

Lipoxin A₄: a novel anti-inflammatory molecule?

Arachidonic acid is metabolised by the cyclooxygenase pathway to the prostaglandins and thromboxane A₂ or via one of the lipoxygenase pathways.¹ Three major lipoxygenase pathways have been identified in mammalian tissue – namely, the 5-, 12-, and 15-lipoxygenases.²⁻⁴ The 5-lipoxygenase pathway metabolises arachidonic acid through two intermediates, 5-hydroperoxyeicosatetraenoic acid (5-HPETE) and leukotriene A₄ (LTA₄), to LTB₄ and the sulphidopeptide leukotrienes LTC₄, LTD₄, and LTE₄.⁵ The sulphidopeptide leukotrienes are potent spasmogens⁶ for non-vascular smooth muscle and may play a part in the pathogenesis of bronchial asthma.⁷⁻¹⁰

The interactions between 5-lipoxygenase and 15-lipoxygenase on arachidonic acid metabolism have recently been studied and a new series of biologically active metabolites described.^{11,12} These molecules have been termed lipoxins. Unlike leukotrienes lipoxins possess a conjugated tetraene structure and the stereochemistries of the two major isomers, lipoxin (LX)A₄ and LXB₄, are 5S, 6R, 5S-trihydroxy 7,9,13-*trans*-11-*cis*-eicosatetraenoic acid and 5S, 14R, 15S-trihydroxy-6,10,12-*trans*-8-*cis*-eicosatetraenoic acid, respectively.^{13,14} It is now established that lipoxins can also be generated by an interaction between the 5- and 12-lipoxygenases, when the 12-lipoxygenase acts with a C-15 specificity.

Lipoxins can be generated by human neutrophils, eosinophils, or platelets from both endogenous or exogenous substrates in vitro.^{11,12,15,16} Furthermore, using gas chromatography and mass spectrometry with selective ion monitoring, LXA₄ has been detected in the bronchoalveolar lavage fluid in patients suffering from pulmonary sarcoidosis, infective bronchopneumonia, asthma, and carcinoma of the lung.¹⁷ It was not detected in normal subjects. In patients with detectable LXA₄ in bronchoalveolar lavage fluid, the ratio of the concentrations of LXA₄ to those of sulphidopeptide leukotrienes ranges from 1.9 to 62 (mean 19.0), indicating that LXA₄ is generated in vivo.

In vitro studies with guinea pig parenchymal lung strips have shown that lipoxins exhibit contractile activity.^{18,19} LXA₄ prepared by total chemical synthesis has been shown to constrict parenchymal strips over a concentration range of 1×10^{-8} to 1×10^{-5} M.¹⁹ The contractile activity of LXA₄ was slow in onset and did not plateau for 20 minutes, and was approximately 10 000 times less potent than that of LTD₄. The contraction was not mediated through secondary generation of cyclooxygenase metabolites or secondary release of sulphidopeptide leukotrienes. The activity of LXA₄ may be elicited via an interaction with an LTD₄ receptor. This suggestion was further supported when it was shown that LXA₄ (1×10^{-7} M) prevented mesangial cell inositol triphosphate generation induced by LTD₄.²⁰ At concentrations of 1×10^{-8} M and 5×10^{-8} M LXA₄ induced the generation of mesangial cell inositol triphosphate which was abolished with a sulphidopeptide leukotriene antagonist SK&F 104353. LXA₄ competed with [³H] LTD₄ for specific binding to cultured rat glomerular mesangial cells. In vivo it antagonised LTD₄-induced falls in glomerular filtration rate. Dahlen and coworkers have reported that LXA₄ at a concentration of 1×10^{-6} M was able to shift the log dose-response curve of LTC₄ on guinea pig lung strip to the right.²¹ Bjork and coworkers showed that LXA₄ at concentrations between 1 M and 30×10^{-6} M produced a dose-dependent contraction of human bronchi and antagonised LTC₄-induced

contractions.²² These studies support the view that LXA₄ may act as a partial agonist at the same or similar site as the sulphidopeptide leukotrienes.

The fact that 15-lipoxygenase is abundant in lung tissue and that LXA₄ has been recovered in the bronchoalveolar lavage fluid of patients with asthma and other lung diseases suggests that LXA₄ may be a potential mediator or modulator of inflammation in the lung. In a recent study eight subjects underwent inhalation challenge with LXA₄,²³ but no effect was seen on specific conductance, rate of airflow at 25% vital capacity (\dot{V}_{25}), blood pressure, pulse, or asthmatic symptoms. There was, however, a significant shift of the specific conductance and \dot{V}_{25} dose-response curve to the right after inhalation challenge with LTC₄ combined with LXA₄ compared with that after inhalation challenge with LTC₄ alone. Thus, LXA₄ may modulate LTC₄-induced airway obstruction in vivo and may act as an endogenous sulphidopeptide leukotriene receptor antagonist.

Further evidence for the anti-inflammatory properties of LXA₄ was suggested by the finding that prior exposure of neutrophils or eosinophils to 10^{-9} – 10^6 M LXA₄ inhibited the chemotactic responsiveness to LTB₄, formyl-methionyl-leucyl-phenylalanine (FMLP), and plasma activating factor in a dose-dependent manner.^{24,25} The finding that LXA₄ attenuated LTB₄-induced neutrophil migration and plasma leakage in the hamster cheek pouch model also supports its putative anti-inflammatory role.²⁶

There is limited information on the mechanisms for the inhibiting effects of LXA₄ on neutrophil functions. The inhibition of chemotactic responses was associated with a concentration-dependent inhibition of phosphoinositide hydrolysis and calcium mobilisation.²⁷ There was no effect on specific binding of [³H] LTB₄ to neutrophils following preincubation with LXA₄, suggesting that the mechanism of the chemotactic factor-induced phosphoinositide hydrolysis was at a post-receptor level. Structure function studies on the mechanism of inhibition of LXA₄ on LTB₄-induced neutrophil migration demonstrated the importance of two adjacent free hydroxy groups in either the R or the S configuration; one hydroxy group has to be in a C-6 position, but the other hydroxy group can be in either the C-5 or the C-7 position for conferment of inhibitory activity.²⁸

Successful elucidation of the mechanism(s) for the inhibitory activity of LXA₄ may provide a novel therapeutic approach in inflammatory diseases.

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