroxone process): a revision of current mechanistic concepts based on thermokinetic and quantum-chemical considerations. Environ Sci Technol 44:3505–3507
- 45. Codorniu-Hernandez E, Kusalik PG (2012) Mobility mechanism of hydroxyl radicals in aqueous solution via hydrogen transfer. J Am Chem Soc 134:532–538
- 46. Codorniu-Hernandez E, Hall KW, Ziemianowicz D, Carpendale S, Kusalik PG (2014) Aqueous production of oxygen atoms from hydroxyl radicals. Phys Chem Chem Phys 16:26094–26102
- 47. Staehelin J, Buhler RE, Hoigne J (1984) Ozone decomposition in water studied by pulse radiolysis. 2. HO and HO4 as chain intermediates. J Phys Chem 88:5999–6004
- 48. Tomono S, Miyoshi N, Sato K, Ohba Y, Ohshima H (2009) Formation of cholesterol ozonolysis products through an ozonefree mechanism mediated by myeloperoxidase-H2O-chloride system. Biochem Biophys Res Comm 383:222–227
- 49. Otsu K, Sato K, Sato M, Ono H, Ohba Y, Katagata Y (2008) Impaired activation of caspase cascade during cell death induced by newly synthesized singlet oxygen generator, 1 buthylnaphthalene-4-propionate-endoperoxide. Cell Biol Int 32: 1380–1387
- 50. Lell B, Viebahn R, Kremsner PG (2001) The activity of ozone against Plasmodium falciparum. Ozone Sci Eng 23:89–93
- 51. Verma P, Biswas S, Mohan T, Ali S, Rao DN (2013) Detection of histidine-rich protein and lactate dehydrogenase of Plasmodium falciparum in malaria patients by sandwich ELISA using in-house reagents. Indian J Med Res 138:977–987