·Review·

Contribution of NMDA receptors to dorsolateral prefrontal cortical networks in primates

Min Wang, Amy F T Arnsten

Department of Neurobiology, Yale Medical School, New Haven, CT 06510, USA Corresponding author: Min Wang. E-mail: Min.wang@yale.edu

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Cognitive disorders such as schizophrenia and Alzheimer's disease are associated with dysfunction of the highly evolved dorsolateral prefrontal cortex (dlPFC), and with changes in glutamatergic N-methyl-*D*-aspartate receptors (NMDARs). Recent research on the primate dlPFC discovered that the pyramidal cell circuits that generate the persistent firing underlying spatial working memory communicate through synapses on spines containing NMDARs with NR2B subunits (GluN2B) in the post-synaptic density. This contrasts with synapses in the hippocampus and primary visual cortex, where GluN2B receptors are both synaptic and extrasynaptic. Blockade of GluN2B in the dlPFC markedly reduces the persistent firing of the Delay cells needed for neuronal representations of visual space. Cholinergic stimulation of nicotinic α7 receptors within the glutamate synapse is necessary for NMDAR actions. In contrast, α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors have only subtle effects on the persistent firing of Delay cells, but contribute substantially to the firing of Cue and Response cells. Systemic administration of the NMDAR antagonist ketamine reduces the persistent firing of Delay cells, but increases the firing of some Response cells. The reduction in persistent firing produced by ketamine may explain why this drug can mimic or worsen the cognitive symptoms of schizophrenia. Similar actions in the medial PFC circuits representing the emotional aspects of pain may contribute to the rapid analgesic and anti-depressant actions of ketamine.

Keywords: glutamate; Alzheimer's disease; schizophrenia; depression; ketamine

Introduction

Glutamate acts at a variety of ionotropic receptors, including α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid receptors (AMPARs), kainate receptors, and N-methyl-*D*-aspartate receptors (NMDARs)^[1]. NMDARs have been of particular interest due to their unique properties: they require depolarization to relieve their Mg⁺⁺ block, and are permeable to Ca⁺⁺ that can initiate second-messenger signaling events, such as mediating neuroplasticity or negative feedback through Ca⁺⁺-sensitive K⁺ channels. NMDARs contain an NR1 subunit and a mixture of NR2A–D subunits that alter the functional properties of the receptor, e.g. NMDARs with NR2A subunits (GluN2A) are more sensitive and have faster kinetics, while those with NR2B subunits (GluN2B) have slower kinetics and can produce increased levels of calcium influx $[2]$. As NMDARs are altered in cognitive disorders such as schizophrenia and Alzheimer's disease, there has been increasing research on these receptors^[3, 4]. The highly-evolved dorsolateral prefrontal cortex (dlPFC) is only found in primates $[5]$ and subserves higher cognitive functions, especially those affected in these mental disorders^[6, 7]. The following review briefly summarizes new data demonstrating the key role of GluN2B receptors in the primate dlPFC, and how their actions in the dIPFC appear to differ from classical findings in the sensory cortex and hippocampus.

NMDAR and AMPAR Actions in Visual Cortex and Hippocampus

There have been extensive studies on the glutamate

192 Neurosci Bull April 1, 2015, 31(2): 191–197 NMDAR and AMPAR mechanisms underlying long-term synaptic plasticity in the primary visual cortex and in CA1 with long-term memory consolidation: it is a momentary, ever-changing pattern of recurrent activation of relatively stable architectural networks, while long-term memory consolidation retains events as structural changes

neurons of the hippocampus $[8-10]$. Long-term plasticity is powerfully regulated by the levels of AMPAR expression: the number of AMPARs inserted into the post-synaptic density can mediate the degree of spine depolarization and thus the NMDAR opening. AMPAR membrane insertion leads to structural synaptic changes such as enlarging the spine head and shortening/thickening of the spine $neck^{[11, 12]}$ to create a stable, mushroom-shaped spine and enduring strengthening of a synaptic connection^[13], and/ or the addition of new spines and synapses^[11]. Synaptic plasticity in the mature visual cortex appears to be governed by GluN2A subunits, which have faster kinetics than GluN2B. GluN2B receptors are expressed in synapses early in development, but many move to extra-synaptic locations in the mature visual cortex and hippocampus^[14]. In the hippocampus, there is some evidence that longterm potentiation (LTP) is mediated by synaptic GluN2A, while long-term depression is mediated by extrasynaptic GluN2B receptors $[8]$. However, this finding is controversial. For example, there is increasing evidence that GluN2B receptors are also important for LTP in hippocampal neurons $[15-17]$. In the mature visual cortex, long-term plastic changes in synapses appear to rely heavily on GluN2A, e.g. a selective GluN2A antagonist inhibits LTP induction, while neither a GluN2B antagonist^[18] nor the over-expression of GluN2B^[19] alters LTP in the visual cortex. The faster kinetics of GluN2A is well-suited to the rapid processing of continuous visual inputs and more faithful neuronal firing to sensory stimulation. Thus, it is appropriate that GluN2A actions predominate in visual cortical synapses.

NMDARs in Primate Dorsolateral Prefrontal Cortex

In contrast to sensory cortex, the dlPFC generates mental representations in the absence of sensory stimulation and these are the foundation of abstract thought. The dlPFC subserves working memory: the ability to keep information in mind and use these representations to provide top-down guidance of behavior, thought, and emotion. Working memory is active and relevant only for a short period of time, usually on the scale of seconds. This capability is a basic building block for more complex dlPFC cognitive operations. Working memory contrasts in synapses. It is not surprising that the circuitry and modulation of working memory differ from those of longterm memory consolidation. The visuo-spatial working memory operations of the dlPFC in monkeys are among the best understood. Much of the data arose from studies using a spatial working memory task termed the oculomotor delayed response (ODR) (Fig. 1). In this task, the monkey fixates on a center spot, while a cue appears briefly in one of eight possible locations. The monkey must remember the cue location over a delay period of several seconds. At the end of the delay period, the monkey makes an eye movement to the remembered location to receive a juice reward. The location of the cue randomly changes from trial to trial, thus requiring constant updating of the contents of working memory. The dlPFC is needed to perform this working memory task, and even small lesions in this area can produce permanent deficits in performance^[20]. Neuronal recordings from the dlPFC in monkeys performing a spatial working memory task have found neurons that fire to the Cue and/or to the Response, but also neurons that are able to maintain spatially-tuned, persistent activity across the delay period $[21]$. This delayrelated persistent activity has been considered to be the neuronal mechanism of working memory due to the following features^[22]: first, this neuronal activity persists

during the time period when a representation needs to be remembered; second, sustained neuronal activity ceases when a memory-guided response has been generated and the representation is no longer needed; third, when activity does not persist throughout the delay period, behavioral performance is compromised; and fourth, the persistent activity is direction-selective. The pioneering work of Goldman-Rakic revealed that this persistent memory-related activity is generated by the recurrent excitation of pyramidal cells interconnecting on dendritic spines in deep layer III of the dIPFC^[23]. Computational models have predicted that this persistent memoryrelated activity requires stimulation of NMDARs rather than $AMPARS^{[24]}$, and that the slow kinetics of GluN2B receptors is particularly well-suited to persistent dlPFC network firing in the absence of sensory stimulation^[25]. In

Fig. 1. The actions of NMDARs on the dlPFC neuronal circuitry underlying spatial working memory in primates. A. The spatial oculomotor delayed response (ODR) task. Trials begin when the monkey fixates on a central point for 0.5 s. A cue is presented in 1 of 8 possible locations for 0.5 s, followed by a 2.5-s delay period. When the fixation point is extinguished, the monkey makes **a saccade to the location of the remembered cue. The position of the cue changes on each trial in a quasi-random manner, thus requiring the constant updating of working memory stores. B. The region of monkey dlPFC where recordings were made. PS,** principal sulcus; AS, arcuate sulcus. C. The deep layer III microcircuits subserving spatially-tuned, persistent firing during the **delay period. B, GABAergic basket cell. D. Working model of a glutamate synapse on a spine in layer III of the dlPFC. Glutamate stimulates NMDAR-NR2B receptors in the post-synaptic density, while AMPARs have only subtle actions. Permissive, depolarizing effects for NMDAR actions appear to be mediated by cholinergic stimulation of nicotinic (nic)-α7Rs, which are also localized in** the synapse. Ca⁺⁺ entry through NMDAR-NR2B may provide negative feedback by facilitating internal Ca⁺⁺ release from the spine apparatus (asterisk); feedforward Ca⁺⁺-cAMP signaling opens nearby K⁺ channels to weaken synaptic efficacy and reduce firing. **E. An example of an individual dlPFC Delay cell under control conditions and following iontophoresis of the NMDAR antagonist** MK801 (25 nA). The rasters and histograms show firing patterns of the neuron's preferred direction and the non-preferred direction **opposite to it. Iontophoresis of MK801 markedly reduced task-related fi ring, which returned towards control levels when delivery** of MK801 was stopped (Recovery; P <0.05). F. Average responses showing the mean + SEM firing patterns of 15 dIPFC Delay cells **for their preferred** *versus* **non-preferred directions under control conditions (blue) and following iontophoresis of MK801 (red).** MK801 markedly suppressed task-related firing, especially for the neurons' preferred direction.

contrast, the faster kinetics of AMPARs leads to dynamic instability and network collapse $[24]$. Consistent with computational predictions, both *in vitro* and *in vivo* studies have found a prominent role of GluN2B neuronal firing in the PFC. Recordings from rat brain slices have shown more extensive expression of GluN2B in the medial PFC than in the primary visual cortex^[26]. A more recent study of the primate dlPFC revealed GluN2B in synapses and that the GluN2B receptor mediates the persistent firing of dlPFC networks in monkeys performing a spatial working

memory task^[27]. Immunoelectron microscopy demonstrated that GluN2B is localized exclusively within the postsynaptic densities of layer III dlPFC excitatory synapses on spines, with no evidence of extra-synaptic labeling. Singleunit recordings coupled with iontophoresis in monkeys performing the ODR task showed that the persistent activity of dlPFC neurons is highly dependent on NMDARs, including GluN2B. Iontophoretic blockade of all NMDARs using the antagonist MK801 completely suppresses taskrelated neuronal firing (Fig. 1). Similarly, blocking GluN2B receptors by iontophoresis of Ro25-6981 produces a marked loss of persistent neuronal firing, and blockade of GluN2A receptors also reduces firing. In contrast, blockade of AMPARs with CNQX/NBQX has only subtle effects on memory-related firing, reducing persistent firing in a small portion of the delay period. AMPAR blockade does alter the firing of sensory neurons in the dIPFC, i.e. it reduces the firing of Cue cells and Post-saccadic Response "feedback" cells. However, the neurons that generate representations of visual space are much more affected by NMDAR than by AMPAR blockade. Interestingly, systemic administration of the NMDAR antagonist ketamine reduces the firing of Delay cells, but increases the firing of Post-saccadic Response neurons (ibid). These results are consistent with the reliance of Delay cells on NMDARs, while the Post-

If AMPARs have little effect on dlPFC Delay neurons, what depolarizes the membrane and relieves the Mq^{+1} block in NMDARs? In the primate dlPFC, these permissive actions appear to be mediated by cholinergic stimulation of nicotinic α7 receptors (nic-α7Rs), rather than AMPARs. Nic-α7Rs are localized in and next to the postsynaptic density in glutamate synapses on spines, and blockade of nic-α7Rs prevents the excitatory actions of NMDA^[28]. As acetylcholine is released during wakefulness but not deep sleep, nic-α7R stimulation may permit conscious thought in the waking state. Thus, in the dlPFC, neuronal networks communicate based on arousal state, while in sensory cortex and the hippocampus, NMDAR actions are based on levels of circuit activity, i.e. glutamate release onto AMPARs. Thus, deficits in either NMDAR or nic-α7R signaling weaken dlPFC function.

saccadic Response cells have a large AMPAR influence.

Finally, Ca^{++} entry through activated NMDARs may contribute to negative feedback to prevent seizures in recurrent excitatory networks. As schematically illustrated in Figure 1, many spines in layer III of the dlPFC contain a spine apparatus, the Ca⁺⁺-storing endoplasmic reticulum extended into the spine that is elaborated near the synapse. Accumulating evidence indicates that feedforward Ca^{++} -cAMP signaling opens nearby K^+ channels on dendritic spines to decrease synaptic efficacy and reduce neuronal firing (reviewed in [29]). Future research is needed to determine whether high levels of Ca⁺⁺ entry through GluN2B receptors activate these intracellular pathways.

Relevance to Mental Illness

A variety of cognitive disorders are associated with altered NMDAR signaling. For example, NMDARs are internalized by β-amyloid oligomers in Alzheimer's disease, and this effect occurs in association with nic-α7Rs^[30]. Schizophrenia is also linked to genetic insults that weaken $NMDAR^{[31, 32]}$ and nic- α 7 $R^{[33]}$ signaling. Post-mortem studies have indicated altered GluN2B expression and trafficking^[3, 34], including links between allelic changes in GluN2B and impaired reasoning in patients with schizophrenia^[35]. There is also accumulating evidence that genetic insults to NMDAR and NMDAR-related synaptic proteins are associated with an increased risk of schizophrenia^[32, 36, 37]. The NMDAR antagonist ketamine has been used to model the cognitive deficits of schizophrenia, reducing the blood oxygenation level-dependent response during the delay period of a working memory task in healthy human individuals^[38, 39] similar to that seen in patients with schizophrenia^[40]. In contrast, the hyperglutamate theories of schizophrenia based on rodent models^[41] likely relate to the increased Post-saccadic Response "feedback" cell firing induced by the systemic administration of NMDA antagonists.

In contrast to schizophrenia, where ketamine worsens the symptoms^[42], acute ketamine treatment rapidly ameliorates the symptoms in some patients with treatmentresistant depression^[43-46], bringing relief within minutes following intra-nasal application $[47, 48]$. The positive response to ketamine in severely depressed patients has been related to their anterior cingulate response to fearful faces before treatment^[49]. Neurons in the anterior cingulate of monkeys have been shown to represent negative emotions such as symbolic punishment^[50], as well as loss of expected rewards^[51]. Thus, it is possible that ketamine treatment is helpful in treating depressive symptoms by reducing the firing of NMDAR-dependent, recurrent excitatory circuits in the anterior cingulate and/or in other ventromedial PFC circuits (e.g. Brodmann's area $25^{[52]}$) that represent negative emotions and instigate mental suffering. Interrupting the activity of these circuits might underlie the immediate beneficial effects of ketamine in some patients, prior to the regrowth of dendritic spines^[53] that may underlie more prolonged beneficial actions. Decreased firing of neurons in the anterior cingulate and area 25 may also underlie the rapid relief of pain by intranasal ketamine (within 5–25 min)^[54], as these medial PFC areas are part of the circuits that process the emotional response to painful events^[55, 56]. Since intra-nasal ketamine relieves physical pain within minutes $[54, 57, 58]$, it thus may relieve "psychic pain" as well. More research is needed to determine whether NMDARs mediate medial PFC circuits in primates similar to their actions in the dlPFC circuits representing visual space.

Conclusion

New research on the primate dlPFC indicates that GluN2B receptors play a prominent role in the generation of mental representations needed for abstract thought. The data suggest that cholinergic actions at nic-α7Rs are permissive for NMDA synaptic activity, and for the dlPFC network representation of visual space. These data underscore why changes in NMDAR or nic-α7R signaling in diseases such as schizophrenia and Alzheimer's disease have such devastating effects on higher cognition. The unique properties of these dlPFC circuits must be considered in order to design effective treatments for cognitive disorders.

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REFERENCES

[1] Dingledine R, Borges K, Bowie D, Traynelis SF. The

glutamate receptor ion channels. Pharmacol Rev 1999, 51: 7–61.

- [2] Erreger K, Dravid SM, Banke TG, Wyllie DJ, Traynelis SF. Subunit-specific gating controls rat NR1/NR2A and NR1/ NR2B NMDA channel kinetics and synaptic signalling profiles. J Physiol 2005, 563: 345-358.
- [3] Kristiansen LV, Bakir B, Haroutunian V, Meador-Woodruff JH. Expression of the NR2B-NMDA receptor trafficking complex in prefrontal cortex from a group of elderly patients with schizophrenia. Schizophr Res 2010, 119: 198–209.
- [4] Kurup P, Zhang Y, Xu J, Venkitaramani DV, Haroutunian V, Greengard P*, et al.* Abeta-mediated NMDA receptor endocytosis in Alzheimer's disease involves ubiquitination of the tyrosine phosphatase STEP61. J Neurosci 2010, 30: 5948–5957.
- [5] Preuss T. Do rats have prefrontal cortex? The Rose-Woolsey-Akert program reconsidered. J Cogn Neurosci 1995, 7: 1–26.
- [6] Goldman-Rakic PS. Working memory dysfunction in schizophrenia. J Neuropsychiatry Clin Neurosci 1994, 6: 348–357.
- [7] Lim HK, Juh R, Pae CU, Lee BT, Yoo SS, Ryu SH*, et al.* Altered verbal working memory process in patients with Alzheimer's disease: an fMRI investigation. Neuropsychobiology 2008, 57: 181–187.
- [8] Liu L, Wong TP, Pozza MF, Lingenhoehl K, Wang Y, Sheng M*, et al.* Role of NMDA receptor subtypes in governing the direction of hippocampal synaptic plasticity. Science 2004, 304: 1021–1024.
- [9] Cho KK, Khibnik L, Philpot BD, Bear MF. The ratio of NR2A/ B NMDA receptor subunits determines the qualities of ocular dominance plasticity in visual cortex. Proc Natl Acad Sci U S A 2009, 106: 5377–5382.
- [10] Lüscher C, Malenka RC. NMDA receptor-dependent longterm potentiation and long-term depression (LTP/LTD). Cold Spring Harb Perspect Biol 2012, 4: pii: a005710.
- [11] Yuste R, Bonhoeffer T. Morphological changes in dendritic spines associated with long-term synaptic plasticity. Annu Rev Neurosci 2001, 24: 1071–1089.
- [12] Tashiro A, Yuste R. Regulation of dendritic spine motility and stability by Rac1 and Rho kinase: evidence for two forms of spine motility. Mol Cell Neurosci 2004, 26: 429–440.
- [13] Araya R, Jiang J, Eisenthal KB, Yuste R. The spine neck filters membrane potentials. Proc Natl Acad Sci U S A 2006, 103: 17961–17966.
- [14] Goebel-Goody SM, Davies KD, Alvestad Linger RM, Freund RK, Browning MD. Phospho-regulation of synaptic and extrasynaptic N-methyl-d-aspartate receptors in adult hippocampal slices. Neuroscience 2009, 158: 1446–1459.
- [15] Baez MV, Oberholzer MV, Cercato MC, Snitcofsky M, Aguirre AI, Jerusalinsky DA. NMDA receptor subunits in the adult

rat hippocampus undergo similar changes after 5 minutes in an open field and after LTP induction. PLoS One 2013, 8: e55244.

- [16] Shipton OA, Paulsen O. GluN2A and GluN2B subunitcontaining NMDA receptors in hippocampal plasticity. Philos Trans R Soc Lond B Biol Sci 2013, 369: 20130163.
- [17] Dupuis JP, Ladépêche L, Seth H, Bard L, Varela J, Mikasova L*, et al.* Surface dynamics of GluN2B-NMDA receptors controls plasticity of maturing glutamate synapses. EMBO J 2014, 33: 842–861.
- [18] Liu XB, Murray KD, Jones EG. Switching of NMDA receptor 2A and 2B subunits at thalamic and cortical synapses during early postnatal development. J Neurosci 2004, 24: 8885– 8895.
- [19] Philpot BD, Weisberg MP, Ramos MS, Sawtell NB, Tang YP, Tsien JZ*, et al.* Effect of transgenic overexpression of NR2B on NMDA receptor function and synaptic plasticity in visual cortex. Neuropharmacology 2001, 41: 762–770.
- [20] Goldman PS, Rosvold HE. Localization of function within the dorsolateral prefrontal cortex of the rhesus monkey. Exp Neurol 1970, 27: 291–304.
- [21] Funahashi S, Bruce CJ, Goldman-Rakic PS. Mnemonic coding of visual space in the monkey's dorsolateral prefrontal cortex. J. Neurophysiology 1989, 61: 331–349.
- [22] Goldman-Rakic PS. The "psychic cell" of Ramón y Cajal. Prog Brain Res 2002, 136: 427–434.
- [23] Goldman-Rakic PS. Cellular basis of working memory. Neuron 1995, 14: 477-485.
- [24] Wang XJ. Synaptic basis of cortical persistent activity: the importance of NMDA receptors to working memory. J Neurosci 1999, 19: 9587–9603.
- [25] Wang XJ. Synaptic reverberation underlying mnemonic persistent activity. Trends in Neurosci 2001, 24: 455–463.
- [26] Wang H, Stradtman GGr, Wang XJ, Gao WJ. A specialized NMDA receptor function in layer 5 recurrent microcircuitry of the adult rat prefrontal cortex. Proc Natl Acad Sci U S A 2008, 105: 16791-16796.
- [27] Wang MJ, Yang Y, Wang CJ, Gamo NJ, Jin LE, Mazer JA*, et al.* NMDA receptors subserve working memory persistent neuronal firing In dorsolateral prefrontal cortex. Neuron 2013, 77: 736–749.
- [28] Yang Y, Paspalas CD, Jin LE, Picciotto MR, Arnsten AFT, Wang M. Nicotinic α7 receptors enhance NMDA cognitive circuits in dorsolateral prefrontal cortex. Proc Nat Acad Sci USA 2013, 110: 12078–83.
- [29] Arnsten AFT, Wang MJ, Paspalas CD. Neuromodulation of thought: Flexibilities and vulnerabilities in prefrontal cortical network synapses. Neuron 2012, 76: 223–239.
- [30] Snyder EM, Nong Y, Almeida CG, Paul S, Moran TH, Choi EY*, et al.* Regulation of NMDA receptor trafficking by

amyloid-beta. Nat Neurosci 2005, 8: 1051–1058.

- [31] Javitt DC. Glutamatergic theories of schizophrenia. Isr J Psychiatry Relat Sci 2010, 47: 4–16.
- [32] Banerjee A, Macdonald ML, Borgmann-Winter KE, Hahn CG. Neuregulin 1-erbB4 pathway in schizophrenia: From genes to an interactome. Brain Res Bull 2010, 30: 132–139.
- [33] Martin LF, Freedman R. Schizophrenia and the alpha7 nicotinic acetylcholine receptor. Int Rev Neurobiol 2007, 78: 225–246.
- [34] Kristiansen LV, Patel SA, Haroutunian VH, Meador-Woodruff JH. Expression of the NR2B-NMDA receptor subunit and its Tbr-1/CINAP regulatory proteins in postmortem brain suggest altered receptor processing in schizophrenia. Synapse 2010, 64: 495–502.
- [35] Weickert CS, Fung SJ, Catts VS, Schofield PR, Allen KM, Moore LT*, et al.* Molecular evidence of N-methyl-D-aspartate receptor hypofunction in schizophrenia. Mol Psychiatry 2013, 18: 1185–1192.
- [36] Schizophrenia Working Group of the Psychiatric Genomics Consortium. Biological insights from 108 schizophreniaassociated genetic loci. Nature 2014, 511: 421–427.
- [37] Fromer M, Pocklington AJ, Kavanagh DH, Williams HJ, Dwyer S, Gormley P, *et al*. De novo mutations in schizophrenia implicate synaptic networks. Nature 2014, 506: 179–184.
- [38] Driesen NR, McCarthy G, Bhagwagar Z, Bloch MH, Calhoun VD, D'Souza DC*, et al.* The impact of NMDA receptor blockade on human working memory-related prefrontal function and connectivity. Neuropsychopharmacology 2013, 38: 2613–2622.
- [39] Anticevic A, Gancsos M, Murray JD, Repovs G, Driesen NR, Ennis DJ*, et al.* NMDA receptor function in large-scale anticorrelated neural systems with implications for cognition and schizophrenia. Proc Natl Acad Sci U S A 2012, 109: 16720–16725.
- [40] Driesen NR, Leung HC, Calhoun VD, Constable RT, Gueorguieva R, Hoffman R*, et al.* Impairment of working memory maintenance and response in schizophrenia: functional magnetic resonance imaging evidence. Biol Psychiatry 2008, 64: 1026–1034.
- [41] Jackson ME, Homayoun H, Moghaddam B. NMDA receptor hypofunction produces concomitant firing rate potentiation and burst activity reduction in the prefrontal cortex. Proc Natl Acad Sci U S A 2004, 101: 8467–8472.
- [42] Malhotra AK, Pinals DA, Adler CM, Elman I, Clifton A, Pickar D*, et al.* Ketamine-induced exacerbation of psychotic symptoms and cognitive impairment in neuroleptic-free schizophrenics. Neuropsychopharmacology 1997, 17: 141–150.
- [43] Berman RM, Cappiello A, Anand A, Oren DA, Heninger GR, Charney DS*, et al.* Antidepressant effects of ketamine in

depressed patients. Biol Psychiatry 2000, 47: 351–354.

- [44] Zarate CAJ, Singh JB, Carlson PJ, Brutsche NE, Ameli R, Luckenbaugh DA*, et al.* A randomized trial of an N-methyl-Daspartate antagonist in treatment-resistant major depression. Arch Gen Psychiatry 2006, 63: 856–864.
- [45] Murrough JW, Iosifescu DV, Chang LC, Al Jurdi RK, Green CE, Perez AM, et al. Antidepressant efficacy of ketamine in treatment-resistant major depression: a two-site randomized controlled trial. Am J Psychiatry 2013, 170: 1134–1142.
- [46] Murrough JW, Perez AM, Pillemer S, Stern J, Parides MK, aan het Rot M*, et al.* Rapid and longer-term antidepressant effects of repeated ketamine infusions in treatment-resistant major depression. Biol Psychiatry 2013, 74: 250–256.
- [47] Clark P. Treatment-refractory depression: A case of successful treatment with intranasal ketamine 10%. Ann Clin Psychiatry 2014, 26: 145.
- [48] Lapidus KA, Levitch CF, Perez AM, Brallier JW, Parides MK, Soleimani L*, et al.* A randomized controlled trial of intranasal ketamine in major depressive disorder. Biol Psychiatry 2014, 76: 970–976..
- [49] Salvadore G, Cornwell BR, Colon-Rosario V, Coppola R, Grillon C, Zarate CAJ*, et al.* Increased anterior cingulate cortical activity in response to fearful faces: a neurophysiological biomarker that predicts rapid antidepressant response to ketamine. Biol Psychiatry 2009, 65: 289–295.
- [50] Seo H, Lee D. Behavioral and neural changes after gains and losses of conditioned reinforcers. J Neurosci 2009, 29: 3627–3641.
- [51] Rushworth MF, Behrens TE. Choice, uncertainty and value in prefrontal and cingulate cortex. Nat Neurosci 2008, 4: 389–397.
- [52] Mayberg HS, Lozano AM, Voon V, McNeely HE, Seminowicz D, Hamani C*, et al.* Deep brain stimulation for treatmentresistant depression. Neuron 2005, 45: 651–660.
- [53] Li N, Lee BT, Liu RJ, Banasr M, Dwyer JM, Iwata M*, et al.* mTOR-dependent synapse formation underlies the rapid antidepressant effects of NMDA antagonists. Science 2010, 329: 959–964.
- [54] Andolfatto G, Willman E, Joo D, Miller PL, Wong WB, Koehn M*, et al.* Intranasal ketamine for analgesia in the emergency department: a prospective observational series. Acad Emerg Med 2013, 20: 1050–1054.
- [55] Vogt BA, Sikes RW. The medial pain system, cingulate cortex, and parallel processing of nociceptive information. Prog Brain Res 2000, 22: 223–235.
- [56] Bushnell MC, Ceko M, Low LA. Cognitive and emotional control of pain and its disruption in chronic pain. Nat Rev Neurosci 2013, 14: 502–511.
- [57] Yeaman F, Meek R, Egerton-Warburton D, Rosengarten P, Graudins A. Sub-dissociative-dose intranasal ketamine for moderate to severe pain in adult emergency department patients. Emerg Med Australas 2014, 26: 237–242.
- [58] McCarty EC, Mencio GA, Walker LA, Green NE. Ketamine sedation for the reduction of children's fractures in the emergency department. J Bone Joint Surg Am 2000, 82-A: 912–918.